Protective effect of methanolic extract of Berberis integerrima Bunge. root on carbon tetrachloride-induced testicular injury in Wistar rats

Fereshteh Rafiee M.Sc., Vahid Nejati Ph.D., Reza Heidari Ph.D., Hossein Ashraf M.Sc.

Department of Biology, Faculty of Science, Urmia University, Urmia, Iran.

Corresponding Author
Fereshteh Rafiee, Department of
Biology, Faculty of Science,
Urmia University, Urmia, Iran.
Email: Fereshteh.rafiee1987@gmail.com
Tel: (+98) 6925222172

Received: 15 July 2014 Revised: 3 May 2015 Accepted: 10 May 2015

Abstract

Background: Tissue protective effect of compounds with antioxidant properties has been demonstrated. The alkaloids found in barberry root are considered as antioxidants.

Objective: According to barberry protective effects in different tissues, in this study, the protective effect of Berberis integerrima Bge. root (MEBIR) was evaluated against CCl4-induced testicular damages in Wistar rats.

Materials and Methods: 40 mature male rats were randomly divided into 5 groups: 1: Normal control, 2: Sham: received CCl4 diluted in olive oil (50% v/v; 1ml/kg bw), intraperitoneally, twice a week for 4 weeks, 3 and 4: Sham rats treated with MEBIR (250 and 500 mg/kg bw) for 28 days, 5: Sham rats treated with silymarin (50 mg/kg bw) for 28 days. After 28 days, serum testosterone level, absolute testis weight, catalase activity, malondialdehyde level, and histological parameters were investigated.

Results: In the treated rats with MEBIR (250 and 500 mg/kg bw) or silymarin (50 mg/kg bw), there was a significant increase in the absolute testis weight, testosterone level, seminiferous tubules diameter (p<0.001), thickness of the epithelium, tubule differentiation index (p<0.001), spermiogenesis index (p<0.001), the activity of catalase, and a significant decrease in interstitial tissue thickness (p<0.001) and malondialdehyde level in comparison with CCl4-treated group. The effect of the MEBIR at dose of 500 mg/kg bw is more than that of the standard drug, silymarin (50 mg/kg bw).

Conclusion: From the results, it is suggested that the protective effects of MEBIR is possibly due to antioxidant effects of its bioactive compounds.

Key words: Berberis, Antioxidant, Carbon tetrachloride, Testis, Rats.

This article extracted from M.Sc. thesis. (Fereshteh Rafiee)

Introduction

ale sexual derangement is caused various agents such alcoholism, drug abuse, smoking, some drugs and toxic chemicals (1). Carbon tetrachloride is one of the compounds that damage testis and other organs such as liver, kidney, brain, and lungs (2-4). Free radicals are produced from metabolism of carbon tetrachloride that binds to cell membrane causing oxidative damages (5). In the body, the construction of reactive oxygen species is an unavoidable result in aerobic organisms leads to the overproduction of unwanted reactions within the body and subsequent tissue damages (6). Antioxidants found in food and the body, even in small amounts, can protect the body against various types of oxidative damage induced by oxygenfree radicals (7).

Berberis integerrima Bge. (Berberidaceae) is an important medicinal plant with yellow wood, obovate leaves, yellow flowers, and oblong red colored fruits. It grows in several regions of Iran, particularly in north and north east of the country (8). For different parts of barberry plant, various properties are listed and these properties have been demonstrated in many studies. Different types of alkaloids are obtained from the root, of which the most Berberine (9). important is Based researches on Barberry root extract and Berberine (its main alkaloid), the following considered: hypoglycemic, effects are hypolipidemic, anti-inflammatory effects, and liver protection (9-12). Majd et al showed that Berberis integerrima Bge. has antioxidant and anticancer properties (13). Furthermore, Ashraf et al showed that Berberis integerrima has antihyperglycemic antihyperlipidemic effects, preventive

therapeutic roles on the serum levels of glucose and lipids, preventive effects on liver injury, and improved renal dysfunction in diabetic rats (14-17).

Given that antioxidant and protective effects of alkaloids in various plants and on multiple tissues have been demonstrated and since barberry plant contains alkaloids, this study was performed for the first time to investigate the protective effect of Berberis integerrima Bge. root extract on carbon tetrachloride-induced toxicity in the testes of male rats.

