The Effect of Gibberellic Acid and Chilling Stratification on Seed Germination of Eastern

Black Walnut (Juglans nigra L.)

P. Parvin^{*1}, M. Khezri², I. Tavasolian², H. Hosseini¹

¹ Department of Horticulture, College of Agriculture, Shahid Bahonar University, Kerman, Iran

² Horticultural Research Institute, Shahid Bahonar University, Kerman, Iran

Received: 15 April 2015 Acc

Accepted: 15 June 2015

Abstract

Eastern black walnut (*Juglans nigra* L.) is used as a rootstock for the Persian walnut (*Juglans regia* L.) in some parts of the world and also has an important role in forestry and wood industry. Due to the deep physiological dormancy, the seed often shows an inconsistent or low germination percentage, making establishment difficult. This experiment was carried out as a completely randomized design with eight treatments and 16 replicates in a controlled greenhouse. The objective of this study was to determine the best treatment of breaking dormancy. Treatment groups consisted of seed priming with GA₃ (400 and 800 ppm) solution for 24 hours, chilling stratification (one month and two months) and the combined treatments of chilling stratification and GA₃. Results showed that the germination rate for separate application of both concentrations of GA₃ and one month chilling treatment was zero, as no seeds germinated. The highest percentage of seed germination (69.27 %) was recorded with the combined treatment of two months chilling and GA₃ (400 ppm). Also, this treatment showed significant differences for morphological, physiological and biochemical parameters compared to other treatments. It was found that the application of the combined treatment of chilling stratification and GA₃ was effective in increasing seed germination percentage and rate as well as improving growth parameters of Eastern black walnut seedlings.

Keywords: Juglans nigra L., Moist-chilling, Seed dormancy, Seed priming.

Introduction

Eastern black walnut (*Juglans nigra* L.) is one of the important species and the largest of the North American walnuts, reaching a height of 45m and a trunk diameter of 2 m. It is native to the deciduous forests of the eastern USA and Canada. Eastern black walnut bears nuts with hard, black shells and stronger flavored kernels than those of *J. regia*. The irregular grooves and ridges on the shell separate it from the other species native to the USA. Eastern black walnut is more resistant to frost than the Persian walnut and now planted in Europe as both a timber species and a rootstock. It is the most valuable

hardwood produced in US and used in various industries (Mc Granahan and Leslie, 2009). The seeds of this species have deep physiological dormancy that is controlled by seed coat and embryo dormancy. Germination of healthy and live seeds may be delayed even in good environmental conditions such as light, oxygen, water and chemicals due to the seed dormancy (Hilhorst, 1995). Therefore, analyzing the causes of dormancy and evaluating methods of breaking dormancy in order to increase seed germination percentage and rate is necessary (Rajabiyan *et al.*, 2007).

* Corresponding author: Email: parisa.parvin35@yahoo.com

Seed germination is a complex process that started with the absorption of water and after a short pause; the enzyme is activated (Matilla and Matilla-Vazquez, 2008). Gibberellins (GAs) are the hormones proposed to control primary dormancy (the form of dormancy that is acquired during seed development) by inducing germination (Hilhorst and Karssen, 1992). Gibberellic acid (GA₃) is an exogenous growth regulator that promotes germination by stimulating the activation of food-mobilizing enzymes (Hartman and Kester, 1983). The use of GA₃ has been studied in fruit culture as a way to increase seed germination and therefore to obtain a uniform seedling size in the nurseries (Hore and Sem, 1993 and Dhupper, 2013). Chilling treatments is often practiced to enhance the germination of dormant seeds (Bello et al., 1998 and Hassan and Fetouh 2014). It is believed that moist-chilling treatment alters the inhibitor-promoter balance. Various dormancy breaking and germination stimulating treatments have been tried with seeds of many fruit species such as papaya (Nagao and Furutani, 1986), persimmon (Taha, 1987), peach (El-Khoreiby and Salem, 1985; El-Dengawy, 1997), pears (Pipinis et al., 2012a) and loquat (El-Dengawy and El-Refaey, 2005; Polat, 1997). Although application of GA3 and moist-chilling treatments seem the most promising techniques in woody species (Powell, 1987 and Nasri et al., 2013), it has not been investigated on breaking dormancy of Eastern black walnuts seeds yet. Therefore, the aim of this study was to determine the effect of priming with GA3 and chilling stratification on improving germination and growth of Eastern black walnut seeds.

