

Review Article

Plasmapheresis- Review Article

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ABSTRACT

Plasmapheresis, which is defined as the removal of plasma, can be either “adjusted plasma” or “exchange of plasma”. The former is defined as selective withdrawal of certain (un)-pathological plasma components in different ways such as perfusion and then returning the remained donor plasma to him, the latter is non-selective removal of all components of plasma to provide blood products for injection into patients or to be used as the input of blood transfusion refinery or to remove the pathogen contained plasma before compensating for the volume losses with an equal volume of plasma or more commonly, replacing plasma with a substitute fluid (colloid or crystalloid) such as albumin. Plasmapheresis was divided generally into two groups:

1- Plasma products by donor plasmapheresis

2- Therapeutic plasmapheresis

Therapeutic plasma exchange or TPE are often attributed to plasma that exit from the body of patient then compensated by any kind of replacement fluid volumes to support neurmolemic situation of patients. Plasmapheresis is currently used as a therapeutic modality in a wide array of conditions. Generally, plasmapheresis is used when a substance in the plasma, such as immunoglobulin, is acutely toxic and can be efficiently removed. Myriad conditions fall under this category, including neurologic, hematologic, metabolic, dermatologic, rheumatologic, and renal diseases, as well as intoxications, that can be treated with plasmapheresis.

Keywords: Plasmapheresis, Exchange the Plasma

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Introduction

Aphaeresis is derived from a Greek word and it means removing or separating (1).

Hemapheresis means bleeding from a donor and removing certain components and returning the remaining components to him (2-3).

There are two kinds of Hemapheresis (4):

1- Cytapheresis is withdrawal of the whole blood cells that have different titles by the type of cell. If it is separated from leukocytes, it is known as "Leukopheresis", if separated from red blood cells, it is "Erythropheresis" and if separated from platelets it is as "Plateletpheresis" or "Plasmapheresis".

2- Plasmapheresis, which is defined as the removal of plasma, can be either "adjusted plasma" or "exchange of plasma". The former is defined as selective withdrawal of certain (un)-pathological plasma components in different ways such as perfusion and then returning the remained donor plasma to him, the latter is non-selective removal of all components of plasma to provide blood products for injection into patients or to be used as the input of blood transfusion refinery or to remove the pathogen contained plasma before compensating for the volume losses with an equal volume of plasma or more commonly, replacing plasma with a substitute fluid (colloid or crystalloid) such as albumin (5-7).

History

Hendon (1902s) was the first to conduct research on the therapy method of Hemaphairesis by withdrawing blood from animals to be washed and reinjected. Fleig (1909) after Hendon used this method for the treatment of human uremia (2). Abel *et al.* (1914) have explained the removal of plasma technique through the treatment of poisoned dogs whose kidneys had been removed. To that effect, in a bid to get more anti-serum from immunized horses for serum therapy, they cor-

rectly did plasmapheresis while preventing blood losses (3, 4).

At the end of 1940s and early 1950s, simultaneously the appearance of plastic advanced blood bag and advanced refrigerated centrifuge, hemapheresis was used more broadly as a treatment method. In 1952, Carifols -Lucas used the manual plasmapheresis procedure for the preparation of blood products. Thus, whole blood has been transfused from a donor, and then the plasma was separated from red blood cells by using centrifuge, and red blood cells after a week bleeding was returned to the donor (1, 8-10). He also reported the usefulness of pheresis in the patients' therapy with hypertension disease. In 1950s, Dr. Edwin Cohn had established the using a continuous flow centrifuge with more than 60 years since it was invented by De Laval for use in the dairy industry to prepare blood products and improve the effectiveness of a therapeutic plasmapheresis during the Second World War. These devices can be separated an on-line blood component in a closed system and in this system, the obtained products were sterile (1, 11-13).

In 1962, Judson provided a device for separation of leukocytes from the whole blood by supporting IBM and NCI and managed to produce 2,990 units of IBM successfully. This device consists of disposable plastic tubes that were sterile previously, and worked with a steady stream of closed system and it was the first device that enables functionality to provide the processing of large volumes of blood. This device is capable of exchanging plasma, collecting platelets, granulocytes, lymphocytes and erythrocytes (1, 14-17).

CS3000 were accessible commercially in the early 1980s, produced by Fenwall. It is a continuous -flow device consisting of two rotating chamber with a special form. In 1988, Cobe Company began selling a new device as SPECTRA. This type of devices is automatically

equipped with computer systems that can platelet aphaeresis with minimal contamination of white blood cell. Moreover, SPECTRA have been moving for medical procedures affairs that are easily transferred to the patient's room (18).

