Original Article



# Protective Effect of *Ricinus communis* Leaves Extract on Carbon Tetrachloride Induced Hepatotoxicity in Albino Rats

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## Abstract

Ricinus communis Linn. (Euphorbiaceae) is a soft wooded tree widely prevalent throughout tropics regions of the world which have a warm temperature. In the Indian system of medicine, the leaves, roots and seed oil of this plant have been used for the treatment of inflammation and liver disorders for a long time. In the present study, the protective effects of ethanol extract of *Ricinus communis L*. leaves on carbon tetrachloride-induced liver damage were investigated in rats. Results were compared with those of silymarin, a standard hepatoprotective drug. It was found that an increase in the activities of serum transaminases and the level of liver lipid peroxidation, protein, glycogen and the activities of acid and alkaline phosphatase in liver induced by CCl4 were significantly inhibited by treatment with Ricinus communis ethanolic extract (250/500mg/kg b.wt). In addition, the depletion of glutathione level and adenosine triphosphatase activity observed in the CCl4-induced rat liver were effectively prevented by treatment with Ricinus communis ethanolic extract (250/500mg/kg b.wt). Histopathological examination further confirmed the hepatoprotective activity of *Ricinus communis* ethanol extract when compared with the CCl4-induced control rats. In conclusion, these results indicate that the ethanol extract of *Ricinus communis* ethanolic extract exhibits hepatoprotective action.

*Keywords:* Carbon tetrachloride; Hepatoprotective effect; Lipid peroxidation; *Ricinus communis*; Serum transaminases.

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### 1. Introduction

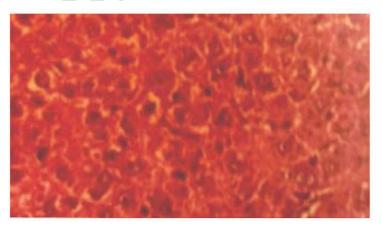
Hepatitis associated with liver cirrhosis has become one of the most prevalent diseases

in the world, which can be induced by virus, alcohol or other toxic chemicals [1]. Carbon tetrachloride, an established hepatotoxin, induces toxicity in rats which closely resembles human cirrhosis [2]. Experimentally, liver diseases have been shown to be produced by the administration of carbon tetrachloride (CCl4), thioacetamide, paracetamol, etc. CCl4-

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induced hepatotoxicity model is frequently used for the investigation of hepatoprotective effect of drugs and plant extracts on experimental animals. CCl4 hepatotoxicity depends on the reductive dehalogenation of CCl4 catalyzed by CYP450 in the liver cell endoplasmic reticulum leading to the generation of an unstable complex of •CCl3 radical. This trichloromethyl radical reacts rapidly with O2 to yield tricholomethyl peroxy radical which is reported as a highly reactive species. This free radical attacks microsomal lipids leading to their peroxidation and also covalently binds to microsomal lipids and proteins ultimately initiating a site of secondary biochemical processes which is the ultimate cause of pathological consequences of CCl4 metabolism [3]. Despite extensive research in the medicine, no drug in the modern system of medicine can be claimed to be effective to cure liver disorders. which in many times become fatal, however, the search for new medicines is still going on [4, 5]. Therefore, many remedies of folk tradition from plant origins are evaluated for its possible antioxidant and hepatoprotective effects against different chemical-induced liver damage in experimental animals.

*Ricinus communis* Linn. (Euphorbiaceae) is a soft wooded tree widely prevalent throughout tropics regions of the world which have a warm temperature [6]. In the Indian system of medicine, the leaves, roots and seed oil of this plant have been used for the treatment of inflammation and liver disorders for a long time [7]. They have been found to be hepatoprotective against galactosamineinduced hepatic damage [8], hypoglycemic [9], laxative [10], diuretic [11] and antibacterial effects [12]. The leaves of R. communis found to contain flavanoids like kaempferol-3-0-beta-D-rutinoside and kaempferol-3-0-beta-D-xylopyranoid and tannins [13, 14]. However, scientific studies on its utility in damaged liver are few and the aim of the present study was to confirm the hepatoprotective effect of *Ricinus communis* extract against CCl4-induced hepatic injury in rats. In addition, its hepatoprotective effect was compared to the effect of silymarin which is known to be effective against CCl4-or acetaminophen-induced liver damages [15].