Materials and Methods

Seed pretreatment

Seeds were collected from a vigor genotype of Eastern black walnut (*Juglans nigra* L.) tree at Botanical Garden, College of Agriculture and Natural Resources, Karaj, Iran. Seeds were collected in mid-October 2013, when the seeds had desiccated to about 12% moisture on a dry weight basis. The seeds were surface sterilized by soaking in 5% sodium hypochlorite solution for 10 minutes and subsequently rinsed thoroughly with sterilized water prior to germination or chilling. This experiment was carried out as completely randomized design in a controlled greenhouse at Shahid Bahonar University of Kerman, Iran, with eight treatments and 16 replicates. Each replicate consisted of four pots with one plant per pot. Seeds were subjected to one of the following treatments: GA₃ (400 ppm); GA₃ (800 ppm); 1 month chilling; 2 months chilling; 1 month chilling + GA_3 (400 ppm); 1 month chilling + GA_3 (800 ppm); 2 months chilling + GA₃ (400 ppm); 2 months chilling + GA₃ (800 ppm). Chilling temperature was set to 4 ± 1 °C. The soaking time in GA₃ solution was 24 hours.

The treated seeds were sown in pots contained cocopeat and perlite sterilized by autoclaving (2:1, respectively). After sowing, the plastic pots were watered regularly and shaded in a greenhouse.

Germination test

Seeds were considered germinated when the radicle reached to half the length of the seed. At the end of germination period (eight weeks), the germination percentage (GP) was calculated using the following formula:

 $GP = \sum G/N*100$

Where GP is the germination percentage, G is the numbers of germinated seeds and N is the numbers of all seeds (Copeland *et al.*, 2001).

Germination rate (GR) was calculated using following equation:

$$Gr = \Sigma n / \Sigma (Dn)$$

Where n is the number of seeds that germinated on day D and D is the number of days counted from the beginning of the test (Copeland *et al.*, 1995).

Morphological and physiological parameters

At the end of experiment, seedlings were cut at soil surface and the roots washed free of soil. After measuring the length of shoot and root, shoot and root fresh weights and root volume were recorded. Root volume was determined by immersing root systems in a container of water placed on a balance. The displaced water (measured in grams) is equal to the volume (measured in cubic centimeters) of the root system in that 1 g of water equals 1 cm³ at room temperature (Burdett, 1979). In order to estimate the dry weight, the samples were oven-dried at 75 °C for 48 hours, until constant weight was reached. Root area was determined using the method of Atkinson (1980):

Y (cm²) = 2{(root length × root volume)}^{0.5} Y (cm²) = root area

The Chlorophyll index of leaf was measured by using Minolta (SPAD) Chlorophyll Meter (SPAD-502 Minolta Sensing Inc., Japan).

Biochemical parameters

The amount of photosynthetic pigment (chlorophyll a, b, total and carotenoids) was determined according to the method of Lichtenthaler *et al.* (1987). The pigment extract was measured vs. a blank of 80% (V/V) acetone at wavelengths of 646.8 nm and 663.2 nm for chlorophyll assays. Reducing sugars was measured by

the method described by Somogyi (1952). The absorbance was measured at 600 nm. Glucose was used as standard solution.

Statistical analysis

Analysis of variance and comparison of means were performed using GLM procedure of SAS (SAS Institute, Inc., 2002). Data were subjected to transformation and normalization where necessary (Zar, 1999). Significant differences among the mean values were compared by Fisher's Least Significant Difference (LSD) ($P \le 0.05$).