Subsequent development of the basic principles established by the people, eventually leading to the construction of several kinds of semi-automatic separation devices for clinical applications in blood that the wider use of plasmapheresis was followed as a method of treatment. Continuation of this trend depends on the development of consumer devices and software whatever needs to be more selective separation of blood components, and other ingredients can be returned to the donor or patient (18, 19).

Plasma pheresis was divided generally into two groups (1, 20):

Plasma products by donor plasmapheresis

Therapeutic plasma pheresis

Plasma Products by Donor Plasmapheresis Description

The donor plasmapheresis interpreted the donated plasma by volunteers to be used in transfusion or conversion to some specific products in plasma processing center (21).

Complications of Donor Plasma

The donor plasma is far less than the whole donor blood cell with the rate of adverse reactions in donors' blood components (22-23).

AFFP: apheresis fresh frozen plasma; 0.5% to 2.5%;

Rare: < 0.5%; very rare: < 0.10%; Frequent: 5% to 20%

Adverse reactions are more in first time donors and it has less vasovagal responses (24).

In contrary the whole blood donors, there are two reasons to reduce the blood volume (hypovolemia) that it has less reduction of blood volume than other donors in donors' plasmapheresis. First, an ejecting of the whole blood volume is often less than whole blood

donors during donor plasmapheresis and or the loss volume will be compensated if it needed alternative and replacement fluids. Second, when plasmapheresis has been done, simultaneously donors have a long relaxation time that it was compared with the whole blood donors and allows to refill the intervascular to the interstitial capillaries with blood (24, 25-27).

There are plasma hematoma and pain of needle in the injection space of the body that is the most common complication in the donor plasma but the incidence of hematoma was not more than whole blood donors.

There are rarely allergic reactions when it was prepared to rub the iodine fluid to skin. It was an important adverse effect associated with anticoagulant citrate when used as anticoagulant in vitro system and it was part of the remaining blood components back to the donor's bloodstream. If donors have a low weight, maybe they have many symptoms of citrate (28-30).

Sometimes there are rare mild reactions to citrate (anesthesia around the mouth, tremor or tingling sensation, tingling, or feeling cold or stuffy nose) and If the patients are faced with these reactions it is not alarming but demonstrated that doctors should control this process to reduce and prevent serious symptoms of citrate toxicity. Sometimes reactions have acute forms due to citrate toxicity (muscle cramps, whole body vibration, nausea, vomiting and tetanus) that are rare in plasmapheresis but potentially have been a serious problem and disorder (31-33).

Therapeutic PlasmaPheresis or Therapeutic Plasma Exchange (TPE)

Description

Therapeutic plasma exchange or TPE are often attributed to plasma that exits from the body of patient then compensated by any kind of replacement fluid volumes to support neurmolemic situation of patients (34). Table 1 shows Indications for Therapeutic Apheresis.

Table 1- Indications for therapeutic apheresis generally accepted for reimbursement by third-party payers (8,9)

Plasma exchange

Ig*G

Myasthenia gravis
Eaton-Lambert syndrome
Goodpasture syndrome
Myeloma with renal failure
Guillain-Barre syndrome
Hemophilia with factor VIII inhibitor
Chronic inflammatory demyelinating
Polyradiculopathy

Immune complex

Glomerulonephritis, rapidly progressive
Rheumatoid vasculitis

Metabolic diseases

Refsum's diseases
Hyperlipoproteinemia, familial
Cholestasis- intractable pruritus

Diseases of unknown cause

Thrombotic thrombocytopenic
Thyroid storm
Scleroderma, refractory
Polymyositis, refractory

Cytapheresis

Acute leukemia-debulking
Hairy cell leukemia- maintenance
Thrombocytosis, symptomatic
Chronic myelogenous leukemia, acute

symptoms

Red cell exchange

Sickle cell diseases

* Immunoglobulin

What features should Pathogenic materials withdrawn by TPE must have?

There are clinical reactions to the withdrawal plasma by TPE, if the pathogen has the following properties (35):

1 – A withdrawal material must be sufficiently large and as big as macromolecules (molecular weight more than 15,000), so that it is not easily

purified by a cheaper techniques such as hemofiltration or unrecoverable high flux hem-dialysis.

