

## Single Nucleotide Polymorphisms as Risk Variants in Crohn's Disease

Fereshteh Izadi<sup>1,\*</sup>

<sup>1</sup> Research Associate, University College London, Division of Biosciences, London, UK

### ABSTRACT

#### Background:

Crohn's disease is the most challenging, common type of autoimmune disorders due to which, intestine is inflamed but its causes have not been defined. Although Crohn's disease has been often thought of as an autoimmune disorder, it can be triggered by whatever that lead to the inflammation in the whole bowel. Henceforth, exploring trade-off among this disease and genomic variants supposedly will enhance the identification of important genes and in turn to the possible therapeutic protocols. The aim of the present study was to identify any association between Crohn's disease and single nucleotide polymorphisms.

#### Materials and Methods:

We retrieved single nucleotide polymorphism of genome-wide association studies (GWAS) on Crohn's disease among which we extracted neighboring genes and expression variants in addition to pathways in which the affected genes are enriched.

#### Results:

Using GWAS data can help to explain the disease incidence. We noticed that genes harboring single nucleotide polymorphisms in Crohn's disease were mainly enriched in interleukin pathways in addition to fatty acid and choline metabolism.

#### Conclusion:

The affected genes obtained by data analysis like fatty acid desaturase (FADS) and solute carrier family 22 (SLC22) could be proposed for future studies in order to clarifying their interactions by confident experiments and cross-sectional studies.

**Keywords:** Crohn's disease, eQTL, Single nucleotide polymorphism

*please cite this paper as:*

Izadi F. Single Nucleotide Polymorphisms as Risk Variants in Crohn's Disease. *Govaresh* 2018;23:183-192.

### INTRODUCTION

The immune system should defend the physiological system against external stimuli homeostatically. Inflammatory bowel disease (IBD) consists of two major forms, Crohn's disease (CD)

and ulcerative colitis (UC), which are immune-mediated diseases that cause the damage to the gastrointestinal tract with an equal prevalence in both sexes (1). UC is a worldwide chronic inflammatory disorder of the colon, which triggers typical ulcers in the mucosa of the rectum and colon. On the other hand, CD is a chronic inflammatory disease of the digestive tract affecting 26-200 per 100,000 in European populations (2). The causes of CD are remained somehow unknown, although it is likely that a disrupted immunological response to gut microbiota in genetically susceptible individuals implicates in CD (3,4). However, there is currently no exact cure for CD and it is controlled through a combination of immunosuppressive agents, nutritional adequacy, or surgery.

It has been well documented that Autophagy-

#### \*Corresponding author:

Fereshteh Izadi, PhD

Department of Genetics, Evolution and Environment,  
University College London, Darwin Building, Gower  
Street, London WC1E 6BT

Telefax: + 44 7846280861

E-mail: izadi1991@yahoo.com

Received: 16 May 2018

Edited: 08 Aug. 2018

Accepted: 09 Aug. 2018

related protein 16-1 (ATG16L1) and Immunity-related GTPase family M protein (IRGM) play role for autophagy in patients with CD (5,6). An exaggerated immune response in patients with CD by the alteration in gut microbial community has been shown to be associated with Toll-like receptor 4 (TLR4), caspase-associated recruitment domain (CARD), interleukin 23R (IL-23R), Signal transducer and activator of transcription 3 (STAT3) and human leukocyte antigen (HLA), tumor necrotic superfamily member 15 (TNFSF15), interferon regulatory factor (IRF) and protein tyrosine phosphatase (PTPN22) genes involved in both innate and adaptive immune reactions, respectively. Most of the many disease-related single nucleotide polymorphisms (SNPs) discovered through the genome-wide association studies (GWAS) are supposed to be influential not only in regulating the expression of neighboring genes but also via another genomic contexts such as outside of protein-coding regions (7,8). GWAS characteristically aim in finding a statistically significant difference in genotypic frequency between a large numbers of individuals with a particular genotype for instance individuals with diseases compared with control population by means of SNP loci distributed in genomes. The SNPs that indicate significant links with disease condition in contrast to normal condition characterize the regions of genomes where likely to be harbored with biomarkers underlying the assessed diseases. Thereof, exploring the impacts of allelic variation on transcriptome using expression quantitative trait locus (eQTL) helps to understand how SNPs lie outside of protein coding regions account for the discovered pathogenic effects. Despite the comprehensive variant mapping by GWAS (9,10) in identifying more than 100 gene loci related to IBD, the pathogenesis of these conditions remains largely unknown. Several genetic and environmental factors have already been reported in correlation with CD, while a number of genome-wide linkage studies suggested different genomic loci as possible candidate genes for susceptibility to IBD (11-15).

Although the association between CD and SNPs has been presented in a number of studies (16-23), data mining methods can be still considered as a powerful approach toward the understanding of the etiology of CD.

In this study, the impact of the SNPs on patients

with CD was investigated. We mainly aimed to elucidate the underlying relationships between CD and SNPs further eQTLs using in silico analytical approaches. The neighbor genes of genomics risk factors and underlying pathways that according to our previous knowledge thought to play important roles in the pathogenesis of CD were subsequently prioritized. Here we tried to provide a systematic view of genome-wide associations implicated in CD.