Materials and methods

Animals

Forty mature male rats (Wistar strain) weighing 220-250 gr were tested in this research study. The animals were housed in polypropylene cages with free access to standard laboratory rat food and faucet water and were maintained in standard conditions of temperature (22±2°C), light cycle (12 hr light/dark), and relative humidity (40-60%). In this study, all animal procedures were carried out using protocols approved by the local ethical committee of Urmia University of Medical Sciences.

Plant collection and extraction

Wild samples of Berberis integerrima Bge. roots were collected from the countryside of Bavanat city (Fars province, Iran). A specimen of plants was submitted at the herbarium of Faculty of Science, Urmia University (Iran), identified and it was by the Botany Department and impounded in the herbarium (No. 9059). Roots (after washing with cold water) were dried in the shade then were powdered by using mechanical grinding. 3 liters of 70% methanol were added to 1000 gr of powder of root to isolate polar and nonpolar compounds. After 72 hr, barberry extract was filtered through Whatman filter paper No.45 and Solvent (methanol 70%) was removed by rotary vacuum evaporator. removing the solvent from the extract, in order to eliminate residual water, the resulting mixture was in the oven for 5 hr at 100°C. The dried extract was stored at 4°C for further in vivo investigations. Specific concentrations of extracts were prepared using normal saline (18).

Acute toxicity study

Acute toxicity study of MEBIR was determined as per the OECD guideline No. 423 (Acute Toxic Class Method). It was observed that MEBIR was found safe up to dose of 2,500 mg/kg of body weight. The rats were observed continuously for 24 hr for behavioral, neurological and then at 24 and 72 hr for any lethality. Test extract was not lethal to the rats even at 2500 mg/kg dose. Hence, 1/10th (250 mg/kg bw) and 1/5th (500 mg/kg bw) of this dose were selected for further study (19).

Experimental design

Forty mature male Wistar rats were randomly divided into 5 groups (n=8) and were treated for four weeks as follows: Group I (Normal control) was treated with normal saline (10 ml/ kg bw). Group II (Sham) was injected intraperitoneally with CCI4 diluted in olive oil (50% v/v) at a dose of 1ml/kg bw twice a week for 4 weeks and received normal saline (10 ml/ kg bw) (20). Group III received CCl4 similar to group two and was treated with MEBIR (dissolved in normal saline) at doses of 250 mg/kg bw. Group IV received CCl4 similar to group two and was treated with MEBIR (dissolved in normal saline) at doses of 500 mg/kg bw. Group V (Standard group) received CCL4 similar to group two and standard drug silymarin at a dose of (50 mg /kg bw, dissolved in normal saline, intragastric (21). Animals received barberry extracts and silymarin daily by gavage for 4 weeks.

Blood collection

At the end of the study, animals were anesthetized with diethyl ether. Blood was collected from their heart by heparinized syringes and kept at 37°C for 30 min. The serum was separated by centrifugation (3000 rpm at 40°C for 15 min) and stored at -30°C to measure testosterone levels.

Assessment of absolute testis weight and histological analysis

The testes of rats were dissected out, cleared of fats and blotted free of blood and weighed with the help of a Sartorius digital balance. The left testes was fixed in 10% buffered neutral formalin and used for histological investigations. After fixation of the testes, the fixed tissues were embedded in paraffin, sectioned (6-8 µm) with a rotary microtome (GmbH, Germany) and stained with Hematoxylin and Eosin. Ultimately, tissue sections were evaluated under the light microscope (Dialux 20 EB) magnification. Furthermore, the right testes were washed in cold normal saline for analysis of tissue homogenates.

Assessment of testosterone

The testosterone level of serum was measured by using radioimmunoassay method and a special kit for rats (WHO/Sigma Asso-RFGC-78/549) (22).

Assessment of catalase (CAT) activity and malondialdehyde (MDA) levels

Homogenate of the testes were prepared (10%). It was centrifuged at 4°C for 10 minutes at 7000 rpm. Supernatant solutions were used to measure CAT activity and malondialdehyde (MDA) levels. The activity of Catalase (CAT) was estimated by the method of Gott (23). MDA as the final product of lipid peroxidation was measured in the testis tissue extracts by the method of Ester Bauer and Cheese man. MDA and thiobarbituric acid react at 90-100°C and produces a pink-colored compound. The absorbance of the tissue samples were read at 532 nm (24).

