

1 RESEARCH ARTICLE

2 Hepatoprotective Activity of *Camellia sinensis* and its
 3 Possible Mechanism of Action

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9 ABSTRACT

10 The present study appraised the hepatoprotective activity of aqueous extract of *Camellia sinensis* leaves
 11 and its possible mechanism of action. Liver damage was induced by intraperitoneal administration of car-
 12 bon tetrachloride/olive oil (50 % v/v, 0.5 ml/kg) in male Wistar rats (150-220g) once daily for 7 days and
 13 the extent of damage was studied by assessing biochemical parameters such as alanine amino trans-
 14 ferase (ALT), aspartate amino transferase (AST), alkaline phosphatase (ALP), total protein and albumin
 15 in serum and concentrations of lipid peroxides (LPO), glutathione (GSH), catalase (CAT) and superoxide
 16 dismutase (SOD) in liver. The aqueous extract of *Camellia sinensis* (100 mg and 200 mg/Kg) were ad-
 17 ministered orally to the animals with hepatotoxicity induced by carbon tetrachloride and its effects on bio-
 18 chemical parameters were compared with those in animals treated with vitamin E (100 mg/Kg). Histopa-
 19 thological studies were also done. *Camellia sinensis* 100 and 200mg/kg results in significant reduction in
 20 serum hepatic enzymes and liver lipid peroxide which were increased by carbon tetrachloride. There was
 21 significant increase in serum total protein, albumin and liver GSH, SOD and CAT when compared to
 22 those in rats treated by carbon tetrachloride. The antioxidant activity of *Camellia sinensis* (100 and
 23 200mg/Kg) were comparable with the effects of vitamin E (100mg/Kg). Histopathological changes (con-
 24 gestion of central vein, centrilobular necrosis and sinusoidal congestion) induced by carbon tetrachloride
 25 were reduced to a moderate extent in *Camellia-sinensis*-treated rats. Taking together, *Camellia sinensis*
 26 protects the liver from carbon-tetrachloride-induced damage. Probable mechanism of its action is its
 27 anti-oxidant property.

28 **Keywords:** *Camellia sinensis*, Antioxidant, Carbon tetrachloride, Hepatoprotective

29 *Camellia sinensis* is a perennial tree belonging to the 47 to reduced anti-oxidant. Antioxidants are compounds
 30 family Theaceae, commonly called as 'Green tea'. It is 48 that protect cell against the damaging effect of reactive
 31 native from the south of China. It appears as a bush of 49 oxygen species such as singlet oxygen, superoxide,
 32 about 2.5 m in the high areas of Asia and China with 50 proxy radicals, hydroxyl radicals and peroxy nitrite. An
 33 warm and humid climates. The leaves are thermogenic, 51 imbalance between antioxidant and reactive oxygen
 34 appetizer, digestive, carminative, diuretic, and useful in 52 species results in oxidative stress, leading to cellular
 35 cardiodynia, hemorrhoids, inflammation and abdominal 53 damage [7]. Catechins are hypothesized to help protect
 36 disorders [1]. It has been previously reported that the 54 against many disease by contributing along with anti-
 37 leaves have used to treat the cancer of duodenum, lung, 55 oxidant vitamin E and enzyme like superoxide dismu-
 38 liver and mammary gland [2-5]. *Camellia sinensis* con- 56 tase, catalase to the total anti-oxidant defense system.
 39 tain many biologically-active polyphenolic flavonol 57 Presence of number of constituents has been reported in
 40 commonly known as catechine which make up 30 % of 58 *Camellia sinensis* catechin (flavanols) especially epigal-
 41 dry weight of its leaves [6]. Free radicals cause oxida- 59 locatechin gallate, epigallocatechin, epicatechin gallate
 42 tion of nucleic acid proteins. Free radical also damage 60 and epicatechin which have been identified as active
 43 biomembranes, reflected by increased lipid peroxida- 61 components responsible for antioxidant property [8].
 44 tion, thereby compromising cell integrity and function. 62 Among the various mechanisms involved in hepato-
 45 During this process, the ability of the body's defense 63 toxic effect of carbon tetrachloride, one is oxidative
 46 system to combat the oxidative stress may diminish due 64 damage through free radical generation and antioxidant