## MATERIALS AND METHODS

Firstly, CD risk genomic variants were retrieved from the National Human Genome Research Institute (NHGRI), European Bioinformatics Institute (EBI) catalog of published GWAS (<https://www.ebi.ac.uk/gwas/home>). The potential regulatory changes triggering by these risk factors including transcription factor binding sites and histone modifications were then investigated in JASPAR (<http://jaspar.genereg.net/>), and TRANSFAC (<http://genexplain.com/transfac/>) databases and Roadmap Epigenomics HM ChIP-seq ([www.roadmapepigenomics.org](http://www.roadmapepigenomics.org)), respectively. Next, the distribution of these risk variants was explored by dbGaP database (<https://www.ncbi.nlm.nih.gov/gap>). The genes neighbor of these risk factors were further functionally enriched in GO molecular labels and biological pathways by DAVID (<https://david.ncifcrf.gov/>) and KEGG (<http://www.genome.jp/kegg/>) databases to find potential impacts of the risk alleles on the affected genes. Additionally, the CD GWAS risk variants were screened through the ExSNP database (<http://www.exsnp.org/DZeQTL>) to determine high-confidence eQTLs at linkage disequilibrium  $r^2 > 0.8$  and at 0.05 centi-Morgan distance to the risk SNPs. We finally took an intersection between the affected genes by these eQTL and curated CD related genes retrieved from DisGeNET v2.0 server (<http://www.disgenet.org/web/DisGeNET/>). These gene-disease associations have been collected from several databases including UniProt, human CTD, PsyGeNET, Orphanet, and the HPO. The shared genes affected by eQTLs were consequently fed into pathwAX web server (<http://pathwax.sbc.su.se/>) to find a network crosstalk of significant pathways. PathwAX contains KEGG pathway information in addition to networks of gene-gene links in model organisms.

**Table 1:** The distribution of genes neighboring Crohn's disease risk variants by dbGaP database. The potential impact of risk variants on the gene products as genotype and potential phenotypes as diseases has been investigated.

Term	Adjusted P value	Z-score	Combined Score	Genes
Crohn's disease	6.88E-27	-2.30498	150.5680109	PUS10,TNFSF15,IL23R,STAT3,MST1,ITLN1,PTPN22,CDKAL,NOD2,FUT2,NELL1,LACC1,ATG16L1,FGFR1OP,BSN,NKX2-3,ZNF365
Colitis, Ulcerative	1.05E-12	-2.00191	64.00450137	IL10,C1ORF106,PUS10,RTEL1,TNFRSF6B,TNFSF15,IL23R,CARD9,MST1,BTNL2,BSN,NKX2-3
Celiac disease	1.96E-11	-2.01583	57.73037652	PUS10,ATXN2,ZMIZ1,TAGAP,LPP,YDJC,PTPN2,BACH2,HLA-DQA1,ICOSLG
Diabetes mellitus, type 1	1.49E-10	-1.95469	51.45321396	NOTCH4,PTPN22,BTNL2,TYK2,RASGRP1,BACH2,INS,IL2RA,HLA-DRA,SH2B3,PTPN2,HLA-DQA1,HLA-DQB1,MICB
Inflammatory bowel diseases	1.62E-09	-2.18517	51.82159741	ATXN2L,CYLD,TEC,TNFSF15,ATG16L1,ZMIZ1,IL23R,NCF4,SAG,NOD2
Psoriasis	9.86E-09	-1.79875	39.07605826	NOS2,NOTCH4,IL23R,LST1,HLAC,PBX2,IL12B,TYK2,TNF,PSORS1C1
Leprosy	3.76E-08	-1.57738	31.91207441	LACC1,TNFSF15,RIPK2,HLA-C,HLA-DRA,HLA-DRB1,PSORS1C1,HLA-DQB1
Multiple sclerosis	5.02E-08	-1.66668	33.01439042	ZMIZ1,IL2RA,NOTCH4,STAT3,CPAMD8,TAGAP,HLA-DRA,BTNL2,HLA-DQA2,PSORS1C1,HLA-DQA1,TNFRSF1A
Vitiligo	7.38E-07	-0.95705	16.27380929	ZMIZ1,RNASET2,IL2RA,PTPN22,BTNL2,LPP

\*The combined score is computed by taking the log of the P values generated by the Fisher exact test and multiplying that by the z-score of the deviation from the expected rank.