2- Relatively removed material has a long half-life so that in vitro removal of such substances into the body is passed much faster than the clearing way of the body called as metabolism.

For example, the IgG with an approximately 21 day's half-life even if it has a treatment with immune-suppressors and stops producing new antibodies after 21 days, its decrease was at 50%.

3- A synthesis of the desired materials has a low speed. Intravascular concentration is often increased and its deletion depends on the speed of their synthesis.

4- Pathogen is an intravascular factor. The ultimate effect of TPE depends on concentration of pathogen, and also the concentration of that substance in any of the vessels and the rate of exchange between these two parts is that if each percentage of both inside and outside of the vessel is greater than the other part and the rate of exchange between the two parts is low, the effectiveness of TPE would be high.

5- The removed materials must be really toxic and resistant to conventional therapy so that its quick elimination from the extracellular fluid by TPE be possible due to the fact that plasma exchange will be time-consuming and costly (35).

At every stage of the TPE process, how much plasma should be removed?

Plasma's volume that should be removed at each stage of TPE and its performance distances, the plasma factor depends on the pathogen (36-38). Once the removed plasma (one volume plasma) has been occurred, an equal volume of plasma clearance was not 100% and about 38% of the material remains in the plasma due to the simultaneous dilution and advanced plasma in the patient during the performance of TPE process by replacement fluids. After exchanging 1.5 volumes of plasma, it remains 22% and after

replacement two volumes are 15% in considered substances (39).

If the high volume of plasma replaced, a smaller fraction of remained substances can be removed until the rate of removing decreased and it is reduced the removed volumes about 1.5-2 equivalent of the plasma volume. It should be noted that the exchange of more than one plasma volume, caused to length the processing time of exchange and decreasing the patient endurance and also increasing the cost (replacing a plasma volume about 1.5-2 hours long and by 2 to 3 equivalent of the volume exchange adds time of the process). According to decreasing the pathogen removal efficiency in volumes greater than 1.5 times of the plasma volume that estimated for each individual and adverse results of increasing the time of performing TPE process caused to increase the volume of exchange, most therapeutic plasma exchange procedures aimed to remove about 1-1.5 plasma volumes in each turns (40-48).

How often should the TPE procedure be performed?

The differences in plasma levels exchange are depends on factors such as the rate of patient

illnesses, the rate of regeneration (re-synthesis) and redistribution (the transfer of desired material to the intravascular) of extra vascular substances (49). For example, the exit of IgG needs to a daily TPE process (due to the high rate of synthesis and entry into the intravascular space outside the vessel) and or in the patient who are awaiting a liver transplant, sometimes TPE is necessary to sustain life every 12 hours and generally, if a amount of new IgG production is low (created during immunosuppressive therapy), that the amount of extra-vascular exchange is about 1-2% in hours against intravascular exchange required 5 times TPE during 7 to 10 days to return 90% of initial body lift immunoglobulin (50-51).

How much plasma exchange is in a TPE process?

General recommendations of AABB for the TPE process indicated that in cases where there is its IgG, the exchange of plasma volume estimated about 1-1.5 equivalent in each person and once every 2-3 days with total exchanging in 3-5 days (52). It should be, of course, noted that the frequency of plasma exchange may change according to the patient’s clinical responses and laboratory testing results. Table 2 shows the final target of plasmapheresis therapy (53).

Table 2- The final target of plasmapheresis therapy (26)

Substance to Remove	Treatment Volume (ml/kg)	Treatment Interval (in hours)	Treatment Endpoint
Autoantibodies	40-60	24-48	Four to six treatments
Immune complexes	40-60	24-48	Treat for response
Paraproteins	40-60	24	Treat for response
Toxins	40-60	24-72	Treat for response
Thrombotic thrombocytopenic purpura/ hemolytic uremic syndrome	40	24	Treat to establish remission
Immunologic rebound	40-60	24-48	Two to three treatment followed by immunosuppressive medication

Which kind of Fluid replacement used in TPE?

Crystalloid fluids, among which the saline fluids are used most widely, would suffice when 500-1000 cc of plasma is removed through a manual plasmapheresis (54). Especially TPE is used to solve hyperviscosity as well. Crystalloid fluids alone is not sufficient when it discharging more than one liter of plasma volume because it caused hypoproteinemia and coagulopathy by prescribing in high volumes in patient.

Therefore, they need to prescribe crystalloid fluid volume for discharging equivalent to 2-3 volumes of plasma that it is for the rapid departure of crystalloid fluids from vessels, and led to hypotension and edema by spreading to the extra-vascular vessels (55-57).

