

Evaluation of Effects of Zingiber officinale on Salivation in Rats

Goli Chamani¹, Mohammad Reza Zarei², Mitra Mehrabani³, and Yousef Taghiabadi⁴

¹ Department of Oral Medicine, Kerman School of Dentistry, Kerman, Iran
² Kerman Neuroscience Research Center, Kerman University of Medical Sciences, Kerman, Iran
³ Department of Pharmacognosy, School of Pharmacy, Kerman University of Medical Sciences, Kerman, Iran
⁴ Private Practice, Kerman, Iran

Received: 2 Aug. 2009; Received in revised form: 18 Apr. 2010; Accepted: 13 Jun. 2010

Abstract- There are some herbal plants in Iranian traditional system of medicine which are believed to be excellent remedies to alleviate the symptoms of xerostomia. The aim of the present study was to evaluate the effect of systemic administration of seven different herbal extracts on the rate of salivation in rats. The extracts of 7 herbs; *Zingiber officinale* Roscoe (Zingiberaceae), *Citrus sinensis* (L.) Osbeck (Rutaceae), *Artemisia absinthium* L. (Asteraceae), *Cichorium intybus* L. (Asteraceae), *Pimpinella anisum* L.(Apiaceae), *Portulaca oleracea* L.(Portulacaceae), *Tribulus terrestris* L. (Zygophyllaceae) were prepared. Nine groups of animals (including negative and positive control groups) were used and seven rats were tested in each group. After the injection of extracts, saliva volume was measured gravimetrically in four continuous sevenminute intervals. The results showed that after injection of ginger extracts salivation was significantly higher as compared to the negative control group and other herbal extracts in all of the four intervals (*P*<0.01). The peak action of the ginger was during the first 7-minute interval and following this, salivation decreased to some extent. The present study suggests that the extract of *Zingiber officianle* can increase the rate of salivation significantly in animal model. Further investigations on different constituents of ginger seem to be essential to identify the responsible constituent for stimulation of saliva secretion.

© 2011 Tehran University of Medical Sciences. All rights reserved. *Acta Medica Iranica* 2011; 49(6): 336-340.

Keywords: Herbal medicine; Salivation; Xerostomia

Introduction

Xerostomia (dry mouth) is the most common longstanding problem for the majority of the patients who suffer from salivary gland diseases. Medications, exposure of the head and neck region to radiation, and connective tissue diseases are the most common causes of salivary gland dysfunction that may in turn lead to xerostomia (1).

Although xerostomia is usually managed symptomatically with saliva substitutes, a large number of potential systemic therapies for chronic xerostomia have emerged during the past decade. More than 24 agents have been proposed as the means of stimulating salivary output systemically (2). Most of the drugs used to increase salivary flow, stimulate saliva secretion through their cholinergic effects on functional acinar cells (3).

Several herbal drugs have been recently developed and used in clinical trials successfully to relieve xerostomia.

Herbal medications claiming to relieve xerostomia include *Cetraria islandica*, Bakumondo-to, LongoVital, hot aqueous extract of the rhizome of *Anemarrhena asphodeloides* and Byakko-ka-ninjin-to (4-8). There are some herbal plants in Iranian traditional system of medicine, which are believed to be excellent remedies to alleviate and help with the symptoms of chronic salivary gland diseases particularly xerostomia. However, lack of scientific papers published on these herbs, makes it difficult to determine either the validity of their effects or their mechanism of action. The present study was designed to evaluate the effect of seven herbal plants widely used to increase the rate of salivation in Iranian folk medicine.

Archive of SID G. Chamani, et al.

Materials and Methods

Animals

In this experimental study, 63 adult male rats (weight: 200-300 g) from NMRI strain were used. The animals were bred and housed at the animal facility of neuroscience research center of Kerman University of Medical Sciences. The animals were kept in a well cross ventilated room with controlled temperature and humidity and a standard 12h light: 12h dark cycle. Standard rodent food and tap water were available. Nine groups of animals (including negative and positive control groups) were used and seven rats were tested in each group. Approval for this study was obtained from the Animal Care Committee at Research Center of Kerman University.

Plants

Rhizome of Zingiber officianale Roscoe (Zingiberaceae), peels of the fruit of Citrus sinensis L. (Rutaceae), aerial parts of Artemisia absinthium L. (Asteraceae), roots of Cichorium intybus L. (Asteraceae), fruits of Pimpinella anisum L. (Apiaceae), seeds of Portulaca oleracea L. (Portulacaceae), and fruits of Tribulus terrestris L. (Zygophyllaceae) were used. The plant materials were identified and authenticated taxonomically by a botanist.