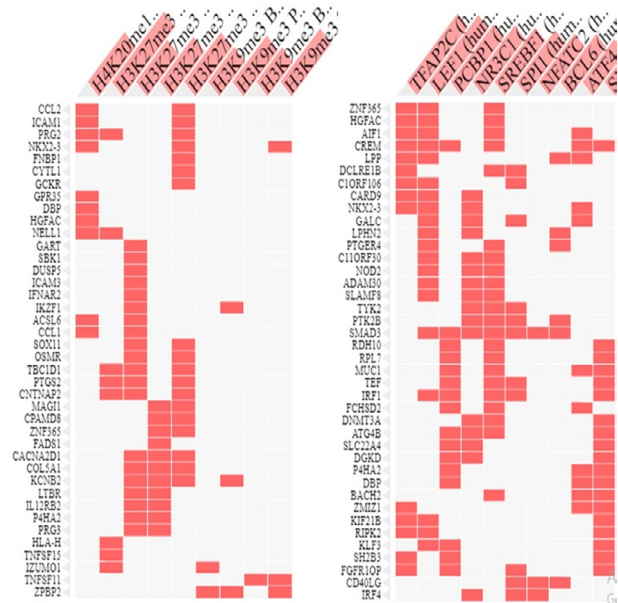
## RESULT

596 SNP-disease associations from 29 CD studies were retrieved from NHGRI at P value 5.00E-08 (supplementary table 1). We only selected SNPs with risk allele frequency > 0.05 of these, 292 mapped neighbor genes were extracted for further investigations. To determine if these genes are related to share diseases, we performed genotype-phenotype interaction analysis by dbGaP database. The genes were immediately grouped as to be related to CD, colitis, ulcerative, celiac disease, diabetes mellitus type 1, and IBD (table 1). The position matrix modelling of JASPAR and TRANSFAC databases showed TEFAP2C, LEF1, and SPI transcription factor binding sites affected by risk alleles at the upstream of 292 unique genes (figure 1a). Moreover, potential histone modifications at H4K20, H3K27, and H3K9 as transcriptional regulatory features were observed (figure 1b). The gene ontology molecular function, categorized the 292 mapped neighbor genes mainly under oncostatine-M, interleukin, chemokine, and CD40 receptors terminologies (figure 2a). Additionally, cytokines, interleukins, interferons, and immune systems were identified as enrichments of pathway themes (figure 2b). We then intersected 292 unique genes affected by risk variants with 899 CD associated genes through the

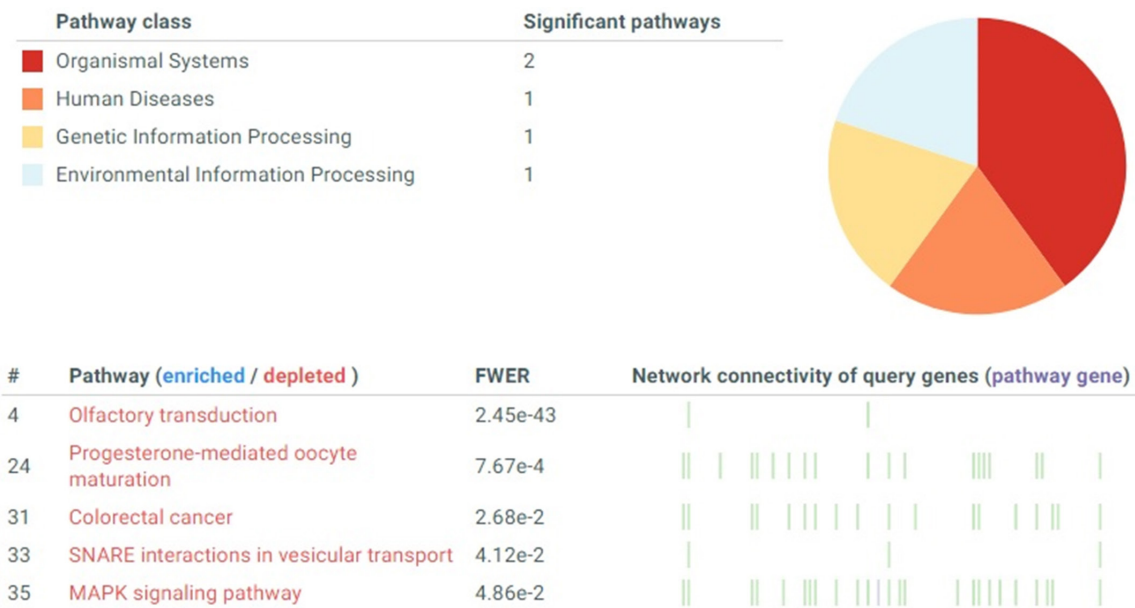
DisGeNET database with at least one evidence from pfam (Supplementary table 2). The common genes were depleted in pathways such as colorectal cancer (figure 3). DisGeNET contains a list of diseases associated genes that has been collected based on the presence of genetic overlaps between diseases. In this list, among the 4,753,986 potential diseases associations, 13,064 diseases were found to share at least one gene with other diseases. Among these associations, CD (mesh: D003424) was significantly associated with mental retardation, skin diseases, tuberculosis, leukemia (b-cell, chronic), osteoporosis (postmenopausal), sepsis, hepatitis b (chronic), and pulmonary disease (chronic obstructive) at P value < 0.05. However, we only focused on colorectal cancer as a depleted pathway (figure 3) to find which gene links have not been significant enough to be related to colorectal cancer whereby these genes are not affected by colorectal cancer. Whereby, CREB5, EIF3C, DUSP5, RXRA, SMNDC1, TAGLN2, AKAP11, ENTPD7, DCLRE1B, CUTC, IFNAR1, ALDH2, USP1, PTK2B, CNTNAP2, ACSL6, HMHA1, TEC, PRDX5, PMM1, RBMX, NHP2L1, UBE2D1, RPL7, and TERF1 were characterized in colorectal depleted pathway. This procedure assures the approximation of CD associated genes involved in molecular networks, which are totally independent from colorectal



**Fig.1:** Gene ontology enrichment and pathway analysis of genes neighboring Crohn's disease risk alleles. (a) Functional classification of biological processes, (b) biological pathways by Enrichr and KEGG databases, respectively with default setting. The bars have been arranged top to down illustrating the number of genes and significance level assigned to each GO molecular terminology and biological pathway.



