

Comparison of Lethal Concentrations (LC_{50-96 H}) of CdCl₂, CrCl₃, and Pb (NO₃)₂ in Common Carp (*Cyprinus carpio*) and Sutchi Catfish (*Pangasius Hypophthalmus*)

Zeynab Abedi¹, Mohammadkazem Khalesi^{2*}, Sohrab Kohestan Eskandari², Hossein Rahmani²

Received: 10.06.2012

Accepted: 15.07.2012

ABSTRACT

Background: The present study compared lethal concentrations (LC_{50-96 h}) of CdCl₂, CrCl₃, and Pb (NO₃)₂ between two scaled and scaleless freshwater fish species: *Cyprinus carpio* (Cyprinidae) and *Pangasius hypophthalmus* (Pangasiidae).

Methods: The experimental fishes were obtained from fish markets/ponds with average lengths and weights of approximately 11.68 ± 1.92 and 9.8 ± 1.9 cm, and 25.92 ± 6.3 and 18.61 ± 3.22 g for *C. carpio* and *P. hypophthalmus*, respectively. The fishes were exposed to different concentrations of cadmium (Cd) (0, 10, 20, 40, 80, 100, 120, 200 mg L⁻¹) lead (Pb) (0, 20, 40, 50, 60, 90, 150 mg L⁻¹), and chromium (Cr) (0, 5, 10, 20, 30, 40 mg L⁻¹) for 96 h. Physicochemical parameters, such as dissolved oxygen, pH, and temperature of aquaria as well as mortality rate of the fishes, were monitored daily.

Results: The 50% lethal concentrations (LC_{50-96 h}) of CdCl₂, CrCl₃, and Pb (NO₃)₂ for *P. hypophthalmus* were found at 64.89, 7.46, and 48.06 mg L⁻¹, and those of CdCl₂, CrCl₃, Pb (NO₃)₂ for *C. carpio* were detected at 84.8, 17.05, and 77.33 mg L⁻¹. The ratios of heavy metal toxicity factors (TF) were greater for common carp compared to those for the catfish.

Conclusion: Our preliminary findings suggest that common carp *C. carpio* with higher LC₅₀ (and LC₁₀₀) values appears to be more tolerant to heavy metals exposure than the catfish (*P. hypophthalmus*). This may be due to the resistance to the heavy metals through protection from the carp's scaled body versus scaleless body of the catfish.

Keywords: Cadmium, Chromium, Lead, Fish Scales, 50% Lethality.

IJT 2012; 672-680

INTRODUCTION

Heavy metals pollution of the aquatic environment has been marked as a serious health concern, which its introduction into these ecosystems happens through various routes such as industrial effluents and wastes, agricultural pesticide run-off, domestic garbage dumps, and mining activities (1). A potential threat for aquatic

organisms is contamination arisen from being exposed to significant amounts of heavy metals, which at high concentrations can cause harmful effects on metabolic, physiological, and biochemical systems of fishes (2-5) together with long-term ecotoxicological effects (6). Thus, these metals are a matter of environmental and health concern because of their toxic

1. Student of M.Sc in Fishery, Faculty of Animal Sciences and Fisheries, Sari Agricultural Sciences and Natural Resources University (SANRU), Sari, Iran.

2. Department of Fisheries, Sari Agricultural Sciences and Natural Resources University (SANRU), Sari, Iran.

* Correspondence Author E-mail: khalesimk@iran.ir

potential to be accumulated in food chain (7).

Toxicity testing has been widely used as a tool to identify suitable organisms as a bio-indicator and to derive water quality standards for chemicals. It is also considered an essential tool for assessing the effects and fate of toxicants in aquatic ecosystems (8). Because toxicity studies quantify an organism's response to a biologically active material (9) and are useful in determining water quality, it is, therefore, crucial to restore and resolve metal pollution through environmental monitoring. Fish absorb dissolved or available metals and can, therefore, serve as a reliable indication of metal pollution in an aquatic ecosystem (10).

