

Review Article

The potential role of regenerative medicine in the management of traumatic patients

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KEY WORDS

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Abstract:

Traumatic injury represents the most common cause of death in ages 1 to 44 years and a significant proportion of patients treated in hospital emergency wards each year. Unfortunately, for patients who survive their injuries, survival is not equal to complete recovery. Many traumatic injuries are difficult to treat with conventional therapy and result in permanent disability. In such situations, regenerative medicine has the potential to play an important role in recovery of function. Regenerative medicine is a field that seeks to maintain or restore function with the development of biological substitutes for diseased or damaged tissues. Several regenerative approaches are currently under investigation, with a few achieving clinical application. For example, engineered skin has gained FDA approval, and more than 20 tissue engineered skin substitutes are now commercially available. Other organ systems with promising animal models and small human series include the central and peripheral nervous systems, the musculoskeletal system, the respiratory and genitourinary tracts, and others. This paper will be a clinically oriented review of the regenerative approaches currently under investigation of special interest to those caring for traumatic patients.

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Introduction

Trauma is the most common cause of death in people between 1 and 44 years and the third most common cause of death overall. Trauma causes more than 150,000 deaths per year in the United States. It has been reported that there were more than 167,000 trauma-related deaths in the US in 2004, but more than 29.6 million trauma patients treated in emergency wards in that year.¹ In the recent wars in Afghanistan and Iraq, significant improvements in personal protective devices have decreased the mortality rate relative to previous wars. Unfortunately, we have seen that survival, per se, does not equal complete recovery.² Thus, trauma-related mortality rates are not a good indicator of the full impact of trauma on our society. The

severity of many traumatic injuries makes them difficult to treat with conventional therapy.

In these situations regenerative medicine has the potential to play an important role. Regenerative medicine and tissue engineering is “an interdisciplinary field which applies the principles of engineering and life sciences towards the development of biological substitutes that aim to maintain, restore or improve tissue function” (Atala et al, 2001).³ As a result of the increasing human life span, the number of patients who need transplant organs has increased.

Issues with rejection and immunosuppressive drug complications have also increased the need for alternative treatments (**regenerative medicine**).www.SID.ir

This article will review the regenerative medicine approaches used in traumatic organ animal models and

human studies. The discussion will be clinically oriented, and as such, a discussion about basic techniques is beyond scope of this article.

Central Nervous System

Previously, scientists believed that the adult central nervous system (CNS) was unable to repair itself after injury or disease⁵ because of the limiting effects of inflammation, cellular loss, cavitations and glial scarring.⁶ However, repair of the CNS is currently one of the most promising fields of regenerative medicine.⁵

Several techniques have been used to create neural tissue in animal models. Spinal cord neural progenitor cells (SCNPCs) were cultured in vitro and transplanted into rat spinal cords after spinal cord injury (SCI). At follow-up, SCNPCs survived, but there was no significant differentiation toward neural morphology or neural functional improvement.⁶ In other studies, human umbilical cord blood (hUCB) cells were used to treat neurological disorders in various animal models. These cells are able to differentiate into several other cell types, including neural cells. Also, they have the ability to produce neurotropic factors that bring about the release of several growth factors. These growth factors stimulate cell survival and angiogenesis and have anti-inflammatory effects. Currently, there are few clinical studies using hUCB because the safety and efficacy has not been determined.⁷ The role of neural supporting cells (astrocytes and oligodendrocytes) in the restoration of neurological function has also been investigated. GRP cell-derived astrocytes (GDAs) were transplanted into transected rat spinal cords. After transplantation, axonal regeneration, neural cell survival, and tissue structure realignment was observed. This study demonstrates a role for astrocyte transplantation therapy in the restoration of neurological function.⁸ In animal models of SCI, transplantation of olfactory ensheathing cells supports axonal remyelination and migration along the spinal cord. This technique is useful for cases in which the anatomy is preserved but axons are physiologically disrupted.⁹ Some substances, such as glial cell line derived neurotrophic factor (GDNF), can increase the ability of a nerve graft placed in a spinal cord transaction gap to promote motor recovery and nerve growth.¹⁰ Stem cells are an ideal way to treat diseases and trauma of the nervous system.

Transplantation of embryonic or adult stem cells can replace damaged cells and aid in the recovery of function.¹¹ In a study by Nori et al. mouse induced pluripotent stem cells (iPSCs) and neural stem cell/progenitor cells (NS/PCs) were transplanted into the spinal cord 9 days after injury. These cells differentiate into all three neural lines and promote re-myelination, axonal re-

growth and recovery of motor function. No evidence of teratoma or other tumor formation was seen.¹² In another study, adipose-derived stromal cells were harvested from mice and humans and injected intravenously into mice with calvarial defects. These mice were compared to controls injected with saline only. Cell migration and bone formation was evaluated by histology and micro-CT scan. Histologic evidence of bone formation was seen in both mouse and human cells, but by CT analysis, only human cells promoted ossification. These results suggest that intravenous injection of human adipose-derived stromal cells may be an effective route for future skeletal regeneration.¹³ Recently, neuroepithelial-like stem cells have been developed from human iPS cells. Transplantation of these cells into a mouse model with SCI improves recovery of hind limb motor function.¹⁴ Studies have investigated the possible benefit of combined cell therapy in SCI models. Ban et al. used activated Schwann cells (ASCs) and bone mesenchymal stem cells (BMSCs) in rats with traumatic SCI. Co-transplantation of these cells led to recovery of hindlimb function, decreased glial scar formation and remyelination of damaged axons.¹⁵ Also, co-transplantation of peripheral nerve tissue with fetal brain tissue has been shown to be more effective for spinal cord reconstruction after injury.¹⁶

