

Mesenchymal Stem Cells: New Aspect in Cell-Based Regenerative Therapy

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ABSTRACT

MSCs are multipotent progenitors which reside in bone marrow. They support hematopoietic stem cells homing, self renewal and differentiation in bone marrow. They can also differentiate into osteoblasts, adipocytes, chondrocytes, myocyates and many other tissues. In vivo, when trauma happens, MSCs operate cell renewal and migrate to the damaged tissues to regenerate that injury. In vitro, MSCs are able to proliferate and differentiate to a variety of cell lineages. This makes them a very hopeful tool for cell-based regenerative therapy for large bone defects, maxillofacial skeletal reconstruction, cardiovascular and spinal cord injury and so many other defects. The most important characteristic that make MSCs an excellent tool for cell replacement is their ability to escape from immune rejection. For therapeutic purposes they usually isolated from human bone marrow or fat and they should proliferate in order to reach an adequate number for implantation. Conventionally DMEM medium supplemented with 10% FBS is used for their expansion, but currently autologous platelet rich products are replaced FBS. Platelet granules contain so many growth factors that can support MSCs proliferation.

Introduction

Two population of multipotent progenitors reside in bone marrow (BM): Hematopoietic stem cells (HSCs) and mesenchymal stem cells (MSCs).^{1,2} In bone marrow MSCs are able to support hematopoiesis by releasing stromal-derived factor-1, Flt-3 ligand and stem cell factor.³ This subset of cells can migrate to damaged tissues⁴ and differentiate into at least three lineages: osteoblasts, adipocytes, and chondrocytes.5-7 Trilineage differentiation considered minimal criteria for their multipotency.8,9

This class of multipotent progenitors first were recognized by Friedenstein et al in France in 1968. 10 Friedenstein and his colleges described an undifferentiated heterogeneous subset of cells which were spindle-shaped, plastic-adherent, non-phagocytic with fibroblast-like morphology. 11,12 Later in 1974, when they cultured a small amount of these cells in basal cell culture medium and saw their ability to generating clonal fibroblast-like colonies, they recognized that these cells had a high potential

of proliferatition, therefore they named them fibroblast colony-forming cells (CFU-F), 11 afterward these cells entitled as mesenchymal stem cells.

In addition to bone marrow, MSCs can be identified in other tissues like fat, muscle, perichondrium, dental pulp and fetal tissues including BM, spleen, lung and liver; 13-15 as well as in other animals like mouse, rat, cat, dog and horse. 16 For experimental and therapeutic purposes, MSCs are usually isolated from human bone marrow and fat. 17,18 Although MSCs are characterized by their ability for differentiation into bone, fat, and cartilage, 5,6 they can also differentiate into other tissues including tendon, muscle, nerve, liver, kidney, pancreas and skin. 16,19

Bone marrow-derived MSCs characteristically lack hematopoietic antigens including CD45, CD34, CD133, CD14, and MHC class II as well as endothelial antigens including CD80, CD34, CD31, vWF. On the other hand, they express stromal cell surface markers and adhesion molecules such as

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CD105, CD106, CD166, CD90, CD73, CD44, CD140b, CD9, CD10, CD13, and CD29. 1,20,21

MSCs have a significant role in lymphopoiesis and myelopoiesis and show extensive immunomodulatory properties too. MSCs arrest B and T lymphocytes in G0/G1 phase of the cell cycle, thus inhibit their responses. MSCs can prevent monocytes from their function as an antigenpresenting cell (APC). Moreover, they improve regulatory T cell expansion. ^{22,23} In vitro, MSCs release IL-6, IL-7, IL-8, IL-11, IL-12, IL-14, IL-15. ²⁴ Self-renewal potential²⁵ and multipotency are MSCs's hallmarks. 12 Bone marrow-derived mesenchymal stem cells (BM-MSCs) are simply isolated from a small aspirate of bone marrow and are able to proliferate ex vivo and generate a variety of cell types.²⁶ In vitro, MSCs can proliferate and differentiate into a desired cell lineage using a specific medium²⁷ with a mixture of growth factors and other elements.²⁸ For instance, if MSCs cultured the presence of dexamethasone, glycerophosphate and ascorbic acid, they will differentiate to osteoblasts.²⁶ Moreover, following implantation of cultured and expanded MSCs into the damaged tissues, they are able to differentiate into mature cells.²⁸ Additionally, MSCs which undergoes genetic modification in vitro can release particular growth factors and cytokines subsequent to tissue implantation.²⁹

