

In Vivo and *In Vitro* Studies on Neutralizing Effects of *Acorus calamus* and *Withania somnifera* root extracts against *Echis carinatus* venom

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ABSTRACT

Neutralization effects of *Acorus calamus* and *Withania somnifera* root extracts were tested against *Echis carinatus* venom. Both plant extracts were effectively neutralized the various pharmacological activities induced by *Echis carinatus* venom. About 0.14 mg of *Acorus calamus* and 0.16 mg of *Withania somnifera* root extracts were able to completely neutralize the lethal activity of 2LD₅₀ of *Echis carinatus* venom. Various pharmacological activities like haemorrhagic, coagulant, edema and phospholipase activities were effectively neutralized by both plant extracts. The above observations confirmed that both plant extracts possess potent snake venom neutralizing compounds, which inhibit the activity of *Echis carinatus* venoms.

Keywords: *Echis carinatus* Venoms, *Acorus calamus*, *Withania somnifera*, Plant extracts, lethality, PLA2

Snake bite is a serious medico-legal problem particularly among the forest workers and agriculturists in rural India. There are over 2000 species of snakes in the world and about 216 species exists in India, of which 52 are Venomous [1]. The common poisonous snakes found in India are Cobra (*Naja naja*), Krait (*Bangarus caeruleus*), Russell's viper (*Daboia russelli*) and Saw Scaled Viper (*Echis Carinatus*) [2]. In India, about 15,000 persons are affected every year by snake envenomation. *Echis carinatus* (Saw-scaled viper) is responsible for a large number of snake bite case, reaching 95% of envenomations in the state of Jammu [3]. Antivenom immunotherapy is the only specific treatment against snake venom envenomation.

Antiserum development in animal is time-consuming, expensive and requires ideal storage condition. Over the years, many attempts have been made for the development of snake venom antagonists especially from plants sources. Extracts from plants have been used among traditional healers, especially in tropical areas where there are plentiful sources for snakebite therapy for a long time [4]. In modern science, there have been many attempts to study these plants to clarify their effectiveness [5, 6]. India has a rich tradition of the usage of medicinal plants. Many Indian medicinal plants are recommended for the treatment of snakebite [7]. Methanolic extracts of

Andrographis paniculata and *Aristolochia indica* plant extracts possess potent snake venom neutralizing capacity and could potentially be used for therapeutic purposes in case of snakebite envenomation [8]. Aqueous extract of *Mimosa pudica* root possesses compounds, which inhibit the activity of *Naja naja* and *Bangarus caeruleus* venoms. He also reported that aqueous extracts of *M. pudica* root possess compounds, which inhibit the activity of *Russell's viper* and *Saw scaled viper* venoms [9, 10]. Aqueous extracts of *Mucuna pruriens* seeds possess compounds, which inhibit the activity of cobra and krait venoms. [11]. The present investigation explored the *Neutralization effects of Acorus calamus* and *Withania somnifera* root extracts against *Echis carinatus* venom using *in vivo* and *in vitro* methods.

MATERIALS AND METHODS

Venom and Experimental animals

The free-dried snake venom powder of *Echis carinatus* was obtained from Irula's Snake Catchers Industrial Co-operative Society Limited, Chennai and was stored at 4°C. Male inbred Swiss albino mice (18-20 gm) were used for the studies of venom toxicity and in the experiments of venom neutralization. Institutional

70 Animal Ethics Committee clearance at Institute of
71 vector control and Zoonoses, Hosur, was obtained to
72 conduct the experiment.

73 Medicinal Plants and Preparation of Extracts

74 *Acorus calamus* and *Withania somnifera* plants were
75 obtained from Nehru Herbal Gardens, Coimbatore and
76 the extracts were prepared by the method of Uhegbu et
77 al. [12] using distilled water as the solvent. Twenty g of
78 powdered sample of the herb was extracted by soaking
79 in 180 mL of distilled water in a beaker, stirred for
80 about 6 min and left overnight. Thereafter, the solution
81 was filtered using filter paper (Whatman No. 1) and the
82 extracts were evaporated to dryness under reduced
83 pressure below 40°C. The plant extracts were expressed
84 in terms of dry weight.