Parts of the plants selected for the present study (20 g) were washed with distilled water to remove dirt and soil, and shade dried. The dried materials submitted for a maceration process. The material was extracted twice with ethanol (80%). The extracts were filtered, pooled, and concentrated at high temperature (+50°C) on a rotary evaporator (Heidolph, Germany). The extracts were suspended in normal saline with ethanol 10% as cosolvent and stored in refrigerator within dark containers.

Saliva collection

Rats were anesthetized using a single intraperitoneal injection of 75 mgkg⁻¹ ketamine (Alfasan, Holland) and 5 mgkg⁻¹ xylazine (Alfasan, Holland). The unconscious rats were kept on a thermal pad to maintain their body temperature at the level of 37°C.

Before saliva collection, the oral cavity was wiped and dried with a cotton pellet and then 4 pre-weighed cotton pellets were inserted into the mouth of each animal: two cotton pellets underneath the tongue and one between the cheek and the teeth on either side. After seven minutes, the cotton pellets were removed and weighed again on a precision weighing balance (Sartorius, Germany). The difference of the weight of the cotton pellets between two determinations was considered as the baseline weight of the saliva secreted. The flow rate of saliva was determined gravimetrically, assuming that the specific gravity of saliva is 1 (i.e. 1 g equals 1ml of saliva).

Following measurement of the baseline secreted saliva, the extracts were injected (10 mgkg⁻¹ body weight) intraperitoneally. The rate of saliva secretion was determined at four continuous seven-minute intervals. The investigator was blinded to all of the injected solutions in this study. The experiment was run in parallel with a negative control (10 mlkg⁻¹ normal saline mixed with co-solvent) and a positive control (4 µmolkg⁻¹ pilocarpine dissolved in distilled water and co-solvent). At the end of the experiments the rats were sacrificed by pentobarbital overdose.

Statistics

All the data were analyzed by the analysis of variance for comparing groups and Bonferroni for multiple comparisons. A value of P<0.05 was considered statistically significant.

Results

The present study showed that the secretion of saliva increased dramatically after injection of extract of Zingiber officianale. The peak action of Zingiber officianale appeared during the first 7-minute interval and after that salivation decreased to some extent. Statistical analysis showed that the mean rate of saliva secretion induced by Zingiber officianale in all of the 7-minute intervals was significantly higher than that of other herbal extracts and negative control (P<0.05) (Table 1 and Figure 1).

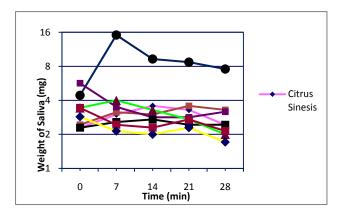


Figure 1. Saliva secretion before and after injection of seven herbal extracts in four continuous seven-minute intervals

Table 1. Saliva secretion (Mean ± SD) before and after injection of seven herbal extracts in four continuous seven-minute intervals

	Time (Minutes)									
Drug	0		7		14		21		28	
	Weight of the Saliva (mg)									
	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Citrus sinensis 1	2.3	0.8	3.0	1.4	3.6	1.7	3.3	1.0	2.4	0.5
Artemisia absinthium ²	2.4	0.8	3.1	1.2	3.0	1.7	3.6	1.3	3.3	1.4
Zingiber officinalis	4.4	1.9	15.1	8.1	9.3	6.6	8.7	6.6	7.6	7.0
Cichorium intybus ³	7.7	5.9	8.4	13.2	6.1	8.8	6.0	8.5	4.1	3.0
Pimpinella anisum ⁴	2.3	0.5	2.6	1.1	2.7	0.8	2.4	0.8	2.4	0.5
Tribulus terrestris 6	3.4	0.5	4.0	0.6	3.3	0.8	2.7	0.5	2.0	0.6
Portulaca oleracea	3.4	1.4	2.4	0.5	2.3	1.0	2.7	0.8	2.1	0.4
Negative control	2.9	1.9	2.1	0.7	2.0	0.8	2.3	1.4	1.7	1.0
Pilocarpine	3.1	0.7	129.3	51.5	201.7	23.1	169.0	28.9	129.9	42.9

- 1. Peels of the fruit of Citrus sinensis L. (Rutaceae)
- 2. Aerial parts of Artemisia absinthium L. (Asteraceae)
- 3. Roots of Cichorium intybus L. (Asteraceae)
- 4. Fruits of Pimpinella anisum L. (Apiaceae)
- 5. Seeds of *Portulaca oleracea* L. (Portulacaceae)
- 6. Fruits of Tribulus terrestris L. (Zygophyllaceae)

The results also indicated that saliva secretion increased after injection of other herbal extracts when compared with the negative control group. However, the difference was not statistically significant. Since the saliva secretion after injection of pilocarpine (positive control) was much higher than extracts of seven herbs, pilocarpine result has not shown in Figure 1. The investigators believe that in order to compare the effect of ginger on salivation with pilocarpine, the dose of pilocarpine should be adjusted in future studies.

