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## In vitro Antimicrobial activity of *Acacia nilotica*, *Ziziphus mauritiana*, *Bauhinia variegata* and *Lantana camara* against some clinical isolated strains

V. Nagumanthri<sup>1</sup>, S. Rahiman<sup>2\*</sup>, B. Ahmad Tantry<sup>3</sup>  
P. Nissankarara<sup>4</sup> and M. Phani kumar<sup>5</sup>

<sup>1</sup>Department of Biotechnology, T. S. R & T. B. K PG College, Andhra Pradesh, India

<sup>2</sup>Department of Biochemistry, College of Medicine, Al Jouf University,  
Saudi Arabia, P.O. Box 2014, Postal code: 75471

<sup>3</sup>Department of Microbiology, College of Medicine, Al Jouf University, Saudi Arabia

<sup>4,5</sup>Department of Biotechnology & Molecular Biology, BioAxis DNA

Research Centre, Hyderabad, India

E-mail: [rahimhi@gmail.com](mailto:rahimhi@gmail.com)

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### Abstract

Plants are potent biochemists; biologically active compounds present in the medicinal plants have always been of great interest to scientists working in this field. Thus, the aim of the current study was to screen the antimicrobial activity of *Acacia nilotica*, *Ziziphus mauritiana*, *Bauhinia variegata* and *Lantana camara* against some selected clinical isolated strains. Although previous studies have documented the antimicrobial properties of these plants, this work is designed to evaluate the specific antibacterial activity of different extracts of these plants against tested microorganisms, in order to know the best extract against specific microorganisms. In this study the fresh parts (leaves, barks & pods) of the test medicinal plant were collected and methanol, ethanol and ethyl acetate extracts were prepared. Antibacterial susceptibility test was done by using Agar diffusion assay method. Statistical analysis was carried out with SPSS 17.0 Windows version. The results of the current study showed that a total of 8 extracts from 4 different plant species were investigated including pods of ethyl acetate extracts of *Lantana camara*, which showed the highest antimicrobial activity against tested clinical isolates (*Bacillus subtilis*  $2\pm 0.1$ mm, *Bacillus circulans*  $2.6\pm 0.2$ mm, *Bacillus sphaericus*  $2\pm 0.1$ mm, *Staphylococcus aureus*  $2.5\pm 0.1$ , and *Serratia liquefaciens*  $2.2\pm 0.1$ mm), followed by its ethyl acetate extracts of leaves. Bark extracts of four tested medicinal plants possess a lower zone on inhibitory activity as compared to the leaves extracts of these plants. Noticeably no antimicrobial activity was found in the methanolic bark extract of *Acacia nilotica* against the tested bacteria except *Bacillus circulans*. The results of the present investigation clearly indicate that the antibacterial activity varies with the species of the plants and plant material used. Thus, the study ascertains the value of plants used in ayurveda, which could be of considerable interest to the development of new drugs. Studies are in progress to further evaluate the mechanisms of action of these *active test* extracts on study organisms associated with certain human diseases.

**Keywords:** Antimicrobial susceptibility; medicinal plants; Agar diffusion; clinical isolates

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### 1. Introduction

Plants are potent biochemists and have been components of phytomedicine since time immemorial; man is able to obtain from them a wondrous assortment of industrial chemicals. Plant based natural constituents can be derived from any part of the plant like bark, leaves, flowers, roots, fruits, seeds, etc [1] i.e. any part of the plant may contain active components. The beneficial medicinal

effects of plant materials typically result from the combinations of secondary products present in the plant. Biologically active compounds present in the medicinal plants have always been of great interest to scientists working in this field. In recent years this interest to evaluate plants possessing antibacterial activity for various diseases is growing [2]. For instance, in developed countries 25% of the medical drugs are based on plants and their derivatives [3]. Even though pharmacological industries have produced a number of new antibiotics in the last three decades, resistance to these drugs by microorganisms has increased [4].