Common carp (*Cyprinus carpio*) accounts for an important farmed species. Hence the optimum culture conditions and caveat on severe aftermath of environmental pollutants including such heavy metals as Cd, Cr, and Pb can effectively help improve cultivation of this valuable species. The scaleless fish, *Pangasius hypophthalmus* (sutchi or striped catfish), is native to the Mekong River drainage, which emerged as a major species for aquaculture purposes research focus, outside of tropical regions of South East Asia, and can be successfully cultured in the western tropics. Briefly, both species are of aquaculture interest, which have achieved commodity status on world seafood markets. Furthermore, development of this catfish culture industry has faced difficulties partly related to the limited knowledge of biology, ecology, and physiology reported in some cultivated stocks (11,12).

Both lethal and sub-lethal concentrations of heavy metals determine the sensitivity of individual organisms across species. Such changes differ from metal to metal, from species to species, and from one experimental condition to another. The exact causes of death due to metal poisoning are multiple and depend on time-concentration (13). The

susceptibility of fish to a particular heavy metal is also a very important factor for LC₅₀ values (14). Conducting 96-h LC₅₀ tests, therefore, makes it possible to measure the susceptibility and survival potential of organisms to particular toxic heavy metals. Because the 96-h LC₅₀ values of fish vary from species to species and from metal to metal (15-17), it would be appealing to scrutinize variations in metal accumulation and toxicity in catfish with that in a species like carp from similar environment for various purposes (18). Therefore, the purpose of this study was to determine and compare the LC₅₀ of CdCl₂, CrCl₃, and Pb (NO₃)₂ in two scaled *C. carpio* and scaleless *P. hypophthalmus* fish species, and to compare the level of sensitivity in each species from exposure to different Cd, Cr, and Pb concentrations.

MATERIALS AND METHODS

P. hypophthalmus was purchased from local aquarium shops and *C. carpio* was obtained from Nasr Fish Culture pond (Sari, Iran) with approximate total length of 8.0-14.0 and 7.0-12 cm, respectively. The fishes were transferred to aquarium room located at SANRU. Prior to toxicity testing, the fishes were acclimatized for one week under laboratory conditions (25±1° C with 12h light: 12h dark). Water quality parameters (TDS = 600 mg L⁻¹, pH = 6.75, EC = 1 ds/m, DO = 5-8 mg L⁻¹) were measured during the experiment. The experimental aquaria were aerated through air stones. Cd, Cr, and Pb heavy metals in the forms of cadmium chloride (CdCl₂.H₂O, BDH), chromium chloride (CrCl₃.6 H₂O, APLICAM), and lead (II) nitrate (Pb (NO₃)₂, MERK) were used as toxicants.

Acute cadmium, lead, and chromium toxicity experiments were performed using different concentrations of Cd (0, 10, 20, 40, 80, 100, 120, and 200 mg L⁻¹), Pb (0, 20, 40, 50, 60, 90, 150 mg L⁻¹), and Cr (0, 5, 10, 20, 30, 40 mg L⁻¹) during 96 h. Metal solutions were prepared by diluting of a stock solution with well water. Each

concentration contained six fish with one replicate each. There was a simultaneous control group (n= 10; with no heavy metal additions) together with the heavy metal treatments, keeping all other conditions alike. The concentration of each heavy metal caused 50% mortality in fish for 96 h was taken as the LC₅₀ value, calculated by Finney's Probit Analysis (in SPSS, V. 16). During the toxicity test, the fishes were not fed. The numbers of dead fish were counted daily and removed immediately from the aquaria. The degree of fish susceptibility to each heavy metal was determined as toxicity factor (TF). This factor is calculated by the ratio of LC₅₀ of a metal in one fish species to that in the other, i.e.: LC₅₀ *C. carpio*/ LC₅₀ *P. hypophthalmus*, at different intervals tested. Then the LC₅₀ of the two species was statistically compared through linear regression and evaluated by one-sample t-test (e.g. 19). To do this, first, the TF ratios were obtained by estimating LC₅₀ of each examined metal for each species at different times. Next, one-sample t-test was employed using SPSS software.

RESULTS

The 50% lethal concentrations (LC₅₀-96h) of CdCl₂, CrCl₃, Pb(NO₃)₂ for *Pangasius hypophthalmus* were 64.89, 7.46, and 48.06 mg L⁻¹, and those for

Cyprinus carpio were 84.8, 17.05, and 77.33 mg L⁻¹, respectively.

