

# Improvement Vase Life of Chrysanthemum (*Dendranthema grandiflorum* L.) Cut Flowers Using Essential Oils of *Geranium*, *Eucalyptus* and *Myrtus*

Siros Bidarigh

Department of Agronomy, Lahijan Branch, Islamic Azad University, Lahijan, Iran.

Received: 28 September 2015

Accepted: 25 November 2015

\*Corresponding author's email: [bidarigh@yahoo.com](mailto:bidarigh@yahoo.com)

## Abstract

This study carried out based on a randomized completely design with 10 treatments geranium, eucalyptus and myrtus extraction, in 3 levels (2, 6 and 10 ml in 600 ml of distilled water) and control (distilled water) with 3 replications in 30 plots and each plot with 4 cut flowers. In this experiment traits such as vase life, water absorption, fresh weight, dry matter percent, petal protein content, petal carotenoids, leaf chlorophyll content, lipid peroxidation (MDA), SOD and POD enzymes activity and bacterial population were measured. The results showed that 2 ml myrtus the per 600 ml was the longest vase life compared to control (9.74 days) increased the vase life of 5.99 days. The highest bacterial population was observed in control plants. Also, the lowest MDA was found in 2 ml myrtus oil.

**Keywords:** *Chrysantemun*, *Eucalyptus*, *Geranium*, *Myrtus*, Vase life.

## INTRODUCTION

Chrysanthemum (*Dendranthema grandiflorum* L.) is one of the most popular cut flowers in the world (Dole and Wilkins, 2004; Hashemi *et al.*, 2013). Studies on postharvest life of chrysanthemum cut flowers have shown that the vase life of this cut flowers is long because it is non-climacteric. However, water stress as a result of vascular occlusion have been reported as an important factor in reducing the quality of chrysanthemum cut flowers (Halevy and Mayak, 1979; Adachi *et al.*, 1999; Nabigol *et al.*, 2005). Microorganisms, especially fungi and bacteria that grow in the vase solution, block the stem end and limit water uptake by cut flowers with the production of chemical compounds.

Disinfectants are used long years to remove microorganisms of vase solution (Roychowdhury and Sarkar, 1995; Nowak and Rudnicki, 1990; Zagory and Reid, 1986). In recent years, the use of natural compounds has been preferred instead of chemicals to control pathogens of fruits and vegetables as well as disinfectants in preservative solution of cut flowers (Burt, 2004; Thanberg *et al.*, 2002; Valero and Frances, 2006; Solgi *et al.*, 2009). Among these materials can be pointed to the active ingredients of medicinal plants or essential oils, which are completely natural and biodegradable. These compounds due to having high concentrations of phenolic compounds, have antimicrobial properties and their use in the vase solution has been evaluated positive on the postharvest life of cut flowers (Bounatirou *et al.*, 2007; Lambert *et al.*, 2001; Mihajilov Krstev *et al.*, 2010).

Solgi *et al.* (2009) reported that the use of essential oils of *Zataria multiflora* and *Thymus vulgaris* as well as their active ingredients in the preservative solution increase the vase life of gerbera cut flowers. Mousavi Bazaz and Tehranifar (2011) found that the effect of essential oils of cumin, mint and thyme on alstroemeria cut flower life is positive. Similar results have been reported for the positive effects of herbal essential oils on the increase of the vase life of gladiolus (Hegazi and Gan, 2009), rose (Jalili Marandi *et al.*, 2011) and cloves (Kazemi and Ameri, 2012) cut flowers.

The aim of this study is to evaluate the effect of essential oils of geranium, eucalyptus and myrtus on the postharvest quality and vase life of chrysanthemum cut flowers.

## MATERIALS AND METHODS

The study of chrysanthemum cut flowers was performed as a randomized complete block design with 10 treatments and 3 replicates. In the laboratory, cut flowers were arranged under water with a height of 50 cm. Essential oils of geranium, myrtus and eucalyptus used in this study were purchased from Barij Essence Company. The used treatments were herbal essential oils of geranium, eucalyptus and myrtus in 3 levels (2, 6 and 10 ml in 600 ml of distilled water with the control treatment that along with 3 % sucrose were permanently used. Chrysanthemum cut flowers were maintained until the end of the experiment under controlled conditions with a 12-hour photoperiod, light intensity of  $12 \mu\text{mol m}^{-2} \text{s}^{-1}$ , relative humidity 60 to 70 % and temperature of  $20 \pm 2^\circ\text{C}$ .