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\*Corresponding author

Received: 27 February 2011 / Accepted: 8 October 2011

Antibiotics are naturally occurring or synthetic organic compounds which inhibit or destroy selective bacteria, generally at low concentrations<sup>5</sup>. Microorganisms have developed resistance to many antibiotics and this has created immense clinical problem in the treatment of infectious diseases [5]. The increase in resistance to microorganisms due to indiscriminate use of antimicrobial drugs forced scientists to search for new antimicrobial substances from various sources including medicinal plants [6]. The antimicrobial efficacy attributed to some plants in treating diseases has been beyond belief. It is estimated that local communities have used about 10% of all flowering plants on Earth to treat various infections, although only 1% have gained recognition by modern scientists [7]. Owing to their popular use as remedies for many infectious diseases, searches for plants containing antimicrobial substances are frequent [8].

*Acacia nilotica* (Mimosaceae) is commonly called Gum Arabica and locally known as Nalla Tumma (Telugu). The powdered bark of the plant with a little salt is used for treating acute diarrhea [9]. *Ziziphus mauritiana* (Rhamnaceae) is locally called Regu (Telugu). Previous studies reveal that the whole plant and leaves are used in the traditional system of medicine as a tonic [10]. *Bauhinia variegata* L. (Caesalpiniaceae), having Kachnar as the common Indian name and locally called Daevakanchanamu (Telugu), is traditionally used in bronchitis, leprosy, tumours and ulcer. *Lantana camara* (Verbanaceae), commonly known as wild or red sage is the most widespread species of this genus and regarded both as a notorious weed and a popular ornamental garden plant [11]. However, it is listed as one of the important medicinal plants of the world [12]. *L. camara* contains lantadenes, the pentacyclic triterpenes which is reported to possess a number of useful biological activities. Several previous reports have described it as antifungal [13] anti proliferative [14]. Moreover, the hydroalcoholic extracts of the leaves have shown an effect on fertility, general reproductive performance, and teratology in rats [15].

Thus, the aim of the current study was to screen the antimicrobial activity of *Acacia nilotica*, *Ziziphus mauritiana*, *Bauhinia variegata* and *Lantana camara* against some selected clinical isolated strains. Although previous studies have been documented on the antimicrobial properties of these plants, this work is designed to evaluate the specific antibacterial activity of different extracts of these plants against tested microorganisms, in order to know the best extract against a specific microorganism.

## 2. Materials and methods

### *Plant material*

The fresh parts (leaves, barks and pods) of the plant were collected from a nearby medicinal plant nursery. The collection was under specialist supervision and these plants are commonly known to everyone. These plants were authenticated by a Botanist. The various plant parts were thoroughly washed, sun-dried for 7-10 days and ground into powder using a laboratory mill prior to analysis.

### *Extraction*

25 g of shade dried powder of plant materials was filled separately in the thimble and extracted successively with 150 ml each of methanol, ethanol and ethyl acetate using a Soxhlet extractor for 48 h. All the extracts were concentrated using rotary flash evaporator. After complete solvent evaporation, each of these solvent extracts was weighed and preserved at 4°C in airtight bottles until further use.

### *Microorganisms*

The clinical isolate cultures of *Bacillus subtilis*, *Bacillus circulans*, *Bacillus sphaericus*, *Staphylococcus aureus* and *Serratia liquefaciens* were obtained from the Microbiology department at BDRC (Bioaxis DNA Reseach Center), Hyderabad, India. The isolates were identified by conventional tests [16, 17]. All the strains were maintained on nutrient agar at 4°C and were sub cultured every month.

### *Determination of antibacterial activity by agar diffusion method*

Sensitivity of different bacterial strains to various extracts was measured in terms of zone of inhibition using agar diffusion assay (ADA) [18]. The plates containing Mueller-Hinton agar were spread with 0.2 ml of the inoculum. Wells (8 mm diameter) were cut out from agar plates using a sterilized stainless steel borer and filled with 0.1 ml of the extract. The plates inoculated with different bacteria were incubated at 37°C up to 48 h and diameter of any resultant zone of inhibition was measured. For each combination of extract and the bacterial strain, the experiment was performed in duplicate and repeated thrice.