**Fig.2:** Functional annotation of SNPs. (Right) the position matrix modelling (PMM) of the influence of SNPs on transcription factor binding sites from JASPAR vertebrate and TRANSFAC. (Left) the influence of SNPs on epigenetic modifications by Roadmap Epigenomics HM ChIP-seq. In, genes are in rows and transcription factors and histones are in columns.



**Fig.3:** PathWAX results for the affected genes by Crohn's disease related eQTLs. The table and pie chart summarize the pathway distribution. The table shows enriched depleted pathways at q-value 0.05 as defined cut-off threshold. Darker shades in colored boxes within the table indicate higher connectivity (links) that a query gene has.

Table 2: Crohn's disease associated eQTLs

Gene(s)	SNP	Risk allele	Chr ID	Context	Intergenic	Risk allele frequency	P value	Region	PubMed ID
FADS1	rs102275	C	11	Non coding transcript exon variant	0	0.341	2.00E-11	11q12	21102463
	rs4246215	?	11	3 prime UTR variant	0		9.00E-07	11q12	28067908
	rs4077515	T	9	Missense variant	0	0.411	1.00E-36	9q34.3	22936669
CARD9	rs10781499	A, ?	9	Synonymous variant	0		6.00E-30	9q34.3	26192919
	rs11145763	C	9	Intron variant	0	0.4	3.00E-08	9q34.3	26301688
	rs4077515	T	9	Missense variant	0		4.00E-06	9q34.3	22936669
SLC22A5	rs2188962	?	5	Intron variant	0		6.00E-36	5q31.1	28067908
SLC22A5/ SLC22A4	rs12521868	T	5	Intron variant	0	0.422	1.00E-20	5q31.1	2110246
GSDMB/ ORMDL3	rs12946510		17	Downstream gene variant	1		1.00E-16	17q12	
	rs12946510		17	Downstream gene variant	1		1.00E-16	17q12	
	rs2872507		17	Intergenic variant	1	0.458	2.00E-09	17q12	
	rs2872507		17	Intergenic variant	1	0.47	5.00E-09	17q12	
ERAP2	rs1363907		5	Intron variant	0		1.00E-14	5q15	
	rs2549794		5	Intron variant	0	0.409	1.00E-10	5q15	
	rs4869313		5	Intron variant	0	0.42	9.00E-08	5q15	
CPEB4	rs359457		5	Intergenic variant	1	0.571	3.00E-12	5q35.2	
	rs17695092		5	Intron variant	0	0.703	5.00E-09	5q35.2	
RNASET2/ FGFR1OP	rs1819333		6	Upstream gene variant	1		2.00E-20	6q27	
	rs2149085		6	Upstream gene variant	1	0.379	8.00E-12	6q27	
	rs1819333		6	Upstream gene variant	1		2.00E-20	6q27	
	rs2149085		6	Upstream gene variant	1	0.379	8.00E-12	6q27	
UBE2L3	rs2266959		22	Intron variant	0		1.00E-15	22q11.2	
IFNGR2	rs2834215		21	Intron variant	0	0.58	3.00E-07	21q22.1	
USP4	rs3197999		3	Missense variant	0		3.00E-23	3p21.3	
FADS2	rs4246215	?	11	3 prime UTR variant	0		9.00E-07	11q12	28067908
SP140	rs6716753	C, ?	2	Intron variant	0		1.00E-13	2q37.1	23128233 28067908
	rs7423615	T	2	Intron variant	0	0.187	3.00E-13	2q37.1	21102463
PTGER4	rs7725052	C	5	Regulatory region variant	1	0.43	8.00E-11	5p13.1	26301688
HLA-C	rs9264942	C	6	Intron variant	0	0.378	5.00E-28	6p21.3	23128233

cancer. We then seek the NHGR CD associations across 12825 eQTL from ExSNP, to find eQTLs. Thereof, 82 SNPs were found to be mutual in NHGR GWAS and ExSNP databases. These SNPs targeted 165 and 51 unique genes from GWAS and ExSNP as eQTLs of these 17 genes where found to be affected as CD associated common eQTLs (table 2). Pathway analysis of the eQTLs showed fatty acid and choline metabolism by implicating FADS2, FADS1,