However, it has the necessary effectiveness for un-transmission of diseases and its low cost. Now saline has been used with albumin as some of (30-40%) the fluid replacement in plasma exchange. Additionally it is useful in the preparation of anticoagulant fluid and instruments (58).

Five percent albumin is used mostly in replacement fluid. For most patients, this liquid plays an important role as hyper-oncotic one and forced to flow this pure fluid to intravascular and make a mild anemia. To prevent from mild anemia must be made 5% albumin to 4-4.5 % albumin by saline. Many patients could tolerate to control this disease by exchanging 25-50% of the volume of saline (59).

An important advantage of albumin is impossible transmission of virus agents'. Albumin prescribed easily to patients that it is not required to test the blood group and without needing to melt or prepare it. The patients have rarely been reported an uncommon albumin reactions but pyogenic materials have fever and prekallikrein activated reactions (60-62).

Currently, based on standard method it is replaced with to use 5% albumin fluid (amount of 60-70% the plasma volume) and remained replaced with crystalloid such as normality saline (63).

FFP is one of another replacement colloid fluid that sometimes is used as replacement fluid for patients with TTP disease which needs replacement therapy of special protein plasma in accessing the metalloproteases von Willebrand. In the TTP treatment, it is formed the replacement volumes as an amount of 60-70% FFP and remained for saline (64).

Synthetic colloid fluids such as HES can also be used to replace all or part of the liquid. It is derived from vegetable starch containing scratch of macromolecules that saline is added to the colloidal fluid. An advantage of this fluid is low cost and non- transmission to others. The disadvantages are allergic reactions to HES that can occur in a small number of patients (64-66). Therefore HES used as a replacement fluid in patient with reaction to albumin or FFP and sometimes religious peoples are not tending to access blood derivation by following plasma exchanges (67).

Enrichment of replacement fluids (saline, albumin, HES) can reduce the incidence of hypocalcaemia due to citrate toxicity with calcium. The volume of calcium should be checked in large volumes (1-2 volumes) in plasmapheresis therapy for several sessions especially after each plasmapheresis session when plasma is used as a replacement therapy that have a large amounts of citrate (68). The best time to control the amount of calcium is a few hours after plasma exchange or in the next morning after and before the replacement of plasmapheresis session (69). Table 3 shows advantages and disadvantages of any replacement fluids in TPE.

Table 3- Advantages & disadvantages of any replacement fluids in TPE*

Replacement fluid	Advantages	Disadvantages
Crystalloids	Low cost Hypoallergenic No viral risk	2-3 volumic required Hypo-oncotic No coagulation factors No immunoglobulins
Albumin	Iso-oncotic No contaminating “inflammatory mediators” No viral risk	High cost No coagulation factors No immunoglobulins
Hydroxyethyl starch	Moderate cost Iso-oncotic No contaminating “inflammatory mediators”	No coagulation factors Long-term residual Levels of HES Contraindicated with renal failure Possible coagulopathy
Plasma	Maintins normal levels of: Immunoglobulins Complement Antithrombin Other proteins	Viral transmission risk Citrate load ABO incompatibility risk Allergic reaction Sensitization

* Therapeutic Plasma Exchange

What type of anticoagulant and how much is needed?

Citrate, heparin, or both combinations can be used during plasmapheresis to prevent the blood circulation outside the body. Patient should be carefully evaluated in terms of the ability against citrate or heparin, to determine the best anticoagulant materials. Other considerations include how to assess intravascular volume of patient, the type and volume of replacement fluid, intravenous access and blood flow rate in the catheter (70-72).

Citrate has been accessed in 4 shapes as follows (73):

ACD-A, ACD-B, sodium citrate and concentration of tri-sodium citrate

ACD-A has been prescribed in the amount of 9:1 to 14:1 that including 3% sodium citrate and also used in the determined rate of anticoagulation (WB/ACD) in blood. In contrast to ACD-A,

ACD-B is including 2% sodium citrate that usually used at the rate of 6:1 to 9:1 from WB/ACD to reduce the citrate toxicity risk.

Unlike heparin, citrate has not any quick metabolisation. It has the exact half-life of approximately 90 minutes and thus leads to have systemic anticoagulant effects in patients (74).