Results

Results showed that neither the seeds treated by GA_3 at 400 ppm and 800 ppm nor one month chilling stratification treatment germinated in the greenhouse. Therefore, data analysis and mean separation were performed for combined treatments. Seed germination percentage and rate were significantly different among treatments. Analysis of variance showed that there were significant differences in terms of morphological, physiological and biochemical parameters (Table 1). It was also found that two months chilling stratification in combination with GA_3 (400 and 800 ppm) resulted significant higher germination percentage and rate than other treatments (Fig. 1).

Table1. Analysis of variance of priming with GA3 and chilling stratification on germination of Eastern black walnut seed (Juglans nigra L.)

16	Germination	Germination	Root	Shoot	Shoot	noot Root Root area	Destaura		Class I FW
dī.	percentage	rate	length	length	length diameter volume Root area	Root area	I otal F w	Shoot Fw	
					MS				
4	2815.20**	179.50**	34.08**	33.60**	0.26 ^{ns}	8.38**	17.36**	9.65**	4.83**
50	2.96	33.53	10.40	6.81	0.13	1.18	3.51	1.92	0.42
df	Root FW	Shoot DW	Root DW	Chlorophyll	Chlorophyll	Chlorophyll	Chlorophyll	Carotenoid	Reducing
ui.	Root I W		Root D II	Index	Total	a	b	Curotenoid	sugars
					MS				
	df. 4 50 df.	df. Germination percentage 4 2815.20** 50 2.96 df. Root FW	df.Germination percentageGermination rate42815.20**179.50**502.9633.53df.Root FWShoot DW	df.Germination percentageGermination rateRoot length42815.20**179.50**34.08**502.9633.5310.40df.Root FWShoot DWRoot DW	df.Germination percentageGermination rateRoot lengthShoot length42815.20**179.50**34.08**33.60**502.9633.5310.406.81df.Root FWShoot DWRoot DWChlorophyll Index	df.Germination percentageGermination rateRoot lengthShoot lengthShoot diameter42815.20**179.50**34.08**33.60**0.26^ns502.9633.5310.406.810.13df.Root FWShoot DWRoot DWChlorophyll IndexChlorophyll TotalMS	df.Germination percentageGermination rateRoot lengthShoot lengthShoot diameterRoot volume42815.20**179.50**34.08**33.60**0.26 ^{BS} 8.38**502.9633.5310.406.810.131.18df.Root FWShoot DWRoot DWChlorophyll IndexChlorophyll TotalChlorophyll a	df.Germination percentageGermination rateRoot lengthShoot lengthShoot diameterRoot volumeRoot area42815.20**179.50**34.08**33.60**0.26^ns8.38**17.36**502.9633.5310.406.810.131.183.51df.Root FWShoot DWRoot DWChlorophyll IndexChlorophyllChlorophyll TotalChlorophyll aChlorophyll b	df.Germination percentageGermination rateRoot lengthShoot lengthShoot diameterRoot volumeRoot areaTotal FW42815.20**179.50**34.08**33.60** 0.26^{ns} 8.38^{**} 17.36^{**} 9.65^{**} 502.9633.5310.40 6.81 0.13 1.18 3.51 1.92 df.Root FWShoot DWRoot DWChlorophyll IndexChlorophyll TotalChlorophyll aChlorophyll bChlorophyll corenoid

Table 1. Continued

Treatment	4	3.22*	1.68**	1.60**	270.75**	271.28**	94.66**	45.91**	34.34**	0.001**
Error	50	1.23	0.09	0.11	0.33	0.42	0.19	0.14	0.09	0.00008
^{ns} , **and * means non-significant, significant at 1% and 5% of probability respectively.										



Fig. 1. The effect of priming with GA₃ and chilling stratification on germination percentage and rate of Eastern black walnut seed (*Juglans nigra* L.). Values are means \pm SE and the means with the same letter are not significantly different at 5% LSD test.

Results showed that two months chilling stratification in combination with GA₃ (400 ppm) significantly improved seedling growth characteristics including seedling length, root length, root volume, root

area, total fresh and dry weights as well as total chlorophyll and reducing sugars compared to other treatments (Tables 2, 3 and 4).