Discussion

Although most clinicians trained in the Western education system are unfamiliar with herbal medications and other forms of alternative medicine (4), alternative medicines are gaining popularity and recognition within the professional community. The results of the present study showed that the rate of salivation increased significantly in response to the injection of *Zingiber officianale* (ginger) extract.

Ginger is one of the most commonly used herbal supplements and its substantial use in folk remedies for different medical conditions has been documented (9). Based on phytochemical studies, ginger is rich in a large number of substances including gingerols and shogaols (10,11). Some of the main pharmacological actions of these compounds include anti-inflammatory, antioxidant and anticarcinogenic activities (10,12,13).

Ginger (*Zingiber officinale* Roscoe, Zingiberacae) has been used for a wide array of unrelated ailments that include vomiting, stomachaches, abdominal spasm, nausea, motion sickness, arthritis, rheumatism, hypertension, ulcerative colitis, fever and infectious diseases (14-20). Extracts and fractions of *Zingiber officinale* have been shown to protect against chemically-induced tissue damage (21). The radio protective effect of ginger extract has also been confirmed (22-24).

As mentioned earlier, ginger (rhizome of Zingiber officinale) has been widely used for centuries in gastrointestinal disorders, particularly dyspepsia, but its precise mode of action was to be elucidated. Ghayur and colleagues showed that aqueous-methanolic extract of ginger 70% (Zo.Cr) exhibits prokinetic activity in rats via activation of post-synaptic muscarinic M3 receptor in rat stomach fundus (25). Prokinetic activity of ginger extract (Zo.Cr) was also confirmed in an *in vivo* test. This study showed that Zo.Cr contains a cholinergic, spasmogenic component evident in stomach fundus preparations which provides a sound mechanistic insight for the prokinetic action of ginger (26).

Although research studies have widely been performed about different pharmacological properties of *Zingiber officianale*, there is little evidence about its effect(s) on saliva secretion in the literature (27). It has been determined that parasympathetic stimulation causes a copious flow of saliva with low outputs of protein and

Archive of SID G. Chamani, et al.

sympathetic nerve stimulation which *per se* causes less fluid secretion, but exocytosis of secretory granules occurs from both acini and granular tubules (28). Since the prokinetic activity of ginger has been confirmed (25, 26), the authors believe that ginger effect on salivation can be attributed to some kind of cholinergic action. However, the exact mechanism is not fully understood and more researches using atropine may help to clear the mechanism of action. Further investigations on different constituents of *Zingiber officianale* in different doses also seem to be essential to identify the responsible constituent and the optimum effective dose for saliva secretion.

Acknowledgement

The authors wish to thank Research Committee of Kerman University of Medical Sciences and Kerman Neuroscience Research Center for financial and executive support of this project.

References

- Kaplan I, Zuk-Paz L, Wolff A. Association between salivary flow rates, oral symptoms, and oral mucosal status. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2008;106(2):235-41.
- Greenberg MS, Glick M, Ship JA. Burket's Oral Medicine. 11th ed. Hamilton, Ontario, Canada: BC Decker Inc.; 2008. p. 215.
- Grisius MM. Salivary gland dysfunction: a review of systemic therapies. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2001;92(2):156-62.
- Kagami H, Horie K, Nishiguchi H, Shigetomi T, Ueda M. Effect of 'bakumondo-to', a Chinese-Japanese herbal medicine, on cultured and dispersed salivary gland cells. J Ethnopharmacol 1996;53(2):89-95.
- Kimura M, Kimura I, Chem FJ. Combined potentiating effect of byakko-ka-ninjin-to, its constituents, rhizomes of Anemarrhena asphodeloides, tomosaponin A-III, and calcium on pilocarpine-induced saliva secretion in streptozocin-diabetic mice. Biol Pharm Bull 1996;19(7):926-31.
- Niijima A, Kubo M, Ichiki H, Komatsu Y, Maruno M, Okada M. Effect of byakko-ka-ninjin-to on the efferent activity of the autonomic nerve fibers innervating the sublingual gland of the rat. J Auton Nerv Syst 1997;63(1-2):46-50.
- Wang Q, Liu H, Qiao N. Effects of traditional Chinese medicine on salivary glands in the patients with head and neck cancer during radiotherapy. Zhongguo Zhong Xi Yi Jie He Za Zhi 1998;18(11):662-4.