Table 1 (A and B) shows lethal concentrations (LC₅₀- 96 h) of Cd, Cr, and Pb in common carp and sutchi (striped) catfish, respectively, at four durations. It is evident that the required concentrations of all the examined heavy metals to reach lethal doses are far greater in common carp compared with those required in the catfish.

Time-response mortality of the two fish species resulting from different concentrations of individual heavy metals (Cd, Cr, and Pb) are presented in Tables 2 to 4. The Cd-response of the sutchi catfish (*P. hypophthalmus*) began at 40 mg L⁻¹ (48 and 72 h), which displayed higher mortalities with rising levels of cadmium especially at 24 and 48 h. The common carp (*C. carpio*) initially responded to a Cd concentration of 80 mg L⁻¹ (96 h), which exhibited rather elevated mortalities with increasing levels of cadmium, especially at 120 and 200 mg L⁻¹. Both species were equal in zero mortality in the control groups.

Table 2 also indicates that LC₁₀₀ values for the common carp (*C. carpio*) and sutchi catfish (*P. hypophthalmus*) were at 120 and 100 mg L⁻¹ of Cd (after 48 h), respectively.

Table1. Lethal concentrations (LC₅₀-96 h, mg L⁻¹) of Cd, Cr, and Pb for common carp (*C. carpio*) (A) and sutchi catfish (*P. hypophthalmus*) (B)

A)				B)			
Duration (h)	Cd	Cr	Pb	Duration (h)	Cd	Cr	Pb
24	287.47	67.84	480.64	24	128.24	27.43	156.4
48	137.45	33.67	228.7	48	83.63	17.57	86.57
72	131.38	21.24	96.19	72	69.74	9.8	62.22
96	84.8	17.05	77.32	96	64.89	7.46	48.06

Table 2. Mortalities of *P. hypophthalmus* and *C. carpio* recorded at different concentrations (mg L⁻¹) of Cd during 96 h

Cd concentration (mg L ⁻¹)		Control	10	20	40	80	100	120	200
Duration (h)									
<i>P. hypophthalmus</i> (sutchi catfish)									
24		0	0	0	0	2	4	4	3
48		0	0	0	1	1	1	2	2
72		0	0	0	1	0	1	0	1
96		0	0	0	0	0	0	0	0
<i>C. carpio</i> (common carp)									
24		0	0	0	0	0	2	3	0
48		0	0	0	0	0	1	3	3
72		0	0	0	0	0	1	0	0
96		0	0	0	0	3	0	0	3

Table 3. Mortalities of *P. hypophthalmus* and *C. carpio* recorded at different concentrations (mg L⁻¹) of Cr during 96 h

Cr concentration (mg L ⁻¹)		Control	5	10	20	30	40
Duration (h)							
<i>P. hypophthalmus</i> (sutchi catfish)							
24		0	2	1	3	0	0
48		0	1	3	1	0	1
72		0	0	2	2	3	3
96		0	0	0	0	2	0
<i>C. carpio</i> (common carp)							
24		0	0	1	1	3	0
48		0	0	0	1	3	1
72		0	0	0	1	1	4
96		0	0	1	0	0	1

Table 4. Mortalities of *P. hypophthalmus* and *C. carpio* recorded at different concentrations (mg L⁻¹) of Pb during 96 h

Pb concentration (mg L ⁻¹)		Control	20	40	50	60	90	150
Duration (h)								
<i>P. hypophthalmus</i> (sutchi catfish)								
24		0	0	0	1	1	0	3
48		0	0	2	0	2	3	2
72		0	0	0	0	2	1	1
96		0	0	0	2	0	2	0
<i>C. carpio</i> (common carp)								
24		0	0	0	0	1	1	0
48		0	0	0	0	2	0	1
72		0	0	0	0	0	2	4
96		0	0	0	1	0	2	0

The mortality of *P. hypophthalmus* initiated at a Cr concentration of 5 mg L⁻¹, which showed greater fish death as the level of chromium was raised. In the common carp, on the other hand, a Cr density of 10 mg L⁻¹ started fish demise and higher Cr levels caused elevated mortalities. No fish death was noticed in the control groups for both species. Table 3 indicates that LC₁₀₀ values for the carp and catfish were at 30 and 10 mg L⁻¹ of Cr (after 72 h), respectively.