Vase life was calculated by counting the days since applying the treatment (first day) until wilting the leaves and flowers (Nabigol *et al.*, 2005). Water uptake of flowers was calculated using the following formula (Hashemi *et al.* 2013):

Water uptake = [initial volume of vase solution - (solution remaining at the last day + average evaporation of room)] / fresh weight of cut flowers at the first day

In order to measure a, b and total chlorophyll and carotenoid of petals, sampling was performed in the fifth day of the test and the process of measuring chlorophyll and carotenoid of petals was performed using Mazumdar and Majumdar method (2003) and finally, the amount of leaf chlorophyll content suggested in mg g<sup>-1</sup> of fresh weight and carotenoids of petals in  $\mu\text{g g}^{-1}$  of fresh weight were calculated using the following formula:

$$\text{chlorophyll a} = 9.93 (A_{660}) - 0.777 (A_{642.5})$$

$$\text{chlorophyll b} = 17.6 (A_{642.5}) - 2.81 (A_{660})$$

$$\text{total chlorophyll} = 7.12 (A_{660}) - 16.8 (A_{642.5})$$

$$\text{carotenoid} = 4.69 + A_{440} - 0.268 \times (20.2) (A_{645}) + (8.02) (A_{663})$$

24 hours after applying the treatments, sampling of the stem end and the vase solution was performed and the bacterial colonies of stem end was counted according to the method of Oraee *et al.* (2011) and bacteria in the vase solution was counted using Liu *et al.* (2009) method. Kjeldahl method was used to measure protein. The level of malondialdehyde (MDA), as a product of peroxidation reaction of membrane fatty acids, was measured using Heath and Parker (1968). In *et al.* (2007) method was used to measure the peroxidase enzyme activity and Giannopolitis and Reis (1997) method was used to measure superoxide dismutase activity.

At the end of the experiment, data analysis was performed using MSTATC software and data mean was compared using LSD method.

## RESULTS AND DISCUSSION

The results of analysis of variance show that the effect of geranium, myrtus and eucalyptus essential oils on all measured traits except POD activity was significant at level of 1%. The effects of treatments on POD activity was significant at 5% level (Table 1).

Table 1. ANOVA effects of various essential oils on the measured traits.

S.O.V	df	Vase life	Dry matter	Water uptake	Carotenoid	Chlorophyll a	Chlorophyll b	Total chlorophyll
Treatments	9	9.20**	11.20**	1.66**	0.031**	2.095**	0.555**	4.705**
Error	20	0.805	1.207	0.237	0.008	0.001	0.009	0.011
CV (%)		6.50	4.52	10.81	1.80	6.66	3.08	1.16

\*\* : Probability 1%

Continues Table1. ANOVA effects of various essential oils on the measured traits.

S.O.V	df	Petal's Protein	Bacterial on vase solution	Bacterial on stem end	POD	SOD	MDA
Treatments	9	5.18**	778**	1654**	0.139*	2.330**	1.890**
Error	20	1.067	108	219	0.042	0.142	0.421
CV (%)		9.28	13.27	12.51	11.11	3.46	4.61

\*\* : Probability 1%

\* : Probability 5%

### Vase life

According to the results of Table 2, it could be stated that the use of eucalyptus, myrtus and geranium essential oils in the vase solution compared to control significantly increased the vase life of chrysanthemum cut flowers. Among all applied treatments, the most durability was related with treatment of 2 ml of myrtus (15.73 days) that has statistically no significant difference with treatments of 10 ml of geranium (14.74 days), 6 ml of myrtus (14.75 days), 2 ml of eucalyptus (15.13 days) and 6 ml of eucalyptus (4.75 days). The control treatment with 9.74 days had the minimum vase life among all treatments (Table 2).

In previous studies, it has been reported that the use of herbal essential oils can delay aging of flower and increase durability through reducing the microbial population of the solution and stem end and improve water uptake (Mousavi Bazaz and Tehranifar, 2011; Solgi *et al.*, 2009). The antimicrobial effect of essential oils on increasing the vase life of cut flowers of cloves (Bayat *et al.*, 2011) and gerbera (Ziyaei Movahed *et al.*, 2010; Solgi *et al.*, 2009) has been reported that is in accordance with the results of current study.

### Water uptake

The result of mean comparison showed that water uptake is significantly increased compared to the control by using herbal essential oils. As the control with 3.19 ml g<sup>-1</sup> F.W. had the lowest water uptake. Among the herbal essential oils, eucalyptus treatments of 6 and 10 ml with

5.39 and 5.14 ml g<sup>-1</sup> F.W., respectively, and 2 and 6 ml of geranium with 5.40 and 5.04 ml g<sup>-1</sup> F.W., respectively, have no statistically significant difference and have the maximum water uptake, Although, 2 and 10 ml of myrtus with 4.52 and 4.51 ml g<sup>-1</sup> respectively, had not different significant with mentioned treatments (Table 2).

Water balance is the most important factor in determining the quality and postharvest life of cut flowers (da Silva, 2003). As mentioned in the statement of results, treatment of chrysanthemum cut flowers with herbal essential oils, improved water uptake compared to the control. In fact, essential oils by controlling bacterial and fungal contamination of the vase solution, prevents obstruction of vascular and causes balance of water relations of cut flowers. Similar results have been reported that the positive effect of herbal essential oils on water uptake of cut flowers of alstroemeria (Mousavi Bazaz and Tehranifar, 2011) and cloves (Bayat *et al*, 2011) and these results are in accordance with the results of current study.