**Figure 1.** Photomicrographies of paraffin-embedded rat liver. A: Normal Architecture of the liver cell was seen in olive oil alone treated (normal) rats; B: Few hepatocytes showing increased cytoplasmic staining and infiltration of mono nuclear cells around the portal triad and in the lobule were observed in liver of CCl4 -induced rats; C: Liver showing minimal inflammatory cellular filtration was observed in Ricinus communis ethanol extract (250 mg/kg b.wt) treated CCl4-induced rats; D: Liver showing marked improvement to normal architecture was observed in Ricinus communis ethanol extract (500 mg/kg b.wt) treated CCl4-induced rats; E: Liver returned to almost normal architecture was observed in silymarin (50 mg/kg b.wt) treated CCl4-induced rats. F: Normal Architecture of the liver cell was seen in Ricinus communis ethanol extract (500mg/kg b.wt) alone treated rats. **Figure 1**A.

 Table 1. Activities of aspartate transaminase (AST) and alanine transaminase (ALT) in serum of normal and experimental rats.

|               | 250           | 0mg/kg/b.wt)            | extract<br>500mg/kg/b.wt)        | 50mg/kg/b.wt)  | extract 500<br>mg/kg/b.wt)   |
|---------------|---------------|-------------------------|----------------------------------|--|--|
| 5.77 237.0±4. | 64 a* 60.     | .0±5.21 b*              | 55.0±5.21 b*                     | 54.9±5.2 b*  | 46.3±3.94  |
| 7.94 120.4±21 | .85 a* 65.    | .2±6.91 b*              | 60.5±6.91 b*                     | 59.5±7.64 b*   | 57.2±6.75  |
|               | 7.94 120.4±21 | 7.94 120.4±21.85 a* 65. | 7.94 120.4±21.85 a* 65.2±6.91 b* | 5.77         237.0±4.64 a*         60.0±5.21 b*         55.0±5.21 b*           7.94         120.4±21.85 a*         65.2±6.91 b*         60.5±6.91 b* | 5.77         237.0±4.64 a*         60.0±5.21 b*         55.0±5.21 b*         54.9±5.2 b* |

vs. Group I (control); b: Group II vs. Group III (CCl4-induced +Ricinus communis extract 250mg/kg/b.wt); Group IV (CCl4-induced +Ricinus communis extract 500mg/kg/b.wt); Group IV (CCl4-induced + silymarin 50mg/kg/b.wt); The symbol represents the stastical significance at: \*p<0.05.

### 2. Material and Methods

#### 2.1. Animals

Male albino rats of Wistar strain (140±10 g b.w.) were obtained from the Tamil Nadu Veterinary College, Chennai, India. They were acclimatized for a week in a light and temperature-controlled room with a 12 hr dark-light cycle and fed with commercial pelleted feed (Water 8.9%, Protein 25.4%, Lipid 4.4%, Carbohydrate 50.3%, Ash 6.9%, and Crude fiber 4.1%) from Hindustan Lever Ltd. (Mumbai, India) and water was made freely available. The animals used in this study were treated and cared for in accordance with the guidelines recommended by the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), Government of India, Ministry of Culture, Chennai, India. Experimental protocol was approved by the departmental ethical committee.

### 2.2. Preparation of the extract

Fresh leaves of R. communis were collected from rural areas around Chennai, India, and it was authenticated by Chief Botanist, VIT University, Vellore, India. The leaves were dried at the room temperature. The dried and powdered leaves of R. communis (1 kg) were extracted with 80% ethanol at room temperature for 24 hr. It was then filtered and the filtrate was evaporated and dried in a vacuum dessicator. The percentage yield of the dried extract was 15% from the initial raw material. This dried extract and a reference drug, silymarin, Micro Labs, Tamil Nadu, India were dissolved in water and administered to the animals by oral intubation method, respectively.