Biomaterial scaffolds create an environment for directed cell growth that may eventually lead to regeneration of specific tissues or even complete organs. Cell behaviors such as migration and proliferation are facilitated by communication of cells with the micro-environment, nearby cells and with the matrix itself. Thus, scaffolds may play an important role in improving some tissue engineering-based therapies.¹⁷ In one human model, human mesenchymal stem cells harvested from the bone marrow and seeded with bioceramics, facilitated bone regeneration after spinal fusion. This method may be an alternative to autologous and allogenic bone fusion.¹⁸

In a human series with SCI (2 male and 2 female) concentrated cellular bone marrow was implanted in an intraspinal lesion at the time of surgery. After 1 year of follow up, three showed improvement. No complication was seen as a result of intraspinal bone marrow cell injection. Considering these results, the potential role of regenerative medicine in the treatment of CNS lesions is no longer speculation.¹⁹

Peripheral Nervous system

The management of injury-associated peripheral nerve defects is difficult because of the absence of suitable autologous nerves for transplantation.²⁰ One

study has found that biomaterial gel made from proteins found in human hair promotes nerve regeneration through activation of Schwann cells. This study and others like it have demonstrated nerve regeneration results comparable to autograft nerve repair.²⁰ In another similar study, the use of keratin gel was a viable guiding material for Schwann cell and axon migration and proliferation, functions that are essential for nerve repair.²¹ It has also been shown that Spidrex conduits, a silk-based biomaterial, promote axonal regeneration and functional recovery, and may be useful for clinical application.²²

As in the CNS, the use of stem cells in both animal models²³ and human models²⁴ of peripheral nerve injury has been reported. After 60 days of follow-up umbilical cord mesenchymal stem cells injected intravenously in a patient with both nonunion and nerve injury resulted in improved nerve reflex, muscle tone and muscle strength. In addition, nerve conduction velocity increased and the fracture gap disappeared.²⁴ These advances may open up exciting new avenues of treatment for patients with peripheral nervous system injury.

Musculoskeletal system

Autologous bone grafts are the gold standard for reconstruction of large bone defects. However, surgical stress to the portion of normal bone being extracted and the amount of extractable bone present limit this method.²⁵ As in other organs, stem cell based therapies have the potential to improve the treatment of osseous defects. Osteoblasts derived from induced adipose-derived stromal cells from rabbits were seeded on chitosan-tricalcium-phosphate-gelatin (CS-TCP-Gel) and chitosan (Cs) scaffolds, that reconstructing reinforced and conventional periosteum respectively. These materials were transplanted into long bone defects in rabbits. The findings of this study showed that reinforced periosteum performs better than conventional periosteum and is more suitable for repair of weight-bearing bones.²⁶ A study by Levi et al. has shown that human adipose-derived stromal stem cells (ASCs) can heal acute calvarial defects in mice, but cannot repair chronic defects. This difference has been attributed to endogenous bone morphogenetic protein (BMP) activation after trauma.²⁷ Thus, local delivery of BMP may enable regeneration of large bone defects without bone grafting.²⁸ Also, short poly-N-acetyl glucosamine (sNAG) fibers have been shown to activate bone regeneration in rabbit femurs.²⁹

Combinations of biological substance may be useful for treatment of overuse injuries. Administration of autologous platelet-rich plasma (PRP) and isolated bone marrow mononucleated cells (BMMNCs) in competition horses showed significant improvement of lameness and

return to competitive status. This suggests that similar application of appropriate cells and growth factors (GFs) may be useful for treatment of human athletes.³⁰ Combinations may have other applications as well. Simultaneous sustained release of an antibiotic and a human growth factor is one viable option for decreasing infection rates and improving vascularization and healing of bone grafts in contaminated open fractures.³¹ Cryptic peptide, derived from the C-terminal telopeptide of collagen 3, is another biological substance shown to have implications in regenerative therapies. In a murine model of limb amputation, cryptic peptide caused bone nodule formation at the site of amputation. In addition, cryptic peptide has the ability to alter stem cell recruitment and differentiation at the injury site, and may serve as a new method for influencing stem cells in that local environment.³² In one human case report, decellularized bovine trabecular bone was seeded with autologous bone marrow cells, cultured for three weeks, and subsequently implanted into 72mm distal tibial defect and fixed with an intramedullary nail. After six weeks, bone formation was seen around the graft, and the patient recovered the ability to walk. At two years no complications have occurred. This graft represents a useful alternative for long bone defect repair.³³