### **Dry matter**

By using herbal essential oils, the percentage of dry matter is increased compared to the control. So that the control treatment with 21.39 % had the lowest dry matter that of course has no statistically significant difference with the treatment of 2 and 10 ml of myrtus and 6 ml eucalyptus (Table 2). The maximum amount of the dry matter was related to the treatment of 6 ml of myrtus with 27.42 %, which has no statistically significant difference with the treatments of 10 ml of geranium and 2 ml of eucalyptus (Table 2). As mentioned in the statement of results, the use of herbal essential oils improved dry matter weight of chrysanthemum cut flower by increasing water relations and fresh weight. Similar results have been reported that the positive effect of herbal essential oils on increasing dry matter of gladiolus cut flowers (Hegazi and Gan, 2009) and these results are in accordance with the results of the this study.

### **Petal's carotenoids content**

Based on the mean of comparison, the amount of carotenoids in petals was increased in all used treatments compared to the control. As seen in Table 2, there is no statistically significant difference between used essential oils in terms of the amounts of carotenoids in petals. The control treatment with 4.77 µg g<sup>-1</sup> F.W has the least amount of carotenoids in petals (Table 2).

Carotenoids and anthocyanins are the main index durability and marketing of cut flowers. In previous studies, it has been found that treatment of cut flowers with a vase solution containing sugar or disinfectants causes maintaining cell turgidity by improving water uptake and thus prevents destruction of important pigments such as carotenoids (Amarjit, 2000; Hassanpour Asil and Karimi, 2010, Edrisi, 2008).

### **Petal's protein content**

The results showed that petal's protein content in the control treatment (9.07 %) was less than other treatments. Maximum petal's protein content was related to treatments of 2 ml of geranium (13.16 %) and 2 ml of eucalyptus (12.91 %) that have no statistically significant difference with treatments of 2, 6 and 10 ml of myrtus and 10 ml of geranium (Table 2).

Researchers reported that herbal essential oils increase vase life through reducing the breakdown of protein. They stated that herbal essential oils have antioxidant properties that prevent activity of oxygen free radicals and damage of proteins caused by free radicals and protect the protein in cut flowers (Kazemi *et al.*, 2014; Rajasekaran *et al.*, 2002). Maintaining and increasing the protein in Lisianthus cut flowers (Kazemi *et al.*, 2014) have been reported with the use of herbal essential oils in the vase solution that these results are in accordance with the results of the present research.

### **Chlorophyll a, b and total**

Based on the results, content f chlorophyll a, b and total has been increased compared to



the control by using essential oils. So that the control treatment has the minimum amount of chlorophyll a, b and total with 4.72, 2.44 and 7.16 mg g<sup>-1</sup> F.W, respectively. The maximum content of chlorophyll a (7.59 mg g<sup>-1</sup> F.W) was obtained for treatment of 2 ml of eucalyptus essential oil. The maximum content of chlorophyll b was obtained for treatment of 2 ml of eucalyptus (3.80), 10 ml of eucalyptus (3.72) and 6 ml of myrtus (3.62). The maximum content of total chlorophyll was also related to treatment of 2 ml of eucalyptus (11.39) (Table 2).

Lise *et al.* (2004) believe that the increase of chlorophyll is because of cells activity and increasing sugar. The researchers reported that essential oils are very effective in maintaining chlorophyll due to their antioxidant properties. Kazemi *et al.* (2014) stated that water stress and obstruction of vessels increase oxygen free radicals in chloroplasts, and so result destruction of chlorophyll molecules and membranes of chloroplasts. The results of this research showed that herbal essential oils with anti-radical property maintain and increase the chlorophyll in lisianthus cut flowers that is in accordance with the results of the present research.

### Bacterial population on vase solution and stem end

Based on the mean comparison, the number of bacterial colonies of the vase solution and stem end is reduced by using essential oils. As seen in Table 2, the highest amount of bacterial colonies in vase solution was found in the control (98.33 colonies per 10 ml agar) that has no statistically significant difference with treatments of 2 and 10 ml of geranium, 10 ml of myrtus and 6 ml of eucalyptus. The minimum bacterial number of vase solution was recorded for treatments of 2 ml of eucalyptus (55 colonies) and 2 ml of myrtus (58.33 colonies) that there was no statistically significant difference between them. However, the maximum bacterial number of stem end was related to the control treatment with 162 colonies that has no statistically significant difference with treatments of 2 ml of geranium (136 colonies) and 10 ml of geranium (137 colonies). The minimum bacterial number of stem end were recorded for two treatments of 2 ml of eucalyptus (88 colonies) and 2 ml of myrtus (89 colonies) that there was no statistically significant difference between them (Table 2).

Table 2. Mean comparison of effect of various essential oils on the measured traits.