#### 2.3. Experimental Design

In this study, all group of rats except group I and group VI (negative control) received CCl4 once only. The animals were divided

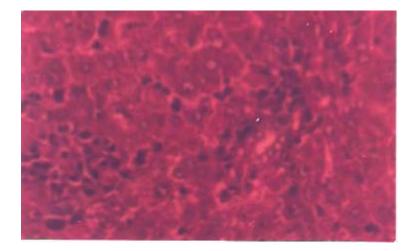


Figure 1B.

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| Parameters Group –I               | Group –II          | Group –III           | Group -IV        | Group-V            | Group -VI         |
|-----------------------------------|--------------------|----------------------|------------------|--------------------|-------------------|
| (IU/L) (Control)                  | (CCl4-induced)     | (CCl4-induced +R.    | (CCl4-induced    | (CCl4-induced      | (R.               |
|                                   |                    | communis extract     | +R. communis     | + silymarin        | communis          |
|                                   |                    | 250mg/kg/b.wt)       | extract          | 50mg/kg/b.wt)      | extract 500       |
|                                   |                    |                      | 500mg/kg/b.wt)   | 1                  | mg/kg/b.wt)       |
| ALP ( $\mu$ moles 9.56± 0.33      | 24.81±1.02a*       | 9.35± 0.55 b*        | 10.0± 1.01 b*    | 11.2 ±1.5 b*       | $10.4 \pm 1.05$   |
| of phenol/min                     |                    |                      |                  |                    |                   |
| mg protein                        |                    |                      |                  |                    |                   |
| ACP ( $\mu$ moles 5.12 $\pm$ 0.32 | 9.37± 0.56 a*      | 6.32± 0.45 b*        | 6.21± 0.39 b*    | 5.8 ±0.41 b*       | $5.4 \pm 0.37$    |
| of phenol/min/                    |                    |                      |                  |                    |                   |
| mg of protein)                    |                    |                      |                  |                    |                   |
| Total ATPase $2.9 \pm 0.12$       | 1.35± 0.10 a*      | 2.6 ±0.09 b*         | 2.1 ±0.22 b*     | 2.45± 0.19 b*      | $2.32 \pm 0.15$   |
| (mg Pi/100g/min)                  |                    |                      |                  |                    |                   |
| Glycogen $11.00\pm 1.26$          | 7.50 ±0.51 a*      | 10.00± 0.63 b*       | 10.33 ±1.03 b*   | 10.50 ±0.83 b*     | $9.83 \pm 1.32$   |
| (mg/100g)                         |                    |                      |                  |                    |                   |
| Protein 185.1±19.2                | 155.2 ±10.9 a*     | $190.5 \pm 14.25b^*$ | 192.5± 18.9 b*   | 189.2±18.5 b       | * 191.2 18.2      |
| (mg/100 g)                        |                    |                      |                  |                    |                   |
| Lipid $164.82 \pm 8.15$           | 237.33± 9.11 a*    | 172.28± 7,89 b*      | 186.66± 16.32 b* | 169.21± 11.63 b* 1 | $166.83 \pm 5.84$ |
| peroxidation (nmol MDA/           |                    |                      |                  |                    |                   |
| mg protein)                       |                    |                      |                  |                    |                   |
| Glutathione $9.57 \pm 0.17$       | $7.52 \pm 0.44$ a* | 9.82± 0.19 b*        | 9.90± 0.21 b*    | 8.90 ± 0.16 b*     | $9.01 \pm 0.17$   |
| (µmol/g)                          |                    |                      |                  |                    |                   |

| Table 2. Effect of R  | communis extract | on CCl4-induced liver | damage in rats |
|-----------------------|------------------|-----------------------|----------------|
| Table 2. Lincer of A. | communis canact  |                       | uamage m rats. |