Table 1. Effect of *Camellia Sinensis* on serum ALT, AST, ALP, Total protein and Albumin in CCl₄- treated rats

Groups	Drug treatment	ALT (U/L)	AST (U/L)	ALP (U/L)	Total Protein g / dl	Albumin g / dl
I	Distilled Water (1ml/kg p.o.)	107.6 ± 9.5	59.5 ± 4.1	249.5 ± 18.2	8.5±0.6	6.2±0.5
II	CCl ₄ (0.5ml/kg i.p.)	378.9 ± 23.7 ^a	254.9±19.3 ^a	586.9 ± 31.6 ^a	5.1±0.4 ^a	2.4±0.3 ^a
III	CCl ₄ + <i>Camellia sinensis</i> (100mg/kg)	131.6 ± 10.6 ^c	71.4 ± 5.6 ^c	267.7 ± 17.8 ^c	6.9±0.8 ^c	4.7±0.3 ^c
IV	CCl ₄ + <i>Camellia sinensis</i> (200mg/kg p.o.)	117.6 ± 6.7 ^b	66.2± 6.1 ^b	258.4 ± 16.2 ^b	7.6±0.6 ^b	5.4±0.5 ^b
V	CCl ₄ + Vitamin E (100mg/kg p.o.)	111.7 ± 8.7 ^b	62.3 ± 5.1 ^b	255.3 ± 24.1 ^b	7.8±0.4 ^b	5.9±0.5 ^b

Values are in Mean ± SEM. Number of animals in each group = 6. ^a p < 0.001 Vs Group I. ^b p < 0.01 Vs Group II. ^c p < 0.05 Vs Group II. (CCl₄: carbon tetrachloride, AST: aspartate aminotransferase, ALT: alanine aminotransferase, ALP: alkaline phosphatase)

property is claimed to be one of the mechanisms of hepatoprotective effect of indigenous substance [9]. *Camellia sinensis* has antioxidant properties [10]. Hence, the objective of the study was to evaluate the hepatoprotective effect of *Camellia sinensis* on carbon tetrachloride-induced hepatotoxicity.

MATERIALS AND METHODS

Drugs and chemicals

Carbon tetrachloride (CCl₄) was obtained from Merck Ltd., Mumbai, India; Thiobarbituric acid (TBA), 5, 5'-dithio-bis-2-nitrobenzodic acid (DTNB) and glutathione (GSH) were obtained from Sigma, USA. Vitamin E was obtained from Hi Media Pvt. Ltd., Mumbai. All chemicals used in the study were of analytical grades.

Plant material

The aerial parts of *Camellia sinensis* were collected from the hills of Ootacamund, South India, in the month of February. The plant samples were identified and authenticated by the botanist, Botanical Survey of India, Agricultural University, Coimbatore, India. The voucher specimen (A 2459) has been deposited in Herbarium.

Extract Preparation

The collected aerial parts of *Camellia sinensis* was washed, air dried, powdered and boiled in sufficient quantity of distilled water for 2 hours and the aqueous extract was filtered, concentrated in vacuo and lyophilized to give a dry extract [11].

Animals

Male Swiss albino mice weighing between 20–25 gm and male wistar Albino rats weighing between 150–220 gm were used. The animals were obtained from animal house, IRT Perundurai Medical College, Erode, Tamilnadu, India. On arrival, the animals were placed at random and allocated to treatment groups in polypropylene cages with paddy husk as bedding. Animals were housed at a temperature of 24±2°C and relative humidity of 30–70 %. A 12:12-h light:dark cycle was followed. All animals were allowed to free access to water and fed with standard commercial pelleted rat chaw (M/s. Hindustan Lever Ltd, Mumbai). All the experimental procedures and protocols used in this study were reviewed by the Institutional Animal Ethics Committee (Regd no: 688/2/C-CPCSEA) and were in accordance with the guidelines of the Committee for the Purpose of Control and Supervision on Experiments on Animals (CPCSEA).