Concerning lead, the initial response of *P. hypophthalmus* was recorded at 40 mg L⁻¹ of Pb (48 h; Table 4). Further concentrations of Pb produced more mortality in the fish. Comparably, *C. carpio* showed primary death at 50 mg L⁻¹ (96 h) of Pb and continued dying with additional levels of Pb. Table 4 also indicates that the carp and catfish showed equal LC₁₀₀ values at 90 mg L⁻¹ of Pb (after 96 h).

TOXICITY FACTOR (TF)

The tested heavy metals were significantly different in TF ratios at different times ($P < 0.05$, Table 5) indicating that *C. carpio* required higher levels of Cd, Pb, and Cr than *P. hypophthalmus* to reach LC₅₀ values. The descending trends of Cd, Pb, and Cr LC₅₀s at different intervals were not significant in *C. carpio* ($P > 0.05$). In *P. hypophthalmus*, Cd and Pb LC₅₀s did not significantly reduce with time ($P > 0.05$) while the decrease in Cr LC₅₀ from 24 to 96 h was significant ($P < 0.05$).

Table 5. Toxicity factors (TFs) in *C. carpio* and *P. hypophthalmus* subjected to the heavy metals at different times

Duration (h)	Cd	Cr	Pb
24	2.31	2.47	3.07
48	1.64	1.91	2.64
72	1.88	2.16	1.54
96	1.3	2.28	1.6

DISCUSSION

This study was done to assess the sensitivity of the scaled *Cyprinus carpio* and the scaleless *Pangasius hypophthalmus* fish species to cadmium, chromium, and lead through determination of acute 96-h LC₅₀ values induced from exposure to different concentrations of the introduced heavy metals. As shown in Table 1 (A and B), *C. carpio* and *P. hypophthalmus* displayed contrasting tolerances to the applied densities of Cd, Cr, and Pb. The data clearly indicate that considerably more densities of the heavy metals are needed to induce dose-response mortality in the common carp than those required in the catfish. This implies that *C. carpio* should be more tolerant of the toxicant burden in comparison with *P. hypophthalmus*. This characteristic, in addition to species-specificity of responses (e.g.15), might have arisen from the fact that because a function of fish skin is to provide an effective protective barrier against environmental chemicals, and as common carp (and scaled fish in general: 20) have thicker epidermis covered by scales, they can effectively shield the animal against extra permeation of toxicants. Also, it is generally accepted that factors such as skin thickness and scale coverage are the determinants in the percutaneous (via skin) uptake rate of toxicants in fish (21). In brown bullhead (*Ameiurus nebulosus*), an important percentage of mercury uptake from the water occurred because of its scaleless permeable skin (22). Further evidence presented suggests protection against heavy metals toxicity by scales in three freshwater species making them tolerant to lethal concentrations of lead or mercury (23). The scales buffered the pH of lead nitrate solution and removed lead (and mercury) from water. The same authors also studied (24) the reduction of lead nitrate toxicity in water due to the presence of scales. They additionally reported reduced toxicity of 27 metals following

filtration through scales and concluded that the keratin in scales may be the most important ectodermal secretion in absorbing metals and in providing protection against their toxic levels. Likewise, the sorption and removal of heavy metals by fish scales was investigated (25, 26, 27); metals accumulation in fish scales is increased during exposure (28). On the other hand, a general conclusion (reviewed by 21) deduced that scaled and scaleless fish showed the same general pattern of response (deterioration and death) to sub-lethal chronic metal concentrations. Similarly, there was not a significant difference between field samples of *Cyprinus carpio* and *Clarias lazera* (scaleless) in Cd concentrations of their comparable tissues (29). The above discrepancies might have occurred because toxicity and accumulation of heavy metals in fish are mainly dependent upon metals concentration and exposure period, although such other factors as water salinity, pH, hardness and temperature, ecological needs, size and age, life cycle, capture season, and feeding habits of fish also play significant roles (4).