Determination of histological parameters

The diameter of the seminiferous tubule was measured using scaled lens; small and large mean diameter of each tubule was calculated using the formula (25). epithelial thickness in a micrometer at 400X magnification (from spermatogonia in basal membrane to the spermatids) was measured using the scaled lens. Also, the thickness of the interstitial tissue was measured using the scaled lens in a micrometer (26). To estimate index tubule differentiation (TDI) and spermiogenesis index (SPI), 100 crossseminiferous sections of tubules randomly analyzed in each rat. TDI is the percentage of seminiferous tubules containing at least four differentiated germ cells (27).

For TDI calculation, seminiferous tubules containing more than three layers of differentiated germinal cells from spermatogonia type A were considered as positive TDI. SPI is the percentage of seminiferous tubules with normal spermiation (28). For SPI calculation, the seminiferous tubules containing sperm and tubules without sperm in cross-sections of seminiferous tubules were identified and counted.

Statistical analysis

Data from experiments are reported as mean±SE. Data were analyzed by one-way ANOVA; a significant difference between groups was determined by the Tukey test.

P<0.05 was considered as the criterion for significant differences.

Results

Absolute testis weight

As shown in figure 1, normal control animals were found to be stable in their testis weight. In sham rats, the testis weight was significantly decreased (p<0.001) in comparison to the normal rats. Rats treated with 250 and 500 mg/kg bw of MEBIR and silymarin (50 mg/kg bw) showed a significant increase in testes weight (71.73%, 115.21%, and 76.08% respectively) as compared with untreated sham rats.

Serum testosterone level

The effect of MEBIR and silymarin on the serum testosterone level of normal and sham rats is shown in figure 2. In sham rats, the serum testosterone level was significantly decreased (p<0.001) in comparison to their normal levels. Administrating 250 and 500 mg/kg bw of MEBIR or silymarin (50 mg/kg bw) lead to significant increase in serum testosterone level in sham rats (189.92%, 233.33%, and 200.77% respectively) as compared with untreated sham rats.

Histological parameters

Table I and figure 3 show the effects of and silymarin on histological parameters of control and experimental groups. The seminiferous tubule diameter, epithelial thickness, tubule differentiation index (TDI), and spermiogenesis index (SPI) in sham in comparison to the normal control were decreased (p<0.001), but interstitial tissue thickness in sham in comparison to the normal control was increased (p<0.001). However, all these parameters, interstitial tissue thickness, were increased significantly in sham rats treated with 250 and 500 mg/kg bw of MEBIR or silymarin (50 mg/kg bw) (19.79%, 21.02% and 17.62% for seminiferous tubule diameter, respectively), (61.98%, 99.17% and 78.51% for epithelial thickness, respectively), (229.34%, 269.98% and 191.31% for TDI, respectively), (and 294.38%, 346.25% and 245.70% for SPI, respectively) compared to the untreated sham rats, while the interstitial tissue thickness was decreased significantly (62.61%, 73.22% and 46.3%. respectively) compared untreated sham rats. Furthermore, the cell connections of seminiferous tubule were disturbed in sham rats while this change was improved in sham rats treated with 250-and 500-mg/kg bw of MEBIR or silymarin (50 mg/kg bw) (Figure 3).

CAT activity and malondialdehyde (MDA) levels

Table II shows the mean values of CAT activity and MDA level of both control and experimental groups after 4 weeks. In sham rats, the CAT activity significantly decreased (p<0.001), but MDA significantly increased

(p<0.001) compared to the normal levels. the CAT activity significantly However, increased in sham rats treated with 250 and 500 mg/kg bw of MEBIR or silymarin (50 mg/kg bw) (106.83%, 134.61% and 98.29% for CAT respectively), compared to the sham while untreated rats. significantly decreased in sham rats treated with 250 and 500 mg/kg bw of MEBIR or silymarin (50 mg/kg bw) (35.46%, 42.38% and 26.40%, respectively) compared untreated sham rats.