Acute Toxicity Studies

Acute toxicity studies were performed according to Organization for Economic Co-Operation and Development (OECD)-423 guidelines [12]. Male Swiss mice selected by random sampling technique were employed in this study. The animals were fasted for 4 hours with free access to water only. *Camellia sinensis* was administered orally at a dose of 5 mg/kg initially. Mortality if any was observed for 3 days. If mortality was observed in two out of three animals, then the dose administered was considered as toxic dose. However, if the mortality was observed in only one animal out of three animals then the same dose was repeated again to confirm the toxic effect. If no mortality was observed, then higher doses (50, 300, 2000 mg/kg) of *Camellia sinensis* were employed for further toxicity studies.

Experimental procedure

The experiment was carried out after obtaining clearance from Institutional Animal Ethics Committee. The animals were divided in to 5 groups of 6 animals each. Group-I, which served as normal control received distilled water (1ml/kg., p.o.), Group-II received equal mixture of CCl₄ and olive oil (50 % v/v, 0.5 ml/kg i.p.) once daily for 7 days [13]. Group-III received equal mixture of CCl₄ and olive oil along with *Camellia sinensis* (100 mg/Kg, p.o.) simultaneously once daily for 7 days. Group-IV received equal mixture of CCl₄

Table 2. Effect of *Camellia sinensis* on liver LPO, GSH, CAT and SOD in CCl₄-treated rats

Groups	Drug treatment	LPO nmol of MDA/mg protein	GSH nmol/mg tissue	CAT nmol of H ₂ O ₂ decom- position/min./mg protein	SOD Units/g protein
I	Distilled Water (1ml/kg p.o.)	4.1 ±0.5	22.5 ±1.9	189.8 ± 11.3	84.6 ± 6.8
II	CCl ₄ (0.5ml/kg i.p.)	14.8 ±1.3 ^a	11.2 ±1.7 ^a	46.2 ±5.6 ^a	42.5 ±3.2 ^a
III	CCl ₄ + <i>Camellia sinen-</i> <i>sis</i> (100mg/kg)	8.4 ±0.7 ^c	17.6 ±1.6 ^c	152.4 ±13.7 ^c	67.7 ±5.7 ^c
IV	CCl ₄ + <i>Camellia sinen-</i> <i>sis</i> (200mg/kg p.o.)	6.1 ±0.5 ^b	18.3 ±1.9 ^b	160.3 ±15.1 ^b	75.3 ±6.9 ^b
V	CCl ₄ + Vitamin E (100mg/kg p.o.)	5.4 ±0.6 ^b	20.9 ±2.2 ^b	168.4 ±11.3 ^b	77.1 ±5.4 ^b

Values are in Mean ± SEM. Number of animals in each group = 6. ^a p < 0.001 Vs Group I. ^b p < 0.01 Vs Group II. ^c p < 0.05 Vs Group II. (CCl₄: carbon tetrachloride, LPO: lipid peroxide, GSH: glutathione, CAT: catalase and SOD: superoxide dismutase)

139and olive oil along with *Camellia sinensis* (200 mg/Kg, 171
140p.o.) simultaneously once daily for 7 days. Group-V
141received equal mixture of CCl₄ and olive oil along with
142vitamin E (100 mg/Kg, p.o.) simultaneously once daily
143for 7 days [14]. On 8th day, the blood was collected by
144direct cardiac puncture under light ether anesthesia and
145serum was separated for various biochemical estima-
146tions. All animals were sacrificed by cervical decapita-
147tion and immediately, the livers were dissected out,
148washed in the ice cold saline and homogenate was pre-
149pared in 0.05 M sodium phosphate buffer (pH 7.0) and
150centrifuged. The supernatant was used for the estimation
151of lipid peroxide (LPO), glutathione (GSH), catalase
152(CAT) and superoxide dismutase (SOD). The activities
153of serum hepatic marker enzymes namely aspartate
154aminotransferase (AST), alanine aminotransferase
155(ALT) and alkaline phosphatase (ALP) were assayed in
156serum using standard kits from Lupin Laboratories and
157Pointe Scientifics. The results were expressed as
158units/litre (U/L). The levels of proteins i.e., total pro-
159teins and albumins were estimated in serum of experi-
160mental animals by earlier method reported [15]. The
161LPO in the liver was determined according to previous
162report [16]. GSH content was estimated in the liver
163homogenate using DTNB [17]. CAT and SOD activity
164was measured in the liver homogenate by the reported
165method [18,19].