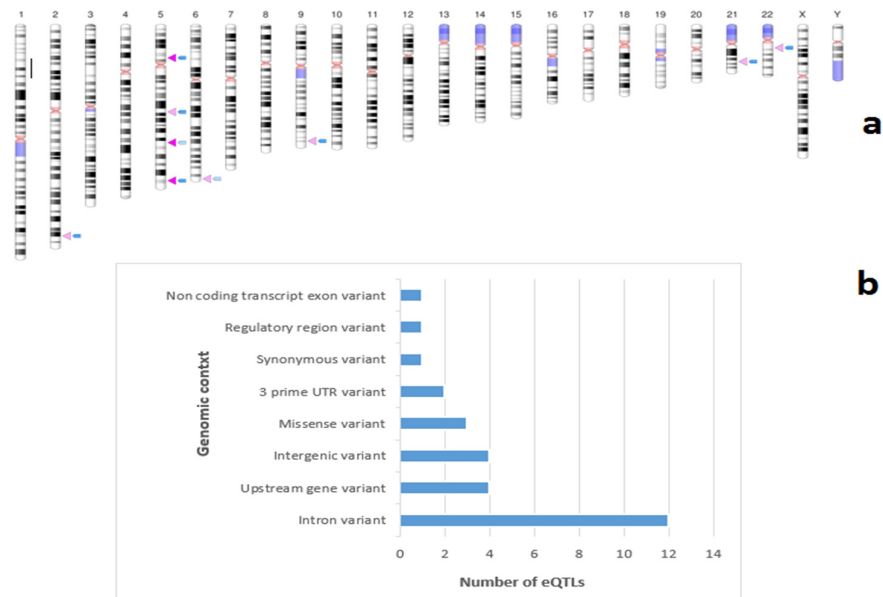
SLC22A4, and SLC22A5 at adjusted  $P$  value  $< 0.05$  (table 3). The genomic distribution of eQTLs and neighbor genes has been illustrated in figure 4.

## DISCUSSION

In this article, 29 GWAS data sources have been employed to investigate the impacts of SNPs in the pathogenesis of CD in addition to elucidating the molecular pathways underlying this disorder

**Table 3: Pathway analysis of 17 Crohn's disease associated genes neighboring the identified eQTLs**

KEGG pathways	Adjusted <i>P</i> value	Z-score	Combined Score	Genes
Biosynthesis of unsaturated fatty acids (hsa01040)	0.00698	-1.6693	14.48661	FADS2, FADS1
Fatty acid metabolism (hsa01212)	0.015368	-1.83498	13.20435	FADS2, FADS1
Choline metabolism in cancer (hsa05231)	0.044669	-1.88791	10.80532	SLC22A4, SLC22A5



**Fig.4: Distribution of eQTLs. (a) The chromosomal distribution of genes neighboring the eQTLs (b) The genomic pattern of eQTLs. The bars have been arranged to illustrate the number of eQTL at each genomic context.**

implicated by the affected neighboring genes. Interestingly, 292 neighbor genes of 596 SNP-CD associations were enriched in chemokines (GO:0008009, GO:0031727, GO:0031731) and interleukins (GO:0004920). Oncostatin-M receptor (GO:0004924) was demonstrated to be the most significant GO biological label (figure 2a). Oncostatin-M has been recently reported with both the pro-inflammatory and anti-inflammatory effects (24,25). In pathways analysis, interferons, interleukins, and immune systems were more noticeable (figure 2b). Accordingly, the levels of specific cytokines and chemokines have been found to be elevated in patients with IBD (26-28). In concordance, the significant pathways relating to interleukins imply on an indispensable role of interleukins in immune-mediated disorders when interleukins have confirmed roles in several immune and inflammatory events that is occurred in autoimmune diseases. Thus, further

studies could help to evaluate the prognostic roles of interleukins in crosstalk of CD and genomic variants. As depicted in figure 2, 596 risk factors trigger potential regulatory changes mostly in binding sites of some transcription factors including TFAP2C at the top in upstream of 292 affected genes (rows of left clustergrammer). Transcription factor AP2C (TFAP2C) has been revealed to play a central role in controlling multiple pathways of estrogen signaling hereon growing evidence proposes that estrogen modulates gut inflammation through the alteration of estrogen receptors in peripheral blood T lymphocytes (29,30).

In the next step, the potential impact of risk variants on the gene products as genotype and potential resulting phenotypes, here CD, has been investigated by dbGaP database. As shown in table 1, 596 SNP of 29 CD studies extracted from NHGRI at *P* value 5.00E-08 have been linked to CD as the

