



Original Article

## Evaluation of Antimicrobial Effects of Gouda Cheese Wax

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### ABSTRACT

**Background and objectives:** Gouda Cheese is regarded as a high quality and one of the most popular cheeses in the world. The defining characteristics of Gouda cheese are its yellow color, great aroma and taste of caramel sweetness. The cheese should be well chilled before waxing, to get better seal. The seal is very important to prevent contamination with molds and putrefactive bacteria. Cheese wax is made from paraffin with additional microcrystalline to make pliable for better seal. The aim of this study was to investigate antibacterial effects of Gouda cheese wax.

**Methods:** Gouda cheese wax samples were collected from four different manufacturers in Iran. The total count of coliforms, *Escherichia coli*, *Salmonella*, coagulase-positive *Staphylococcus* and mold and yeast on the samples was determined. The antimicrobial activities of Gouda cheese wax against *E. coli*, *S. aureus*, *Saccharomyces cerevisiae*, *Aspergillus brasiliensis* and *Salmonella enterica* were investigated by determining minimum bactericidal concentration and minimum inhibitory concentration.

**Results:** The results indicated that all Gouda cheese wax samples were prepared in accordance with the national standards. In addition, the examined wax samples had no antimicrobial properties against the tested microorganisms.

**Conclusions:** The wax used in production of Gouda cheese in Iran has no antimicrobial properties.

**Keywords:** *Escherichia coli*, Cheese, *Salmonella*.

## INTRODUCTION

Gouda cheese is one of the most popular cheeses worldwide (1, 2). This semi-hard to hard cheese was first introduced in the Netherlands and was prepared from pasteurized cow's milk. Aged Gouda cheese have a strong flavor with a sharp yet sweet taste at the same time. Various microbial and chemical changes occur in the preparation of Gouda cheese. At the final stage of production, it is waxed with paraffin-based ingredients and then dried (1, 2). Production of this cheese include milk pasteurization, coagulation, cutting the curds, releasing the whey, moulding of cheese, brining, coating and maturing and storing (3-5).

There are International and National Standards on Gouda cheese that define the sensory, microbial, chemical, physical and packaging properties of this product. These standards also determine the microbial limitations for coliforms, *Escherichia coli*, *Salmonella*, coagulase-positive *Staphylococcus*, mold and yeast (4, 5).

According to Centers for Disease Control and Prevention in the US, a total of 113 outbreaks caused by cheese consumption were reported during 1998–2015, resulting in 2418 illnesses, 291 hospitalizations and 18 deaths (6).

Gouda cheese wax is obtained from petroleum derivatives. This type of wax can be recovered and used in a wide range of desirable applications. Paraffin and microcrystalline are widely used in the food industry because of their neutral properties. Paraffin with microcrystalline are used in packaging of various hard and semi-hard cheeses, such as Edam cheese and Gouda cheese. Edible paraffin and microcrystalline waxes are used to provide strength and waterproof properties (7-9). The function of the coating is to protect the cheese from microbial contamination while allowing moisture to evaporate. This coating is semi-permeable, which allows cheese to breathe and continue the maturation process (10).

Research plays an important role in increasing the longevity and improving the quality and safety of cheese products (11). The aim of this study was to evaluate antimicrobial effect of Gouda cheese wax and to determine the microbial contamination of Gouda cheese with and without wax.

## MATERIALS AND METHODS

Samples of Gouda cheese wax were collected from four Iranian dairy manufacturers between April and June 2018. [Table 1](#) shows the specifications of the collected samples.

Culture media including violet red bile lactose agar, mannitol salt agar, lauryl sulfate broth, SS agar and yeast extract glucose chloramphenicol agar were purchased from Merck Co. (Germany) to culture coliforms, *S. aureus*, *E. coli*, *Salmonella* and mold and yeast, respectively (12). The standard strains *E. coli* ATCC 19118, *S. aureus* ATCC 6538, *Saccharomyces cerevisiae* PTCC 5074, *Aspergillus brasiliensis* and *Salmonella enterica* PTCC 1709 were obtained from the Ibresco Co. (Iran).

According to the Codex C-5; 2013 and Institute of Standards and Industrial Research of Iran (ISIRI) 9013, all collected samples were examined for microbial enumeration of coliforms, *E. coli*, *Salmonella* and coagulase-positive *Staphylococcus* based on ISO 4832, ISO 11866-1, ISO 6785, ISO 6888-3 and ISO 6611, respectively (4, 5, 12-19).

The MIC of Gouda cheese wax against the tested microorganisms was determined using the broth microdilution method. First, serial dilutions were prepared from the wax in microbial growth media. The tested microorganisms were added to the wax dilutions and incubated for growth. The examined wax was serially diluted in sterile purified water. Next, 5 ml of each wax dilution were added to 5 ml of 2× concentrated media. Two media control tubes were prepared by mixing 5 ml sterile purified water with 5 ml of 2× media (14, 15).

All tubes were inoculated with 0.05 ml of the test microorganisms and were incubated at  $37 \pm 2$  °C for 16-20 hours. Finally, absorption at 620 nm was read by a spectrophotometer (UNIQO UV2100, USA) to determine bacterial growth.