Repair of articular cartilage is difficult, as it is both avascular and aneural.³⁴ Recently tissue engineering of cartilage has entered into clinical practice.³⁵ Autologous chondrocytes were recently implanted into articular cartilage defects in multiple human series. After long-term follow-up, results were excellent with patients returning to normal activity levels.^{36,37,38} Hyaluronan-based scaffolds seeded with autologous chondrocytes have been implanted in the knees of 53 patients with up to 7 years follow up. Hyalograft C autografts resulted in clinical improvement in young healthy patients with single cartilage defects, but results were poor in patients with osteoarthritis.³⁹ Autologous mesenchymal stem cells harvested from the iliac crest and cultured with growth factors have also been implanted into an adult human knee, resulting in an increase in meniscal cartilage volume.⁴⁰

Damaged tendons rarely heal completely. Recently, regenerative medicine has made complete repair possible by means of an extracellular matrix derived from porcine small intestinal submucosa. This technique may aid in the organization of host tissue⁴¹ and allow tendon regeneration with stem cells.^{42,43} Three hundred British orthopedic surgeons specializing in knee surgery were invited to participate in an online questionnaire about tissue engineering of the anterior cruciate liga-

ment. Overall, most surgeons would be prepared to use this technique if it were an improvement over the current technique.⁴⁴

Skin

Regenerative science has rapidly improved the clinical management of wounds. Currently there are more than 20 skin substitutes commercially available. These may contain allogenic or autogenic living cells or no living cells.⁴⁵ The first product commercially available was an autologous epidermal substitute.⁴⁶ Later, Kuroyanagi developed an allogenic cultured dermal substitute. This substitute consists of a 2-layered spongy matrix of hyaluronic acid and atelocollagen containing fibroblasts. The skin substitute releases numerous substances necessary for wound healing.⁴⁶ Platelet-derived growth factor is one substance necessary for wound repair, participating in chemotaxis, cell proliferation, etc.^{47,48} Topical therapy with platelet gel and PG-derived GF also has a potential role in wound healing.⁴⁸ Tissue engineered skin has now been approved by the United States Food and Drug Administration (FDA). In some human reports, tissue engineered skin was used to treat dermal atrophy secondary to traumatic avulsion^{49,50} and for the treatment of wounds in the premature neonate in order to prevent hypertonic wound extravasations.⁵¹

Use of stem cells for the treatment of skin damage has been reported. Adipose-derived stem cells^{52,53} and bone marrow-derived stem cells⁵⁴ with or without dermal substitute were used in animal and human model with positive results. Adipose-derived stem cells exhibit antioxidant effects, increase vascularity and collagen synthesis and improve skin regeneration.^{52,53} These cells can usually still be found in deep layers of skin wounds in patients with major traumatic injuries, such as extensive burns. Thus, adipose-derived stem cells may be obtained from discarded skin from burn victims.⁵⁵ Application of stem cells for the management of radiation burns has been reported in animal models via systemic transplantation⁵⁶ and local cell therapy in human cases.^{57,58} Such therapies may provide new hope in the treatment of radiation burns and radiotherapy complications.

Delayed wound healing (two weeks after the injury) is usually associated with poor aesthetics and impaired function. In order to eliminate the need for cell culture before treatment, an innovative cell isolation technique has been used in which a biopsy of healthy skin is exposed to two enzymes and the desired cells transferred to ringer's lactate solution. After wound debridement, the cells are immediately transplanted with a spraying mechanism.^{59,60}

Another novel method for improving wound healing is the application of an active dressing that supplies cell support and provokes regeneration by wound irrigation.⁶⁰

Respiratory Tract

Lung injuries can cause significant mortality and morbidity.⁶¹ There are several sources of adult stem cells in respiratory tract, which contribute to repair of the respiratory tract lining. To date, great progress has been made in the field of tracheal bioengineering. One method employed biodegradable three-dimensional scaffolds for cell attachment, cell differentiation and extracellular matrix production and finally whole tracheal regeneration. Using this method the world's first bioengineered trachea was produced.⁶²

Genitourinary system

Traumatic events can lead to organ damage or even loss within the genitourinary tract. Several regenerative medicine strategies for replacing these organs are currently under investigation.⁶³ Renal tissue is one the most difficult to regenerate and most efforts for kidney regeneration have focused on cell therapy.³ In one study, culture of primary renal cells in three-dimensional collagen-based scaffolds promoted regeneration of tubule- and glomeruluslike structures that had positive staining for Tamm-Horsfall protein.⁶⁴ Unfortunately, due to the complexity of both the anatomy and function of the kidney, regenerative medicine does not yet play a prominent role in the treatment of patients with traumatic renal injuries.