166 Statistical analysis

167 The values were expressed as mean ± SEM. The sta-
168tistical analysis was carried out by one way analysis of
169variance (ANOVA) followed by Dunnet's t-test. P val-
170ues <0.05 were considered significant.

RESULTS

172 Acute Toxicity Studies

173 All the doses (5, 50, 300, 2000 mg/kg, p.o.) of
174 *Camellia sinensis* employed for acute oral toxicity stud-
175ies were found to be non-toxic. *Camellia sinensis* did
176not produce any mortality even at the highest dose
177(2000 mg/kg, p.o.) employed. Two sub-maximal doses
178(100 and 200 mg/kg, p.o.) which were found to be safe
179were employed for further pharmacological investiga-
180tions.

181 Biochemical estimations

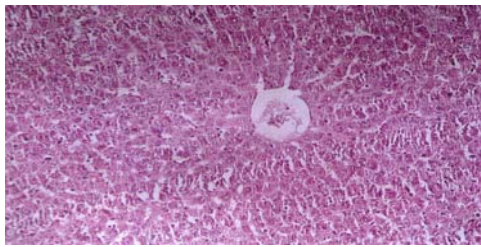
182 The results of hepatoprotective activity of *Camellia*
183 *sinensis* on CCl₄-treated rats are shown in Tables 1 and 2.

184 Serum hepatic enzymes

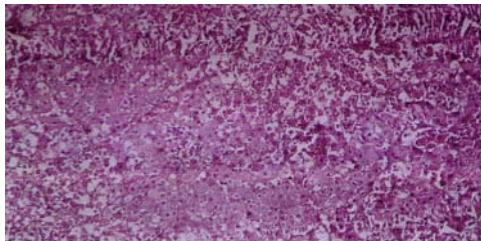
185 The hepatic enzymes ALT, AST and ALP in serum
186was significantly increased in CCl₄ treated animals
187when compared to control (p<0.001). The *Camellia*
188 *sinensis* treatment (200 mg/kg) significantly (p<0.01)
189increased the levels of hepatic enzymes when compared
190to CCl₄-treated animals. The *Camellia sinensis* treat-
191ment (100 mg/kg) less significantly (p < 0.05) increased
192the levels of hepatic enzymes when compared to CCl₄-
193treated animals. Vitamin E (100 mg/kg)-treated animals
194also showed significant (p<0.01) increase in the levels
195of hepatic enzymes when compared to CCl₄-treated
196animals.

197 Serum proteins

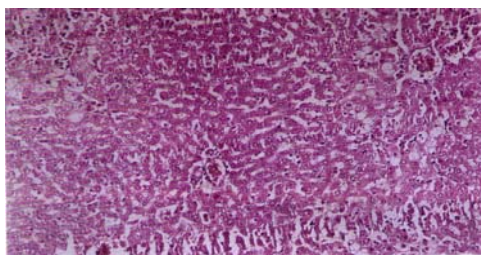
198 There was a significant (p<0.001) decrease in the se-
199rum total protein and albumin levels with CCl₄ treat-
200ment in group II when compared to control group I;



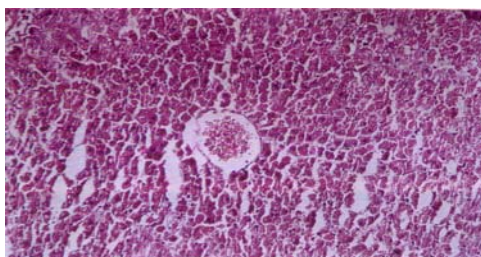
(A) Transverse section of the liver of control rats, showed normal hepatic cells with well preserved cytoplasm, prominent nucleus and nucleolus and central vein.