These last characteristics make MSCs a very promising candidate for clinical applications of autograft regenerating therapies in tissues damaged by trauma, aging or acute and chronic diseases.² Furthermore, MSCs are appropriate therapeutic tool to repair large bone defects 30,31 and maxillofacial skeletal reconstruction;³² as well as cell replacement skeletal reconstruction; as well as tell replacement therapy in diabetes, 33 spinal cord injury, 34 cardiovascular, 35 neurological, 34 pulmonary 36 and immunological diseases. 23 Over and above, MSCs have a supportive function in co-transplantation with hematopoietic stem cells^{17,18,37,38} by secreting stromal-derived factor-13 Flt-3 ligand and stem cell factor, ²⁴ along with expressing extra-cellular matrix proteins including fibronectin, Laminin and vimentin³⁹ which have a crucial role in HSC homing in bone marrow niche. 38,40 One of the most important properties that make MSCs an excellent tool for cellbased therapeutic strategies is their ability to escape from immune rejection; therefore, HLA-matching is not that much important for their implantation and HLA-mismatched donors can be selected too. 22,23 MSCs comprise a remarkably rare population of unfractionated bone marrow and most tissues like fat.²³ It is absolutely difficult to determine the exact number of MSCs in bone marrow, due to different techniques of bone marrow aspiration and MSC isolation. However, it is estimated that MSCs are about 0.001% of mononucleated cells in BM,³⁰ although the number of them decrease with age. 41 For

clinical purposes we need a large-scale of MSCs, therefore amplification procedure must be done to generate plenty of cells.

Optimized condition for MSCs expansion consist of low glucose α-DMEM (Dulbecco's Modified Eagle Medium) as basal media, 10% fetal bovine serum (FBS) as protein supplement supporter and penicillin/streptomycin solution to prevent bacterial contaminations. As recommended by Meuleman et al, enriching media with some synthetic growth factors or anti-oxidants will decrease the instance between MSCs isolation and clinical application.⁴² Nowadays, some guidelines suggest to avoid utilize animal derivatives like FBS, synthetic growth factors and allogeneic materials for therapeutic application since they raise the risks of pathogen transmission, infection and immune responses. Hence, autologous alternatives like platelet lysate(PL), 43,44 platelet rich plasma(PRP) and platelet rich fibrin(PRF) 48-50 can be employed. These three products are rich in platelets⁵¹ which contain a variety of growth factors including platelet-derived growth factor (PDGF), fibroblast growth factor (FGF), insulin-like growth factor (IGF), transforming growth factor (TGFb), platelet factor 4 (PF-4), and platelet-derived epidermal growth factor (PDEGF).⁵² These growth factors enhance and accelerate MSCs proliferation in vitro and in vivo. 45,53

Conclusion

MSCs are capable to expand in vitro and differentiate into a variety of cell lineages either in vitro or in vivo after implantation. 4-6 As a result they can be considered as a hopeful tool for cell-based replacement therapy. 26-33 For this purpose, first they should be expanded in vitro to reach an adequate number for clinical approaches. Conventionally we use FBS for their proliferation in cell culture medium.39 Since FBS is a bovine derived additive and may cause infectious, prion transmission or may raise immune responses; it cannot be an appropriate supplement for cell culture medium when transplantation expanded MSCs is our final intention.40 Because platelets have a variety of growth factors which may enhance MSCs proliferation, platelet rich products would be an excellent alternative for FBS replacement. 49 Nowadays using autologous platelet rich products including platelet lysate(PL), ^{43,44} platelet rich plasma(PRP) ⁴⁵⁻⁴⁷ and platelet rich fibrin(PRF) ⁴⁸⁻⁵⁰ for MSCs expansion become more general.⁵³ Employing autologous platelet rich products for MSCs expansion is a convenient, non-toxic, safe and cheap therapeutic method that promotes using MSCs for cell therapy.

