Published online 2014 October 11.

Research Article

The Effects of Hydroalcoholic Extract of Apium graveolens Leaf on the Number of Sexual Cells and Testicular Structure in Rat

Wesam Kooti^{1,2}; Esrafil Mansouri^{3,*}; Maryam Ghasemiboroon⁴; Mahmoud Harizi⁴; Damoon Ashtary-Larky^{4,5}; Reza Afrisham⁴

¹Department of Immunology, School of Medicine, Kurdistan University of Medical Sciences, Sanandaj, IR Iran

²Student Research Committee, Kurdistan University of Medical Sciences, Sanandaj, IR Iran ²Cellular and Molecular Research Center, Department of Anatomical Sciences, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran

⁴ Student Research Committee, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran
⁵ Department of Clinical Biochemistry, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran

*Corresponding author: Esrafil Mansouri, Cellular and Molecular Research Center, Department of Anatomical Sciences, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran. Tel: +98-9111183028, E-mail: esrafilmansori@yahoo.com

Received: January 12, 2014; Revised: March 11, 2014; Accepted: July 4, 2014

Background: Use of medicinal plants with high antioxidant properties could be effective to increase fertility and improvement of disorders such as hormonal imbalance, impotency, oligospermia and immotile sperm. Celery (Apium graveolens) is rich in antioxidant agents. The leaf and stems of celery contain phenols, furanocoumarin and luteolin. Apigenin is one of the main flavonoids of celery leaf. Objectives: This study aimed to investigate the effects of hydroalcoholic extract of celery on histological properties of testis and number of sexual cells in male rats.

Materials and Methods: Thirty-two male Wistar rats were divided into four groups of eight rats each. Control, did not receive any medication; sham, received normal saline; and two groups received celery extract orally in dosages of 100 and 200 mg/kg/BW once every two days for 60 days. At the end, animals were anesthetized, and caudal part of the right epididymis was used for sperm counting. After fixation of testis, tissue sections were prepared and studied microscopically to evaluate morphometric (lumen diameter, number of primary spermatocyte and sertoli cell) and histological changes. Data was analyzed by one-way ANOVA test using SPSS15 software. P < 0.05 was considered as statistically significant.

Results: There was a significant increase in the number of sperms, sertoli cells, and primary spermatocyte (P < 0.05) in groups receiving extract; however, structural changes were not observed in the groups.

Conclusions: It seems that celery increases spermatogenesis in male rats, but has no destructive effects on testicular tissue.

Keywords: Apium graveolens; Testicular Structure; Rat; Extract

1. Background

Infertility is one of the most common health problems in the world, which involves approximately 15% of couples. Infertility can be present in both genders, but about 50% of infertility is associated with male factor. Decreased sperm count and motility and deformity of sperm are the most important factors of male infertility (1). Various chemical drugs are available to treat infertility; however, researchers are looking for drugs with less adverse effects and toxicity. In developing countries, traditional medicine is important in maintaining health of population (2). Celery is used as food in some countries, which also has medicinal properties. On the other hand, due to adverse effects of synthetic drugs, it is necessary to conduct scientific researches to assess medicinal properties of various plants. Celery (Apium graveolens) belongs to the parsley (umbelliferae) species from the Apiaceae family, with a height of 100 cm, and a strong scent and fleshy and solid stems (3). Celery seeds contain 2% to 3% essential fat. This fat contains mostly limonene (typically 60%), selenin (10%), furocoumarin, furocoumarin glycosides, palmitic acid and federonoid (4). Celery contains vitamin C and other compounds such as phalides and coumarins, which are health-enhancing compounds. Different parts of this plant owe a wide spectrum of biological, pharmacological and therapeutic effects including anti-rheumatism, sedative, blood pressure lowering, antifungal, analgesic, anti-inflammatory, detoxification, anti-spasmodic, anti-bacterial, anti-contractions and antiepileptic (5). Recently, several studies reported increasing prevalence of infertility. Infertility can be due to consumption of natural plant compounds (phytoestrogens) if consumed in high amounts. These compounds can affect the reproductive system and reduce fertility (6). Based on available reports, plants such as celery contain phytoestrogens, which can be effective in fertility and reproductive system (7). Plasma membrane of sperm is susceptible to oxi-