Treatments	Vase life (day)	Dry matter (%)	Water uptake (ml g <sup>-1</sup> F W)	Carotenoid (µg g <sup>-1</sup> F W)	Petal's protein (%)	Chlorophyll a (mg g <sup>-1</sup> F W)	Chlorophyll b (mg g <sup>-1</sup> F W)	Total chlorophyll (mg g <sup>-1</sup> F W)	Bacteria in vase solution (colonies per 10 ml agar)	Bacteria in the stem end (colonies per 10 ml agar)
Control	9.71 <sup>d</sup>	21.39 <sup>e</sup>	3.19 <sup>c</sup>	4.77 <sup>b</sup>	9.07 <sup>c</sup>	4.72 <sup>j</sup>	2.44 <sup>e</sup>	7.16 <sup>g</sup>	98.33 <sup>a</sup>	162 <sup>a</sup>
G <sub>1</sub>	13.73 <sup>bc</sup>	25.05 <sup>bc</sup>	5.40 <sup>a</sup>	5.00 <sup>a</sup>	13.16 <sup>a</sup>	6.02 <sup>f</sup>	3.08 <sup>bc</sup>	9.10 <sup>d</sup>	94.00 <sup>a</sup>	136 <sup>ab</sup>
G <sub>2</sub>	13.42 <sup>bc</sup>	24.67 <sup>bcd</sup>	5.04 <sup>a</sup>	5.02 <sup>a</sup>	10.03 <sup>bc</sup>	6.35 <sup>d</sup>	3.22 <sup>b</sup>	9.57 <sup>c</sup>	66.66 <sup>bc</sup>	111 <sup>bc</sup>
G <sub>3</sub>	14.74 <sup>ab</sup>	25.68 <sup>ab</sup>	4.04 <sup>bc</sup>	5.10 <sup>a</sup>	11.71 <sup>ab</sup>	5.72 <sup>g</sup>	2.96 <sup>cd</sup>	8.73 <sup>e</sup>	94.00 <sup>a</sup>	137 <sup>ab</sup>
M <sub>1</sub>	15.73 <sup>a</sup>	21.80 <sup>e</sup>	4.52 <sup>ab</sup>	5.09 <sup>a</sup>	11.47 <sup>ab</sup>	6.25 <sup>e</sup>	3.21 <sup>b</sup>	9.46 <sup>c</sup>	58.33 <sup>c</sup>	89 <sup>d</sup>
M <sub>2</sub>	14.75 <sup>ab</sup>	27.42 <sup>a</sup>	3.80 <sup>bc</sup>	5.12 <sup>a</sup>	11.24 <sup>ab</sup>	7.01 <sup>b</sup>	3.62 <sup>a</sup>	10.46 <sup>b</sup>	72.33 <sup>bc</sup>	114 <sup>bc</sup>
M <sub>3</sub>	12.13 <sup>c</sup>	22.73 <sup>de</sup>	4.51 <sup>ab</sup>	4.04 <sup>a</sup>	11.46 <sup>ab</sup>	5.72 <sup>g</sup>	2.87 <sup>d</sup>	8.60 <sup>ef</sup>	92.00 <sup>a</sup>	126 <sup>b</sup>
O <sub>1</sub>	15.13 <sup>ab</sup>	25.60 <sup>ab</sup>	5.39 <sup>a</sup>	5.04 <sup>a</sup>	12.91 <sup>a</sup>	7.59 <sup>a</sup>	3.80 <sup>a</sup>	11.39 <sup>a</sup>	55.00 <sup>c</sup>	88 <sup>d</sup>
O <sub>2</sub>	14.75 <sup>ab</sup>	23.13 <sup>cde</sup>	5.14 <sup>a</sup>	5.02 <sup>a</sup>	10.00 <sup>bc</sup>	5.51 <sup>h</sup>	2.87 <sup>d</sup>	8.51 <sup>f</sup>	85.33 <sup>ab</sup>	121 <sup>bc</sup>
O <sub>3</sub>	13.75 <sup>bc</sup>	25.04 <sup>bc</sup>	3.99 <sup>bc</sup>	5.12 <sup>a</sup>	10.17 <sup>bc</sup>	6.93 <sup>c</sup>	3.72 <sup>a</sup>	10.66 <sup>b</sup>	68.66 <sup>bc</sup>	97 <sup>cd</sup>

In each column, means with the similar letters are not significantly different at 5% level of probability using LSD test.

Essential oils of <i>Geranium</i>		Essential oils of <i>Myrtus</i>		Essential oils of <i>Eucalyptus</i>	
G <sub>1</sub> : 2 ml in 600 ml of distilled water		M <sub>1</sub> : 2 ml in 600 ml of distilled water		O <sub>1</sub> : 2 ml in 600 ml of distilled water	
G <sub>2</sub> : 6 ml in 600 ml of distilled water		M <sub>2</sub> : 6 ml in 600 ml of distilled water		O <sub>2</sub> : 6 ml in 600 ml of distilled water	
G <sub>3</sub> : 10 ml in 600 ml of distilled water		M <sub>3</sub> : 10 ml in 600 ml of distilled water		O <sub>3</sub> : 10 ml in 600 ml of distilled water	

Anjum *et al.* (2001) reported that addition of a suitable disinfectant to the preservative solution of cut flowers prevents growth of microorganisms and increases water uptake and therefore causes more durability of cut flowers. The positive impact of herbal essential oils on reducing microbial population of solution and stem end of cut flowers of gladiolus (Hegazi and Gan, 2009), cloves (Kazemi and Ameri, 2012) and rose (Shanan, 2012) has been reported is in accordance with the results of the present research.