most significant and later with the other types of autoimmune-diseases including colitis ulcerative, celiac disease, diabetes mellitus type1, IBD, psoriasis, leprosy, multiple sclerosis, and vitiligo. Regarding diabetes mellitus type1, TNF- $\alpha$  has been shown in developing insulin resistance during obesity (31) and in autoimmune destruction of pancreatic  $\beta$ -cells (32) that could explain the effects of TNF- $\alpha$  antagonism on glucose metabolism. Systemic changes of microbiota products may have promoted insulin resistance (33). Regarding psoriasis, the most important susceptibility locus is known to be located on the gene encoding TNF- $\alpha$  providing new venues to the associations of autoimmune-diseases (34). HLA-DR-DQ, RIPK2, CCDC122-LACC1, and NOD2 have been detected as leprosy susceptibility, related to the NOD2 signaling pathway and are CD susceptibility loci. NOD2 is the first recognized susceptibility locus in CD, which has been reported to trigger an immune response to bacterial cell wall. However, in this study HLAs seem to be the most frequent susceptibility loci based on the dbGaP database (table 1). Accordingly, predisposing genetic factors, and HLAs contribute to the genetic pathogeny of autoimmune disorders such as IBD at 30%-50% (35,36). As the main type of chronic IBD, CD has been shown to be associated with specific HLA class I and II (37). Keeping with these, probably genomic risk variants are targeting a circuit of interferons coding genes responsible for inflammation development (38). In the light of previous studies, SNPs found in a cohort of 29 GWAS likely discover susceptibility variants for pathways contributing in intestinal inflammation whereby anti-TNF therapy could be suggested as a clinical target. Further, 82 CD-associated mutual SNPs identified in the NHGR GWAS and ExSNP databases targeting 165 and 51 unique genes, respectively were introduced as eQTLs of those 17 genes were considered as CD associated eQTLs (table 2). The pathway analysis of these 17 genes affected by eQTLs exhibited fatty acid and choline metabolism pathways with the implication of FADS2, FADS1, SLC22A4, and SLC22A5 at adjusted  $P$  value  $< 0.05$  (table 3). Serum levels of choline and its derivatives are lower in patients with IBD compared with healthy individuals as a result of pro-inflammatory cytokine levels (39). Regarding the fatty acid metabolism, accumulating evidence

has highlighted the co-existence of non-alcoholic fatty liver disease and IBD, which supposedly is contributed by agents such as a severe alteration in intestinal permeability, gut dysbiosis, and chronic inflammatory response (40). As illustrated in figure 4a, most of the aforementioned eQTLs have been located on 5q mainly distributed through the intronic regions (figure 4b). These results suggest that eQTLs especially ones located in intronic regions could practically contribute in inflammatory events. In the present study, fatty acid desaturase 1 (FADS1) with rs102275, rs4246215, and rs4077515 has been identified as a gene neighboring 3 eQTLs associated to CD and in conjugation with FADS2 contribute to the biosynthesis of unsaturated fatty acids (hsa01040) and fatty acid metabolism (hsa01212) pathways. A number of FADs play a pivotal role in the production of omega-6 and omega-3 polyunsaturated fatty acid (PUFAs), for instance eicosapentaenoic, docosahexaenoic acids, and arachidonic acid obtained by the dietary contains fatty acid (41,42). The SNPs within the FADs may impact on the FADS2 expression and consequently the levels of PUFAs within cells and the inflammatory signaling through eicosanoids, and these pathways have been suggested as resolving mediators in chronic inflammatory disorders like atherosclerosis and other immune-mediated disorders (43). The diet-eQTL links could be biologically relevant to a sort of gene-environment interactions underlying CD. SLC22A5 and SLC22A4 risk factors including rs12521868 and rs2188962 as intronic variants were related to choline metabolism in cancer pathway (hsa05231). These genes are members 4 and 5 of the solute carrier family 22 (SLC22A4 [OMIM 604190] and SLC22A5 [OMIM 605956]) encode the organic cation transporters (OCTN) 1 and OCTN2 located through the IBD susceptibility locus IBD5 on chromosome 5q31 have been proposed to be related to CD by several genome screenings. Seemingly, decreasing the OCTN activity or expression may cause the alteration in gut bacterial community, which in turn generates susceptibility to CD (44-46).

Taken together, a number of neighboring genes of genomic risk variants were highlighted in this study whose interactions with each other may contribute to the development of CD. This is mainly occurred through the genes involved in chemokine and cytokine

pathways, which could be a relevance of the necessity in declining the expression of these chemicals to inhibit inflammatory responses in susceptible individuals as a therapeutic strategy.

However, this analysis is challenged by the disadvantage of inevitable overestimation in computational approaches, thus, employing more stringent parameters in predicting the in silico relationships would be apparently helpful in acquiring more reliable results and overcoming any inaccuracy coming from the nature of computational methodologies at the first come.

### CONCLUSION

The main goal of this analysis was exploiting interplay between gene and SNPs, which are thought to be influential in the pathogenesis of CD. Utilizing GWAS data with pooling information of regulatory interactions can help to discover underlying mechanisms and enlighten more molecular underpinnings of inflammatory conditions. We observed that the identified affected genes by genomic risk factors, mainly guided to interleukin signaling pathway and fatty acid and choline metabolism. To summarize, selected genes viz. FADs and SLC22s could be taken into account for future detection and therapeutic targets by experimental investments.

### ACKNOWLEDGEMENT

We thank Dr. Nooshin Omranian, scientific staff in Systems Biology and Mathematical Modelling Group, Max Planck Institute for Molecular Plant Physiology, Potsdam, Germany for her precious guidance.

### CONFLICT OF INTEREST

The authors declare no conflict of interests related to this work.