Conflict of Interest

The authors report no conflicts of interest.

References

- 1. Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, et al. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999;284(5411):143-7.
- 2. Bianco P, Riminucci M, Gronthos S, Robey PG. Bone marrow stromal stem cells: nature, biology, and potential applications. *Stem Cells* 2001;19(3):180-92.
- 3. Ponomaryov T, Peled A, Petit I, Taichman RS, Habler L, Sandbank J, et al. Induction of the chemokine stromal-derived factor-1 following DNA damage improves human stem cell function. *J Clin Invest* 2000;106(11):1331-9.
- 4. Omidkhoda A, Mozdarani H, Movasaghpoor A, Fatholah AA. Study of apoptosis in labeled mesenchymal stem cells with superparamagnetic iron oxide using neutral comet assay. *Toxicol In Vitro* 2007;21(6):1191-6.
- 5. Caplan AI. Mesenchymal stem cells. *J Orthop Res* 1991;9(5):641-50.
- 6. Owen M, Friedenstein AJ. Stromal stem cells: marrow-derived osteogenic precursors. *Ciba Found Symp* 1988;136:42-60.
- 7. Majumdar MK, Thiede MA, Mosca JD, Moorman M, Gerson SL. Phenotypic and functional comparison of cultures of marrow-derived mesenchymal stem cells (MSCs) and stromal cells. *J Cell Physiol* 1998;176(1):57-66.
- 8. Horwitz EM, Le Blanc K, Dominici M. Clarification of the nomenclature for MSC: The International Society for Cellular Therapy position statement. *Cytotherapy* 2005;7:393-5.
- 9. Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini F, Krause D, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 2006;8(4):315-7.
- 10. Friedenstein AJ, Petrakova KV, Kurolesova AI, Frolova GP. Heterotopic of bone marrow. Analysis of precursor cells for osteogenic and hematopoietic tissues. *Transplantation* 1968;6(2):230-47.
- 11. Friedenstein AJ, Deriglasova UF, Kulagina NN. Precursors for fibroblasts in different populations of hematopoietic cells as detected by the in vitro colony assay method. *Exp Hematol* 1974;2:83-92.
- 12. Friedenstein AJ, Gorskaja JF, Kulagina NN. Fibroblast precursors in normal and irradiated mouse hematopoietic organs. *Exp Hematol* 1976;4(5):267-74.
- 13. Arai F, Ohneda O, Miyamoto T, Zhang XQ, Suda T. Mesenchymal stem cells in perichondrium express activated leukocyte cell adhesion molecule and participate in bone marrow formation. *J Exp Med* 2002;195(12):1549-63.
- 14. Im GI, Shin YW, Lee KB. Do adipose tissuederived mesenchymal stem cells have the same