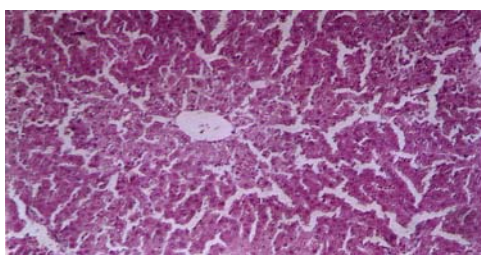
(B) Transverse section of the liver of CCl₄ treated animals showing hydropic changes in centrilobular hepatocytes with single cell necrosis surrounded by neutrophils, congestion of central vein and sinusoids were seen with acute inflammatory cells infiltrating sinusoids mainly in central zone.



(C) Transverse section of the liver, after simultaneous treatment of *Camellia sinensis* (100mg/kg) and CCl₄ treated animals showing mild fatty change and mild sinusoidal congestion.



(D) Transverse section of the liver, after simultaneous treatment of *Camellia sinensis* (200mg/kg) and CCl₄ treated animals showing residual hepatocellular necrosis with cords of regeneration hepatocytes.



(E) Transverse section of the liver, after simultaneous treatment of Vitamin E and CCl₄ treated animals showing mild central venous congestion and mild fatty vacuolation.

Fig 1. Histopathological studies of *Camellia sinensis* and vitamin E on CCl₄-treated rats

201 which was significantly ($p < 0.01$) reversed with the 212 animals also showed significant ($p < 0.01$) decrease in the
202 treatment of *Camellia sinensis*. 213 levels of LPO when compared to CCl₄-treated animals.

203 *Lipid peroxidation*

214 *Glutathione, catalase and super oxide dismutase*

204 The LPO level in liver was significantly increased 215 In order to find the possible mechanism by which
205 ($p < 0.001$) in CCl₄-treated animals when compared to 216 *Camellia sinensis* prevents hepatic damage caused by
206 control. Treatment with *Camellia sinensis* at 200 mg/kg 217 CCl₄, investigation on levels of GSH, SOD and CAT
207 showed significant ($p < 0.01$) decrease in LPO level 218 was carried out. The levels of GSH, SOD and CAT in
208 when compared to CCl₄-treated groups. Treatment with 219 liver homogenate were significantly decreased
209 *Camellia sinensis* at 100 mg/kg showed less significant 220 ($p < 0.001$) in CCl₄-treated animals when compared to
210 ($p < 0.05$) decrease in LPO level when compared to 221 control. Treatment with *Camellia sinensis* (200 mg/ kg)
211 CCl₄-treated groups. Vitamin E (100 mg/kg)- treated 222 showed significant ($p < 0.01$) rise in GSH, SOD and CAT

223 levels when compared to CCl₄-treated groups. Treat-279 oxidative degradation of membrane lipids of endoplas-
224 ment with *Camellia sinensis* (100 mg/ kg) dose showed280 mic reticulum which are rich in polyunsaturated fatty
225 less significant ($p<0.05$) rise in GSH, SOD and CAT281 acids. This leads to formation of lipid peroxides, which
226 levels when compared to CCl₄-treated groups. Vitamin282 in turn gives products like melanodialdehyde (MDA)
227 E (100 mg/kg)-treated animals also showed significant283 that cause damage to the membrane. This lipid peroxi-
228 ($p<0.01$) rise in the levels of GSH, SOD and CAT when284 dative degradation of biomembrane is one of the princi-
229 compared to CCl₄-treated animals.285

230 Histopathological studies

231 The results of histopathological studies of *Camellia*
232 *sinensis* on CCl₄-treated rats are shown in Fig 1. In con-
233 trol rats, liver sections showed normal hepatic cells with
234 well-preserved cytoplasm, prominent nucleus and nu-
235 cleolus and central vein. In CCl₄-treated rats, liver there
236 were hydropic changes in centrilobular hepatocytes with
237 single cell necrosis surrounded by neutrophils. Conges-
238 tion of central vein and sinusoids were seen with acute
239 inflammatory cells infiltrating sinusoids mainly in cen-
240 tral zone. In *Camellia sinensis* (100mg/kg) and CCl₄-
241 treated rats, liver sections showed mild fatty change and
242 mild sinusoidal congestion. In *Camellia sinensis*
243 (200mg/kg) and CCl₄ treated rats, liver sections showed
244 residual hepatocellular necrosis with cords of regenerat-
245 ing hepatocytes. In Vitamin E and CCl₄ treated rats,
246 there was mild central venous congestion and mild fatty
247 vacuolation.