Comparison of the numbers of dead fishes at different levels of the heavy metals indicate that lethality of the metals lies in order of $Cr > Pb > Cd$. Accordingly, chromium was more toxic than cadmium and lead to both fishes and *P. hypophthalmus* showed more sensitivity than *C. carpio* to all the metals. This corroborates the review of (14) stating that the acute toxicity of chromium to fish reveals the differences in the 96h-LC₅₀ values between fish species, which can be attributed to the complicated metal-induced changes in the physiology and survival of aquatic organisms under metallic stress.

Lead has been reported as the most toxic element to aquatic invertebrates, algae and fish at the lowest concentration (30, 31), the toxicity of which is strongly influenced by lower water hardness (32).

Moreover, the concentration of heavy metals in fish is related to several factors such as physicochemical properties of the water (e.g. water hardness: 33) and the presence of other ions in the environment (34, 35). The high total water hardness (TDS: ca. 600 mg L⁻¹) in this study, therefore, could have probably reduced Pb toxicity rendering it an intermediate lethality. Similar order of toxicities and bioaccumulations ($Cr > Pb > Cd$) have been reported in two freshwater and five coastal fish species (36, 37).

Cadmium belongs to the most toxic water contaminants. Lethal values (96 h LC₅₀) for fish range from 0.5 µg dm⁻³ (38) to 21.1 mg dm⁻³ (39) depending both on intrinsic factors such as fish species or age, and on environmental conditions (40). Exposure of *C. carpio* to 0.560 mg L⁻¹ cadmium [Cd (NO₃)₂] killed all fish in 8 days (41) whereas 1.5 – 6 mg L⁻¹ of lead (PbNO₄) for two months caused no mortality in this species (42). In catfish *Heteropneustes fossilis*, LC₅₀ of cadmium chloride was found to be 50.41 mg L⁻¹ (4), which tends to proximate that (>60 mg L⁻¹) detected in this study. Reported LC₁₀₀ of Cd as 86.32 mg L⁻¹ (43) is almost near that (100 mg L⁻¹) found in this study (43).

The ratios of heavy metal toxicity factor (TF) calculated in here were greater for common carp compared to those for the catfish. Conversely, relatively higher copper TF ratio was found for the catfish *Clarias gariepinus* (TF= 1.19) as opposed to that estimated in tilapia *Oreochromis niloticus* (TF= 1.0) (20). This disparity might be due to the physicochemical characteristics of the test media (39, 44), species and ages, and their sensitivity rates to the tested heavy metals.

CONCLUSION

The development and use of toxicity tests provide types of data regarding toxic responses of different fish species, which could be more effectively used in predictive toxicology and risk assessment. The common carp (*C. carpio*) with higher

LC₅₀ (and LC₁₀₀) values appears to be more tolerant to heavy metals exposure than sutchi catfish (*P. hypophthalmus*). This attribute may be due to the increased resistance to the heavy metals through protection from the carp's scaled body versus scaleless body of the catfish. However, further studies are recommended to confirm it because various factors influence toxicant responses in natural and controlled ecosystems.

ACKNOWLEDGEMENTS

The authors are grateful to Samira Kiani, Zeynab Darvish, and Mojgan Ghanbari for their help during the experimental procedure.