Copyright @ 2014, School of Pharmacy, Ahvaz Jundishapur University of Medical Sciences; Published by DOCS. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (http://creativecommons.org/licenses/by-nc/4.0/) which permits copy and redistribute the material just in noncommercial usages, provided the original work is properly cited.

dative damage due to large amounts of unsaturated fatty acids, finally leading to decreased motility and viability of sperm. Antioxidant compounds increase sperm function and can improve fertility. Furthermore, a study indicated that celery has a protective effect on testes against sodium valproate (8) and di (2-ethylhexyl) phthalate (9). Studies demonstrated that celery protects testes from functional and structural damages and sperm from toxicity induced by atrazine (10) and quinine sulfate (11).

2. Objectives

The present study was performed to assess the effects of hydroalcoholic extract of celery leaf on the number of sperm and testes structure in male rats.

3. Materials and Methods

This experimental study was performed in Physiology Research Center of Ahvaz Jundishapur University of Medical Sciences (AJUMS). Thirty-two male Wistar rats (weight range of 170-220 grams) were obtained from the central animal house of AJUMS. Animals were housed in plastic cages with 12/12h light/dark cycle at 21 ± 2 °C. These conditions were maintained constant throughout the experiments. The study was performed in accordance with the principles of laboratory care established by the Ethics Committee of Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran.

3.1. Extraction

To prepare the extract, leaves of celery was prepared from agricultural lands of Hamidiye (Khoozestan, Iran) and specified by an expert in Pharmacognosy department of pharmacy faculty of AJUMS. The leaves were shade-dried and milled to fine powder using a mechanical grinder. Powder (50 g) was soaked in 200 mL of 70% ethanol (Merck, Germany) for three days with occasional shaking. The solution was passed through filter paper and evaporated in oven at 40°C for 48 hours. Extracted powder was kept at 4°C until use. Concentrations of 100 and 200 mg/kg/BW were prepared from obtained powder of celery using normal saline as solvent (Daru Pakhs, Iran) (12). Hydroalcoholic extract and solvent administrated by gavage once every two days for 60 days. The duration of spermatogenesis in Wistar rats is 52 days according to the literature (13, 14).

3.2. Experimental Design

In this study, animals were randomly divided into four groups of eight rats each (15). The groups were divided as follows: group I as control did not receive any medication; group II as sham received 1 mL solvent of normal saline; group III received hydroalcoholic extract of celery (100 mg/kg); group IV received hydroalcoholic extract of Celery (200 mg/kg) (9). At the end of the experimental, rats were anaesthetized and killed by IP injection of ketamine (60 mg/kg) and xylazine (10 mg/kg) (Alfasan, Holland). Then, testes and epididymis were carefully separated. Testes were weighed by a digital scale (Sartorius-PT120-Germany) and fixed by Bouin's fixative, dehydrated through increasing concentrations of ethanol and embedded in paraffin. Then five sections with thickness of 5 µm (an interval of 100 µm) in each animal were stained with hematoxylin-eosin. The sections were studied regarding morphometric and histopathologic changes (10 fields in each section) (16) by Motic software (Micro-Optic Industrial Group CO., LTD., UK)(17) and light microscope, respectively (Olympus, 3H-Z-Japan). Sperms were extracted from the caudal portion of epididymis by fragmentation of epididymis into 1 mL in normal saline. After five minutes incubation at 37°C, sperm suspension was diluted with a ratio of 1:100. This diluted solution was used for sperm count using Neubauer's slide. Therefore, a drop of solution was poured on the slide, and lamella was placed on it; sperms were counted by light microscope in corner areas (18).

3.3. Statistical Analysis

All data were expressed as mean \pm SE. Data was analyzed by One-way ANOVA and LSD tests using SPSS for windows (version 15, IBM, USA). P < 0.05 was considered as statistically significant.

4. Results

4.1. Testis Index

There was no significant difference between the groups regarding testis index (P > 0.05) (Table 1).

Table 1. Testicular Index ^a				
Group	Control	Sham	Experimen- tal Group 1, 100 mg/kg	Experimen- tal Group 2, 200 mg/kg
Testicular index, g	0.63±0.01	0.65± 0.01	0.66± 0.01	0.65± 0.01

 $^{\rm a}$ Comparison of Mean \pm SE of testis index between the experimental (100 and 200 mg/kg hydro alcoholic extract of celery) and control groups.