## Malondialdehyde (MDA)

As can be seen in Fig. 1, the concentration of MDA was maximum in the control treatment (14.96 nmol g<sup>-1</sup> F.W.). However, it has no statistically significant difference with treatments of 2, 6 and 10 ml of geranium and eucalyptus essential oils. The minimum concentration of MDA was related to the treatments of 6 ml of myrtus with 12.65 nmol g<sup>-1</sup> F.W. and then 10 ml of myrtus (13.09 nmol g<sup>-1</sup> F.W.) that these treatments were the best ones for this trait (Fig. 1).

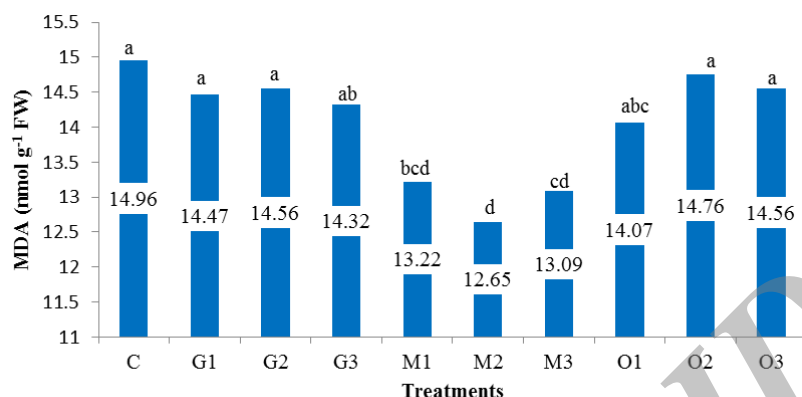


Fig. 1. Effect of various essential oils on the amount of MDA.

Essential oils of <i>Geranium</i>			Essential oils of <i>Myrtus</i>			Essential oils of <i>Eucalyptus</i>		
G <sub>1</sub> : 2 ml in 600 ml of distilled water	M <sub>1</sub> : 2 ml in 600 ml of distilled water	O <sub>1</sub> : 2 ml in 600 ml of distilled water	G <sub>2</sub> : 6 ml in 600 ml of distilled water	M <sub>2</sub> : 6 ml in 600 ml of distilled water	O <sub>2</sub> : 6 ml in 600 ml of distilled water	G <sub>3</sub> : 10 ml in 600 ml of distilled water	M <sub>3</sub> : 10 ml in 600 ml of distilled water	O <sub>3</sub> : 10 ml in 600 ml of distilled water

Palma *et al.* (2002) believe that the herbal essential oils by preventing the activity of oxygen species causes reducing the lipid peroxidation in cell membrane and reducing the concentration of MDA. The researchers believe that treatment of cut flowers with sugar and disinfectants by reducing the stresses applied on flowers causes to reduce the accumulation of MDA (Jin *et al.*, 2006). Kazemi and Ameri (2012) reported that the positive effect of herbal essential oils of thyme and lavender on the stability of the membrane and reduction of MDA in clove cut flowers that is in accordance with the results of the present research.

## Antioxidant enzymes (peroxidase and superoxide dismutase)

Based on the results, the maximum SOD activity was associated with the control treatment with 13.14 IU g<sup>-1</sup> F.W. By using essential oils, the amount of SOD has been significantly reduced, so that the minimum SOD activity was recorded for treatments of 6 ml of geranium (10.12 IU g<sup>-1</sup> F.W.) and 10 ml of myrtus (10.09 IU g<sup>-1</sup> F.W.) that there was no significant difference between them statistically (Fig. 2).

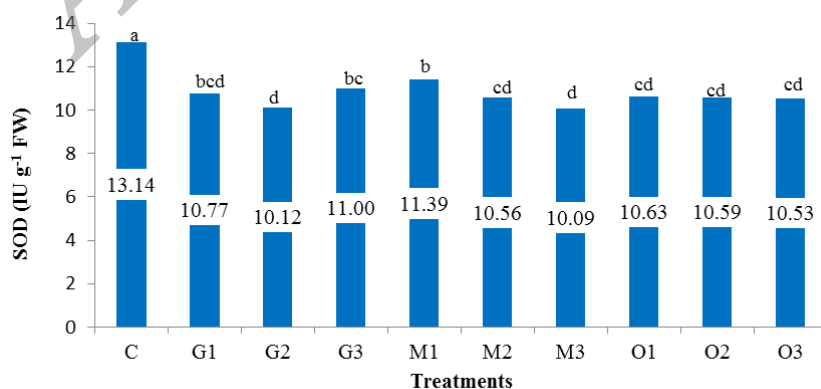


Fig. 2. Effects of various essential oils on the activity of SOD.