- osteogenic and chondrogenic potential as bone marrow-derived cells? *Osteoarthr Cartilage* 2005;13:845-53.
- 15. Campagnoli C, Roberts IA, Kumar S. Identification of mesenchymal stem/progenitor cells in human first-trimester fetal blood, liver, and bone marrow. *Blood Rev* 2001:98:2396-402.
- 16. Hardy S, Maltman D, Przyborski S. Mesenchymal stem cells as mediators of neural differentiation. *Curr Stem Cell Res Ther* 2008;3(1):43-52.
- 17. Horwitz EM, Prockop DJ, Fitzpatrick LA. Transplantability and therapeutic effects of bone marrow-derived mesenchymal cells in children with osteogenesis imperfecta. *Nat Med* 1999;5:309-13.
- 18. Koc ON, Gerson SL, Cooper BW, Dyhouse SM, Haynesworth SE, Caplan AI, et al. Rapid hematopoietic recovery after coinfusion of autologous-blood stem cells and culture-expanded marrow mesenchymal stem cells in advanced breast cancer patients receiving high-dose chemotherapy. *J Clin Oncol* 2000;18(2):307-16.
- 19. Arthur A, Zannettino A, Gronthos S. The therapeutic applications of multipotential mesenchymal/stromal stem cells in skeletal tissue repair. *J Cell Physiol* 2009;218(2):237-45.
- 20. Soncini M, Vertua E, Gibelli L, Zorzi F, Denegri M, Albertini A, et al. Isolation and characterization of mesenchymal cells from human fetal membranes. *J Tissue Eng Regen Med* 2007;1(4):296-305.
- 21. Schallmoser K, Bartmann C, Rohde E, Reinisch A, Kashofer K, Stadelmeyer E, et al. Human platelet lysate can replace fetal bovine serum for clinical-scale expansion of functional mesenchymal stromal cells. *Transfusion (Paris)* 2007;47(8):1436-46.
- 22. Siegel G, Schafer R, Dazzi F. The immunosuppressive properties of mesenchymal stem cells. *Transplantation* 2009;87(9 Suppl):S45-9.
- 23. Dazzi F, Marelli-Berg FM. Mesenchymal stem cells for graft-versus-host disease: close encounters with T cells. *Eur J Immunol* 2008;38(6):1479-82.
- 24. Horwitz EM, Maziarz RT, Kebriaei P. MSCs in hematopoietic cell transplantation. *Biol Blood Marrow Transplant* 2011;17(1 Suppl):S21-9.
- 25.Moriscot C, De Fraipont F, Richard MJ, Marchand M, Savatier P, Bosco D, et al. Human bone marrow mesenchymal stem cells can express insulin and key transcription factors of the endocrine pancreas developmental pathway upon genetic and/or microenvironmental manipulation in vitro. *Stem Cells* 2005;23(4):594-603.

- 26. Takamine Y, Tsuchiya H, Kitakoji T, Kurita K, Ono Y, Ohshima Y, et al. Distraction osteogenesis enhanced by osteoblastlike cells and collagen gel. *Clin Orthop Relat Res* 2002(399):240-6.
- 27.Jiang Y, Jahagirdar BN, Reinhardt RL, Schwartz RE, Keene CD, Ortiz-Gonzalez XR, et al. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature* 2002;418(6893):41-9.
- 28. Caplan AI. Review: mesenchymal stem cells: cell-based reconstructive therapy in orthopedics. *Tissue Eng* 2005;11(7-8):1198-211.
- 29. Colleoni S, Donofrio G, Lagutina I, Duchi R, Galli C, Lazzari G. Establishment, differentiation, electroporation, viral transduction, and nuclear transfer of bovine and porcine mesenchymal stem cells. *Cloning Stem Cells* 2005;7(3):154-66.
- 30. Kitoh H, Kitakoji T, Tsuchiya H, Mitsuyama H, Nakamura H, Katoh M, et al. Transplantation of marrow-derived mesenchymal stem cells and platelet-rich plasma during distraction osteogenesis--a preliminary result of three cases. *Bone* 2004;35(4):892-8.
- 31. Prockop DJ. Marrow stromal cells as stem cells for nonhematopoietic tissues. *Science* 1997;276(5309):71-4.
- 32. Schilephake H. Bone growth factors in maxillofacial skeletal reconstruction. *Int J Oral Maxillofac Surg* 2002;31(5):469-84.
- 33. Li L, Xie T. Stem cell niche: structure and function. *Annu Rev Cell Dev Biol* 2005;21:605-31.
- 34. Parr AM, Tator CH, Keating A. Bone marrow-derived mesenchymal stromal cells for the repair of central nervous system injury. *Bone Marrow Transplant* 2007;40(7):609-19.
- 35. Amado LC, Saliaris AP, Schuleri KH, St John M, Xie JS, Cattaneo S, et al. Cardiac repair with intramyocardial injection of allogeneic mesenchymal stem cells after myocardial infarction. *Proc Natl Acad Sci U S A* 2005;102(32):11474-9.
- 36. Rojas M, Xu J, Woods CR, Mora AL, Spears W, Roman J, et al. Bone marrow-derived mesenchymal stem cells in repair of the injured lung. *Am J Respir Cell Mol Biol* 2005;33(2):145-52.
- 37. Delalat B, Pourfathollah AA, Soleimani M, Mozdarani H, Ghaemi SR, Movassaghpour AA, et al. Isolation and ex vivo expansion of human umbilical cord blood-derived CD34+ stem cells and their cotransplantation with or without mesenchymal stem cells. *Hematology* 2009;14(3):125-32.
- 38. Akbari AA, Mozdarani H, Akhlaghpoor S, Pourfatollah AA, Soleimani M. Evaluation of the homing of human CD34+ cells in mouse bone marrow using clinical MR imaging. *Pak J Biol Sci* 2007;10(6):833-42.