4.2. Morphometric Studies

Number of Sertoli cells increased significantly in group receiving extract (200 mg/kg) (17.16 \pm 0.6) compared to control group (10.99 \pm 0.47) (P < 0.05) (Figure 1). Similarly, the results showed that lumen diameter was significantly reduced in groups III and IV (82.77 \pm 7.03 and 71.18 \pm 5.53, respectively) compared to the control group (P < 0.05) (105.3 \pm 5.6) (Figure 2). The mean number of primary spermatocytes was significantly increased in groups III and IV (61.5 \pm 2.5 and 63.5 \pm 6.5, respectively) compared to the control group (P < 0.05) (105.3 \pm 5.6) (Figure 2).

Figure 1. Comparison of Mean \pm SE of Sertoli Cells Between the Experimental (100 and 200 mg/kg Hydroalcoholic Extract of Celery) and Control Groups (n = 8)



Number of Sertoli cells increased significantly in groups receiving extract (200 mg/kg); * (P < 0.05).

Figure 3. Comparison of Mean ± SE of Primary Spermatocytes Between the Experimental (100 and 200 mg/kg Hydro-Alcoholic Extract of Celery) and Control Groups (n = 8)



Number of primary spermatocytes increased significantly in experimental groups (100 and 200 mg/kg); * (P < 0.05).

Figure 2. Comparison of Mean \pm SE of Lumen Diameter Between the Experimental (100 and 200 mg/kg Hydro-Alcoholic Extract of Celery) and Control Groups (n = 8)



Lumen diameter decreased significantly in experimental groups (100 and 200 mg/kg); $^{\circ}$ (P < 0.05).

Figure 4. Comparison of Mean \pm SE of Sperm Count Between the Experimental (100 and 200 mg/kg Hydro-Alcoholic Extract of Celery) and Control Groups (n = 8)



Sperm count increased significantly in experimental groups (100 and 200 mg/kg); * (P < 0.05).

4.3. Sperm Count

Administration of hydroalcoholic extract of celery with dosages of 100 (77.9 \pm 5.8) and 200 mg/kg (89.68 \pm 5.5) increased sperm count compared to the control group (56.78 \pm 4.9) (P < 0.05) (Figure 4).

4.4. Histological Study

Germinal epithelium of seminiferous tubules and interstitial tissue in the control and sham groups were normal and tissue damage was not observed (Figures 5A and B). Moreover, we did not observe any changes in arrange-

www.SID.ir

ment and structure of epithelial cells in seminiferous tubules and interstitial tissue in groups receiving hydroalcoholic extract of celery compared to groups I and II; only the germinal epithelium was thicker in these groups (Figures 5C and D).





I, control; II, sham (solvent of normal saline); III and IV, experimental (100 and 200 mg/kg hydroalcoholic extracts of celery). Germinal epithelium of seminiferous tubules and interstitial tissue in the control and sham groups were normal. Germinal epithelium was thicker in groups III and IV.