Regeneration of the ureter has been performed in animal models at several centers. Use of small-intestinal sub-mucosal as a ureteral replacement material⁶⁵ and transplantation of urothelial and smooth cells on tubular polymer scaffolds⁶⁶ have both been done in canine models with good results. However, the relatively small incidence of ureteral injury and the large expense involved in ureteral regeneration currently limit the translation of these techniques to humans.³

A number of animal studies and human clinical experiences have shown the value of bladder tissue engineering.⁶⁷ Urothelial progenitor cells can be harvested from the bladder, especially the bladder neck and trigone areas.³ Additionally, amniotic-derived, bone marrow-derived and urine-derived stem cells have the ability to differentiate into bladder tissue.^{3,68} Still, the use of biomaterial-based scaffolds seeded with autologous urothelial and smooth muscle cells is the most efficacious method for engineering bladder tissue.⁶⁷ Using this method Atala et al. constructed neobladders

for seven patients with myelomeningocele. After 22-61 months of follow-up biopsies of the neobladders showed normal structure and phenotype.⁶⁹

Various methods have been proposed for urethral regeneration.³ Acellular collagen matrices harvested from bladder submucosa have been used experimentally and clinically for onlay urethral substitution with good results.^{70,71} More than 200 patients with urethral stricture have been successfully treated with this technique, eliminating the need for an autologous graft.³ Histological examination of the implanted matrices showed typical urethral stratified epithelium.⁷¹ Acellular collagen matrices have had poor results in patients needing tubularized graft, and in these patients collagen matrices seeded with cells demonstrated improved performance.⁷² In one observational study five boys with urethral stricture underwent urethral biopsies, and muscle and epithelial cells were isolated and seeded onto tubularized synthetic scaffolds. The tissue-engineered tubularized urethra was then implanted into the patients. After 71 months median follow up, the urethras had normal function and anatomy. This technique represents a valid option in patients requiring difficult urethral reconstruction.⁷³

Several studies have shown the feasibility of tissue engineering of the corpora cavernosa. Smooth muscle and endothelial cells seeded on collagen matrices have created a corpora cavernosa-like tissue structure in a rat model.⁷⁴ Also, acellular collagen matrices derived from rabbit corpora and a biodegradable polymer scaffold were seeded with human corpus cavernosa muscle and endothelial cells. Using these scaffolds, Falke et al were able to form corporal tissue in vivo.^{75,76} Recently, neocorpora were engineered for total penile corporal body replacement in a rabbit model. The bioengineered corpora were similar to the native tissue structurally and functionally.⁷⁷ These results are promising for future patients with penile trauma.

Vaginal epithelial smooth muscle cells can be cultured in vitro⁷⁸ and with cell seeded scaffolds, it is possible to engineer a vagina for total vaginal replacement.^{79,80}

Other Organs

Stem cells play an important role in retinal regeneration. Studies have shown that stem cells have the potential to create a functional retina and form neural circuitry sufficient for vision.^{81,82} In one study ocular limbal cells cultured with keratinocyte stem cells from the corneal epithelium were grafted in 116 patients suffering from chemical destruction of limbus.⁸³ The results suggest a promising potential for the restoration of vision in such patients. Regeneration of the orbital wall with autologous bone marrow stromal cells has also been demonstrated in a canine model.⁸⁴ Regeneration of auricular cartilage, oral mucosa, tooth and periodontal tissue, and vocalcords are under investigation in several studies.^{85,86,87,88,89}

Conclusion

Trauma is an important health issue, particularly for young people needing replacement tissues or organs. Regenerative medicine has an essential role in the treatment of these patients. As discussed above, various tissues and organs are in different stages of investigation, with some already in clinical use and others in preclinical trials and experimental studies. As more products pass the FDA approval process and attain clinical applicability, regenerative medicine will hopefully be a valuable option for traumatic patients who need organ replacement and repair.

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References

1. Cothren CC, Biffl WL, Moore EE. Trauma. In: Brunicaudi FC, Andersen DK, Billiar TR, Dunn DL, Hunter JG, Matthews JB (eds): Schwartz 's Principle of Surgery, Ninth Edition. New York: McGraw-Hill, 2010.
2. Dean W. The Armed Forces Institute of Regenerative Medicine: a collaborative approach to Department of Defense-relevant research. *Regen Med.* 2011 Nov;6(6 Suppl):71-4.
3. Atala A. Regenerative Medicine in Urology: Stem Cells, Tissue Engineering, and Cloning. In: Wein AJ, Kavoussi LR, Novick AC, Partin AW, Peters CA (eds): Campbell-Walsh Urology, Tenth Edition. Philadelphia, PA: Saunders Elsevier, 2011.
4. Oslon JL, Atala A, Yoo JJ. Tissue engineering: current strategies and future directions. *Chonnam Med J.* 2011 Apr;47(1):1-13.
5. Okano H, Yoshizaki T, Okada S. Regeneration of central nervous system: its concept and strategy. *Rinsho Shinkeigaku.* 2003 Nov;43(11):824-6.
6. Webber DJ, Bradbury EJ, McMahon SB, Minger SL. Transplanted neural progenitor cells survive and differentiate but achieve limited functional recovery in the lesioned adult rat spinal cord. *Regen Med.* 2007 Nov;2(6):929-45