Table I. Effect of MEBIR and silymarine on histological parameters in sham rats

Group	Treatment	Dose (mg/kg)	TDI (%)	SPI (%)	Epithelial thickness (µm)	Interstitial tissue thickness (µm)	Seminiferous tubules diameter (µm)
1	N+C	10 ml/kg	90.68 ± 1.86	88.92 ± 2.64	46.60 ± 1.50	6.64 ± 0.58	259.40 ± 3.21
2	SH	10 ml/kg	$23.72 \pm 2.75^{\#}$	$18.16 \pm 1.86^{\#}$	$24.20 \pm 2.08^{\#}$	24.50 ± 0.84 #	$211.20 \pm 3.92^{\#}$
3	SH+MEBIR	250	78.12 ± 2.78^c	71.62 ± 3.54^{c}	39.20 ± 2.22^a	$9.16 \pm 0.58^{\circ}$	253.00 ± 5.66^c
4	SH+MEBIR	500	87.76 ± 2.20^{c}	81.04 ± 4.11^{c}	48.20 ± 2.26^{c}	$6.78 \pm 0.49^{\circ}$	255.60 ± 6.75^c
5	SH+Sily	50	$69.10 \pm 5.26^{\circ}$	$62.78 \pm 4.60^{\circ}$	43.20 ± 4.25^{b}	13.06 ± 1.61^{c}	248.40 ± 7.07^{c}

Values are presented as mean±SE. One-way ANOVA followed by Tukey test. (n=8)

#: p<0.001 sham rats were compared with normal control rats.

a: p<0.05, b: p<0.01, and c: p<0.001 sham rats treated with MEBIR or silymarin were compared with untreated sham rats.

SPI: Spermiogenesis index N+C: Normal control MEBIR: Methanolic extract of Berberis integerrima root

SH: Sham Sily: Silymarin TDI: Tubule differentiation index

Table II. Effect of MEBIR and silymarin on MDA and CAT in sham rats

Group	Treatment	Dose(mg/kg)	MDA (nmol/mg tissue)	CAT (u/min)
1	N+C	10 ml/kg	30.67 ± 2.36	5.77 ± 0.53
2	SH	10 ml/kg	$58.93 \pm 2.88^{\#}$	$2.34 \pm 0.21^{\#}$
3	SH+MEBIR	250	38.03 ± 3.57^{b}	4.84 ± 0.55^a
4	SH+MEBIR	500	33.95 ± 2.11^{c}	5.49 ± 0.39^b
5	SH+Sily	50	43.37 ± 3.34^a	4.64 ± 0.22^a

Values are presented as mean \pm SE. MEBIR. Methanolic extract of Berberis integerrima root. (n=8) One-way ANOVA followed by Tukey test; #p<0.001 sham rats were compared with normal control rats. ap<0.05, bp<0.01 and cp<0.001 sham rats treated with MEBIR or silymarin were compared with sham rats. CAT: catalase MDA: malondialdehyde N+C: normal control SH: share the shame of the s

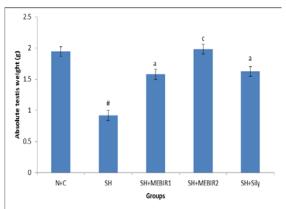


Figure 1. MEBIR 1 and 2: Methanolic extract of Berberis integer Rima root (250 and 500 respectively), N+C: Normal control, SH: Sham, SH+Sily: Sham rats treated with silymarin. Values are presented as mean±SEM; n=8 in each group. One-way ANOVA followed by Tukey test. #: p<0.001 sham rats were compared with normal control rats. a: p <0.05 and cp<0.001 sham rats treated with MEBIR 1 and 2 or silymarin were compared with untreated sham rats.

Sily: silymarin

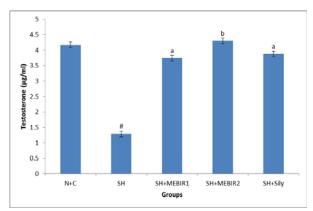


Figure 2. MEBIR 1 and 2: Methanolic extract of Berberis integerrima root (250 and 500 respectively), N+C: Normal control, SH: Sham, SH+Sily: Sham rats treated with silymarin. Values are presented as mean±S.E.M. n=8 in each group. One-way ANOVA followed by Tukey test. #: p<0.001 sham rats were compared with normal control rats. a: p<0.05 and b: p<0.01, sham rats treated with MEBIR 1 and 2 or silymarin were compared with untreated sham rats.

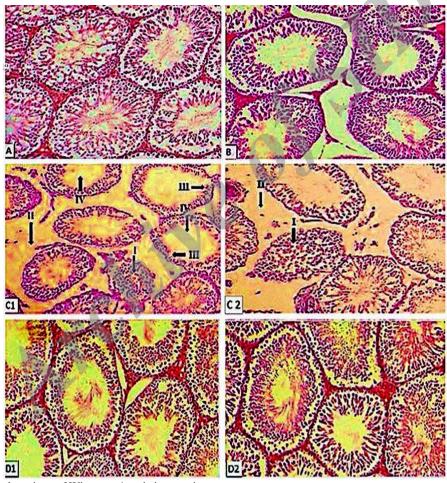


Figure 3. Histological sections of Wistar rats' testis in experiment groups.