### *Statistical analysis*

All the tests were conducted in triplicate. The data of all the parameters were statistically

analyzed and expressed as mean  $\pm$  S.D with the aid of SPSS 17.0 Windows version.

### 3. Result and Discussion

Plants are an important source of potentially useful structures for the development of new chemotherapeutic agents. The first step towards this goal is the *in vitro* antibacterial activity assay [19]. The potential for developing antimicrobials from higher plants appears rewarding as it will lead to the development of a phytomedicine to act against microbes. Plant-based antimicrobials have enormous therapeutic potential as they can serve the purpose with lesser side effects that are often associated with synthetic antimicrobials [20]. Continued further exploration of plant-derived antimicrobials is needed today.

A total of 8 extracts from 4 different plant species were investigated. Extracts of the different parts of the tested medicinal plants used in this study were shown in Table 1. The Antibacterial susceptibility by means of disk diffusion method showed that the 4 plant extracts tested exhibited an antimicrobial effect against *Bacillus circulans*, *Bacillus circulans*, *Bacillus circulans*, *Staphylococcus aureus* and *Serratia liquefaciens* (Table 2). Barks extracts of the four tested medicinal plants possess a lower zone on inhibitory activity as compared to the leaf extracts of these plants.

A total of 8 extracts from 4 different plant species were investigated; pods ethyl acetate extracts of *Lantana camara* showed the highest antimicrobial activity against *Acacia nilotica* and was found to give the most potent antimicrobial extract (Table 2). It is reported to have antimicrobial, antihyperglycemic and antiplasmodial properties [21-23]. Noticeably no antimicrobial activity was found in methanolic bark extract of *Acacia nilotica* against the tested bacteria except *Bacillus ciurlans* (Table 2).

When tested with disk diffusion method, *Lantana camara* ethyl acetate extracts of pods possess significant antimicrobial activity against tested clinical isolates (*Bacillus subtilis*  $2\pm 0.1$ mm, *Bacillus circulans*  $2.6\pm 0.2$ mm, *Bacillus sphaericus*  $2\pm 0.1$ mm, *Staphylococcus aureus*  $2.5\pm 0.1$ , and *Serratia liquefaciens*  $2.2\pm 0.1$ mm), followed by its ethyl acetate extracts of leaves. *L. camara* has been studied extensively for their antibacterial properties [24-26]. *L. camara* possess many important biological activities viz., antipyretic, antimicrobial, antimutagenic, antimicrobial, fungicidal, insecticidal, nematicidal, and others [24-26]. Lantadenes present in all *L. camara* is believed to be responsible for almost all the biological activities [27].

Therefore, antibacterial activities of *L. camara* leaf and flower extracts reported here might be due to the presence of some of these chemical constituents, particularly lantadenes and thesides in the extracts. Bhakta and Ganjewala [28] have recently confirmed the presence of phenolics, anthocyanins and roanthocyanidins in *L. camara* leaves which could also be responsible for the antibacterial properties of the *L. camara* reported here. Though, the mechanism of the action of these chemical constituents is not yet fully known, it is clear that the effectiveness of the extracts largely depends on the type of solvent used. Perhaps it is one of the reasons behind the differences in the antibacterial activities of the plants. Moreover, the effectiveness of the extracts varies with its concentration and the kind of bacteria used in the study. These differences in the susceptibility of the test organisms to the different extracts might be due to the variation in the rate at which active ingredients penetrate their cell wall and cell membrane structures [29, 30]. Thus, *S. aureus* was found to be resistant to all the extracts, which is most probably due to its outer membrane. Nevertheless, it is the ability of the active principle of the extracts that disrupt the permeability barrier of cell membrane structures and thus inhibit the bacterial growth [29, 30].