7. Park DH, Lee JH, Borlongan CV, Sanberg PR, Chung YG, Cho TH. Transplantation of umbilical cord blood stem cells for treating spinal cord injury. *Stem Cell Rev.* 2011 Mar;7(1):181-94.
8. Noble M, Davies JE, Mayer-Pröschel M, Pröschel C, Davies SJ. Precursor cell biology and the development of astrocyte transplantation therapies: lessons from spinal cord injury. *Neurotherapeutics.* 2011 Oct;8(4):677-93.
9. Bartolomei JC, Greer CA. Olfactory ensheathing cells: bridging the gap in spinal cord injury. *Neurosurgery.* 2000 Nov;47(5):1057-69.
10. Guzen FP, de Almeida Leme RJ, de Andrade MS, de Luca BA, Chadi G. Glial cell line-derived neurotrophic factor added to a sciatic nerve fragment grafted in a spinal cord gap ameliorates motor impairments in rats and increases local axonal growth. *Restor Neurol Neurosci.* 2009;27(1):1-16.
11. Orlacchio A, Bernardi G, Orlacchio A, Martino S. Stem cells: an overview of the current status of therapies for central and peripheral nervous system diseases. *Curr Med Chem.* 2010;17(7):595-608.
12. Nori S, Tsuji O, Okada Y, Toyama Y, Okano H, Nakamura M. Therapeutic potential of induced pluripotent stem cells for spinal cord injury. *Brain Nerve.* 2012 Jan;64(1):17-27.
13. Levi B, James AW, Nelson ER, Hu S, Sun N, Peng M, et al. Studies in adipose-derived stromal cells: migration and participation in repair of cranial injury after systemic injection. *Plast Reconstr Surg.* 2011 Mar;127(3):1130-40.
14. Fujimoto Y, Abematsu M, Falk A, Tsujimura K, Sanosaka T, Juliandi B, et al. Treatment of a mouse model of spinal cord injury by transplantation of human induced pluripotent stem cell -derived long-term self-renewing neuroepithelial-like stem cells. *Stem Cells.* 2012 Jun;30(6):1163-73.
15. Ban DX, Ning GZ, Feng SQ, Wang Y, Zhou XH, Liu Y, et al. Combination of activated Schwann cells with bone mesenchymal stem cells: the best cell strategy for repair after spinal cord injury in rats. *Regen Med.* 2011 Nov;6(6):707-20.
16. Zurita M, Vaquero J, Oya S. Grafting of neural tissue in chronically injured spinal cord: influence of the donor tissue on regenerative activity. *Surg Neurol.* 2000 Aug;54(2):117-25.
17. Tabesh H, Amaabediny G, Nik NS, Heydari M, Yosefifard M, Siadat SO, et al. The role of biodegradable engineered scaffolds seeded with Schwann cells for spinal cord regeneration. *Neurochem Int.* 2009 Feb;54(2):73-83.
18. Barbanti Brodano G, Mazzoni E, Tognon M, Griffoni C, Manfrini M. Human mesenchymal stem cells and biomaterials interaction: a promising synergy to improve spine fusion. *Eur Spine J.* 2012 May;21 Suppl 1:S3-9.
19. Attar A, Aytan M, Ozdemir M, Ozgencil E, Bozkurt M, Kaptanoglu E, et al. An attempt to treat patients who have injured spinal cords with intraleisional implantation of concentrated autologous bone marrow cells. *Cytherapy.* 2011 Jan;13(1):54-60.