Normal control: The seminiferous tubules at all parameters are in normal situation and all spermatogenic cell lines can be seen, (C1 and C2): Sham, (D1 and D2): Sham rats treated with 250 and 500 mg/kg bw of MEBIR respectively, (B): Sham rats treated with 50 mg/kg bw of silymarin. The seminiferous tubules are in normal situation similar to group A but still in some areas the increase at the interstitial tissue thickness can be seen. (I): Cellular connections of seminiferous tubule are disturbed and reduction in the seminiferous tubules diameter is visible. (II): Interstitial tissue. (III): TDI. (IV): SPI (magnification: ×400).

Discussion

In the present study, the activity of CAT enzyme and low level of MDA were inhibited

by the effects of carbon tetrachloride on the testis, which is likely a direct toxic effect on the tissue and probably influence on gonadal reply to FSH and LH, that ultimately leading to reduced testosterone production (29). Furthermore, treated rats with MEBIR or with silymarin, showed a significant decrease in the level of MDA, and significant increase in the activity of CAT.

In the present study, CCl4-induced toxicity in rats was similar to the study reported by Khan and Ahmed (30). One of the common criteria for the diagnosis of tissue oxidative damage is the peroxidation of lipids in the form of aldehyde products such as MDA, which is one of the main reasons in the toxicity induced by carbon tetrachloride (31). Similar to the present study, Danladi et al showed that CCl4-induced toxicity increase in MDA level in the testes of the rats, and Curtis et al showed that, there was a reduction in the activity of CAT in sham rats (29, 32). In consonance with the present study, significant low MDA level was seen in groups treated with barberry extract, confirmed that the extracts are rich in antioxidants (9), and the use of barberry extracts prevented the decrease in CAT enzyme, which may be due to the elimination of free radicals by barberry extract. This leads to the preservation and conservation of this enzyme. This result is in other agreement with researches antioxidant and elimination properties of barberry plant (13, 18).

It has been illustrated that, the reduction of CAT activity and increase in the MDA level, in toxicity induced by CCL4 in the testes of the rats, were improved by post administration of Launaea procumbens, and it is suggested that this protective effect is possibly due to antioxidant effects of its bioactive compounds (33). Moreover, present study showed that, the level of testosterone, absolute testis weight, SPI, TDI, seminiferous tubule diameter, and epithelial thickness decreased, and interstitial tissue thickness increased in sham rats.

We found that, treated rats with MEBIR or silymarin, showed a significant decrease in the interstitial tissue thickness, and a significant increase in the absolute testis weight, SPI, TDI, seminiferous tubules diameter and the level of testosterone, while the effect of the atrophy was not observed. Moreover, in the treated rats with MEBIR or silymarin, a close connection between the seminiferous tubules and different lines of sex cells in the seminiferous tubules, and the natural integrity in the testis were clearly

observed. This effect is probably due to the antioxidant properties of barberry.

It has been shown that, cigarette smoke leads to reduction in seminiferous tubules diameter and seminiferous epithelial height. This reduction was improved supplementation of honey, and it is suggested that this protective effect is due to antioxidant capacities of honey (34). On the other hand, Shalizar Jalali et al. showed that reduction in SPI, TDI, serum level of testosterone and absolute testis weight, in cyclophosphamideinduced toxicity, improved towards normality, by administration of Crataegus monogyna due to its antioxidant properties (35).

Moreover, Pashaeian et al. showed that after immobilization stress in rat, reducing interstitial tissue thickness in ginseng treatment group was observed (36). From the result of this study, and other findings about antioxidant properties of silymarin, it can be firmly claimed that the antioxidant activity of silymarin leads to improve the change observed in sham rats (37). Detrimental effects of carbon tetrachloride on the testes well documented (30).Carbon tetrachloride is deposited in various tissues such as liver, brain, and testes because of the solubility in lipid and crossing of the cell membranes (38). Detoxification enzymes of the cytochrome P450 cause the production of secondary active and toxic metabolites, which can damage the testis and other body tissues (39).