Essential oils of <i>Geranium</i>			Essential oils of <i>Myrtus</i>			Essential oils of <i>Eucalyptus</i>		
G <sub>1</sub> : 2 ml in 600 ml of distilled water	M <sub>1</sub> : 2 ml in 600 ml of distilled water	O <sub>1</sub> : 2 ml in 600 ml of distilled water	G <sub>2</sub> : 6 ml in 600 ml of distilled water	M <sub>2</sub> : 6 ml in 600 ml of distilled water	O <sub>2</sub> : 6 ml in 600 ml of distilled water	G <sub>3</sub> : 10 ml in 600 ml of distilled water	M <sub>3</sub> : 10 ml in 600 ml of distilled water	O <sub>3</sub> : 10 ml in 600 ml of distilled water

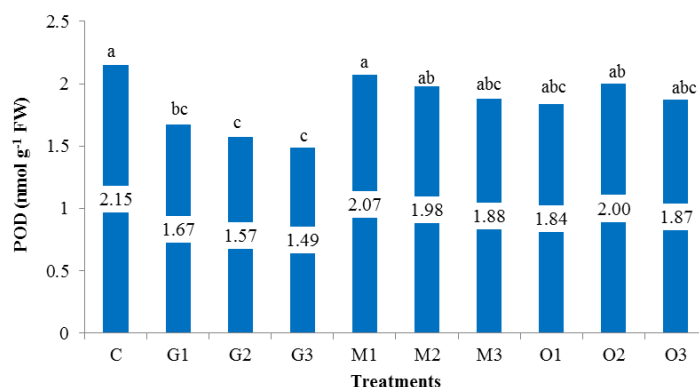


Fig. 3. Effects of various essential oils on the activity of POD.

Essential oils of <i>Geranium</i>	Essential oils of <i>Myrtus</i>	Essential oils of <i>Eucalyptus</i>
G <sub>1</sub> : 2 ml in 600 ml of distilled water	M <sub>1</sub> : 2 ml in 600 ml of distilled water	O <sub>1</sub> : 2 ml in 600 ml of distilled water
G <sub>2</sub> : 6 ml in 600 ml of distilled water	M <sub>2</sub> : 6 ml in 600 ml of distilled water	O <sub>2</sub> : 6 ml in 600 ml of distilled water
G <sub>3</sub> : 10 ml in 600 ml of distilled water	M <sub>3</sub> : 10 ml in 600 ml of distilled water	O <sub>3</sub> : 10 ml in 600 ml of distilled water

Study of the results of POD activity show that among the used essential oils, the concentrations of 2, 6 and 10 ml of geranium has only decreased POD activity significantly. As can be seen in Fig. 3, the maximum POD activity was associated with the control treatment with 2.15 nmol g<sup>-1</sup> F.W. that has no statistically significant difference with treatments of 2, 6 and 10 ml of myrtus and eucalyptus. The minimum POD activity in the treatment of 10 ml of geranium (1.49 nmol g<sup>-1</sup> F.W.) and 6 ml of geranium (1.57 nmol g<sup>-1</sup> F.W.) that there was no significant difference between them statistically (Fig. 3).

Among the various treatments, the control treatment had the highest antioxidant enzymes activity. Xiao Zheng and Huang (2002) reported that after harvesting cut flowers from the stock plant and transferring them to a vase solution, flowers exposed to stress especially water stress. Upadhyaya and Panda (2004) believe that the use of disinfectant in the vase solution reduces the production of oxygen free radicals by reducing water stress and thus they reduce the oxidation of membrane lipids and thereby prevent the wilting of cut flowers.

### Litrature Cited

- Adachi, M., Kawabata, S. and Sakiyama, R. 1999. Changes in carbohydrate content in cut chrysanthemum [*Dendranthema grandiflorum* (Ramat) Kitamura] 'Shuhou-no-Chikara' stems kept at different temperature during anthesis and senescence. *Journal of the Japanese Society for Horticultural Science*, 68: 505-512.
- Amarjit, B. 2000. Plant growth regulation agriculture and horticulture. Food Product Press. 5:147-165.
- Anjum, M. A., Naveed, F., Sahakeel, F. and Amin, S. 2001. Effect of some chemicals on keeping quality and vase life of tuberose (*Polianthus tuberosa* L.) cut flower. *Journal of Research (Science)*, Bahauddin Zakariya University, Multan, Pakistan, 12 (1): 1-7.
- Bayat, H., Azizi, M., Shoor, M. and Mardani, H. 2011. Effect of ethanol and essential oils on extending vase life of carnation cut flower (*Dianthus caryophyllus* cv. 'Yellow Candy'). *Notulae Scientia Biologicae*, 3(4): 100-104.
- Bounatirou, S., Simitis, S., Miguel, M. G., Faleiro, L., Rejeb, M. N., Neffati, M., Costa, M. M., Figueiredo, A. C., Barroso, J. G. and Pedro, L. G. 2007. Chemical composition antioxidant and antibacterial activities of the essential oils isolated from tunisian *Thymus capitatus*. *Link Food Chemistry*. 105: 146-155.
- Burt, S. 2004. Essential oils: their antibacterial properties and potential applications in foods. *International Journal of Food Microbiology*, 94:223-253.
- Da Silva, J.A. 2003. The cut flower: postharvest consideration. *Journal of Biological Sciences*. 3(4): 406-442.
- Dole, J.M. and Wilkins, F.H. 1999. Floriculture principles and species. Prentice Hall Pub., New Jersey, USA.
- Edrisi, B. 2008. Postharvest physiology of cut flowers. Payame Digar- Arak. 150 pages.