Treatments*	Stem length (cm)	Stem diameter (mm)	Root length (cm)	Root volume (cm ³)	Root area (cm ²)
2 months chilling	20.45±2.53b**	3.40±0.39ab	20.45±2.50b	7.18±1.2b	17.61±1.1 ab
1 month chilling+ GA ₃ 400ppm	19.99±2.90b	3.32±0.35ab	22.24±2.89ab	5.36±0.92c	15.33±1.35 c
1 month chilling+ GA ₃ 800ppm	21.86±1.62b	3.56±0.4ab	20.31±3.28b	6.63±1.20b	16.32±1.99 bc
2 months chilling+ GA ₃ 400ppm	24.58±3.06a	3.66±0.38a	24.58±3.61a	7.63±0.67a	18.64±2.29 a
2 months chilling+ GA ₃ 800ppm	21.36±2.67b	3.30±0.24b	23.07±3.66ab	6.27±1.27bc	16.88±2.28 bc

Table 2. The effect of GA ₃ and chilling stratification	on morphological parameters of Eastern bla	ck walnut seedling (Juglans nigra L.)
--	--	---------------------------------------

* Application of GA3 at both concentrations (400 and 800 ppm) and also one month chilling stratification had no germination percentage, therefore, the analysis of mean comparison was carried out among other treatments. ** Values are means \pm SE and the means with the same letter are not significantly different at 5% LSD test.

Table 3. The effect of GA ₃ and chilling stratification on physiological	parameters of Eastern black walnut seedling (Juglans nigra L.)

Treatments*	Total FW (g)	Shoot FW (g)	Root FW (g)	Shoot DW (g)	Root FW (g)	Chlorophyll Index
2 months chilling	12.25±1.42b**	4.88±0.96b	7.19±0.78b	1.38±0.36b	1.53±0.37b	37.70±0.39d
1 month chilling+ GA ₃ 400ppm	11.80±1.66b	4.57±0.43b	7.08±1.20b	1.29±1.27b	1.35±0.32b	34.59 ±0.77e
1 month chilling+ GA ₃ 800ppm	12.44±1.20b	4.67±0.68b	7.59±1.22ab	1.52±0.24b	1.37±0.31b	39.99 ±0.40c
2 months chilling+ GA ₃ 400ppm	14.23±0.96a	6.21±0.34a	8.46±0.98a	2.26±0.27a	2.28±0.30a	47.06 ±0.49a
2 months chilling+ GA ₃ 800ppm	13.03±1.56b	5±0.65b	7.58±1.27ab	1.52±0.34b	1.55±0.39b	44.04 ±0.70b

* Application of GA3 at both concentrations (400 and 800 ppm) and also one month chilling stratification had no germination percentage, therefore, the analysis of mean comparison was carried out among other treatments. ** Values are means \pm SE and the means with the same letter are not significantly different at 5% LSD test.

Table 4. The effect of GA ₃ ar	nd chilling stratification on bioch	emical parameters of Eastern black	walnut seedling (Juglans nigra L.)
	0	1	

Treatments*	Total Chlorophyll (mg/g FW)	Chlorophyll a (mg/g FW)	Chlorophyll b (mg/g FW)	Carotenoid (mg/g FW)	Reducing sugars (mg/g FW)
2 month chilling	20.09 ±0.94d**	13.95 ±0.46d	6.13 ±0.48d	5.76±0.33d	$0.11 \pm 0.009 b$
1 month chilling+ GA400ppm	16.24 ±0.36e	13.95 ±0.55e	4.80 ±0.33e	3.13±0.37e	$0.11 \pm 0.008 b$
1month chilling+ GA800ppm	22.90 ±0.62c	15.26 ±0.38c	7.64 ±0.31c	5.76±0.33c	$0.12\pm0.008b$
2 months chilling+ GA400ppm	29.33 ±0.68a	19.17 ±0.42a	10.16 ±0.41a	7.90±0.28a	$0.14\pm0.011a$
2 months chilling+ GA800ppm	25.17 ±0.48b	16.94 ±0.31b	8.22 ±0.33b	6.68±0.19b	$0.11 \pm 0.008 b$

Application of GA₃ at both concentrations (400 and 800 ppm) and also one month chilling stratification had no germination percentage, therefore, the

analysis of mean comparison was carried out among other treatments. ** Values are means \pm SE and the means with the same letter are not significantly different at 5% LSD test.