Decomposition of carbon tetrachloride leads to the production of trichloromethyl and trichloromethyl peroxyl free radicals (40). It appears that the effect of free radicals causes the peroxidation of membrane lipid and unsaturated fatty acids as well as the reduction of antioxidant enzymes (38). It has been suggested that, one of the major causes of testicular damage is oxidative stress caused by production of free radicals, which causes lipid peroxidation, change in enzymatic activity and finally induces necrosis (41, 42).

CAT is an important enzyme that has been widely distributed in animal tissue, decomposes hydrogen peroxide and protects tissues against highly active hydroxyl radicals (43). Thus, the reduction of CAT activity may lead to some destructive effects of superoxide anion and hydrogen peroxide radicals. A significant decrease in testosterone leads to

pathological changes in the testis tissue (44). The reduction of the seminiferous tubule diameter in sham rats indicates that the atrophy of these tubules leads to morphological and spermatogenesis disturbance in the testis. In fact, there is a positive relationship between the seminiferous tubules diameter and spermatogenesis activity (45).

The root and stem bark of barberry contain alkaloids various such as: Berberine. oxvacanthine. bermamine. palmatine. jatrorrhizine, columbamine, and berberubine which have antioxidant, anti-inflammatory, and therapeutic properties. Plant alkaloids are the important antioxidant compounds. Considering the fact that barberry roots are rich in alkaloids and given that antioxidant properties of barberry root have been proven in many studies, we can say that the antioxidant activity of barberry root is probably related to its alkaloid compounds (9, 17, 46, 47)9).

Conclusion

Based on our results, the barberry root could protect the testes of rats against CCl4-induced toxicity. Furthermore, the effect of barberry root extract (especially at doses of 500 mg/kg bw) was more than that of the standard drug, silymarin (50 mg/kg bw). Thus it could be suggested that, the extract of barberry root probably by its antioxidant properties protected the rat's testes against the toxic effects of carbon tetrachloride.

Acknowledgments

This study was financially supported by Vice Chancellor of Research, Faculty of Sciences, Urmia University. The authors would like to express thanks to Mrs. Farnad, Biochemistry Laboratory, Biology Department, Urmia University, Iran, for providing laboratory facilities.

Conflict of interest

There is no conflict of interest in this paper.

References

 Khaki A, Fathiazad F, Nouri M, Khaki AA, Ghanbari Z, Ghanbari M, et al. Anti-oxidative Effects of Citro-

- flavonoids on Spermatogenesis in Rat. *Afr J Pharm Pharmaco* 2011; 5:721–725.
- Abraham P, Wilfred G, Cathrine SP. Oxidative damage to lipids and proteins of the lungs, testis and kidney of rats during CCl4 intoxication. *Clinical Acta* 1999; 289:177–179.
- Mahmoud SM, Moneim AAE. The Protective Effect of Pomegranate (Punica granatum) Juice against Carbon Tetrachloride-Induced Oxidative Stress in Brain Tissue of Adult Male Albino Rats. *Life Sci* 2013; 10:151-158.
- Ganie SA, Haq E, Hamid A, Qurishi Y, Mahmood Z, Zargar BA, et al. Carbon tetrachloride induced kidney and lung tissue damages and antioxidant activities of the aqueous rhizome extract of Podophyllum hexandrum. BMC Complement Altern Med 2011; 11:1-10.
- Parola M, Leonarduzzi G, Biasi F, Albano E, Biocca ME, Poli G, et al. Vitamin E dietary supplementation protects against carbon tetrachloride-induced chronic liver damage and cirrhosis. *Hepatology* 1992; 16: 1014-1021.
- Qian ZJ, Jung WK, Kim SK. Free radical scavenging activity of a novel antioxidative peptide purified from hydrolysate of bullfrog skin, Ran catesbeiana shaw. Bioresour Technol 2008; 99:1690-1698.
- Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacogn Rev* 2010; 4: 118-126.
- 8. Arayne MS, Sultana N, Bahadur SS. The berberis story: Berberis vulgaris in therapeutics. *Pak J Sci* 2007; 20: 83-92.
- Ivanovska N, Phlipov S. Study on the antiinflammatory action of Berberis vulgaris root extract, alkaloid fractions and pure alkaloid. *Int J Ethnopharmacol* 1999; 64: 161-166.
- Yin I, Hu R, Chen M, Tang J, Li F, Yang Y, et al. Effects of Berberine on glucose metabolism in vitro. Metabolism 2002: 51: 1439-1443.
- Doggrell SA. Berberine-a novel approach to cholesterol lowering. Expert Opinion Investin Drugs 2005; 14: 683-685.
- 12. Roy A, Kumar Sahu R, Gupta R, Pandey P. Hepato protective activity of Berberis coriaceae on liver damage induced by ccl4 in rats. *Pharmacology* online 2011; 3: 838-842.
- 13. Majd A, Mehrabian S, Mostafai H, Rahmani H. Antioxidant and anticancer effect of aqueous extract of berberis integerrima. *J Biol Sci* 2008; 1:31-38.
- Ashraf H, Heidari R, Nejati V. Antihyperglycemic and Antihyperlipidemic Effects of Fruit Aqueous Extract of Berberis integerrima Bge. in Streptozotocin-induced Diabetic Rats. *Iran J Pharm Res* 2014; 13: 1313-1318.
- 15. Ashraf H, Heidari R, Nejati V, Ilkhanipoo M. Preventive Effect of Berberis Integerrima on the Serum Levels of Glucose and Lipids in Streptozotocin (STZ)-Induced Diabetes in Rats. J Fasa Univ Med Sci 2012; 2: 148-155.
- 16. Ashraf H, Zare S. Preventive Effects of Aqueous Extract of Berberis integerrima Bge. Root on Liver Injury Induced by Diabetes Mellitus (Type 1) in Rats. *Iran J Pharm Res* 2015; 14: 335-343.
- 17. Ashraf H, Heidari R, Nejati V, Ilkhanipoor M. Aqueous extract of Berberis integerrima root improves renal dysfunction in streptozotocin induced diabetic rats. *Avicenna J Phytomed* 2013; 3: 82-90.