When these particular characteristics are replaced with non-plasma fluids during plasmapheresis (e.g. albumin 5% or HES) it is difficult to remove the coagulation factors in plasma and their replacement. Heparin can be used alone or in combination with ACD-A and or ACD-B. When they use the combination of heparin and citrate to create effective anticoagulant effects, heparin is needed less and less ACD volumes. Therefore, this combination reduced the incidence of systemic toxicity citrate anticoagulation due to minimizing the heparin consumes alone and also reduces the whole volumes of the injected fluid during TPE process (75).

In vitro, what kind of tests need during the TPE process?

Laboratory studies are based on the ultimate goal of therapy and pursuing of the reduction factor. Plasmapheresis can be evaluated and studied an early CBC consideration, electrophoresis serum protein, electrolytes and coagulant factors (76). More detailed laboratory analysis is required when desired a large number of low- frequency plasma exchange. We should give several hour chances to body to follow the necessary laboratory TPE tests until to shift extra- and intra-vascular fluids to reach equilibrium and bleeding has to be taken especially in biochemistry (77).

TPE Indications

The appropriate use of therapeutic plasma exchange (TPE) based on two American societies

for aphaeresis (ASFA) and American Association of Blood Bank (AABB) organizations rules are as follows (78-83):

Before reading the table, it is necessary to be familiar with the following concepts:

Group I: The standard acceptable therapy of TPE in first-line for these diseases.

Group II: There is sufficient evidence to suggest the efficacy of plasma exchange therapy as second-line or adjunctive therapy.

Group III: There are inconclusive evidence in efficacy of plasma exchange therapy with uncertain risks in final therapy way.

Group IV: There is lack of efficacy in controlled TPE trials.

Table 4, 5, 6 shows Plasmepheresis Therapy Indications.

Table 4- Plasmepheresis therapy indications

Disease	Procedure	Indication Category
Neurologic disorders		
Chronic inflammatory demyelinating Polyradiculoneuropathy (CIDP)	Plasma exchange	I
Acute inflammatory demyelinating Polyradiculoneuropathy (AIDP or Gullian-Barre syndrome)	Plasma exchange	I
Myasthenia gravis	Plasma exchange	I
Lambert-Eaton myasthenia gravis	Plasma exchange	II
Multiple sclerosis and related disorders	Plasma exchange	III
Acute fulminant central nervous system demyelination		III
Relapsing or progressive	Plasma exchange	III
Paraneoplastic Neurologic syndrome	Immunoabsorption	III
Paraproteinemic polyneuropathies	Plasma exchange	I
Demyelinating polyneuropathy	Immunoabsorption	III
Ig ⁺ G/Ig ⁺ A	Plasma exchange	II
Polyneuropathy with Ig ⁺ M	Immunoabsorption	III
(±Waldenstrom's macroglobulinemia)	Plasma exchange	II
Cryoglobulinemia with polyneuropathy	Plasma exchange	III
Multiple myeloma with polyneuropathy	Plasma exchange	III
POEMS syndrome	Plasma exchange	IV
Systemic (AL) amyloidosis	Plasma exchange	III
Inflammatory myopathies	Leukapheresis	IV
Polymyositis or dermatomyositis	Plasma exchange	III
	Leukapheresis	IV
	Plasma exchange	III
Inclusion- body myositis		
Rasmussen encephalitis	Plasma exchange	III
Stiff-person syndrome	Plasma exchange	III