The values are expressed as mean  $\pm$  S.D.; Each group consists of six animals.; Comparisons were made as follows: a: Group II (CCl4-induced) vs. Group I (control); b: Group II vs. Group III (CCl4-induced +*Ricinus communis* extract 250mg/kg/b.wt); Group IV (CCl4-induced +*Ricinus communis* extract 500mg/kg/b.wt); Group-V(CCl4-induced + silymarin 50mg/kg/b.wt); The symbol represents the stastical significance at: \*p<0.05.

into six groups of six animals each and were treated as follows:

• Group I: normal control (vehicle olive oil only)

• Group II: CCl4 (3 ml/kg/b.wt i.p. in olive oil (1:1, v/v), single administration).

• Group III: CCl4 + *R. communis* ethanol extract (250 mg/kg/b.wt, p.o. for five days).

• Group IV: CCl4 + *R. communis* ethanol extract (500 mg/kg/b.wt, p.o. for five days).

• Group V: CCl4 + silymarin (50

mg/kg/b.wt, p.o. for five days).

• Group VI: negative control (*R. communis* ethanol extract 500 mg/kg/b.wt, p.o. for five days).

At the end of the experimental period, all the animals were sacrificed under ether anesthesia. Blood and liver samples were collected. The blood was allowed to clot for 30 min.; serum was separated by centrifuging at 37 °C which was used for biochemical estimations.

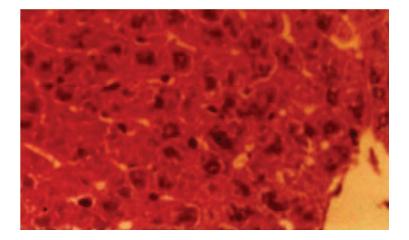
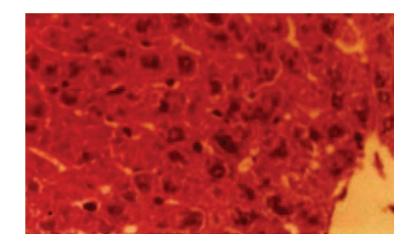


Fig 1C



### Fig 1D.

2.4. Assessment of hepatoprotective activity The activities of serum glutamate pyruvate transaminase (SGPT) and serum glutamate oxaloacetate transaminase (SGOT) were assayed by the method of Reitman and Frankel [16]. Estimation of activities of alkaline phosphatase (ALP), acid phosphatase (ACP) by the method of King [17], and total ATPases [18], protein [19], and glycogen content [20], lipid peroxide [21] and glutathione level [22] were also carried out in liver tissue to assess the damage caused by CCl<sub>4</sub>.

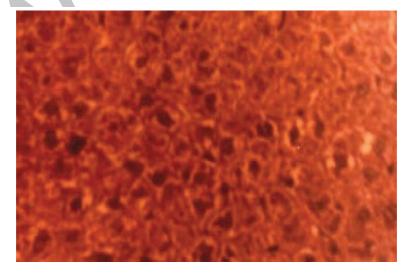
# 2.5. Histopathological studies

Immediately after sacrifice, a portion of the liver was fixed in 10% formalin. The washed

tissue was dehydrated in descending grades of isopropanol and finally cleared in xylene. The tissue was then embedded in molten paraffin wax and cut into 5 µm thick sections in a rotary microtome. The sections were then stained with haematoxylin and were studied for histopathological changes, i.e. necrosis, fatty changes and lymphocyte infiltration. Histological damages were scored as follows: 0; absent; +: mild; ++: moderate; +++: severe.

## 2.6. Statistical analysis

The results were expressed as mean±S.D. and statistical analysis was performed using ANOVA to determine significant differences between the groups, followed by Student's



Newman-Keul's test. p < 0.05 was the implied significance.

### 3. Results

Administration of  $CCl_4$  to rats caused a significant elevation in serum aspartate, and alanine aminotransmainases (Table 1). However, the elevated SGOT and SGPT activity observed in  $CCl_4$ -treated rats was found to be significantly decreased by prior administration of *Ricinus communis* ethanol extract.