- Giannopolitis, C. and Ries, S. 1997. Superoxid desmutase. I: occurence in higher plant. *Plant Physiology*, 59: 309–314.
- Halevy, A.H. and Mayak, S. 1979. Senescence and postharvest physiology of cut flowers, part 1. *Horticultural Reviews*. 1: 204–236.
- Hashemi, M., Mirdehghan, S. H. and Farahmand, H. 2013. The effects of thymol, menthol and eugenol on quality and vase life of chrysanthemum cut flowers. *Iran Agricultural Research*, 32(2):55-70.
- Hassanpour Asil, M. and Karimi, M. 2010. Efficiency of benzyladenine reduced ethylene production and extended vase life of cut *Eustoma* flowers. *Plant Omics Journal*, 3(6): 199-203.
- Heath, R. L. and Parker, L. 1968. Photoperoxidation in isolated chloroplasts: I. Kinetics and stiochiometry of fatty acid peroxidation. *Archives of Biochemistry and Biophysics*, 125: 189-198.
- Hegazi, M. A. and Gan, E. K. 2009. Influences of some essintial oils vase life of *Gladiolus hybrid* L. 'Spikes'. *International Journal for Agro Veterinary and Medical Sciences*. 3:19-24.
- In, B. C., Motomura, S., Inamoto, K., Doi, M. and Mori, G. 2007. Multivariate analysis of realation between preharvest environmental factors, postharvest morphological and physiological factors and vase life of cut Asomi Red Roses. *Japanese Society for Horticultural Science*. 76: 66-72.
- Jalili Marandi, R., Hassani, A., Abdollahi, A. and Hanafi, S. 2011. Application of *Carum copticum* and *Satureja hortensis* essential oils and salicylic acid and silver thiosulphate in increasing the vase life of cut rose flowers. *Journal of Medicinal Plants Research*, 5(20):5034-5038.
- Jin, J., Ningwei, S. H., Nan, M., Jinhe, B. and Junping . C. 2006. Regulation of ascorbate peroxidase at the transcript level is involved in tolerance to postharvest water deficit stress in the cut rose 'Samanta'. *Postharvest Biology and Technology*. 40: 236-243.
- Kazemi, M. and Ameri, A. 2012. Response of vase life carnation cut flower to salicylic acid, silver nano particles, glutamine and essential oil. *Asian Journal of Animal Science*, 6(3): 122-131.
- Kazemi, S., Hassanpour Asil, M. and Ghasemnezhad, M. 2014. Physiological effects of some essential oils in comparison with 8-hydroxyquinoline in cut lisianthus flowers (*Eustoma grandiflorum* L.). *Iranian Journal of Horticultural Science*, 45(2): 185-195.
- Lambert, R. J. W., Skandamis, P. N., Coote, P. J. and Nychas, G. J. E. 2001. A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol and carvacrol. *Journal of Applied Microbiology*, 91:453- 462.
- Lise, A., Michelle, H. and Serek, M. 2004. Reduced water availability improves drought tolerance of potted miniature roses: Is the ethylene pathway involved. *Journal of Horticultural Science & Biotechnology*, 79 (1):1-13.
- Liu, J., Zhang, Z., Joyce, D. C., He, S., Cao, J. and Lv, P. 2009. Effect of postharvest nanosilver treatments on cut flowers. *Acta Horticulturae*, 847: 245-250.
- Mazumdar, B. C. and Majumdar, K. 2003. Methods on physicochemical analysis of fruits. [www. Sundeep books.com](http://www.Sundeepbooks.com). 187p.
- Mihajilov Krstev, T., Radnovic, D., Kitic, D., Stojanovic Radic, Z. and Zlatkovic, B. 2010. Antimicrobial activity of *Satureja hortensis* L. essential oil against pathogenic microbial strains. *Archives of Biological Science Belgrade*, 62:159-166.
- Mousavi Bazaz, A. and Tehranifar, A. 2011. Effects of ethanol, methanol and essential oils as novel agents to improve vase life of *Alstroemeria* flowers. *Journal of Biodiversity and Environmental Sciences*, 5(14):41-46.
- Nabigol, A., Naderi, R., Babalar, M. and Kafi, M. 2005. Increasing vase life of chrysanthemum cut flowers by using floral preservatives and recutting. *Iranian Society for Horticultural Science*, 7(4): 207-216.
- Nowak, J. and Rudnicki, R. M. 1990. Postharvest handling and storage of cut flowers, florist greens and potted plants. Timber Press, Portland, Oregon. USA.
- Orace, T., Asgharzadeh, A., Kiani, M. and Orace, A. 2011. The role of preservative compounds on number of bacteria in the end of stems and vase solution of cut *Gerbera*. *Journal of Ornamental and Horticultural Plants*, 1(3): 161-166.