20. Sierpinski P, Garrett J, Ma J, Apel P, Klorig D, Smith T, et al. The use of keratin biomaterials derived from human hair for the promotion of rapid regeneration of peripheral nerves. *Biomaterials.* 2008 Jan;29(1):118-28.
21. Lin YC, Ramadan M, Van Dyke M, Kokai LE, Philips BJ, Rubin JP, et al. Keratin gel filler for peripheral nerve repair in a rodent sciatic nerve injury model. *Plast Reconstr Surg.* 2012 Jan;129(1):67-78.
22. Huang W, Begum R, Barber T, Ibba V, Tee NC, Hussain M, et al. Regenerative potential of silk conduits in repair of peripheral nerve injury in adult rats. *Biomaterials.* 2012 Jan;33(1):59-71.
23. Walsh SK, Kumar R, Grochmal JK, Kemp SW, Forden J, Midha R. Fate of stem cell transplants in peripheral nerves. *Stem Cell Res.* 2012 Mar;8(2):226-38.
24. Xue G, He M, Zhao J, Chen Y, Tian Y, Zhao B, et al. Intravenous umbilical cord mesenchymal stem cell infusion for the treatment of combined malnutrition nonunion of the humerus and radial nerve injury. *Regen Med.* 2011 Nov;6(6):733-41.
25. Jimi E, Hirata S, Osawa K, Terashita M, Kitamura C, Fukushima H. The current and future therapies of bone regeneration to repair bone defects. *Int J Dent.* 2012;2012:148261.
26. Guo H, Li X, Yuan X, Ma X. Reconstruction of radial bone defects using the reinforced tissue-engineered periosteum: an experimental study on rabbit weightbearing segment. *J Trauma Acute Care Surg.* 2012 Feb;72(2):E94-100.
27. Levi B, James AW, Nelson ER, Peng M, Wan DC, Commons GW, et al. Acute skeletal injury is necessary for human adipose-derived stromal cell-mediated calvarial regeneration. *Plast Reconstr Surg.* 2011 Mar;127(3):1118-29.
28. Yano K, Namikawa T, Uemura T, Hoshino M, Wakitani S, Takaoka K et al. Regenerative repair of bone defects with osteoinductive hydroxyapatite fabricated to match the defect and implanted with combined use of computer-aided design, computer-aided manufacturing, and computer-assisted surgery systems: a feasibility study in a canine model. *J Orthop Sci.* 2012 Jul;17(4):484-9
29. Muise-Helmericks RC, Demcheva M, Vournakis JN, Seth A. Poly-N-acetyl glucosamine fibers activate bone regeneration in a rabbit femur injury model. *J Trauma.* 2011 Aug;71 (2 Suppl 1):S194-6.
30. Torricelli P, Fini M, Filardo G, Tschon M, Pischedda M, Pacorini A et al. Regenerative medicine for the treatment of musculoskeletal overuse injuries in competition horses. *Int Orthop.* 2011 Oct;35(10):1569-76.
31. Wenke JC, Guelcher SA. Dual delivery of an antibiotic and a growth factor addresses both the microbiological and biological challenges of contaminated bone fractures. *Expert Opin Drug Deliv.* 2011 Dec;8(12):1555-69.
32. Agrawal V, Kelly J, Tottey S, Daly KA, Johnson SA, Siu BF, et al. An isolated cryptic peptide influences osteogenesis and bone remodeling in an adult mammalian model of digit amputation. *Tissue Eng Part A.* 2011 Dec;17(23-24):3033-44.