85 Neutralization effects of *Acorus calamus* and 86 *Withania somnifera* root extracts against *Echis* 87 *carinatus* venom

88 In vivo neutralization assays

89 Lethal toxicity

90 The median lethal dose (LD₅₀) of *Echis carinatus*
91 venom was determined according to the method
92 developed by Theakston and Reid [13]. Various
93 concentrations of venom in 0.2 ml of physiological
94 saline was injected into the tail vein of mice (18-20
95 gms), using groups of 3-5 mice for each venom dose.
96 The LD₅₀ was calculated with the confidence limit at
97 50% probability by the analysis of deaths occurring
98 within 24 h of venom injection. The anti-lethal
99 potentials of *Acorus calamus* and *Withania somnifera*
100 root extracts were determined against 2LD₅₀ of *Echis*
101 *carinatus* venom. Various amount of Plant extracts were
102 mixed with 2LD₅₀ of venom sample and incubated at
103 37°C for 30 min and then injected intravenously into
104 mice. 3-5 mice were used for each antivenom dose.
105 Control mice received same amount of venom without
106 antivenom (Plant extracts). The median Effective Dose
107 (ED₅₀) was calculated from the number of deaths within
108 24h of injection of the venom/antivenom mixture. The
109 ED₅₀ was expressed as µl antivenom/mouse and
110 calculated by probit analysis.

111 Edema- forming Activity

112 The Minimum edema-forming dose (MED) of *Echis*
113 *carinatus* venom was determined by the method of
114 Lomonte et al. and Camey et al. [14, 15]. Group of four
115 mice were injected subcutaneously in the right footpad
116 with various amounts of venom (0.25µg - 10µg)
117 dissolved in 50 µl of phosphate-buffered saline (PBS),
118 pH 7.2. The left footpad received 50 µl of PBS alone
119 (control). Edema was calculated as percentage of
120 increase in the thickness of the right foot injected with
121 venom compared to the left foot. The thickness of each
122 footpad was measured every 30 min after venom
123 injection with a low-pressure spring caliper [16].
124 Minimum edema-forming dose (MED) was the venom

125 dose that induced 30% edema within 6 h of venom
126 injection when compared to control. The ability of
127 *Acorus calamus* and *Withania somnifera* root extracts in
128 neutralizing the edema were carried out by pre-
129 incubating the constant amount of venom and various
130 dilutions *Acorus calamus* and *Withania somnifera* root
131 extracts and incubated for 30 minutes at 37°C. Then,
132 groups of four mice (18-20 g) were injected
133 subcutaneously in the right footpad with 50 µl of the
134 mixtures, containing venom/plant extracts, whereas the
135 left footpad received 50µl of PBS alone. Control mice
136 were injected with venom in the right footpad and 50 µl
137 of PBS in the left footpad. One hour after injection,
138 edema was evaluated as described by Yamakawa et al.
139 [17]. Edema was expressed as the percentage increase in
140 thickness of the right footpad compared to the right
141 footpad of the control mice.

142 Haemorrhagic activity

143 The minimum haemorrhagic dose (MHD) of *Echis*
144 *carinatus* venom was determined by the method
145 described by Theakston and Reid [13]. The minimum
146 haemorrhagic dose was defined as the least amount of
147 venom which when injected intradermally (i.d.) into
148 mice, it resulted in a haemorrhagic lesion of 10 mm
149 diameter in 24 h. Neutralization of the haemorrhagic
150 activity was estimated by mixing a fixed amount of
151 venom with various amounts of *Acorus calamus* and
152 *Withania somnifera* root extracts. The mixture of plant
153 extract and venom was incubated at 37°C for 1 h and
154 0.1 ml of the mixture was injected intradermally into
155 mice. The haemorrhagic lesion was estimated after 24 h.

156 In vitro neutralization assays

157 Phospholipase activity

158 Phospholipase A₂ activity was measured using an
159 indirect hemolytic assay on agarose-erythrocyte-egg
160 yolk gel plate by the methods described by Gutierrez et
161 al. [18]. Increasing concentrations of *Echis carinatus*
162 venom was added to 3-mm wells in agarose gels (0.8%
163 in PBS, pH 8.1) containing 1.2% sheep erythrocytes,
164 1.2% egg yolk as a source of lecithin and 10 mM CaCl₂.
165 Slides were incubated at 37°C overnight and the
166 diameters of the hemolytic halos were measured.
167 Control wells contained 15 µl of saline. The minimum
168 indirect hemolytic dose (MIHD) corresponds to a
169 concentration of venom, which produced a hemolytic
170 halo of 11 mm diameter. The efficacy of *Acorus*
171 *calamus* and *Withania somnifera* root extracts in
172 neutralizing the phospholipase activity was carried out
173 by mixing constant amount of venom with various
174 amount of plant extracts and incubated for 30 min at
175 37°C. Then, aliquots of the mixtures (10 µl) were added
176 to wells in agarose-egg yolk-sheep erythrocyte gels.
177 Control samples contained venom without Plant
178 extracts. Plates were incubated at 37°C for 20 h.
179 Neutralization expressed as the ratio mg antibodies/mg