Alterations in the activities of acid phosphatase, alkaline phosphatase, adenosine triphosphatase, glycogen and protein content, lipid peroxidation and glutathione level of liver after single exposure to  $CCl_4$  were expressed in Table 2. Reduced glutathione and enhanced lipid peroxide level were seen in the CCl<sub>4</sub>-treated group, whereas the drug treated groups showed a significant rise in glutathione level with the reduction in lipid peroxidation level, when compared to the CCl<sub>4</sub>-treated group. In CCl<sub>4</sub>-treated rats, the activities of acid phosphatase and alkaline phosphatase were found to be increased, whereas a decreased glycogen and protein content and adenosine triphosphatase activity was observed when compared to the normal control groups. Administration of R.

*communis* ethanol extract to the  $CCl_4$ -treated rats modulates the above alterations to near normal levels.

As shown in Figure 1A-F and Table 3, liver sections from the control group (Figure 1A) and *R. communis* ethanol extract (500 mg/kg. b.wt) alone treated rats (Figure 1F) showed normal lobular architecture and normal hepatic cells. The liver section from animals given  $CCl_4$  showed increased cytoplasmic staining and infiltration of mono nuclear cells around the portal triad and in the lobule (Figure 1B). The histological pattern of the liver of the  $CCl_4$ -induced rats treated with *R. communis* ethanol extract (Figure 1C and 1D) and silymarin (Figure 1E) showed marked improvement and return to normal architecture, respectively.

# 4. Discussion

 $CCl_4$ -induced hepatic injury is often used as a model for hepatoprotective drug screening and the extent of the hepatic damage is assessed by the level of release cytoplasmic transaminases (SGOT and SGPT) in circulation [23]. The present study revealed a significant increase in the activities of SGOT and SGPT in serum (Table 1) and elevated acid and alkaline phosphatases (ALP) (Table 2) in liver on exposure to  $CCl_4$ , indicating

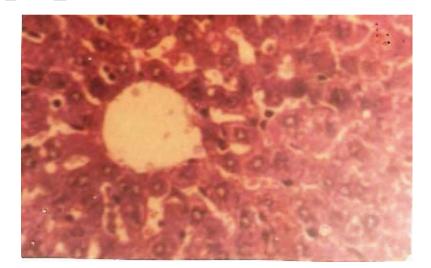


Fig 1F.

| Microscopic observation   |          |               |                            |
|---|----------|---------------|----------------------------|
| Groups  | Necrosis | Fatty changes | Infiltration of lymphocyte |
| Control   | 0        | 0             | 0                          |
| CCl <sub>4</sub> (3ml/kg/b.wt, i.p.)  | +++      | +++           | +++                        |
| CCl <sub>4</sub> (3ml/kg/b.wt, i.p.) +  | ++       | ++            | ++                         |
| <i>R. communis</i> ethanol<br>extract (250mg/kg/b.wt, p. o)<br>CCl <sub>4</sub> (3ml/kg/b.wt, i.p.) + | +        | +             | +                          |
| <i>R. communis</i> ethanol<br>extract (500mg/kg/b.wt, p. o)<br>CCl <sub>4</sub> (3ml/kg/b.wt, i.p.) + |          |               |                            |
| silymarin (50mg/kg/b.wt, p. o.)   | +        | 0             | +                          |
| Negative control ( <i>Ricinus</i>   | +        | 0             | +                          |
| <i>communis</i> ethanol extract 500mg/kg/b.wt, p. o   |          |               |                            |

Table 3. Effect of the *R. communis* ethanol extract on histopathological damages induced by CCl<sub>4</sub> injection in rats.