- Palma, J.M., Sandalio, L.M., Corpas, F.J., Romero, M.C., McCarthy, I. and Río, L.A. 2002. Plant proteases, protein degradation, and oxidative stress: role of peroxisomes. *Plant Physiology and Biochemistry*, 40 (6-8): 521-530.
- Rajasekaran, L., Stiles, A. R. and Caldwell, C.D. 2002. Stand establishment in processing carrots: Effects of various temperature regimes on germination and the role of salicylates in promoting germination at low temperatures. *Canadian Journal of Plant Science*, 82: 443–450.
- Roychowdhury, N. and Sarkar, S. 1995. Influence of chemicals on vase life of gladiolus. *Acta Horticulture*, 4(5):389-391.
- Shanan, N. 2012. Applications of essential oils to prolong the vase life of rose (*Rosa hybrid* L. cv. 'Grand') cut flowers. *Journal of Horticultural Science and Ornamental Plants*, 4(1):66-74.
- Solgi, M., Kafi, M., Taghavi, T. S. and Naderi, R. 2009. Essential oils and silver nano particles (SNP) as novel agents to extend vase life of gerbera (*Gerbera jamesonii* cv. 'Dune') flowers. *Postharvest Biology and Technology*. 53: 155-158.
- Thanberg, R. L., Tran, T. T., Bennett, R. W., Matthewes, R. N. and Belay, N. 2002. Microbial evaluation of selected fresh produce obtained at retail markets. *Journal of Food Protection*. 65:677-682.
- Upadhyaya, H. and Panda, S. K. 2004. Responses of *Camellia sinensis* to drought and rehydration. *Biologia Plantarum*, 48 (4): 597-600.
- Valero, M. and Frances, E. 2006. Synergistic bactericidal effect of carvacrol, cinnamaldehyde or thymol and refrigeration to inhibit *Bacillus cereus* in carrot broth. *Food Microbiology*. 23:68-73.
- Xiao Zheng, L. and Huang, B. 2002. Cytokinin effects on creeping bentgrass response to heat stress. *Crop Science Society of America*, 42(2): 466-472.
- Zagory, D. and Reid, M. S. 1986. Role of vase solution microorganisms in the life of cut flowers. *American Society for Horticultural Science*, 111:154-158.
- Ziyaei Movahed, Z., Kafi, M., Khalighi, A., Azizi, M. and Sharifi, R. 2010. Investigation of the possibility in replacing natural ingredients (essential oil and extracts of Australian Cheese wood) instead of anti-bacterial chemicals ingredients in preservative solution of the gerbera cut flower. *Iranian Journal of Horticultural Science*, 41, 337-345. (In Farsi).

# افزایش عمر پس از برداشت گل بریده داودی (*Dendranthema grandiflorum L.*) به کمک اسانس‌های ژرانیوم، اکالیپتوس و مورد

سیروس بیدریغ

گروه زراعت، واحد لاهیجان، دانشگاه آزاد اسلامی، لاهیجان، ایران

تاریخ تایید: ۴ آذر ۱۳۹۴

تاریخ دریافت: ۶ مهر ۱۳۹۴

\* ایمیل نویسنده مسئول: [bidarigh@yahoo.com](mailto:bidarigh@yahoo.com)

این مطالعه با هدف بررسی اثر اسانس‌های گیاهی بر عمر گلجایی و کیفیت پس از برداشت گل بریده داودی بر پایه طرح کاملاً تصادفی با ۱۰ تیمار شامل اسانس گیاهی ژرانیوم، مورد و اکالیپتوس در ۳ سطح (۲، ۶ و ۱۰ میلی‌لیتر در ۶۰۰ میلی‌لیتر آب مقطر) همراه با تیمار شاهد (آب مقطر)، در ۳ تکرار و ۳۰ پلات به صورت دائمی انجام شد. صفاتی نظیر عمر گلجایی، جذب آب، درصد ماده خشک، میزان پروتئین گلبرگ، کاروتنوئید گلبرگ، کلروفیل برگ، غلظت MDA، فعالیت آنزیم‌های SOD و POD و جمعیت میکروبی اندازه‌گیری شدند. نتایج نشان داد که تیمار ۲ میلی‌لیتر اسانس مورد در ۶۰۰ میلی‌لیتر آب مقطر بیشترین عمر گلجایی را داشت که نسبت به شاهد (۹/۷۴ روز) ۵/۹۹ روز عمر گلجایی را افزایش داد. شاهد بیشترین جمعیت میکروبی را داشت. کمترین تجمع MDA نیز برای تیمار ۲ میلی‌لیتر مورد ثبت شد.

چکیده

کلید واژگان: اکالیپتوس، داودی، ژرانیوم، عمر گلجایی، مورد.