### REFERENCES

- Pascual V, Dieli-Crimi R, López-Palacios N, Bodas A, Medrano LM, Núñez C. Inflammatory bowel disease and celiac disease: Overlaps and differences. *World J Gastroenterol* 2014;20:4846-56.
- Liu JZ, Anderson CA. Genetic studies of Crohn's disease: Past, present and future. *Best Pract Res Clin Gastroenterol* 2014;28:373-86.
- Loftus EV Jr. Clinical epidemiology of inflammatory bowel disease: Incidence, prevalence, and environmental influences. *Gastroenterology* 2004;126:1504-17.
- Khor B, Gardet A, Xavier RJ. Genetics and pathogenesis of inflammatory bowel disease. *Nature* 2011;474:307-17.
- Henderson P, Stevens C. The Role of Autophagy in Crohn's Disease. *Cells* 2012;1:492-519.
- Homer CR, Richmond AL, Rebert NA, Achkar JP, McDonald C. ATG16L1 and NOD2 interact in an autophagy-dependent, anti-bacterial pathway implicated in Crohn's disease pathogenesis. *Gastroenterology* 2010;139:1630-41.e2.
- Hindorf LA, Sethupathy P, Junkins HA, Ramos EM, Mehta JP, Collins FS, et al. Potential etiologic and functional implications of genome-wide association loci for human diseases and traits. *Proc Natl Acad Sci USA* 2009;106:9362-7.
- Maurano MT, Humbert R, Rynes E, Thurman RE, Haugen E, Wang H, et al. Systematic localization of common disease-associated variation in regulatory DNA. *Science* 2012;337:1190-5.
- Anderson CA, Boucher G, Lees CW, Franke A, D'Amato M, Taylor KD, et al. Meta-analysis identifies 29 additional ulcerative colitis risk loci, increasing the number of confirmed associations to 47. *Nat Genet* 2011;43:246-52.
- Franke A, McGovern DP, Barrett JC, Wang K, Radford-Smith GL, Ahmad T, et al. Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nat Genet* 2010;42:1118-25.
- Halme L, Paavola-Sakki P, Turunen U, Lappalainen M, Farkkila M, Kontula K. Family and twin studies in inflammatory bowel disease. *World J Gastroenterol* 2006;12:3668-72.
- Shivananda S, Lennard-Jones J, Logan R, Fear N, Price A, Carpenter L, et al. Incidence of inflammatory bowel disease across Europe: is there a difference between north and south? Results of the European Collaborative Study on Inflammatory Bowel Disease (EC-IBD). *Gut* 1996;39:690-7.
- Hiatt RA, Kaufman L. Epidemiology of inflammatory bowel disease in a defined northern California population. *West J Med* 1988;149:541-6.
- Farrokhyar F, Swarbrick ET, Irvine EJ. A critical review of epidemiological studies in inflammatory bowel disease. *Scand J Gastroenterol* 2001;36:2-15.
- Mathew CG, Lewis CM. Genetics of inflammatory bowel disease: progress and prospects. *Hum Mol Genet* 2004;13 Spec No 1:R161-8.
- Pascual V, Dieli-Crimi R, López-Palacios N, Bodas A, Medrano LM, Núñez C. Inflammatory bowel disease and celiac disease: Overlaps and differences. *World J Gastroenterol* 2014;20:4846-56.
- Iezzi LE, Medeiros BA, Feitosa MR, Ribeiro de Almeida