Syndrome's chorea/pediatric autoimmune neuropsychiatric associated with streptococcal infections (PANDAS)	Plasma exchange	II
Hematologic disease	Plasma exchange (recipient)	II
ABO-incompatible hematopoietic cell transplant	Erythrocytapheresis	II
RErythrocytosis/polycythemia vera	Cytapheresis	I
Leukocytosis and thrombocytosis	Plasma exchange	I
Thrombotic thrombocytopenic purpura	Plasma exchange	I
Post-transfusion purpura	Red cell exchange	I
Sickle cell diseases	Plasma exchange	II
Myeloma/paraproteins/hyperviscosity	Plasma exchange	II
Myeloma/acute renal failure	Plasma exchange	III
Coagulation factor inhibitors	Photopheresis	I
Aplastic anemia/pure red cell aplasia	Leukapheresis	III
Cutaneous T-cell lymphoma	Plasma exchange	III
Hemolytic disease of the newborn	Plasma exchange	III
Platelet alloimmunization and refractoriness	Immunoabsorption	III
Malaria/babesiosis	Red cell exchange	III
Renal and metabolic disease	Plasma exchange	I
Antiglomerular basement membrane antibody disease (Goodpasture's syndrome)	Plasma exchange	II
Rapidly progressive glomerulonephritis	Plasma exchange	III
Hemolytic-uremic syndrome	Plasma exchange	IV
Renal transplantation	Plasma exchange	III
Rejection	Plasma exchange	III
Presensitization	Plasma exchange	III
Recurrent focal glomerulosclerosis	Plasma exchange	III
Heart transplant rejection	Photopheresis	III
Acute hepatic failure	Plasma exchange	III
Familial hyper cholesterolemia	Selective adsorption	I
Overdose poisoning	Plasma exchange	II
Phytanic acid storage disease (Refsum's)	Plasma exchange	III
Autoimmune and rheumatic disease	Plasma exchange	I
Cryoglobulinemia	Plasma exchange	II
Idiopathic thrombocytopenia purpura	Immunoabsorption	II
Raynaud's phenomenon	Plasma exchange	III
Vasculitis	Plasma exchange	III
Autoimmune hemolytic anemia	Plasma exchange	III
Rheumatoid arthritis	Immunoabsorption	II
Scleroderma/progressive systematic sclerosis	Lymphoplasmapheresis	II
Systematic lupus erythematosus	Plasma exchange	IV

Category I = Standard acceptable therapy

Category II = sufficient evidence to suggest efficacy usually as adjunctive therapy

Category III = inconclusive evidence of efficacy or uncertain risk/benefit ratio

Category IV = lack of efficacy in controlled trials.

*Immunoglobulin

Table 5- Indications of plasmapheresis therapy based on ASFA and AABB classification in neurologic diseases

Disease	Procedure	Indication Category
Guillain-Barre syndrome	Plasma exchange	I
Chronic inflammatory demyelinating Polyradiculoneuropathy	Plasma exchange	I
Polyneuropathy with IgG/IgA monoclonal protein	Plasma exchange	II
Polyneuropathy with IgM monoclonal protein	Plasma exchange	I
Myasthenia gravis	Plasma exchange	III
Stiff-person syndrome	Plasma exchange	II
Lambert-Eaton myasthenia syndrome	Plasma exchange	III
Paraneoplastic Neurologic syndrome	Plasma exchange	III
Polymyositis or dermatomyositis	Leukapheresis	IV
Multiple sclerosis	Plasma exchange	III
Idiopathic inflammatory demyelinating disease	Plasma exchange	II
Refsum`s disease	Plasma exchange	III
Rasmussen`s encephalitis	Plasma exchange	III
Sydenham`s chorea	Plasma exchange	II
PANDAS	Plasma exchange	II

PANDAS = Pediatric Autoimmune Neuropsychiatric Disorders Associated with Streptococcal Infections

Table 6- Indications of plasmapheresis therapy based on ASFA and AABB classification in blood disease and disproteinemia

Disease	AABB category	ASFA category
ABO-incompatible marrow transplant	II	II
Aplastic anemia	III	III
Autoimmune hemolytic anemia	III	III
Coagulation factor inhibitors	II	II
Cryoglobulinemia	II	II
HELLP syndrome (postpartum)	NR*	NR**
Hemolytic-uremic syndrome	III	III
Hyperviscosity/multiple myeloma	II	II
Immune thrombocytopenia purpura	II+	II+
Platelet alloimmunization	III	III
Post-transfusion purpura	I	I
Pure red cell aplasia	III	III
Red cell alloimmunization	III	III
Thrombocytopenic purpura	I	I

*Disorder not ranked by either AABB or ASFA.

**This disorder is ranked only in context of staphylococcal protein A

Immuniadsorption

AABB = American Association of Blood Banks; ASFA = American Society for Apheresis; Category I = Standard acceptable therapy; Category II = sufficient evidence to suggest efficacy usually as adjunctive therapy; Category III = inconclusive evidence of efficacy or uncertain risk/benefit ratio; Category IV = lack of efficacy in controlled trails; HELLP = Hemolysis, elevated liver enzymes, and low platelet.

Conclusion

Plasmapheresis, which is defined as the removal of plasma, can be either “adjusted plasma” or “exchange of plasma”. Plasmapheresis is currently used as a therapeutic modality in a wide array of conditions. Generally, plasmapheresis is used when a substance in the plasma, such as immunoglobulin, is acutely toxic and can be efficiently removed. Myriad conditions fall under this category, including neurologic, hematologic, metabolic, dermatologic, rheumatologic, and renal diseases, as well as intoxications, that can be treated with plasmapheresis.