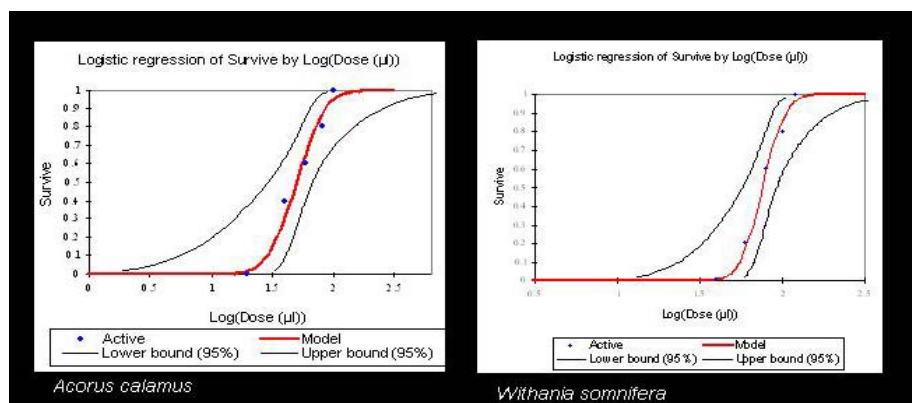


Fig 1. Dose response curve for Neutralization of Lethality by *Acorus calamus* and *Withania somnifera* root extracts against *Echis carinatus* venom in experiments involving preincubation of venom (2 x LD₅₀) and various concentrations of antivenoms (Plant extracts). The median effective dose for *Echis carinatus* venom was 0.14 mg for *Acorus calamus* and 0.16 mg for *Withania somnifera* root extracts

Table 1. Neutralization of *Echis carinatus* venom induced lethality by *Acorus calamus* and *Withania somnifera* root extracts

Plant Extracts	Concentration of <i>Echis carinatus</i> venom (µg)	Neutralization of venom by Plant extracts (ED ₅₀ in mg)
<i>Acorus calamus</i>	24 (2LD ₅₀)	0.14 mg
<i>Withania somnifera</i>	24 (2LD ₅₀)	0.16 mg

180 venom able to reduce 50% the diameter of the 210 with varying amount of *Acorus calamus* and *Withania*
181 hemolytic halo when compared to the effect induced by 211 *somnifera* plant extracts at 37°C for 1 h. After
182 venom alone. 212 incubation, the mixture was applied to the wells in the

183 Procoagulant activity

184 The procoagulant activity was done according to the 215
185 method described by Theakston and Reid [13] modified 216
186 by Laing *et al.* [19]. Various amounts of venom 217
187 dissolved in 100 µl PBS (pH 7.2) was added to human 218
188 citrated plasma at 37°C. Coagulation time was recorded 219
189 and the minimum coagulant dose (MCD) was 220
190 determined as the venom concentration which induced 221
191 clotting of plasma within 60 seconds. Plasma incubated 222
192 with PBS alone served as control. In neutralization 223
193 assays, constant amount of venom was mixed with 224
194 various dilutions of plant extracts. The mixtures were 225
195 incubated for 30 min at 37°C. Then, 0.1 ml of mixture 226
196 was added to 0.3 ml of citrated plasma and the clotting 227
197 times were recorded. In control tubes, plasma was 228
198 incubated with either venom alone or plant extracts 229
199 alone. Neutralization was expressed as effective dose 230
200 (ED), defined as the ratio µl antivenom (plant 231
201 extracts)/mg venom at which the clotting time was 232
202 increased three times when compared with clotting time 233
203 of plasma incubated with two MCD of venom alone. 234

204 Fibrinolytic activity

205 A modified plaque assay was used [16]. The 235
206 minimum fibrinolytic concentration was defined as the 236
207 concentration of venom that induced a fibrinolytic halo 237
208 of 10 mm diameter. Neutralization experiments were 238
209 performed by incubating a constant amount of venom 239

214 fibrinolytic halos were measured.