Discussion

Results showed that Eastern black walnut seeds display an endogenous dormancy that might be removed by priming with moist-chilling and GA_3 for a certain period of time. It was found that seeds treated with GA_3 at two concentrations (400 ppm and 800 ppm) for 24 hours and one month chilling treatment alone showed zero germination percentage. A similar trend was observed by Conner *et al.* (2008) on 'Fry' muscadine seed.

The effect of GA_3 and stratification on enhancing growth could be attributed to the solubility of fats and sugars due to stratification plus the increasing of gibberellins synthesizing enhanced the growth. In addition, the improving effect of GA_3 and stratification on seed germination may reflect on enhancing the shoot parameters. These results are in agreement with Dahkaei (2009) on *Danae racemosa*, Rawat *et al.* (2010) on *Punica granatum* and Hassan and Fetouh (2014) on seeds of *Magnolia grandiflora*.

The effect of GA_3 and stratification on root parameters followed the same trend of its effect on shoots. The promotion effect of GA_3 and stratification on root parameters might be explained through the role of GA_3 and stratification in enhancing gibberellins synthesis which also leads to increase the growth and root branching and overall increased roots fresh weight (Penfield *et al.* 2005). The results of the present experiment are in agreement with Rawat *et al.* (2010) on *Punica granatum* and Hassan and Fetouh (2014) on *Magnolia grandiflora* seeds.

It was also found that GA_3 (400 ppm) combined with two months chilling stratification increased the chlorophyll and carotenoid contents and reducing sugars. These results are in accordance with the findings of Amooaghaie (2009) on *Ferula ovina* seeds. The increase of reducing sugars could be explained by the possible conversion of fatty acids to sugars and the conversion of soluble sugars to reducing sugars. It is also possible that insoluble sugars were converted to soluble and reducing sugars during seed germination and seedling growth and development. Previous studies showed a preference of sugars to fatty acids for energy production during germination of oil seed crops (To *et al.*, 2002; Tonguc *et al.*, 2012).

It has been reported that seeds of the Persian walnut (Juglans regia L.) treated with two months of chilling had higher germination percentage and rate compared to one month chilling treatment and significantly improved seedling characteristics (Vahdati et al. 2012). Hassan and Fetouh (2014) and Pipinis et al., (2011b) reported a similar trends on seeds of Magnolia grandiflora and Cercis siliquastrum seeds, respectively. However, in the present study, one month of chilling treatment showed no impact on improving seed germination while combined treatment of two months of chilling and GA₃ (400 ppm) showed the highest seed percentage and rate. According to Khan (1977) and Hassan and Fetouh (2014), stratification affects metabolic processes including changes in hormones, disappearance of ABA, activation of GA₃ and initiation of germination. Previous studies have shown that after-ripening and moist (warm or cold) stratification affect metabolic and physiological changes in seeds that involve both the embryo and its covering layers (Leubner-Metzger, 2005; Bair et al., 2005 and Shakarishvili et al., 2013). It was also indicated that seed stratification causes a rapid decline in the abscisic acid (ABA) content and ABA sensitivity and increases GA3 sensitivity of imbibed dormant seeds (Gubler et al., 2005).

Exogenous application of GA_3 has been reported to be effective in breaking dormancy and substituting for the chilling requirement in seeds of many species (Karam and Al-Salem, 2001; Pipinis *et al.*, 2011a; 2012; Smiris *et al.*, 2006 and Dhupper, 2013). Ghayyad *et al.* (2010) reported that GA_3 is effective in shortening the chilling requirement. However, in the present study, the application of GA_3 treatments separately showed zero germination percentage. Since the highest percentage of seed germination and seedling growth parameters were recorded for a two months chilling period combined with GA_3 (400 ppm), it may be possible that higher concentration of GA_3 (800 ppm) could not shorten the chilling period.