The MBC was determined as the lowest concentration of cheese wax required to kill the tested microorganisms (20, 21). After determining the MIC, the microorganisms were cultured on a culture medium of Muller's Hilton-Agar sterile, and the plates were placed in a 35 °C incubator for 16 to 18 hours.

Table 1- Technical specifications of the collected Gouda cheese wax samples

Manufacturers	A	B	C	D
Number of samples	15	15	15	15
Health Code (Ministry of Health)	✓	✓	✓	✓
ISO 9001: 2008	✓	✓	✓	✓
ISO 22000: 2005	✓	✓	✓	✓
HACCP	✓	✓	✓	✓
Standard Logo	✓	✓	✓	✓
Production date and expiration	✓	✓	✓	✓

ISO: International organization for standardization

HACCP: Hazard Analysis Critical Control Point

## RESULTS

All wax samples collected from Gouda cheese met the required standards (Table 2).

The results showed that of the tested cheese wax samples had no effects on the growth of

the microorganisms. In other words, the collected samples of Gouda cheese wax had no inhibitory effect on *E. coli*, *S. aureus*, *S. cerevisiae*, *A. brasiliensis* and *S. enterica*.

Table 2- The results of contamination of wax of Gouda cheese samples

Gouda cheese wax samples	Total coliforms (Cfu/g)	<i>E. coli</i> in one gram	Coagulase-positive <i>Staphylococcus</i> in one gram	Mold and Yeast (Cfu/g)	<i>Salmonella</i> in 25 grams	
Manufacturer A	1	10>	Negative	Negative	0.3×10 <sup>2</sup>	Negative
	2	10>	Negative	Negative	0.2×10 <sup>2</sup>	Negative
	3	10>	Negative	Negative	0.4×10 <sup>2</sup>	Negative
	4	10>	Negative	Negative	0.6×10 <sup>2</sup>	Negative
	5	10>	Negative	Negative	0.5×10 <sup>2</sup>	Negative
Manufacturer B	1	10>	Negative	Negative	0.1×10 <sup>2</sup>	Negative
	2	10>	Negative	Negative	0.1×10 <sup>2</sup>	Negative
	3	10>	Negative	Negative	0.3×10 <sup>2</sup>	Negative
	4	10>	Negative	Negative	0.3×10 <sup>2</sup>	Negative
	5	10>	Negative	Negative	0.5×10 <sup>2</sup>	Negative
Manufacturer C	1	10>	Negative	Negative	0.1×10 <sup>2</sup>	Negative
	2	10>	Negative	Negative	0.1×10 <sup>2</sup>	Negative
	3	10>	Negative	Negative	0.3×10 <sup>2</sup>	Negative
	4	10>	Negative	Negative	0.4×10 <sup>2</sup>	Negative
	5	10>	Negative	Negative	0.1×10 <sup>2</sup>	Negative
Manufacturer D	1	10>	Negative	Negative	0.7×10 <sup>2</sup>	Negative
	2	10>	Negative	Negative	0.4×10 <sup>2</sup>	Negative
	3	10>	Negative	Negative	0.1×10 <sup>2</sup>	Negative
	4	10>	Negative	Negative	0.1×10 <sup>2</sup>	Negative
	5	10>	Negative	Negative	0.2×10 <sup>2</sup>	Negative

The results showed that of the tested cheese wax samples had no effects on the growth of the microorganisms. In other words, the collected samples of Gouda cheese wax had no inhibitory effect on *E. coli*, *S. aureus*, *S. cerevisiae*, *A. brasiliensis* and *S. enterica*.

## DISCUSSION

Cheese is vulnerable to microbiological deterioration throughout storage, distribution, processing and ripening due to its high water extensive microorganism development at cheese surface, which considerably reduces the

content and favorable pH for microbial growth (22). Conditions during ripening often promote cheese quality (23, 24). The cheese surface is an open ecosystem with abiotic properties that may be suitable for growth of various microorganisms. Maintaining hygienic conditions during ripening and storage plays a significant role in microbial contamination of cheese surface (6, 25). Cheese wax is a special coating used for preventing contamination, moisture retention and shaping cheese. Waxing cheese is a simple method to protect hard cheese from airborne bacteria, unwanted mold and excessive drying. This process is generally applied on semi-hard and hard cheeses. Hard cheese wax is a food-grade, paraffin-based wax. We found that wax samples from Gouda cheese had no inhibitory effect on potential pathogens in Gouda cheese, which is in agreement with two previous studies (11, 23).

Coating is a traditional method of maintaining the quality of cheese during shelf life. Cheese can be contaminated with various microorganisms during initial preparation, processing, packaging and storage. Thus, it can be concluded that antimicrobial compounds can improve cheese quality and shelf life (6, 26).

Our findings indicated that wax had no role in the prevention of microbial contamination of Gouda cheese. Some studies have investigated production of wax with antimicrobial properties. For instance, Taherkhani et al. used Kermanian Black Cuminin extract in Gouda cheese wax to induce antimicrobial properties

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#### CONCLUSION

The Gouda cheese wax samples collected from Iran have no antimicrobial properties. It is recommended to use Gouda cheese coating with antimicrobial properties to minimize the risk of contamination and spread of microorganisms.

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#### CONFLICT OF INTEREST

The authors declare no conflict of interest.

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