248

DISCUSSION

249 CCl₄ is one of the most commonly used hepatotox-309
250 ins in the experimental study of liver diseases. [20].310
251 Assessment of liver function can be made by estimating311
252 the activities of serum and liver tissue enzymes origi-312
253 nally present or absent in cytoplasm. During hepatic313
254 damage, there may be imbalance in these enzyme levels314
255 with the extent of liver damage. The altered levels of315
256 these enzymes in CCl₄-treated rats in the present study316
257 corresponded to the extensive liver damage induced by317
258 the toxin.318

259 The serum ALT, AST and ALP are reliable markers319
260 of liver function. They were significantly increased in320
261 CCl₄-treated groups. On the other hand, in group III321
262 animals which were treated with *Camellia sinensis* (100322
263 mg/kg, p.o.), the activity of ALT, AST and ALP had323
264 decreased significantly ($p<0.05$) and in group IV, ani-324
265 mals which were treated with *Camellia sinensis* (200325
266 mg/kg, p.o.), the activity of ALT, AST and ALP had326
267 decreased significantly ($p<0.01$). Simultaneous treat-327
268 ment of *Camellia sinensis* and CCl₄ caused significant328
269 recovery from the damage induced by CCl₄ treatment.329

270 Histopatholoical studies showed that CCl₄ caused
271 centrilobular necrosis, congestion of central vein and
272 sinusoids. *Camellia sinensis* administration exhibited
273 protection against CCl₄-induced hepatotoxicity, which
274 and green tea [22] protects the liver against CCl₄.333
275 confirmed the results of biochemical studies. The results
276 induced hepatic damage in rats which support hepato-334
277 protective activity of *Camellia sinensis*. The hepatotoxic335
278 *sinensis* in CCl₄-treated rats protects liver damage. The
279 effects of CCl₄ are largely due to its active metabolite,336
280 biochemical evaluation indicates the hepatoprotective
281 trichloromethyl radical [23]. These activated radicals337
282 effects of *Camellia sinensis* may be due to its antioxi-
283 dant property.338

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344 **REFERENCES**

345 1. Warriar PK, Nambiar VPK, Ramankutty C. Indian Medicinal
346 Plants A compendium of 500 species. Vol. 1, Orient Longman
347 Pvt Ltd, India. 1994; P. 349-351.

348 2. Yamane T, Nakatani H, Kikuoka N, Matsumoto H, Iwata Y,
349 Kitao Y. Inhibitory effects and toxicity of green tea polyphenols
350 for gastrointestinal carcinogenesis. *Cancer* 1996; 77: 1662-7.

351 3. Xu Y, Ho CT, Amin SG, Han C, Chung FC. Inhibition of to-
352 bacco specific nitrosamine induced lung tumorigenesis in A/J
353 mice by green tea and its major polyphenol as antioxidants.
354 *Cancer Res* 1992; 35: 3875-9.

355 4. Cao J, Xu Y, Chen J, Klauning JE. Chemo preventive effects of
356 green tea and black tea on pulmonary and hepatic carcinogene-
357 sis. *Fundam Appl Toxicol* 1996; 29: 244-0.

358 5. Tanake H, Hirose M, Kawabe M, Sano M, Takesada Y, Hagi-
359 wara A. Post initiation inhibitory effects of green tea catechins
360 on 7,12-dimethylbenz(a) anthracene induced mammary gland
361 carcinogenesis in female Sprague Dawley rats. *Cancer Lett*
362 1997; 116: 47-52.

363 6. Ahmad N, Mukhtar H. Green tea polyphenols and cancer; bio-
364 logic mechanism and practical implications. *Nutr Rev* 1999; 57:
365 78-83.

366 7. Skrzydlewska E, Ostrowska J, Farbisxewsk R, Michalak. Protec-
367 tive effect of green tea against lipid peroxidation in the rat liver,
368 blood and brain. *Phytomedicine* 2002; 9: 232-8.

369 8. Graham HN. Green tea composition, consumption and polyphe-
370 nol chemistry. *Prev Med* 1992; 21: 334-50.