The antimicrobial potency of plants is believed to be due to tannins, saponins, phenolic compounds, essential oils and flavonoids [31]. It has been proposed that the mechanism of the antimicrobial effects involves the inhibition of various cellular processes, followed by an increase in plasma membrane permeability and finally, ion leakage from the cells [32]. In our study *B. subtilis* was found to be highly susceptible to *L. camara* ethyl acetate extracts of pods and leaves, similar to one of the early studies [33]. *Z. mauritiana* ethanol leaf extract was active against *S. aureus* supported by a previous study [34]. The ascending sequence of maximum antimicrobial activity against test microorganisms were as follows: *Lantana camara*, *Bauhinia variegata*, *Acacia nilotica* and *Ziziphus mauritiana* (Table 2).

Medicinal plants can be poisonous if wrong plant parts or wrong concentrations are used [35]. Herbal medicines are assumed to be harmless. Although herbal extracts need to be assured for quality control and efficacy for a particular dose, the results of present study clearly indicate that the antibacterial activity varies with the species of the plants and plant material used. Thus, the study ascertains the value of plants used in ayurveda, which could be of considerable interest to the development of new drugs. Studies are in progress to further evaluate the mechanisms of action of

these *active test* extracts on study organisms associated with certain human diseases

**Table 1.** Extracting solvents and parts of the medicinal plant used for antimicrobial activity

Medicinal plant	Family	Part used	Extracting Solvent
<i>Acacia nilotica</i>	Mimosaceae	Leaves & Barks	Methanol & Ethanol
<i>Ziziphus mauritiana</i>	Rhamnaceae	Leaves & Barks	Methanol & Ethanol
<i>Bauhinia variegata</i>	Caesalpiniaceae	Leaves & Barks	Ethanol
<i>Lantana camara</i>	Verbanaceae	Leaves & Pods	Ethyl acetate

**Table 2.** Zone of inhibitory activity (in millimeter) of different plant extracts against clinical isolates/Microorganisms

Microorganism/Clinical isolate strain	Zone of Inhibition (mm diameter)											
	<i>Acacia nilotica</i>				<i>Ziziphus mauritiana</i>				<i>Bauhinia variegata</i>		<i>Lantana camara</i>	
	Leaves		Bark		Leaves		Bark		Leaves	Bark	Leaves	Pods
	E	M	E	M	E	M	E	M	E	E	EA	EA
<b>Bacillus circulans</b>	0.7±0.0	1±0.1	1.5±0.2	0.6±0.0	0.6±0.1	Nil	Nil	Nil	1.3±0.2	1.1±0.0	2.3±0.2	2±0.1
<b>Bacillus subtilis</b>	1.1±0.1	1.3±0.2	1±0.0	Nil	1.7±0.3	0.6±0.0	0.9±0.0	1±0.1	0.7±0.1	0.4±0.0	2.4±0.1	2.6±0.2
<b>Bacillus sphericus</b>	1.1±0.0	0.8±0.0	1.7±0.1	Nil	0.9±0.0	1.1±0.1	Nil	Nil	1.1±0.1	Nil	1.8±0.1	2±0.1
<b>Staphylococcus aureus</b>	0.4±0.0	1±0.1	1±0.1	Nil	0.6±0.0	1±0.0	Nil	Nil	1.3±0.1	0.5±0.1	1.5±0.2	2.5±0.1
<b>Serratia liquefaciens</b>	1.3±0.1	0.9±0.1	1±0.0	Nil	1±0.1	Nil	0.6±0.0	0.8±0.1	1.1±0.0	0.6±0.0	2.4±0.1	2.2±0.1