- 18. Eidi A. Zarin Ghalam J. Rezazade Sh. Adeli R. Hepatoprotective effect of Berberis vulgaris L. extract on CCI4-induced toxicity in rats. Kowsar Med J 2011; 16: 169-173.
- 19. OECD 2002. Acute oral toxicity. Acute oral toxic class method guideline 423 adopted 23.03.1996. In: Eleventh Addendum to the, OECD, guidelines for the testing of chemicals organization for economical co-operation and development, Paris, June, 2000.
- 20. Abdel-Wahhab KGE, Khadrawy YA, Mannaa FAE. Aged garlic extract enhances paraoxonase 1 activity and suppress oxidative stress in CCI4 intoxicated rats. Comunicata Scientiae 2012; 3: 55-63.
- 21. Osman M, Ahmed M, Mahfouz S, Elaby S. Biochemical Studies on The Hepatoprotective Effects of Pomegranate and Guava Ethanol Extracts. N Y Sci J 2011; 4: 2741.
- 22. Dehkhargani SF, Malekinejad H, Shahrooz R, Sarkhanloo RA. Detrimental Effect of Atrazine on Testicular Tissue and Sperm quality: Implication for Oxidative stress and Hormonal Alterations. Iran J Toxicol 2011; 5: 426-435.
- 23. Gott L. A simple method for determination of serum catalase activity and revision of reference range. Clin Chim Acta 1991; 196: 143-151.
- 24. Esterbauer H, Cheesman KH. Determination of aldehydic lipid peroxidation products: Malonaldehyde and 4-hydroxynonenal. Methods Enzymol 1990; 186: 407-421.
- 25. Vendramini V, Sasso-Cerri E, Miraglia SM. Amifostine reduces the seminiferous epithelium damage in doxorubicin-treated prepubertal rats without improving the fertility status. Reprod Biol Endocrinol 2010; 8: 3.
- 26. Abdullah Nejad A, Goals A, Dabiri SH, Javadi AB. Effects of garlic juice on diabetes-induced testicular damage in rats. Iran J Endocrinol Metab 2009; 11: 443-453.
- 27. Malekinegad H, Mirzakhani N, Razi M, Cheraghi H, Alizadeh A, Dardmeh F. Protective effects of Melatonin and Glycyrrhiza glabra Extract on Ochratoxin A-Induced Detrimental impact on Testes in Mature Male Rats. Hum Exp Toxicol 2010; 29: 110-123.
- 28. Rezvanfar MA, Sadrkhanlou AR, Ahmadi A, Shojaei-Sadee H, Rezvanfar M, Mohammadirad A, et al. Protection of cyclophosphamide-induced toxicity in reproductive tract histology, sperm characteristic, and DNA damage by an herbal source; evidence for role of free radical toxic stress. Hum Exp Toxicol 2008; 27: 901-910.
- 29. Danladi J, Ahmed SA, Akpulu SP, Owolagba GK, Iduh MU, Mairiga AA. Protective effect of cool extraction of Black Seed (Nigella Sativa) oil against CCI4-Induced oxidative damages in wistar rats testis. IOSR-J Pharm Bio Sci 2013; 5: 68-74.
- 30. Khan MR, Ahmed D. Protective effects of (Digera muricata L.) Mart. on testis against oxidative stress of carbon tetrachloride in rat. Food Chem Toxicol 2009; 47:1393-1399.
- 31. Wang T, Sun NL, Zhang WD, Li HL, Lu GC, Yuan BJ. et al. Protective effects of dehydrocavidine on carbon tetrachloride-induced acute hepatotoxicity in rats. J Ethnopharmacol 2008; 117: 300-308.
- 32. Curtis SJ, Mortiz M, Sondgrass PJ. Serum enzyme derived from liver cell fractions. The response of