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References

1. Crookston KP, Richter DM. Teaching and learning apheresis medicine: The Bermuda Triangle in Education. *J Clin Apher* 2010; 25(6):338-46.
2. Witt V. Training courses for pediatric apheresis on site; how apheresis technology transfer can be performed. *Transfus Apher Sci* 2010; 43(2):223-5.
3. Gahar LR, Visser O. The ins and outs of hemapheresis units in the Netherlands: practice and organization. *Transfus Apher Sci* 2013; 48(2):201.
4. Sadeghi M, Daniel V, Wang H, Zeier M, Schemmer P, Mehrabi A, et al. Plasmapheresis adjusts inflammatory responses in potential kidney transplant recipients. *Transplantation* 2013; 95(8):1021-9.
5. Makroo RN, Walia RS, Aneja S, Bhatia A, Chowdhry M. Preoperative predictors of blood component transfusion in living donor liver transplantation. *Asian J Transfus Sci* 2013; 7(2):140-6.
6. Hunt EA, Jain NG, Somers MJ. Apheresis therapy in children: an overview of key technical aspects and a review of experience in pediatric renal disease. *J Clin Apher* 2013; 28(1):36-47.
7. Federici AB, Vanelli C, Arrigoni L. Transfusion

- issues in cancer patients. *Thromb Res* 2012; 129 Suppl 1:S60-5.
8. Brand A. Passenger leukocytes, cytokines, and transfusion reactions (editorial). *N Eng J Med* 1994; 331: 670-1.
9. Smith Jw, Gilcher RO. Red blood cells, plasma, & other new apheresis derived blood products: Improving product quality & donor utilization. *Transfus Med Rev* 1999; 13: 118-23.
10. Owens MR, Sweeney JD, Tahhan RH, Fortkolt P. Influence of type of exchange fluid on survival in therapeutic apheresis for TTP. *J Clin Apheresis* 1995; 10: 178-82.
11. McLeod BC, Price TH, Owen H, Ciavarella D, Sniecinski I, Randels MJ, et al. Frequency of immediate adverse effects associated with apheresis donation. *Transfusion* 1998; 38(10):938-43.
12. Davenport A. What are the anticoagulation options for intermittent hemodialysis? *Nat Rev Nephrol* 2011; 5; 7(9):499-508.
13. Osion PR, Cox C, McCoullough J. Laboratory and clinical effects of the infusion of ACD solution during plateletpheresis. *Vox Sanguinis* 1987; 33: 79-87.
14. Strauss RG. Effects on donors of repeated leukocyte losses during plateletpheresis. *J Clin Apheresis* 1994; 9: 130-4.
15. Powel L. Intense plasmapheresis in the pregnant Rh sensitized woman. *Am J Obstet Gynecol* 1968; 101(2):153-70.
16. Isojima S, Hisano M, Suzuki T, Sago H, Murashima A, Yamaguchi K. Early plasmapheresis followed by high-dose γ -globulin treatment saved a severely Rho-incompatible pregnancy. *J Clin Apher* 2011;26(4):216-8.
17. Clarke CA, Elson CJ, Bradley J, Donohoe WT, Lehane D, Hughes-Jones NC. Intensive plasmapheresis as a therapeutic measure in Rhesus-immunized woman. *Lancet* 1970 18;1(7651):793-8.
18. Pusey CD, Levy JB. Plasmapheresis in immunologic renal disease. *Blood Purif* 2012;33(1-3):190-8.
19. Jones JV, Cumming RH, Bacon PA, Evers J, Fraser ID, Bothamley J, et al. Evidence for a therapeutic

effect of plasmapheresis in patient with systemic lupus erythematosus. *Q J Med* 1979; 48(192):555-76.

20. Nollet KE. Blood banking and transfusion medicine in extreme or resource-limited conditions. *Transfus Apher Sci* 2013 Sep 14. doi:pii: S1473-0502(13)00285-1. 10.1016/j.transci.2013.09.002.

21. Coppo P, Bussel A, Charrier S, Adrie C, Galicier L, Boulanger E, *et al.* High-dose plasma infusion versus plasma exchange as early treatment of thrombotic thrombocytopenic purpura/hemolytic-uremic syndrome. *Medicine (Baltimore)* 2003; 82(1):27-38.

22. Ibrahim RB, Balogun RA. Medications and therapeutic apheresis procedures: are we doing our best? *J Clin Apher* 2013; 28(1):73-7.