5. Discussion

Historically, plants have been considered as a valuable source; however, they are mostly used for food consumption rather than their therapeutics effects. The current study showed that hydroalcoholic extract of celery has useful effects on spermatogenesis and testis in male rats. In this study, we observed a slightly increase of testes index in groups III and IV, but was not statistically significant. In addition, we did not observe any changes or destructive effects on testicular tissue. The results indicated that used dosages of hydroalcoholic extract of celery had no clear effect on these parameters. Celery contains flavonoids and phenolic acids with anti-inflammatory effects; also, apigenin and epiein as main flavonoids of celery owe anti-inflammatory properties (19). Apigenin as an antioxidant inhibits the production of hydrogen peroxide and IgE, which is responsible for inflammation and allergic responses (20). Apigenin has inhibitory effects on cyclooxygenase and lipoxygenase (21). Therefore, celery is able to reduce harmful effects of free radicals on cells and prevents cell death and loss of weight or tissue volume due to its antioxidant and anti-inflammatory properties. These results are in agreement with some previous investigations (9). Besides, our results indicated that Sertoli cells and primary spermatocytes increased and lumens diameter reduced in experimental groups. Our result is similar to those reported in previous studies (10, 11). A previous study suggested that hydroalcoholic extract of celery seed increased testosterone secretion (22). The level of testosterone as the most important androgenic hormone is effective in the evolution and proliferation of germ cells and spermatid differentiation. Likely, celery directly affects Sertoli cells by stimulating secretion of testosterone (23). Sertoli cells have a major role in differentiation and development of spermatogenesis. These cells promote caryokinesis karyokinesis, cytokinesis and differentiating of spermatozoa with producing growth factors such as activin in the presence of calcium ion. Furthermore, Sertoli cells secrete tubule fluid, which helps to nourish sexual cells and plays an important role in their supporting (24). In addition to the shape and quality of sex cells, blood concentration of reproductive hormones is an important parameter to evaluate function of the reproductive system. Testosterone affects seminiferous tubules and induces spermatogenesis (25). Our findings indicated that administration of hydroalcoholic extract of celery at both dosages increased sperm count. In a similar study by Hamza et al. it was shown that celery extract decreased toxic effects of sodium valproate and increased all of spermatogenic cells lineages (8). Sperms count is one of the important factors of fertility, and decrease of sperm count can decrease the probability of a successful pregnancy. Researchers suggested that spermatogenesis and maturation of sexual cells depend on protection of cytotoxic and pathologic lesions that threaten these events (26, 27). Previous studies demonstrated that induced damage by free radicals and oxidative stress could cause various disorders, such as infertility (28, 29). Spermatogenic cells, unlike other cells are very sensitive to oxidative damage, which is due to high amounts of poly-unsaturated fatty acids in their plasma membrane and very low levels of cytoplasmic antioxidants (30). Peroxidation of membrane fatty acids leads to loss of membrane fluidity and decreases its enzyme activity and ion channels leading to gradual loss of the ability of sperm to bind to the oocyte. The main effect of lipid peroxidation in all cells, especially sperm is disturbance of structure and function of organelles or cell membranes (ion transport processes, fluidity and permeability, metabolic gradients). These changes can affect the structure and function of sperm (31). In normal conditions, a balance exists between reactive species oxygen and free radicals production and antioxidant defense system. An imbalance between free radical production and antioxidant defenses leads to injuries induced by free radicals. Flavonoids are secondary metabolites of plant compounds with powerful antioxidant properties, and cannot be synthesized in the body and must be received through diet. Celery is a strong antioxidant due to flavonoids such as apiein and apigenin (32, 33). Antioxidant compounds are able to protect cell membranes against damage (34). Antioxidants affect hypothalamic-pituitary-testicular axis directly or indirectly, thus increasing sperm count and fertility (35, 36). Moreover this herb contains vitamins E and C (32, 33). In an experimental study, it was shown that these vitamins improve sperm parameters such as count and motility (37, 38). Vitamin E is a powerful antioxidant with a supporting role in quality and quantity of sperm, fertilization and fertility in humans. This vitamin is abundant in Sertoli cells, spermatogonia, and round spermatid. Therefore, increasing sexual cells due to protective effect of vitamins and compounds of celery is justified leading to reduction in diameter of seminiferous tubules lumen. The results of the present study confirmed that hydroalcoholic extract of celery leaf could increase spermatogenesis in rats. This increase was more pronounced at higher dosage. Perhaps this plant could be used to treat infertility in men. However, it is suggested to perform further experimental and clinical studies on total extracts of this plant and its exact mechanisms on spermatogenesis.

Acknowledgements

We would like to gratitude Mrs. Noorbehbahani, from the Laboratory of Biochemistry Department who helped us and the Research Consultation Center (RCC) for their technical support.

Authors' Contributions

Study concept, design and critical revision of the manuscript for important intellectual content: Esrafil Mansouri, Wesam Kooti, Maryam Ghasemiboron; drafting of the manuscript and advisor and conducting the experiments: Mahmoud Harizi, Reza Afrisham and Damoon Ashtary-Larky.