It was found that GA increases the growth potential of embryo and promotes germination and is necessary to overcome the mechanical restraint conferred by the seed covering layers by weakening of the tissues surrounding the radicle (Finch-Savage and Leubner-Metzger, 2006).

The combination of chilling stratification and GA_3 pretreatment has been reported to improve germination in *Prunus* species (Imani *et al.*, 2011) and cherry seeds (Al-Absi, 2010). Moreover, GA_3 and chilling stratification affect physiological and metabolic activities of seeds resulting in early germination, which was previously reported (Amooaghaie, 2010; Zeinalabedini, 2009 and Pipinis *et al.*, 2012b).

The combination of GA_3 (400 ppm) and chilling might be more effective in bringing a hormonal shift that not only enhanced germination but also sped it up. Such results are in accordance with those of Nasri *et al.* (2013), Amooaghaie (2009) and El-Dengawy (1997) on peach seeds and Chin *et al.* (1992) on kiwi fruit seeds. They concluded that the combination between a suitable moist-chilling period and an effective level of GA_3 would considerably enhance seed germination, which is in accordance with the present study.

In conclusion, it was found that the combined treatments of two months of chilling followed by soaking in GA_3 (400 ppm) solution for 24 hours might be recommended for improving the seed germination process and improving growth characteristics of the Eastern black walnut seedlings.

Acknowledgments

The authors thank Shahid Bahonar University.

References

- Al-Absi KM (2010) The effects of different pre-sowing seed treatments on breaking the dormancy of Mahaleb cherries, (*Prunus mahaleb* L.) seeds. Seed Science and Technology. 38, 332-340.
- Amooaghaie R (2009) The effect mechanism of moistchilling and GA₃ on seed germination and subsequent seedling growth of *Ferula ovina Boiss*. The Open Plant Science Journal. 3, 22-28.
- Amooaghaie R (2010) Effect of gibberellin and stratification on stimulating seed germination and subsequent seedling growth, of "Japanese medlar". Journal of Biology. 23, 299-308. [In Persian].
- Atkinson D (1980) The distribution and effectiveness of the roots of tree crops. Horticulture Review. 2, 424-490.
- Bair0 NB, Meyer SE, Allen PS (2005) A hydrothermal after-ripening time model for seeds dormancy loss in *Bromus tectorum* L. Seed Science Research. 16, 17–28.
- Bello IA. Hatterman-Valentini H, Owen MDK (1998) LITELLS of stratification, temperature, and oxygen on woolly cup grass (*Eriochloa villosa*) seed dormancy. Weed Science. 46, 526-529.
- Burdett AN (1979) A nondestructive method for measuring the volume of intact plant parts. Canadian Journal of Forest Research. 9, 120-122.
- Conner PJ (2008) Effects of stratification, germination, temperature and pretreatment with gibberellic acid and hydrogen peroxide on germination of 'Fry' muscadine (*Vitis rotundifolia*) seed. Hort Science. 43, 853–856.
- Copeland LO, Mc Donald MB (1995) Principals of seed science and Technology. Third Edition. Chapman and Hall, New Tork. Pp 236.