E: Ethanol extract; M: Methanol extract; EA: Ethyl acetate extract

## References

- [1] Gordon, M. C. & David, J. N. (2001). Natural product drug discovery in the next millennium. *Pharm Bio*, 139, 8-17.
- [2] Clark, A. M. & Hufford, C. D. (1993). Disco and development of novel prototype antibiotics for opportunistic infections related to the acquired immunodeficiency syndrome. In Human Medical Agents from Plants, *American Chemical Society (ACS Symposium series 534)* (228-241). Washington, D.C. 534.
- [3] Principe, P. (1991). Monetising the pharmacological benefits of plants. US Environmental protection Agency, Washington, D. C. EPA/600/D-91/142 (NTIS PB91218958).
- [4] Cohen, M. L. (1992). Epidemiology of drug resistance: implications for a post-antimicrobial era. *Sci*, 257, 1050-1055.
- [5] Brooks, G. F., Carroll, K. C., Butel, J. S. & Morse, S. A. (2007). *The Growth and death of microorganism*. In Jawetz, Melnick and Adelberg's Medical Microbiology 24th ed McGraw-Hill Toronto, p. 57.
- [6] Davis, J. (1994). Inactivation of antibiotics and the dissemination of resistance genes. *Science*, 264, 375-382.
- [7] Karaman, I., Sahin, P., Gulluce, M., Ogutun, H., Songul, M. & Adiguzed, A. (2003). Antimicrobial activity of aqueous and methanol extracts of *Juniperus oxycedrus* L. *J. Ethnopharmacol*, 2837, 1-5.
- [8] Kafaru, E. (1994). *Immense help formative workshop*. In Essential Pharmacology; 1st Ed.: Lagos, Nigeria: Elizabeth Kafaru Publishers.
- [9] Betoni, J. E. C., Mantovani, R. P., Barbosa, L. N., Di-Stasi, L. C. & Fernandes, A. (2006). Synergism between plant extract and antimicrobial drugs used on *Staphylococcus aureus* diseases. *Mem. Inst. Oswaldo Cruz*, 101, 387-390.
- [10] Gill, L. S. (1992). *Ethanomedical uses of plants in Nigeria*. University of Benin Press, Benin City Nigeria.
- [11] Kirtikar, K. R. & Basu, B. D. (1993). *Indian Medicinal Plants, vol. II*. International Book Publisher, Dehradun.
- [12] Sharma, O. P., Dawra, R. K. & Makkar, H. P. S. (1987). Isolation and partial purification of *Lantana (Lantana camara* L.) toxins. *Toxicology Letters*, 37, 165-172.
- [13] Ross, I. A. (1999). *Medicinal plants of the world*. Chemical constituents, traditional and modern medical uses. New Jersey, Humana Press., xi, 415.
- [14] Kumar, V. P., Neelam, S. C. & Harish, P. (2006). Search for antibacterial and antifungal agents from selected Indian medicinal plants. *Journal of Ethopharmacology*, 107, 182-188.
- [15] Nagao, T., Abe, F. & Kinjo, J. (2002). Antiproliferative constituents in plants: Flavones from the leaves of *Lantana montevidensis* Briq. and consideration of structural relationship. *Biochemical Pharmaceutical Bulletin*, 25, 875-879.
- [16] Mello, F. B., Jacobus, D., Carvalho, K. & Mello, J. R. B. (2005). Effects of *Lantana camara* (Verbenaceae) on general reproductive performance and teratology in rats. *Toxicology*, 45, 459-466.