371 9. Deleve LD, Kaplowitz N. Mechanisms of drug induced liver
372 disease. *Gastroenterol Clin North Am* 1995; 24 : 787-810.

373 10. Lin YL, Cheng CY, Lin YP, Lau YW, Jaun IM, Lin JK. Hypol-
374 ipidemic effect of green tea leaves through induction of antioxi-
375 dant and Phase II enzymes including superoxide dismutase, cata-
376 lase and glutathione S- transferase in rats. *J Agric Food Chem*
377 1998; 46 : 1893-9.

378 11. Lee H-T, Seo E-K, Suk J-C, Shim C-K. Prokinetic activity of an
379 aqueous extract from dried immature fruit of Poncirus trifoliata
380 (L). *Raf J Eth Pharmacol* 2005; 102: 131-6.

381 12. Ecobichon DJ. The Basis of Toxicology Testing. CRC Press,
382 New York. 1997 ;P. 43-86.

383 13. Rao PGM, Rao SG, Kumar V. Effect of hepatoguard against
384 carbon tetrachloride induced liver damage in rats. *Fitoterapia*
385 1993; 64: 108-13.

386 14. Sheweita SA, Abd El-Gabar M, Bastawy M. Carbon tetrachlo-
387 ride-induced changes in the activity of Phase II drug metaboliz-
388 ing enzymes in the liver of male rats: role of antioxi-
389 dants.*Toxicol* 2001; 165: 217-24.

390 15. Gowenlock AH, McMurray JR, McLauchlan DM. Plasma Pro-
391 teins. In, Varley's Practical Clinical Biochemistry, CBS Publish-
392 ers, New Delhi. 1988; 6: 401-35.

393 16. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in ani-
394 mal tissues by thiobarbituric acid reaction. *Ann Biochem* 1979;
395 95: 351-8.

396 17. Buetler E, Duron O, Kelly BM. Improved method for determina-
397 tion of blood glutathione. *J Lab Clin Med* 1963; 61: 882-90.

398 18. Bergmeyer HV, Gawehn K, Grassik M. Methods of enzyme
399 analysis. In: Bergmeyer HV, Chemie V, Weinhein S (Eds). Aca-
400 demic press, New York. 1994; P.348.

401 19. Misra HP, Fridovich I. The role of super oxide anion in the auto
402 oxidation of epinephrine and a simple assay for super oxide dis-
403 mutase. *J Biol Chem* 1972; 247: 3170-85.

404 20. Johnson DE, Kroening C. Mechanism of early Carbon tetrachlo-
405 ride toxicity in cultured rat hepatocytes. *Pharmacol Toxicol*
406 1998; 83: 231-9.

407 21. Takashi H, Miho G, Hiroyuki S, Naomi O, Mayumi O, Kazuki
408 K. Yellow tea is more potent than other types of tea in suppress-
409 ing liver toxicity induced by carbon tetrachloride in rats. *Phyto-*
410 *ther Res* 2007; 21: 668-70.

411 22. Almurshed KS. Protective effect of black and green tea against
412 carbontetrachloride induced oxidative stress in rats. *Saudi Med J*
413 2006; 27:1804-9.

414 23. Srivastava SP, Chen NO, Holtzman JL. The Invitro NADPH
415 dependent inhibition by CCl₄ of the ATP-dependent calcium up-
416 take of hepatic microsomes from male rats. Studies on the
417 mechanism of inactivation of the hepatic microsomal calcium
418 pump by the CCl₄ radical. *J Biol Chem* 1990; 265: 8392-9.

419 24. Cotran RS, Kumar V, Robbins SL. Cell injury and cellular
420 death. In: Robbins Pathologic Basic of Disease, Prism Book
421 Pvt. Ltd. 1994; 5: 379-430.

422 25. Kaplowitz N, Aw TY, Simon FR, Stolz A. Drug induced hepato-
423 toxicity. *Ann Intern Med* 1986; 104: 826-39.

424 26. Ashok Shenoy K, Somayaji SV, Bairy KL. Hepatoprotective
425 effects of *Ginkgo biloba* against carbon tetrachloride induced
426 hepatic injury in rats. *Ind J Pharmacol* 2001; 33: 260-6.

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