REFERENCES

- Merian E. Metals and their compounds in the environment: occurrence, analysis and biological relevance: VCH Verlagsgesellschaft mbH; 1991.
- Heath AG. Water pollution and fish physiology: CRC; 1995.
- Folmar LC. Effects of chemical contaminants on blood chemistry of teleost fish: a bibliography and synopsis of selected effects. Environmental Toxicology and Chemistry. 1993;12(2):337-75.
- Yang H, Rose NL. Distribution of mercury in six lake sediment cores across the UK. The Science of the total environment. 2003;304(1-3):391-404.
- Canli M, Atli G. The relationships between heavy metal (Cd, Cr, Cu, Fe, Pb, Zn) levels and the size of six Mediterranean fish species. Environmental Pollution. 2003;121(1):129-36.
- Strmac M, Braunbeck T. Isolated hepatocytes of rainbow trout (*Oncorhynchus mykiss*) as a tool to discriminate between differently contaminated small river systems. Toxicology in vitro. 2000;14(4):361-77.
- Elnimr T. Evaluation of some heavy metals in *Pangasius hypophthalmus* and *Tilapia nilotica* and the role of acetic acid in lowering their levels. International Journal of Fisheries and Aquaculture. 2011;3(8):151-7.
- Shuhaimi-Othman M, Nadzifah Y, Ahmad A. Toxicity of Copper and Cadmium to Freshwater Fishes. Proceedings of World Academy of Science, Engin. Technol 2010;65:869-71.
- Alderdice D. The detection and measurement of water pollution-biological assays. Canadian Fisheries Report. 1967(9):33-9.
- Nussey G, Van Vuren J, Du Preez H. Bioaccumulation of aluminium, copper, iron and zinc in the tissues of the moggel from Witbank Dam, Upper Olifants River Catchment (Mpumalanga). South African Journal of Wildlife Research. 1999;29(4):130-44.
- Hung L, Lazard J, Mariojouis C, Moreau Y. Comparison of starch utilization in fingerlings of two Asian catfishes from the Mekong River (*Pangasius bocourti* Sauvage, 1880, *Pangasius hypophthalmus* Sauvage, 1878). Aquaculture Nutrition. 2003;9(4):215-22.
- Michael V. Aquaculture of Tilapia and Pangasius; A Comparative Assessment .Available from: http://caribefish.com/portal/index.php?option=com_content&view=article&id=71%3Aaquaculture-of-tilapia-and-pangasius-a-comparative-assessment&catid=19%3Aaquaculture-information&Itemid=76&lang=en.
- Velma V, Vutukuru S, Tchounwou PB. Ecotoxicology of hexavalent chromium in freshwater fish: a critical review. Reviews on environmental health. 2009;24(2):129.
- Das KK, Banerjee SK. Cadmium toxicity in fishes. Hydrobiologia. 1980;75(2):117-21.
- Gill TS, Pant JC. Mercury-induced blood anomalies in the freshwater teleost. Water, Air, & Soil Pollution. 1985;24(2):165-71.
- Kirubakaran R, Joy K. Inhibition of testicular 3 beta-hydroxy-delta 5-steroid dehydrogenase (3 beta-HSD) activity in catfish *Clarias batrachus* (L.) by mercurials. 1988; 26:907-8.
- Veena K, Radhakrishnan C, Chacko J. Heavy metal induced biochemical effects in an estuarine teleost. Indian journal of marine sciences. 1997;26(1):74-8.
- Crafford D. Uptake of selected metals in tissues and organs of *Clarias gariepinus* (sharp-tooth catfish) from the Vaal River