References

- Karamzade A, Mirzapour H, Kheirollahi M. Genetics Aspects of Male Infertility. J Isfahan Med Sc. 2013;31(246):1149–62.
- Austin DF. Ipomoea littoralis (Convolvulaceae)—taxonomy, distribution, and ethnobotany. Econ Bot. 1991;45(2):251–6.
- Khare CP. Indian Medicinal Plants: An Illustrated Dictionary:New York, USA: Springer Science; 2007.
- Kitajima J, Ishikawa T, Satoh M. Polar constituents of celery seed. Phytochem. 2003;64(5):1003–11.
- Blumental M, Cladbery A, Brinkman J, Newton MA. Herbal medicine: expanded commission E Monographs. *Integ Med Communicat.* 2000:78–83.
- Csupor-Loffler B, Hajdu Z, Zupko I, Rethy B, Falkay G, Forgo P, et al. Antiproliferative effect of flavonoids and sesquiterpenoids from Achillea millefolium s.l. on cultured human tumour cell lines. *Phytother Res.* 2009;23(5):672–6.
- Kooti W, Ghasemiboroon M, Asadi-Samani M, Ahangarpoor M, Zamani M, Amirzargar A, et al. The Effect of Halcoholic Extract of Celery Leaves on the Delivery Rate (Fertilization and Stillbirths), the Number, Weight and Sex Ratio of Rat off Spring. Adv Environ Biol. 2014;8(10):824-30.
- Hamza AA, Amin A. Apium graveolens modulates sodium valproate-induced reproductive toxicity in rats. J Exp Zool A Ecol Genet Physiol. 2007;307(4):199–206.
- 9. Madkour NK. The beneficial role of celery oil in lowering of di(2ethylhexyl) phthalate-induced testicular damage. *Toxicol Ind Health*. 2012.

- 10. Abarikwu SO, Pant AB, Farombi EO. The protective effects of quercetin on the cytotoxicity of atrazine on rat Sertoli-germ cell coculture. *Int J Androl.* 2012;**35**(4):590–600.
- 11. Farombi EO, Ekor M, Adedara IA, Tonwe KE, Ojujoh TO, Oyeyemi MO. Quercetin protects against testicular toxicity induced by chronic administration of therapeutic dose of quinine sulfate in rats. *J Basic Clin Physiol Pharmacol*. 2012;**23**(1):39–44.
- Kooti W, Ghasemiboroon M, Ahangarpoor A, Hardani A, Amirzargar A, Asadi-Samani M. The effect of hydro-alcoholic extract of celery on male rats in fertility control and sex ratio of rat offspring. J Babol Univ Med Sci. 2014;16(4):43-9.
- Freitas F, Cordeiro-Mori F, Sasso-Cerri E, Lucas S, Miraglia S. Alterations of spermatogenesis in etoposide-treated rats: a stereological study. *Interciencia*. 2002;27(1):227-35.
- Momen HR, Eskandari N. Effect of vitamin E on sperm parameters and DNA integrity in sodium arsenite-treated rats. *Iran J Reprod Med.* 2012;**10**(3):249–56.
- Najafizadeh P, Dehghani F, Panjeh Shahin M, Hamzei Taj S. The effect of a hydro-alcoholic extract of olive fruit on reproductive argons in male sprague-dawley rat. *Iran J Reprod Med.* 2013;11(4):293–300.
- Gholampour F, Owji SM, Javadifar TS, Bahaoddini A. Long term Exposure to Extremely Low Frequency Electromagnetic Field Affects Sex Hormones Level and Structure of Testis in Rats. Inter J Zool Res. 2012;8(3):130–6.
- Takzaree N, Mortazavi H, Hassanzadeh G, Safaye S, Hossini M. Male rat spermatogenesis influenced by Achillea millefolium L . Tehran Univ Med J. 2013;70(11):684–90.
- Khaki A, Prove T. Effect of ciprofloxacin on caudal epididymis sperm quality and apoptosis. UMJ. 2008;19(1):29–35.
- Mencherini T, Cau A, Bianco G, Della Loggia R, Aquino RP, Autore G. An extract of Apium graveolens var. dulce leaves: structure of the major constituent, apiin, and its anti-inflammatory properties. J Pharm Pharmacol. 2007;59(6):891-7.
- 20. Sultana S, Ahmed S, Jahangir T, Sharma S. Inhibitory effect of celery seeds extract on chemically induced hepatocarcinogenesis: modulation of cell proliferation, metabolism and altered hepatic foci development. *Cancer Lett.* 2005;**221**(1):11–20.
- 21. Silvan AM, Abad MJ, Bermejo P, Villar A. Effects of compounds extracted from Santolina oblongifolia on TXB(2) release in human platelets. *Inflammopharmacology*. 1998;**6**(3):255–63.
- 22. Kerishchi P, Nasri S, Amin Gh, Tabibian M. The effects of Apium graveolens extract on sperm parameters and H-G hormonal axis in mice. Hamadan Iran: Proceedings of the International Congress of Physiology and Pharmacology; 2011.
- 23. McLachlan RI, O'Donnell L, Meachem SJ, Stanton PG, de Kretser DM, Pratis K, et al. Identification of specific sites of hormonal regulation in spermatogenesis in rats, monkeys, and man. *Recent Prog Horm Res.* 2002;**57**:149–79.
- 24. Grootegoed JA, Siep M, Baarends WM. Molecular and cellular mechanisms in spermatogenesis. *Baillieres Best Pract Res Clin Endocrinol Metab.* 2000;**14**(3):331–43.
- Wang C, Zhang Y, Liang J, Shan G, Wang Y, Shi Q. Impacts of ascorbic acid and thiamine supplementation at different concentrations on lead toxicity in testis. *Clin Chim Acta*. 2006;370(1-2):82-8.
- Modaresi M, Messripour M, Rajaei R. Effect of cinnamon extract on the number of spermatocyte and spermatozoa cells in mice. *IJMAP*. 2010;26(1):83-90.
- 27. Nikravesh MR, Jalali M, Mohamadi Sh. Effects of Crude Onion Extract on Murine Testis. *J Reprod Infertility*. 2009;**10**(41):239–44.
- 28. Santos FW, Graca DL, Zeni G, Rocha JB, Weis SN, Favero AM, et al. Sub-chronic administration of diphenyl diselenide potentiates cadmium-induced testicular damage in mice. *Reprod Toxicol.* 2006;**22**(3):546–50.
- 29. Allen JA, Diemer T, Janus P, Hales KH, Hales DB. Bacterial endotoxin lipopolysaccharide and reactive oxygen species inhibit Leydig cell steroidogenesis via perturbation of mitochondria. *Endocrine*. 2004;**25**(3):265-75.
- Gebreegziabher Y, Marcos E, McKinon W, Rogers G. Sperm characteristics of endurance trained cyclists. Int J Sports Med. 2004;25(4):247-51.
- 31. Aitken RJ. Free radicals, lipid peroxidation and sperm function. *Reprod Fertil Dev.* 1995;7(4):659–68.