33. Hesse E, Kluge G, Atfi A, Correa D, Haasper C, Berding G, et al. Repair of a segmental long bone defect in human by implantation of a novel multiple disc graft. *Bone*. 2010 May;46(5):1457-63.
34. Nelson L, Fairclough J, Archer CW. Use of stem cells in the biological repair of articular cartilage. *Expert Opin Biol Ther*. 2010 Jan;10(1):43-55.
35. Ochi M, Adachi N, Nobuto H, Yanada S, Ito Y, Agung M. Articular cartilage repair using tissue engineering technique--novel approach with minimally invasive procedure. *Artif Organs*. 2004 Jan;28(1):28-32.
36. Saris DB, Vanlauwe J, Victor J, Almqvist KF, Verdonk R, Bellemans J, et al. Treatment of symptomatic cartilage defects of the knee: characterized chondrocyte implantation results in better clinical outcome at 36 months in a randomized trial compared to microfracture. *Am J Sports Med*. 2009 Nov;37 Suppl 1:10S-19S.
37. Kasemkijwattana C, Kesprayura S, Chaipinyo K, Chanlalit C, Chansiri K. Autologous chondrocytes implantation for traumatic cartilage defects of the knee. *J Med Assoc Thai*. 2009 May;92(5):648-53.
38. Vasiliadis HS, Danielson B, Ljungberg M, McKeon B, Lindahl A, Peterson L. Autologous chondrocyte implantation in cartilage lesions of the knee: long-term evaluation with magnetic resonance imaging and delayed gadolinium-enhanced magnetic resonance imaging technique. *Am J Sports Med*. 2010 May;38(5):943-9.
39. Nehrer S, Dorotka R, Domayer S, Stelzeneder D, Kotz R. Treatment of full-thickness chondral defects with hyalograft C in the knee: a prospective clinical case series with 2 to 7 years' follow-up. *Am J Sports Med*. 2009 Nov;37 Suppl 1:81S-87S.
40. Centeno CJ, Busse D, Kisiday J, Keohan C, Freeman M, Karli D. Regeneration of meniscus cartilage in a knee treated with percutaneously implanted autologous mesenchymal stem cells. *Med Hypotheses*. 2008 Dec;71(6):900-8.
41. Gilbert TW, Stewart-Akers AM, Simmons-Byrd A, Badylak SF. Degradation and remodeling of small intestinal submucosa in canine Achilles tendon repair. *J Bone Joint Surg Am*. 2007;89:621-30.
42. Uysal AC, Mizuno H. Tendon regeneration and repair with adipose derived stem cells. *Curr Stem Cell Res Ther*. 2010 Jun;5(2):161-7.
43. Cohen S, Leshansky L, Zussman E, Burman M, Srouji S, Livne E, et al. Repair of full-thickness tendon injury using connective tissue progenitors efficiently derived from human embryonic stem cells and fetal tissues. *Tissue Eng Part A*. 2010 Oct;16(10):3119-37.
44. Rathbone S, Maffulli N, Cartmell SH. Most British surgeons would consider using a tissue-engineered anterior cruciate ligament: a questionnaire study. *Stem Cells Int*. 2012;2012:303724.
45. Auger FA, Lacroix D, Germain L. Skin substitutes and wound healing. *Skin Pharmacol Physiol*. 2009;22(2):94-102.
46. Kuroyanagi Y. Regenerative medicine for skin. *Nihon Ronen Igakkai Zasshi*. 2006 May;43(3):326-9.
47. Sánchez-González DJ, Méndez-Bolaina E, Trejo-Bahena NI. Platelet-rich plasma peptides: key for regeneration. *Int J Pept*. 2012;2012:532519.
48. Rozman P, Bolta Z. Use of platelet growth factors in treating wounds and soft-tissue injuries. *Acta Dermatovenerol Alp Panonica Adriat*. 2007 Dec;16(4):156-65.
49. Maier JP, Lippitt C, Mooney EK. Use of tissue-engineered skin in the dermal atrophy patient with traumatic avulsion injuries. *Ann Plast Surg*. 2002 Jul;49(1):67-72.
50. Kuroyanagi Y, Yamada N, Yamashita R, Uchinuma E. Tissue-engineered product: allogeneic cultured dermal substitute composed of spongy collagen with fibroblasts. *Artif Organs*. 2001 Mar;25(3):180-6.
51. Onesti MG, Carella S, Maruccia M, Marchese C, Fino P, Scuderi N. A successful combined treatment with dermal substitutes and products of regenerative medicine in a patient affected by extravasation injury from hypertonic solution. *In Vivo*. 2012 Jan-Feb;26(1):139-42.
52. Kim WS, Park BS, Sung JH. The wound-healing and antioxidant effects of adipose-derived stem cells. *Expert Opin Biol Ther*. 2009 Jul;9(7):879-87.
53. Meruane MA, Rojas M, Marcelain K. The use of adipose tissue-derived stem cells within a dermal substitute improves skin regeneration by increasing neoangiogenesis and collagen synthesis. *Plast Reconstr Surg*. 2012 Jul;130(1):53-63.
54. Yoshikawa T, Mitsuno H, Nonaka I, Sen Y, Kawanishi K, Inada Y, et al. Wound therapy by marrow mesenchymal cell transplantation. *Plast Reconstr Surg*. 2008 Mar;121(3):860-77.
55. Natesan S, Wrice NL, Baer DG, Christy RJ. Debrided skin as a source of autologous stem cells for wound repair. *Stem Cells*. 2011 Aug;29(8):1219-30.
56. Shi C, Cheng T, Su Y, Mai Y, Qu J, Lou S, et al. Transplantation of dermal multipotent cells promotes survival and wound healing in rats with combined radiation and wound injury. *Radiat Res*. 2004 Jul;162(1):56-63.
57. Lataillade JJ, Doucet C, Bey E, Carsin H, Huet C, Clairand I, et al. New approach to radiation burn treatment by dosimetry-guided surgery combined with autologous mesenchymal stem cell therapy. *Regen Med*. 2007 Sep;2(5):785-94.
58. Bey E, Duhamel P, Lataillade JJ, de Revel T, Carsin H, Gourmelon P. Treatment of radiation burns with surgery and cell therapy. A report of two cases. *Bull Acad Natl Med*. 2007 Jun;191(6):971-8.
59. Gerlach JC, Johnen C, Ottoman C, Bräutigam K, Plettig J, Belfekroun C, et al. Method for autologous single skin cell isolation for regenerative cell spray transplantation with non-cultured cells. *Int J Artif Organs*. 2011 Mar;34(3):271-9.
60. Plettig J, Johnen CM, Bräutigam K, Zeilinger K, Borneman R, Gerlach JC. Active wound dressing with artificial capillaries for temporary wound irrigation and skin cell supply. *Artif Organs*. 2012 Apr;36(4):446-9.