- System - Chromium, copper, iron, manganese and zinc. 2001. Available from: <http://www.readperiodicals.com/201104/2343235511.html>
19. Ezeonyejiaku C, Obiakor M, Ezenwelu C. Toxicity of copper sulphate and behavioral locomotor response of tilapia (*Oreochromis niloticus*) and catfish (*Clarias gariepinus*) species. *Online J Anim Feed Res* 2011;1(4):130-4.
 20. Mckim JM, Lien GJ. Toxic responses of the skin. In: Schlenk D, Benson WH, editors. *Target organ toxicity in marine and freshwater teleosts*, vo. London: Taylor & Francis; 2001. p. 165-230.
 21. Ferreira J, Schoonbee H, Smit G. The uptake of the anaesthetic benzocaine hydrochloride by the gills and the skin of three freshwater fish species. *Journal of Fish Biology*. 1984;25(1):35-41.
 22. Rose J, Hutcheson MS, West CR, Pancorbo O, Hulme K, Cooperman A, et al. Fish mercury distribution in Massachusetts, USA lakes. *Environmental Toxicology and Chemistry*. 1999;18(7):1370-9.
 23. Coello W, Khan M. Protection against heavy metal toxicity by mucus and scales in fish. *Archives of environmental contamination and toxicology*. 1996;30(3):319-26.
 24. Coello WF, Khan MAQ. Effect of keratin on heavy metal chelation and toxicity to aquatic organisms. *Environmental Toxicology and Risk Assessment*. In: Little EE, Delonay AJ, Greenberg BM, editors. West Conshohocken: American Society for Testing and Materials; 1998. p. 418.
 25. Varanasi U, Markey D. Uptake and release of lead and cadmium in skin and mucus of coho salmon (*Oncorhynchus kisutch*). *Comparative Biochemistry and Physiology Part C: Comparative Pharmacology*. 1978;60(2):187-91.
 26. Mustafiz S. The Application of Fish Scales in Removing Heavy, Metals from Energy-Produced Waste Streams: The Role of Microbes. *Energy sources*. 2003;25(9):905-16.
 27. El-Sheikh AH, Sweileh JA. Sorption of Trace Metals on Fish Scales and Application for Lead and Cadmium Pre-concentration with Flame Atomic Absorption Determination. *Jordan J Chem* 2008;3(1):87-97.
 28. Sauer GR, Watabe N. Ultrastructural and histochemical aspects of zinc accumulation by fish scales. *Tissue and Cell*. 1989;21(6):935-43.
 29. Al-Weher S. Levels of Heavy Metal Cd, Cu and Zn in Three Fish Species Collected from the Northern Jordan Valley, Jordan. *Jordan J Biol Sci*. 2008;1(1):41-6.
 30. Jorma KM. The Accumulation and excretion of heavy metals in organisms. In: Krenkel, editor. *Heavy metals in the aquatic environment*. Tennessee: Pergamon Press; 1973. p. 155-162.
 31. Eisler R. *Handbook of chemical risk assessment*. New York: Lewis Publishers; 2000.
 32. USEPA. *Ambient water quality criteria for lead*. Washington. 1980.
 33. Bryan GW. Some aspects of heavy metal tolerance in aquatic organisms. In: Lockwood APM, editors. *Effects of pollutants on aquatic organisms*. Cambridge: Cambridge University Press; 1976. p. 7-34.
 34. Giesy Jr JP, Wiener JG. Frequency distributions of trace metal concentrations in five freshwater fishes. *Transactions of the American Fisheries Society*. 1977;106(4):393-403.
 35. Wepener V, Vuren J, Preez H. Effect of manganese and iron at a neutral and acidic pH on the hematology of the banded tilapia (*Tilapia sparrmanii*). *Bulletin of environmental contamination and toxicology*. 1992;49(4):613-9.
 36. Yousafzai AM, Chivers DP, Khan AR, Ahmad I, Siraj M. Comparison of Heavy Metals Burden in Two Freshwater Fishes *Wallago attu* and *Labeo dyocheilus* With Regard to Their Feeding Habits in Natural Ecosystem. *Pakistan Journal of Zoology*. 2010;42(5):537-44.
 37. Lakshmanan R, Kesavan K, Vijayanand P, Rajaram V, Rajagopal S. Heavy metals accumulation in five commercially important fishes of Parangipettai, Southeast Coast of India. *Adv J Food Sci Technol*. 2009;1(1):63-5.
 38. Cusimano RF, Brakke DF, Chapman GA. Effects of pH on the toxicities of cadmium, copper, and zinc to steelhead trout (*Salmo gairdneri*). *Canadian Journal of Fisheries*

- and Aquatic Sciences. 1986;43(8):1497-503.
39. Lin HC, Dunson WA. The effect of salinity on the acute toxicity of cadmium to the tropical, estuarine, hermaphroditic fish, *Rivulus marmoratus*: a comparison of Cd, Cu, and Zn tolerance with *Fundulus heteroclitus*. Archives of environmental contamination and toxicology. 1993;25(1):41-7.
40. Witeska M. Changes in the common carp blood cell picture after acute exposure to cadmium. Acta Zoologica Lituanica. 2001;11(4):366-71.
41. Iger Y, Lock R, Meij JCA, Wendelaar Bonga S. Effects of water-borne cadmium on the skin of the common carp (*Cyprinus carpio*) Archives of environmental contamination and toxicology. 1994;26(3):342-50.
42. Iger Y, Abraham M. Effects of lead pollution on carp skin. In: Billard R, Pauw N, eds. Proceedings of the 1989 European Aquaculture Society Conference. European Aquaculture Society Press; 1989.
43. Singh A, Jain DK, Kumar P. Determination of LC₅₀ of cadmium chloride in *Heteropneustes fossilis*. GERF Bull Biosci. 2010;1(1):21-4.
44. Solbe JF. The toxicity of zinc sulphate to rainbow trout in very hard water. Water Res 1974;8:389-91.

Archive of SID