- 39. Zhang C, Zhang X, Chen XH. Granulocyte-colony stimulating factor-mobilized mesenchymal stem cells: A new resource for rapid engraftment in hematopoietic stem cell transplantation. *Med Hypotheses* 2011;76(2):241-3
- 40. Peled A, Petit I, Kollet O, Magid M, Ponomaryov T, Byk T, et al. Dependence of human stem cell engraftment and repopulation of NOD/SCID mice on CXCR4. *Science* 1999;283(5403):845-8.
- 41.Mueller SM, Glowacki J. Age-related decline in the osteogenic potential of human bone marrow cells cultured in three-dimensional collagen sponges. *J Cell Biochem* 2001;82(4):583-90.
- 42. Meuleman N, Tondreau T, Delforge A, Dejeneffe M, Massy M, Libertalis M, et al. Human marrow mesenchymal stem cell culture: serum-free medium allows better expansion than classical alpha-MEM medium. *Eur J Haematol* 2006;76(4):309-16.
- 43. Lucchini G, Introna M, Dander E, Rovelli A, Balduzzi A, Bonanomi S, et al. Platelet-lysate-expanded mesenchymal stromal cells as a salvage therapy for severe resistant graft-versus-host disease in a pediatric population. *Biol Blood Marrow Transplant* 2010;16(9):1293-301.
- 44. Doucet C, Ernou I, Zhang Y, Llense JR, Begot L, Holy X, et al. Platelet lysates promote mesenchymal stem cell expansion: a safety substitute for animal serum in cell-based therapy applications. *J Cell Physiol* 2005;205(2):228-36.
- 45.Landesberg R, Roy M, Glickman RS. Quantification of growth factor levels using a simplified method of platelet-rich plasma gel preparation. *J Oral Maxillofac Surg* 2000;58(3):297-300; discussion -1.
- 46. Marx RE, Carlson ER, Eichstaedt RM, Schimmele SR, Strauss JE, Georgeff KR. Platelet-rich plasma: Growth factor enhancement for bone grafts. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 1998;85(6):638-46.
- 47.Eppley BL, Woodell JE, Higgins J. Platelet quantification and growth factor analysis from platelet-rich plasma: implications for wound healing. *Plast Reconstr Surg* 2004;114(6):1502-8.
- 48. Anitua E, Sánchez M, Nurden AT, Nurden P, Orive G, Andía I. New insights into and novel applications for platelet-rich fibrin therapies. *Trends biotechnol* 2006;24(5):227-34.
- 49. Dohan DM, Choukroun J, Diss A, Dohan SL, Dohan AJ, Mouhyi J, et al. Platelet-rich fibrin (PRF): A second-generation platelet concentrate. Part I: Technological concepts and evolution. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2006;101(3):e37-44.
- 50. Dohan Ehrenfest DM, De Peppo GM, Doglioli P, Sammartino G. Slow release of growth factors and thrombospondin-1 in Choukroun's plateletrich fibrin (PRF): a gold standard to achieve for

- all surgical platelet concentrates technologies. *Growth Factors* 2009;27(1):63-9.
- 51. Dohan Ehrenfest D, Rasmusson L, Albrektsson T. Classification of platelet concentrates: from pure platelet-rich plasma (P-PRP) to leucocyte- and platelet-rich fibrin (L-PRF). *Trends Biotechnol* 2009;27(3):158-67.
- 52.Harrison P, Cramer EM. Platelet alpha-granules. *Blood Rev* 1993;7(1):52-62.
- 53. Lucarelli E, Beccheroni A, Donati D, Sangiorgi L, Cenacchi A, Del Vento AM, et al. Platelet-derived growth factors enhance proliferation of human stromal stem cells. *Biomaterials* 2003;24(18):3095-100.