- Copeland LO, Mc Donald MB (2001) Principles of Seed Science and Technology. Dordrecht, the Netherlands: Kluwer Academic Publishers.
- Dahkaei MNP (2009) Effect of Gibberellic acid, temperature and cold moist stratification on seed germination of Danae racemosa. Acta Horticulturae. 813, 445-452.
- Dhupper R (2013) Effect of gibberellic acid on seed germination and seedling growth behaviour in three desert tree species. The Journal of Biological Chemistry research. 30(1), 227-232.
- El-Dengawy EFA (1997) Physiological and biochemical studies on seeds dormancy and germination process in deciduous fruit trees. Ph.D. Thesis. Fac. Agric. Mansoura University, Egypt.
- El-Dengawy El-Refaey FA (2005) Promotion of seed germination and subsequent seedling growth of loquat (*Eriobotrya japonica*, Lindl) by moistchilling and GA₃ applications. Scientia Horticulture. 105, 331–342.
- El-Khoreiby AMK, Salem TA (1985) Effect of stratification and GA₃ on seed germination and subsequent seedling growth of apricot and peach. Bulletin of Faculty of Agriculture, Cairo University Egypt. 36, 299–309.
- Finch-Savage WE, Leubner-Metzger G (2006) Seed dormancy and the control of germination. New Phytologist. 171, 501-523.
- Ghayyad M, Kurbysa M, Napolsy G (2010) Effect of endocarp removal, gibberelline, stratification and sulfuric acid on germination of Mahaleb (*Prunus mahaleb* L.) seeds. American-Eurasian Journal of Agricultural and Environmental Science. 9, 163-168.
- Gubler F, Millar AA, Jacobsen JV (2005) Dormancy release. ABA and pre-harvest sprouting. Current Opinion in Plant Biology. 8, 183–187.

- Hartman HT, Kester DE (1983) Plant propagation principles and practices. 4th Ed. Prentice Hall, Englewood Cliffs, NJ.
- Hassan FA, Fetouh MI (2014) Seed germination criteria and seedling characteristics of *Magnolia* grandiflora L. trees after cold stratification treatments. International Journal of Current Microbiology and Applied Sciences. 3(3), 235-241.
- Hilhorst HWM (1995) A critical update on seed dormancy. I. Primary dormancy. Seed Science Research. 5, 61-73.
- Hilhorst HWM, Karssen CM (1992) Seed dormancy and germination: the role of abscisic acid and gibberellins and the importance of hormone mutants. Plant Growth Regulation. 11, 225-238.
- Hore JK, Sem SK (1993) Viability of papaya (*Carica papaya* L.) seeds under different pre-storage treatments. Environment and Ecology. 11, 273-275.
- Imani A, Rasouli M, Tavakoli R, Zarifi E, Fatahi R, Barba-Espín G, Martínez-Gómez P (2011) Optimization of seed germination in *Prunus* species combining hydrogen peroxide or gibberellic acid pre-treatments with stratification. Seed Science and Technology. 39(1), 204-207.
- Karam NS, Al-Salem MM (2001) Breaking Dormancy in Arbutus and rachne L. by Stratification and Gibberellic acid. Seed Science and Technology. 29, 51-56.
- Khan AA (1977) Seed dormancy: changing concepts and theories. In: Khan AA (Ed.), the Physiology and Biochemistry of Seed Dormancy and Germination. North-Holland, Amsterdam. pp. 29-50.
- Leubner-Metzger G (2005) 1, 3 Glucanase gene expression low-hydrated seeds as a mechanism

for dormancy release during tobacco afterripening. Plant Journal. 41, 133–145.

- Lichtenthaler HK (1987) Chlorophylls and Carotenoids Pigments of Photosynthetic Biomemberanes. Methods in Enzymology. 148, 350-382.
- Matilla AJ, Matilla-Vazquez MA (2008) Involvement of ethylene in seed physiology. Plant Science. 175, 87-97.
- Mc Granahan GH, Leslie C (2009) Breeding Walnuts. In: Jain SM, Priyadarshan PM (Eds.). Breeding plantation tree crops: Temperate species. Springer Science. 249-273.
- Nagao MA, Furutani SC (1986) Improving germination of papaya seed by density separation, potassium nitrate and gibberellic acid. HortScience. 21, 1439–1440.
- Nasri F, Ghaderi N, Mohammadi J, Mortazavi SN, Koshesh Saba M (2013) The effect of gibberellic acid and stratification on germination of alstroemeria (*Alstroemeria ligtu* hybrid) seed in vitro and in vivo conditions. Journal of Ornamental Plants. 3(4), 221-228.
- Penfield S, Josse EM, Kannangara R, Gilday AD, Halliday KJ, Graham IA (2005) Cold and light control seed germination through the bHLH transcription factor spatula. Current Biology. 15, 1998-2006.
- Pipinis E, Milios E, Smiris P (2011a) Effect of sulphuric acid scarification, cold moist stratification and gibberellic acid on germination of *Paliurus spinachristi* Mill. Seeds. Forestry Ideas. 17(1), 45-52.
- Pipinis E, Milios E, Mavrokordopoulou O, Gkanatsiou CH, Aslanidou M, Smiris P (2012a) Effect of Pretreatments on Seed Germination of (*Prunus mahaleb* L.). Notulae Botanicae Horti Agrobotanic. 40, 183-189.
- Pipinis E, Milios E, Smiris P, Gioumousidis CH (2011b) Effect of acid scarification and cold moist