Jundishapur J Nat Pharm Prod. 2014;9(4):e17532

www SID ir

- Fazala SS, Ansarib MM, Singlac RK, Khand S. Isolation of 3-n-Butyl Phthalide & Sedanenolide from Apium graveolens Linn. *IGJPS*. 2012;2(3):258–61.
- Fazal SS, Singla RK. Review on the Pharmacognostical & Pharmacological Characterization of Apium Graveolens Linn. *IGJPS*. 2012;2(1):36–42.
- Bouayed J, Bohn T. Exogenous antioxidants Double-edged swords in cellular redox state: Health beneficial effects at physiologic doses versus deleterious effects at high doses. Oxid Med Cell Longev. 2010;3(4):228–37.
- Shittu LAJ, Bankole MA, Oguntola JA, Ajala O., Shittu RK, Ogundipe OA, et al. Sesame leaves intake improve and increase epididymal spermatocytes reserve in adult male Sprague Dawley rat. Sci Res

Essays. 2007;2(1):319-24.

- Shittu IAJ, Shittu RK, Adesite SO, Ajala MO, Bankole MA, Benebo AS, et al. Sesame radiatum Phytoestrogens Stimulate Spermatogenic Activity and Improve Sperm Quality in Adult Male Sprague Dawley Rat Testis. Inter J Morphol. 2008;26(3).
- Kooti W, Mansori E, Ghasemiboroon M, Harizi M, Amirzargar A. Protective effects of celery (Apium Graveolens) on testis and cauda epididymal spermatozoa in rat. *Iran J Reprod Med*. 2014; 12(5):365-6.
- Nouri M, Ghasemzadeh A, Farzadi L, Shahnazi V, Ghaffari-Novin M. Vitamins C, E and lipid peroxidation levels in sperm and seminal plasma of asthenoteratozoospermic and normozoospermic men. *Iran J Reprod Med.* 2008;6(1):1–5.