 Pedersen A, Gerner N, Palmvang I, Høier-Madsen M. LongoVital in the treatment of Sjögren's syndrome. Clin Exp Rheumatol 1999;17(5):533-8.

- El-Abhar HS, Hammad LN, Gawad HS. Modulating effect of ginger extract on rats with ulcerative colitis. J Ethnopharmacol 2008;118(3):367-72. Epub 2008 May 15.
- Masuda Y, Kikuzaki H, Hisamoto M, Nakatani N. Antioxidant properties of gingerol related compounds from ginger. Biofactors 2004;21(1-4):293-6.
- Jolad SD, Lantz RC, Chen GJ, Bates RB, Timmermann BN. Commercially processed dry ginger (Zingiber officinale): composition and effects on LPS-stimulated PGE2 production. Phytochemistry 2005;66(13):1614-35.
- Grzanna R, Lindmark L, Frondoza CG. Ginger: an herbal medicinal product with broad anti-inflammatory actions. J Med Food 2005;8(2):125-32.
- 13. Shukla Y, Singh M. Cancer preventive properties of ginger: a brief review. Food Chem Toxicol 2007;45(5):683-90. Epub 2006 Nov 12.
- 14. Langner E, Greifenberg S, Gruenwald J. Ginger: history and use. Adv Ther 1998;15(1):25-44.
- Afzal M, Al-Hadidi D, Menon M, Pesek J, Dhami MS. Ginger: an ethnomedical, chemical and pharmacological review. Drug Metabol Drug Interact 2001;18(3-4):159-90.
- Ficker CE, Arnason JT, Vindas PS, Alvarez LP, Akpagana K, Gbéassor M, De Souza C, Smith ML. Inhibition of human pathogenic fungi by ethnobotanically selected plant extracts. Mycoses 2003;46(1-2):29-37.
- 17. Ghayur MN, Gilani AH, Afridi MB, Houghton PJ. Cardiovascular effects of ginger aqueous extract and its phenolic constituents are mediated through multiple pathways. Vascul Pharmacol 2005;43(4):234-41. Epub 2005 Sep 12.
- Ghayur MN, Gilani AH. Ginger lowers blood pressure through blockade of voltage-dependent calcium channels. J Cardiovasc Pharmacol 2005;45(1):74-80.
- 19. White B. Ginger: an overview. Am Fam Physician 2007;75(11):1689-91.
- 20. Ali BH, Blunden G, Tanira MO, Nemmar A. Some phytochemical, pharmacological and toxicological properties of ginger (Zingiber officinale Roscoe): a review of recent research. Food Chem Toxicol 2008;46(2):409-20. Epub 2007 Sep 18.
- 21. Yemitan OK, Izegbu MC. Protective effects of Zingiber officinale (Zingiberaceae) against carbon tetrachloride and acetaminophen-induced hepatotoxicity in rats. Phytother Res 2006;20(11):997-1002.
- 22. Jagetia GC, Baliga MS, Venkatesh P, Ulloor JN. Influence of ginger rhizome (Zingiber officinale Rosc) on survival, glutathione and lipid peroxidation in mice after wholebody exposure to gamma radiation. Radiat Res 2003;160(5):584-92.

A Effects of zingiber officinalis on salivation in rats

- 23. Jagetia G, Baliga M, Venkatesh P. Ginger (Zingiber officinale Rosc.), a dietary supplement, protects mice against radiation-induced lethality: mechanism of action. Cancer Biother Radiopharm 2004;19(4):422-35.
- 24. Sharma A, Haksar A, Chawla R, Kumar R, Arora R, Singh S, Prasad J, Islam F, Arora MP, Kumar Sharma R. Zingiber officinale Rosc. modulates gamma radiationinduced conditioned taste aversion. Pharmacol Biochem Behav 2005;81(4):864-70.
- 25. Ghayur MN, Khan AH, Gilani AH. Ginger facilitates cholinergic activity possibly due to blockade of muscarinic autoreceptors in rat stomach fundus. Pak J Pharm Sci 2007;20(3):231-5.
- 26. Ghayur MN, Gilani AH. Pharmacological basis for the medicinal use of ginger in gastrointestinal disorders. Dig Dis Sci 2005;50(10):1889-97.
- 27. Blumberger W, Glatzel H. The physiology of spices and condiments. The action of ginger on the amount and condition of the saliva. Nutr Dieta Eur Rev Nutr Diet 1964;64:181-92.
- 28. Garrett JR, Suleiman AM, Anderson LC, Proctor GB. Secretory responses in granular ducts and acini of submandibular glands in vivo to parasympathetic or sympathetic nerve stimulation in rats. Cell Tissue Res 1991;264(1):117-26.