61. Siniscalco D, Sullo N, Maione S, Rossi F, D'Agostino B. Stem cell therapy: the great promise in lung disease. *Ther Adv Respir Dis*. 2008 Jun;2(3):173-7.
62. Chistiakov DA. Endogenous and exogenous stem cells: a role in lung repair and use in airway tissue engineering and transplantation. *J Biomed Sci*. 2010 Dec 7;17:92.
63. Atala A, Koh C. Applications of tissue engineering in the genitourinary tract. *Expert Rev Med Devices*. 2005 Jan;2(1):19-26.
64. Joraku A, Stem KA, Atala A, Yoo JJ. In vitro generation of three-dimensional renal structures. *Methods*. 2009 Feb;47(2):129-33.
65. Jaffe JS, Ginsberg PC, Yanoshak SJ, Costa LE Jr, Ogbolu FN, Moyer CP, et al. Ureteral segment replacement using a circumferential small-intestinal submucosa xenogenic graft. *J Invest Surg*. 2001 Sep-Oct;14(5):259-65.
66. Yoo JJ, Satar N, Retik AB, Atala A. Ureteral replacement using biodegradable polymer scaffolds seeded with urothelial and smooth muscle cells. *J Urol*. 1995;153:375A.
67. Atala A. Tissue engineering of human bladder. *Br Med Bull*. 2011;97:81-104.
68. Bharadwaj S, Liu G, Shi Y, Markert C, Andersson KE, Atala A, et al. Characterization of urine-derived stem cells obtained from upper urinary tract for use in cell-based urological tissue engineering. *Tissue Eng Part A*. 2011 Aug;17(15-16):2123-32.
69. Atala A, Bauer SB, Soker S, Yoo J, Retik AB. Tissue-engineered autologous bladders for patients needing cystoplasty. *Lancet*. 2006 Apr 15;367(9518):1241-6.
70. Forraz N, Wright KE, Jurga M, McGuckin CP. Experimental therapies for repair of the central nervous system: stem cells and tissue engineering. *J Tissue Eng Regen Med*. 2013 Jul;7(7):523-36.
71. El-Kassaby AW, Retik AB, Yoo JJ, Atala A. Urethral stricture repair with an off-the-shelf collagen matrix. *J Urol*. 2003 Jan;169(1):170-3.
72. De Filippo RE, Yoo JJ, Atala A. Urethral replacement using cell seeded tabularized collagen matrices. *J Urol*. 2002 Oct;168(4Pt 2):1789-92.
73. Raya-Rivera A, Esquiliano DR, Yoo JJ, Lopez-Bayghan E, Soker S, Atala A. Tissue-engineered autologous urethras for patients who need reconstruction: an observational study. *Lancet*. 2011 Apr 2;377(9772):1175-82.
74. Kwon TG, Yoo JJ, Atala A. Autologous penile corpora cavernosa replacement using tissue engineering technique. *J Urol*. 2002 Oct;168(4Pt2):1754-8.
75. Falke G, Yoo JJ, Kwon TG, Moreland R, Atala A. Formation of corporal tissue architecture in vivo using human cavernosal muscle and endothelial cells seeded on collagen matrices. *Tissue Eng*. 2003 Oct;9(5):871-9.
76. Kershan RT, Yoo JJ, Moreland RB, Krane RJ, Atala A. Reconstitution of human corpus cavernosum smooth muscle in vitro and in vivo. *Tissue Eng*. 2002 Jul;8(3):515-24.
77. Chen KL, Eberli D, Yoo JJ, Atala A. Bioengineered corporal tissue for structural and functional restoration of the penis. *Proc Natl Acad Sci USA*. 2010 Feb 23;107(8):3346-50.
78. De Filippo RE, Yoo JJ, Atala A. Engineering of vaginal tissue in vivo. *Tissue Eng*. 2003 Apr;9(2):301-6.
79. Dorin RP, Atala A, De Filippo RE. Bioengineering a vaginal replacement using a small biopsy of autologous tissue. *Semin Reprod Med*. 2011 Jan;29(1):38-44.
80. De Filippo RE, Bishop CE, Filho LF, Yoo JJ, Atala A. Tissue engineering a complete vaginal replacement from a small biopsy of autologous tissue. *Transplantation*. 2008 Jul;86(2):208-14.
81. Enzmann V, Yolcu E, Kaplan HJ, Ildstad ST. Stem cells as tool in regenerative therapy for retinal degeneration. *Arch Ophthalmol*. 2009 Apr;127(4):563-71.
82. Viczian AS, Solessio EC, Lyou Y, Zuber ME. Generation of functional eyes from pluripotent cells. *PLoS Biol*. 2009 Aug;7(8):e1000174.
83. De Luca M, Pellegrini G, Green H. Regeneration of squamous epithelia from stem cells of cultured grafts. *Regen Med*. 2006 Jan;1(1):45-57.
84. Xiao C, Zhou H, Ge S, Tang T, Hou H, Luo M, et al. Repair of orbital wall defects using biocoral scaffolds combined with bone marrow stem cells enhanced by human bone morphogenetic protein-2 in a canine model. *Int J Mol Med*. 2010 Oct;26(4):517-25.
85. Nayyer L, Patel KH, Esmaili A, Rippel RA, Birchall M, O' toole G, et al. Tissue engineering: revolution and challenge in auricular cartilage reconstruction. *Plast Reconstr Surg*. 2012 May;129(5):1123-37.
86. Kiyokawa K, Kiyokawa M, Takagi M, Rikimaru H, Fukaya T. New regenerative treatment for tooth and periodontal bone defect associated with posttraumatic alveolar bone crush fracture. *J Craniofac Surg*. 2009 May;20(3):780-3.
87. Mehra P, Miner J, D'Innocenzo R, Nadershah M. Use of 3-d stereolithographic models in oral and maxillofacial surgery. *J Maxillofac Oral Surg*. 2011 Mar;10(1):6-13.
88. Li H, Zhang D, Wen Y, Wang C. Growth of compound layer tissue engineered oral mucosa and its clinical application in hetero-transplantation. *Zhongguo Xue FU Chong Jian Wai Ke Za Zhi*. 2006 Feb;20(2):177-80.
89. Kumai Y, Kobler JB, Herrera VL, Zeitels SM. Perspectives on adipose-derived stem/stromal cells as potential treatment for scarred vocal folds: opportunity and challenges. *Curr Stem Cell Res Ther*. 2010 Jun;5(2):175-81.