- carbon tetrachloride intoxication in rats. Gastroentrology 1972; 62: 84-92.
- 33. Khan RA. Protective effects of Launaea procumbens on rat testis damage by CCI4. Lipids Health Dis 2012; 11:103.
- 34. Mohamed M, Sulaiman SA, Jaafar H, Salam Sirajudeen KN. Antioxidant Protective Effect of Honey in Cigarette Smoke-Induced Testicular Damage in Rats. Int J Mol Sci 2011; 12: 5508-5521.
- 35. Shalizar Jalali A, Hassanzadeh SH, Malekinejad H. Chemoprotective effect of Crataegus monogyna aqueous extract against cyclophosphamide-induced reproductive toxicity. Vet Res Forum 2011; 2: 266-273.
- 36. Pashaeian N, Safavi SE, Gharamaleki NM. Study of the effect of ginseng on testicular function after immobilization stress in rat. Int J Biosci 2015; 6: 184-
- 37. Pradeep K, Mohan CVR, Gobianand K, Karthikeyan S. Silvmarin modulates the oxidant-antioxidant during diethylnitrosamine imbalance oxidative stress in rats. Eur J Pharmacol 2007; 560: 110-116.
- 38. Szymonik-Lesiuk S, CzechowskA G, Stryjecka-Zimmer M, Słomka M, Madro A, Celiński k, et al. Catalase, superoxide dismutase, and glutathione peroxidase activities in various rat tissues after carbon tetrachloride intoxication. J Hepatobiliary Pancreat Surg 2003; 10: 309-331.
- 39 Jeong TC. Pretreatment of male BALB/c mice Biononepotentiates thioacetamide -induced hepatotoxicity. Toxicol Lett 1999; 105: 39-45.
- 40. Janbaz KH, Saeed S, Gilani AH. Protective effect of Paracetamol and CCI4- induced hepatotoxicity inrodents. Fitoterapia 2002; 73: 557-564.
- 41. Yousef MI, Salama AF. Propolis protection from reproductive toxicity caused by aluminium chloride in male rats. Food Chem Toxicol 2009; 47: 1168-1175.
- 42. Bruckner JV, Ramanathan R, Lee KM, Muralidhara S. Mechanisms of circadian rhythmicity of carbon tetrachloride hepatotoxicity. J Pharmacol Exp Ther 2002; 300: 273-281.
- 43. Chance B, Greenstein DS, Roughton RJW. The mechanism of catalase action. Steady-state analysis. Arch Biochem Biophys 1952; 37: 301-321.
- 44. Stefanovic A, Stevuljevic JK, Spasic S, Stanojevic NB, Bujisic N. The influence of obesity on the oxidative stress status and the concentration of leptin in type 2 diabetes mellitus patients. Diabetes Res Clin Pract 2008; 79: 156-163.
- 45. Predes FS, Monterio JC, Paula T, Dmatta P. Evalution of rat testes treated with arctium lappa 1: Morphometric study. Braz J Morphol Sci 2007; 24:
- 46. Ashraf H, Heidari R, Nejati V, Ilkhanipoor M. Effects of aqueous extract of Berberis integrrima root on some physiological parameters in streptozotocininduced diabetic rats. Iran J Pharm Res 2013; 12: 425-434.
- 47. Taheri S, Zarei A, Changizi Ashtiyani S, Rezaei A, Zaheiri S. Evaluation of the effects of hydroalcoholic extract of Berberis vulgaris root on the activity of liver enzymes in male hypercholesterolemic Avicenna J Phytomed 2012; 2: 153-161.