0; absent; +: mild; ++: moderate; +++: severe; rats were injected (i.p.) with *R. communis* ethanol extract for five days after the injection of CCl4 (3ml/kg/b.wt, i.p.). Histopathological damages were assessed as explained under materials and methods.

considerable hepatocellular injury. Administration of *R. communis* ethanol extract at two different dose levels attenuated the increased levels of marker enzymes, observed in  $CCl_4$ -induced rats. The hepatoprotective effect of the *R. communis* ethanol extract was further supported by the limited extent of histological changes (Figure 1), in *R. communis* treated  $CCl_4$ -induced rats. This is in agreement with the commonly accepted view that serum levels of transaminases return to normal with healing of hepatic parenchyma and the regeneration of hepatocytes [24].

CCl<sub>4</sub> is believed to be metabolized by microsomal CYP450 in the liver to a highly reactive trichloromethyl free radical, which can start a chain of reactive free radical formation resulting in peroxidation of lipids and damage to proteins and cell components [5]. The level of lipid peroxide is a measure of membrane damage and alterations in structure and function of cellular membranes. Glutathione protect cells against electrophilic attacks provided by xenobiotics such as free radicals and peroxides. The elevation of MDA levels, which is one of the end products of lipid peroxidation in the liver tissue, and reduction of hepatic GSH levels are important indicators in CCl<sub>4</sub>-intoxicated rats [25]. In this study, enhanced liver lipid peroxidation and reduced glutathione level were seen in Group II  $CCl_4$  treated rats (Table 2). However, *R. communis* extract treatment, prevented glutathione depletion with reduction in lipid peroxidation level in  $CCl_4$ -induced rats when compared to  $CCl_4$ -treated control group. The *R. communis* extract with the potentiality to scavenge the free radicals contains flavanoids and tannins. Flavanoids and tannins have been reported to have anti-peroxidative effects [26]. Moreover recently *R. communis* root extract was also found to have free radical scavenging activity [27]. Thus, this finding suggests that the *R. communis* extract was effective in bringing about functional improvement of hepatocytes.

The decreased hepatic glycogen content, and adenosine triphosphatase activity observed in  $CCl_4$  induced rats (Table 2) in our study agrees with previous reports [28]. Glycogen is the main source of energy in the liver [29], so the reduced glycogen content observed in  $CCl_4$  induced rats may be due to excess requirement of energy in liver.  $CCl_4$  intoxication is capable of initiating cell injury and cellular sites for free radical generation include mitochondria, endoplasmic reticulum and plasma membrane. A significant fall in the activity of adenosine triphosphatase observed in  $CCl_4$  induced rats may be due to the structural and functional disorganization of mitochondria assembly. Upon *R. communis* extract treatment to  $CCl_4$  induced rats, the above said changes were found to be restored significantly when compared to  $CCl_4$  induced rats.

The ability of a hepatoprotective drug to reduce the injurious effects or to preserve the normal hepatic physiological mechanisms, which have been disturbed by a hepatotoxin, is the index of its protective effects [30]. It is reported earlier that fresh leaves and aqueous extract of R. communis offered protection against CCl<sub>4</sub>-induced hepatic damage in albino rats [31]. In our study, our results demonstrated that the possible hepatoprotective mechanisms of the ethanol extract of *R*. *communis* leaves on CCl<sub>4</sub>-induced hepatic damage in rats. Histopathological examinations also show that R. communis ethanol extract (250/500mg/kg/b.wt) at both levels of dosage offer hepatoprotection. In addition, this plant extract up to an oral dose of 1 g/kg was found to be devoid of any lethal effects and no apparent behavioral change was observed.

### 5. Conclusions

In conclusion, the present study has demonstrated that the ethanolic extract of R. communis has hepatoprotective effect against CCl<sub>4</sub>-induced hepatotoxicity in rats. The beneficial effect of the R. communis ethanol extract may be due to the presence of some flavanoids that may have membrane stabilizing and antiperoxidative effects. Thus, this result suggests that the flavanoids and tannins present in the *R. communis* ethanol extract might efficiently increase the regenerative and reparative capacity of the liver. Although R. communis ethanol extract has comparable hepatoprotective effect with silymarin in our study, clarification of the hepatoprotective mechanism and the active components of the R. communis extract need

### further investigation.

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Conflict of Interest Statement:

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