stratification on the germination of *Cercis* siliquastrum L. seeds. Turkish Journal of Agriculture and Forestry. 35, 259-264.

- Pipinis E, Milios E, Kiamos N, Mavrokordopoulou O, Smiris P (2012b) Effects of stratification and pretreatment with gibberellic acid on seed germination of two Carpinus species. Seed Science and Technology. 40, 21-31.
- Polat AA (1997) Determination of germination rate coefficients of loquat seeds and their embryos stratified in various media for different durations. Turkish Journal of Agriculture and Forestry. 21, 219–224.
- Powell L (1987) The hormonal control of bud and seed dormancy in woody plants. In: Davies, P.J. (Ed.), Plant Hormones and Their Role in Plant Growth and Development. Martinus Nijhof Publishers, Dordrecht. Pp. 539- 552.
- Rajabiyan T, Saboora A, Hassani B, FallahHosseini H (2007) Effect of gibberellic acid on seed germination and cold acetone (*Ferula assafoetida* L.). Quarterly Scientific-Research of Medicinal and Aromatic Plants. 23, 391-404.
- Rawat JMS, Tomar YK, Rawat V (2010) Effect of stratification on seed germination and seedling performance of wild pomegranate. Journal of American Science. 6(5), 97-99.
- SAS Institute, Inc. (2002) SAS User's guide: Statistics. Version 9.1. Gray, N.C.
- Shakarishvili N, Asieshvili L, Eradze N, Siradze M (2013) Effect of stratification on seed germination and epicotyl dormancy in *Arbutus* andrachne l. Bulletin of the Georgian National Academy of Sciences. 7(1), 79-82.
- Smiris P, Pipinis E, Aslanidou M, Mavrokordopoulou O, Milios E, Kouridakis A (2006) Germination study on Arbutus unedo L. (Ericaceae) and Podo cytisus caramanicus Boiss and Heldr

(Fabaceae). Journal of Biological Research. 5, 85-91.

- Somogy M (1952) Note on sugar determitation. Journal of Biochemistry. 195, 19-29.
- Taha FA (1987) Effect of plant growth regulators on seed germination and seedling characters of persimmon root-stock (*Diospyros kaki* L.). Egyptian Journal of Horticulture. 14, 15–20.
- To JPC, Reiter WD, Gibson SI (2002) Mobilization of seed storage lipid by Arabidopsis seedlings is retarded in the presence of exogenous sugars. BMC Plant Biology. 2, 4-10.
- Tongue M, Elkoyunu R, Erbas S, Karakurt Y (2012) Changes in seed reserve composition during germination and initial seedling development of safflower (*Carthamus tinctorius* L.). Turkish Journal of Biology. 36, 107-112.

- Vahdati K, Aslani Aslamarz A, Rahemi M, Hassani D, Leslie CH. (2012) Mechanism of seed dormancy and its relationship to bud dormancy in Persian walnut. Environmental and Experimental Botany. 75, 74-82.
- Zar JH (1999) Biostatistical analysis. 4th Ed. Prentice Hall, NJ.
- Zeinalabedini M, Majourhat K, Khayam-Nekoui M, Hernández JA, Martínez-Gómez P (2009) Breaking seed dormancy in long-term stored seeds from Iranian wild almond species. Seed Science Technology. 37, 267-275.