215 Statistical Analysis

216 Statistical evaluation was performed using XL stat
217 2008 and SPSS 10 Softwares. The *p values* < 0.005 was
218 considered statistically significant.

RESULTS

220 Neutralization effects of *Acorus calamus* and
221 *Withania somnifera* root extracts were tested against
222 *Echis carinatus* venom by *in vivo* and *in vitro* methods.
223 The lethal toxicity (LD₅₀) of *Echis carinatus* venom was
224 assessed using Balb/c strain mice. About 12 µg of *Echis*
225 *carinatus* venom was found to be LD₅₀ for mice
226 (weight: 18 g). The neutralization of lethality was done
227 by pre-incubating constant amount of venom with
228 various dilutions of *Acorus calamus* and *Withania*
229 *somnifera* root extracts prior to injection. We found that
230 0.14 mg of *Acorus calamus* and 0.16 mg of *Withania*
231 *somnifera* root extracts were able to completely
232 neutralize the lethal activity of 2LD₅₀ of *Echis carinatus*
233 venom (Table 1, Fig 1). In edema forming activity, 7µg
234 of Saw-scaled viper venom induced edema within 3 h
235 which is considered as 100% activity. The edema was
236 reduced up to 20% when 4000 µl of plant extracts/mg
237 venom was given. There was no further reduction in the
238 percentage of edema even when there was an increase in
239 anti-venom dose (Fig 2). In the case of hemorrhagic

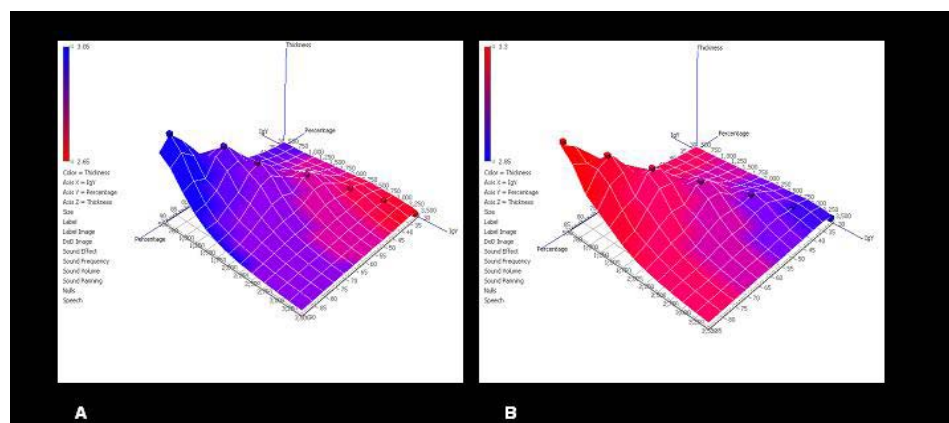


Fig 2. Neutralization of Edema induced by *Echis carinatus* venom by A) *Acorus calamus* and B) *Withania somnifera* root extracts in experiments with pre-incubation. Various mixtures of venom and antivenoms were incubated and tested in the foot pad assay. Edema was assessed 1 h after injection and expressed as percentage. Edema induced in control mice (venom alone) was considered as 100% activity. Results presented as mean \pm SE (N=3). $p < 0.005$ at all antivenoms/venom ratios.

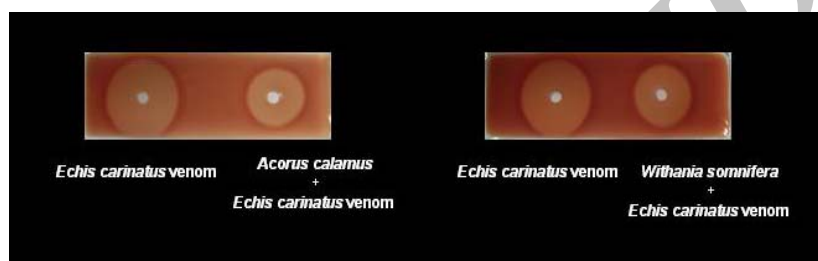


Fig 3. Neutralization of Phospholipase activity by *Acorus calamus* and *Withania somnifera* root extracts against *Echis carinatus* venom.