- [17] Ewing, H. & Martin, J. (1974). *Enterobacteriaceae*. In Manual of clinical microbiology, 2nd ed., edited by E. H. Lennette, E. H. Spaulding and J. P. Truant, Washington., p. 109.
- [18] Cowans, T. & Steelk, J. (1961). Diagnostic tables for the common medical bacteria. *J. Hyg. Camb*, 59, 357.
- [19] Tona, L., Kambu, N., Ngimbi, K. C. & Vlietinck, A. J. (1998). Antiamoebic and phytochemical screening of some Congolese medicinal plants. *J. Ethnopharmacol*, 61, 57-65.
- [20] Iwu, M. W., Duncan, A. R. & Okunji, C. O. (1999). *New antimicrobials of plant origin*. In: Janick J. ed. Perspectives on New Crops and New Uses. Alexandria, VA: ASHS Press, pp. 457-462.
- [21] Sotohy, S. A., Muller, W. & Ismail, A. A. (1995). In vitro effect of Egyptian tannin-containing plants and their extracts on the survival of pathogenic bacteria. *Dtsch. Tierarztl. Wochenschr*, 102, 344-348.
- [22] Meena, P. D., Kaushik, P., Shukla, S., Soni, A. K., Kumar, M. & Kumar, A. (2006). Anticancer and Antimutagenic Properties of *Acacia nilotica* (Linn) on 7, 12- dimethylbenz(a) anthracene-induced Skin Papillomagenesis in Swiss Albino Mice. *Asian Pac. J. Cancer Prev*, 7, 627-632.
- [23] El-Tahir, A., Satti, G. M. & Khalid, S. A. (1999). Antiplasmodial Activity of Selected Sudanese Medicinal Plants with Emphasis on *Acacia nilotica*. *Phytother. Res*, 13, 474-478.
- [24] Siddiqui, B. S., Raza, S. M., Begum, S. & Siddiqui, S. (1995). Pentacyclic triterpenoids from *Lantana camara*. *Phytochemistry*, 38, 681-685.
- [25] Mello, F. B., Jacobus, D., Carvalho, K. & Mello, J. R. B. (2005). Effects of *Lantana camara* (Verbenaceae) on general reproductive performance and teratology in rats. *Toxicology*, 45, 459-466.
- [26] Verma, R. K. & Verma, S. K. (2006). Phytochemical and termiticidal study of *Lantana camara* var. *aculeata* leaves. *Fitoterapia*, 77, 466-468.
- [27] Barre, J. T., Bowden, B. F., Coll, J. C., De Jesus, J., De La Fuente, V., Janairo, G. C. & Ragasa, C. Y. (1997). Abioactive triterpenoid from *Lantana camara*. *Phytochemistry*, 4, 321-324.
- [28] Bhakta, D. & Ganjewala, D. (2009). Effect of leaf positions on total phenolics, flavonoids and proanthocyanidins content and antioxidant activities in *Lantana camara* (L). *Journal of Scientific Research*, 1, 363-369.
- [29] Nikaido, H. & Vaara, M. (1985). Molecular basis of bacterial outer membrane permeability. *Microbiology Review*, 1, 1-32.
- [30] Priya, K. & Ganjewala, D. (2007). Antibacterial Activities and Phytochemical Analysis of Different Plant parts of *Nyctanthes arbor-tristis* L. *Research Journal of Phytochemistry*, 1, 61-67.
- [31] Aboaba, O. & Efuwape, B. M. (2001). Antibacterial Properties of Some Nigerian Species. *Bio. Res. Comm*, 13, 183-188.
- [32] Walsh, S. E., Maillard, J. Y., Russel, A. D., Catrenich, C. E., Charbonneau, A. L. & Bartolo, R. G. (2003). Activity and Mechanism of Action of Selected Biocidal Agents on Gram -positive and -negative Bacteria. *J. Appl. Microbiol*, 94, 240-247.
- [33] Deepak, G., Silviya, S. K. & Hayat, K. (2009). Biochemical compositions and antibacterial activities of *Lantana camara* plants with yellow, lavender, red and white flowers. *EurAsia J BioSci*, 3, 69-77.
- [34] Abalaka, M. E., Daniyan, S. Y. & Mann, A. (2010). Evaluation of the antimicrobial activities of two *Ziziphus* species (*Ziziphus mauritiana* L. and *Ziziphus spinachristi* L.) on some microbial pathogens. *African Journal of Pharmacy and Pharmacology*, 4(4), 135-139.
- [35] Frohne, D. (1999). Plant intoxicants - is there a risk from herbal preparation? *J. hytotherap*, 20, 201-202.