- A, Parra RS, Ribeiro de Rocha J, et al. Association between celiac disease and Crohn's disease – a challenge to the coloproctologist. *J Coloproctol* 2012;32:329-33.
18. Tamboli CP, Neut C, Desreumaux P, Colombel JF. Dysbiosis as a prerequisite for IBD. *Gut* 2004;53:1057.
  19. Schreiber S. Of mice and men: what to learn about human inflammatory bowel disease from genetic analysis of murine inflammation. *Gastroenterology* 2005;129:1782-4.
  20. Xavier RJ, Podolsky DK. Unravelling the pathogenesis of inflammatory bowel disease. *Nature* 2007;448:427-34.
  21. Jostins L, Ripke S, Weersma RK, Duerr RH, McGovern DP, Hui KY, et al. Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* 2012;491:119-24.
  22. Frank DN, Robertson CE, Hamm CM, Kpadeh Z, Zhang T, Chen H, et al. Disease phenotype and genotype are associated with shifts in intestinal-associated microbiota in inflammatory bowel diseases. *Inflamm Bowel Dis* 2011;17:179-84.
  23. Rehman A, Sina C, Gavrilova O, Hasler R, Ott S, Baines JF, et al. Nod2 is essential for temporal development of intestinal microbial communities. *Gut* 2011;60:1354-62.
  24. Son HJ, Lee SH, Lee SY, Kim EK, Yang EJ, Kim JK, et al. Oncostatin M Suppresses Activation of IL-17/Th17 via SOCS3 Regulation in CD4+ T Cells. *J Immunol* 2017;198:1484-91.
  25. Beigel F, Friedrich M, Probst C, Sotlar K, Göke B, Diegelmann J, et al. Oncostatin M Mediates STAT3-Dependent Intestinal Epithelial Restitution via Increased Cell Proliferation, Decreased Apoptosis and Upregulation of SERPIN Family Members. *PLoS One* 2014;9:e93498.
  26. Singh UP, Singh NP, Murphy EA, Price RL, Fayad R, Nagarkatti M, et al. Chemokine and cytokine levels in inflammatory bowel disease patients. *Cytokine* 2016;77:44-9.
  27. Parronchi P, Romagnani P, Annunziato F, Sampognaro S, Becchio A, Giannarini L, et al. Type 1 T-helper cell predominance and interleukin-12 expression in the gut of patients with Crohn's disease. *Am J Pathol* 1997;150:823-32.
  28. Sasaki S, Yoneyama H, Suzuki K, Suriki H, Aiba T, Watanabe S, et al. Blockade of CXCL10 protects mice from acute colitis and enhances crypt cell survival. *Eur J Immunol* 2002;32:3197-205.
  29. Khalili H, Higuchi LM, Ananthkrishnan AN, Manson JE, Feskanich D, Richter JM, et al. Hormone Therapy Increases Risk of Ulcerative Colitis but not Crohn's Disease. *Gastroenterology* 2012;143:1199-206.
  30. Pierdominici M, Maselli A, Varano B, Barbati C, Cesaro P, Spada C, et al. Linking estrogen receptor  $\beta$  expression with inflammatory bowel disease activity. *Oncotarget* 2015;6:40443-51.
  31. Hotamisligil GS, Shargill NS, Spiegelman BM. Adipose expression of tumor necrosis factor-  $\alpha$ : direct role in obesity-linked insulin resistance. *Science* 1993;259:87-91.
  32. Yang XD, Tisch R, Singer SM, Cao ZA, Liblau RS, Schreiber RD, et al. Effect of tumor necrosis factor  $\alpha$  on insulin-dependent diabetes mellitus in NOD mice. I. The early development of autoimmunity and the diabetogenic process. *J Exp Med* 1994;180:995-1004.
  33. Donath MY, Shoelson SE. Type 2 diabetes as an inflammatory disease. *Nat Rev Immunol* 2011;11:98-107.
  34. Sherlock ME, Walters T, Tabbers MM, Frost K, Zachos M, Muise A, et al. Infliximab-induced psoriasis and psoriasiform skin lesions in pediatric Crohn disease and a potential association with IL-23 receptor polymorphisms. *J Pediatr Gastroenterol Nutr* 2013;56:512-8.
  35. Mohan M, Chow CT, Ryan CN, Chan LS, Dufour J, Aye PP, et al. Dietary Gluten-Induced Gut Dysbiosis Is Accompanied by Selective Upregulation of microRNAs with Intestinal Tight Junction and Bacteria-Binding Motifs in Rhesus Macaque Model of Celiac Disease. *Nutrients* 2016;8. pii: E684.
  36. Mahdi BM. Role of HLA typing on Crohn's disease pathogenesis. *Ann Med Surg (Lond)* 2015;4:248-53.
  37. Ebert EC. IL-15 converts human intestinal intraepithelial lymphocytes to CD94+ producers of IFN- $\gamma$  and IL-10, the latter promoting Fas ligand-mediated cytotoxicity. *Immunology* 2005;115:118-26.
  38. Rauch I, Müller M, Decker T. The regulation of inflammation by interferons and their STATs. *JAK-STAT* 2013;2:e23820.
  39. Sagami S, Ueno Y, Tanaka S, Fujita A, Niitsu H, Hayashi R, et al. Choline Deficiency Causes Colonic Type II Natural Killer T (NKT) Cell Loss and Alleviates Murine Colitis under Type I NKT Cell Deficiency. *PLoS One* 2017;12:e0169681.
  40. Chao CY, Battat R, Al Khoury A, Restellini S, Sebastiani G, Bessissow T. Co-existence of non-alcoholic fatty liver disease and inflammatory bowel disease: A review article. *World J Gastroenterol* 2016;22:7727-34.
  41. Lattka E, Eggers S, Moeller G, Heim K, Weber M, Mehta D, et al. A common FADS2 promoter polymorphism increases promoter activity and facilitates binding of transcription factor ELK1. *J Lipid Res* 2010;51:182-91.
  42. Martinelli N, Consoli L, Olivieri O. A 'desaturase hypothesis' for atherosclerosis: Janus-faced enzymes in omega-6 and omega-3 polyunsaturated fatty acid metabolism. *J Nutrigenet Nutrigenomics* 2009;2:129-39.
  43. Serhan CN, Clish CB, Brannon J, Colgan SP, Chiang N, Gronert K. Novel functional sets of lipid-derived mediators with antiinflammatory actions generated from

omega-3 fatty acids via cyclooxygenase 2-nonsteroidal antiinflammatory drugs and transcellular processing. *J Exp Med* 2000;192:1197-204.

44. Ma Y, Ohmen JD, Li Z, Bentley LG, McElree C, Pressman S, et al. A genome-wide search identifies potential new susceptibility loci for Crohn's disease. *Inflamm Bowel Dis* 1999;5:271-8.
45. Rioux JD, Silverberg MS, Daly MJ, Steinhart AH, McLeod RS, Griffiths AM, et al. Genomewide search in Canadian families with inflammatory bowel disease reveals two novel susceptibility loci. *Am J Hum Genet* 2000;66:1863-70.
46. Peltekova VD, Wintle RF, Rubin LA, Amos CI, Huang Q, Gu X, et al. Functional variants of OCTN cation transporter genes are associated with Crohn disease. *Nat Genet* 2004;36:471-5.