240activity, 8 μ g of venom produced a hemorrhagic spot of 203
24110 mm diameter (MHD). Both Plant extracts were able
242to neutralize the hemorrhage induced by the venom.
243In phos
244pholipase activity (PLA₂), 10 μ g of *Echis carinatus*
245venom was able to produce 11-mm hemolytic halo,
246which is considered to be 1 U (U/10 μ g). *Acorus*
247*calamus* and *Withania somnifera* root extracts were
248capable of inhibiting PLA₂-dependent hemolysis of
249sheep RBC's induced by *Echis carinatus* venom in a
250dose-dependent manner (Table 2, Fig 3). The minimum
251coagulant dose (MCD) was determined and we found
252that 120 μ g of Saw-scaled viper venom clotted human
253citratd plasma within 60 s. In the neutralization assay,
254the absence of clot formation shows the neutralizing
255ability of both plant extracts. High concentration of
256venom caused rapid clotting that required very high
257concentration of antivenom to neutralize. The
258fibrinolytic effect was effectively antagonized by the
259both plant extract. The ED₅₀ of *Acorus calamus* and
260*Withania somnifera* root extracts against *Echis*
261*carinatus* venom were found to be 0.5 and 0.8 mg
262respectively.

DISCUSSION

264 The most efficient treatment for snake bite
265envenomation is the specific heterologous serum. Anti-
266venom against snakes bites are lacking in the rural areas
267of coastal region. Antiserum is the only therapeutic
268agent and its development from animal source is time-
269consuming and expensive. Although, use of plants
270against the effects of snakes bite has been long
271recognized, more scientific attention has been given for
272the last 20 years [20]. Many Indian medicinal plants are
273recommended for the treatment of snakebites [7]. In the
274present study, we checked the antivenom potential of
275*Acorus calamus* and *Withania somnifera* root extracts
276against *Echis carinatus* venom. It is essential to
277understand the pharmacological action of snake venom
278in order to devise a rational treatment for snakebite. The
279neutralization ability of snake antivenoms is still
280assessed by the traditional in vivo lethality assay
281(minimum effective dose ED₅₀), comparable to those
282used for bacterial antitoxins, usually performed in mice
283[21]. Thus, various pharmacological activities like
284lethality, edema-forming activity, hemorrhagic activity,
285phospholipase activity (PLA₂) and pro-coagulant

Table 2. Phospholipase activity of *Echis carinatus* venom and its neutralization by *Acorus calamus* and *Withania somnifera* root extracts

Plant extracts	Dose of <i>Echis carinatus</i> venom (μ g)	Neutralization of venom by plant extracts (ED ₅₀ in mg)
<i>Acorus calamus</i>	10 (1 Unit)	0.10 mg
<i>Withania somnifera</i>	10 (1 Unit)	0.12 mg

- activity caused by *Echis carinatus* venom were carried out. Neutralization of these pharmacological effects was carried out using *Acorus calamus* and *Withania somnifera* root extracts. Neutralization studies can be performed by incubating venom and plant extracts prior to testing (pre-incubation method). The results showed that the both plant extracts were capable of neutralizing the lethality induced by the venom. The *Echis carinatus* venom showed the presence of PLA₂ enzymes by means of producing hemolytic haloes in indirect hemolytic assays. Both plant extracts were capable of inhibiting PLA₂-dependent hemolysis of sheep RBCs in a dose-dependent manner. The medicinal plants *Thea sinensis* Linn and *Cordia verbenacea* were effectively neutralized the phospholipase A₂ activity induced by snake venoms [22, 23]. Edema-forming activity was assessed for *Echis carinatus* venom and both plant extracts were found to be effective in neutralization of edema induced by venoms. There was a significant decrease in the edema (footpad thickness) when there was an increase in the antivenom (plant extract) concentration. Procoagulant activity induced by *Echis carinatus* venom was studied using human citrated plasma and *Acorus calamus* and *Withania somnifera* root extracts were found to be effective in the neutralization of procoagulant activity.
- The present experimental results indicate that *Acorus calamus* and *Withania somnifera* root extracts were effective in neutralizing the main toxic and enzymatic effects of *Echis carinatus* venom. The antivenom properties of both plant extracts were potent enough to neutralize the lethality and various pharmacological activities of venom. The result from this preliminary study indicates that both plant extracts could be used for therapy in patients with snakebite envenomation. Further investigations are needed for identification and purification of the active components involved in the neutralization of the snake venom.
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