23. Williams ME. Starting a new therapeutic apheresis service: medical directorship and other matters. *J Clin Apher* 2013; 28(1):11-5.

24. Kawasaki Y, Ono A, Ohara S, Suzuki Y, Suyama K, Suzuki J, *et al.* Henoch-Schönlein purpura nephritis in childhood: pathogenesis, prognostic factors and treatment. *Fukushima J Med Sci* 2013; 59(1):15-26.

25. Dau PC. Immunologic rebound. *J Clin Apher* 1995; 10(4):210-7.

26. Rummeler S, Maier K, Barz D. Therapeutic apheresis in transplantation medicine, experience with cardiac and lung transplantation in Jena. *Atheroscler Suppl* 2013;14(1):33-8.

27. Dyer M, Neal MD, Rollins-Raval MA, Raval JS. Simultaneous extracorporeal membrane oxygenation and therapeutic plasma exchange procedures are tolerable in both pediatric and adult patients. *Transfusion* 2013; 10.

28. Orlin JB, Berkman EM. Partial plasma exchange using albumin replacement: Removal & recovery of normal plasma constituents. *Blood* 1980; 56: 1055-9.

29. Mcleod Bc, Sasseti RJ, Stefoski D, Davis FA. Partial plasma protein replacement in therapeutic plasma exchange. *J Clin Apher* 1983;1(2):115-8.

30. Waldmann TA, Strober W. Metabolism of Immunoglobulins. *Prog Allergy* 1969;13:1-110.

31. Michaud M, Pourrat J. Cryofibrinogenemia. *J Clin Rheumatol* 2013; 19(3):142-8.

32. Wells JV, Fundenberg HH. Metabolism of radioiodinated IgG in patients with abnormal serum IgG levels. *Hypergammaglobulinemia. Clin Exp Immunol* 1971;9(6):775-83.

33. Schwartz J, Winters JL, Padmanabhan A, Balogun RA, Delaney M, Linenberger ML, *et al.* Guidelines on the use of therapeutic apheresis in clinical practice-evidence-based approach from the Writing Committee of the American Society for Apheresis: the sixth special issue. *J Clin Apher* 2013; 28(3):145-284.

34. Sanna G, Bertolaccini ML, Khamashta MA. Neuropsychiatric involvement in systemic lupus erythematosus: current therapeutic approach. *Curr Pharm Des* 2008; 14(13):1261-9.

35. Philip J, Sarkar RS, Pathak A. Adverse events associated with apheresis procedures: Incidence and relative frequency. *Asian J Transfus Sci* 2013; 7(1):37-41.

36. Dau PC. Immunologic rebound. *J Clin Apheresis*.1995; 10: 210-7.

37. Derksen RH, Schuurman HJ, Meyling FH, Struyvenberg A, Kater L, *et al.* The efficacy of plasma exchange in the removal of plasma components. *J Lab Clin Med.* 1984; 104(3):346-54.

38. Junghans RP. IgG biosynthesis. No "Immunoregulatory feedback". *Blood* 1997; 90: 3815-8.

39. Yu Z, Lennon VA. Mechanism of intravenous immunoglobulin therapy in antibody-mediated autoimmune disease. *N Eng J Med* 1999; 340: 227-8.

40. Poisson JL, Low A, Park YA. The treatment of nephrogenic systemic fibrosis with therapeutic plasma exchange. *J Clin Apher* 2013; 28(4):317-20.

41. Cortese I, Cornblath DR. Therapeutic plasma exchange in neurology: 2012. *J Clin Apher* 2013; 28(1):16-9.

42. Lewis EJ, Hunsicker LG, Lan SP, Rohde RD, Lachin JM. A controlled trial of plasmapheresis therapy in severe lupus nephritis. *N Engl J Med* 1992; 326(21):1373-9.

43. Loo CY, Mohamed Said MS, Mohd R, Abdul Gafor AH, Saidin R, Halim NA. Immunoabsorption and plasmapheresis are equally efficacious as adjunctive therapies for severe lupus nephritis. *Transfus Apher Sci*

2010; 43(3):335-40.

44. Pool M, Mcleod BC. Pyrogen reactions to human serum albumin during plasma exchange. *J Clin Apher* 1995; 10(2):81-4.

45. Stefanutti C, Julius U. Lipoprotein apheresis: state of the art and novelties. *Atheroscler Suppl* 2013; 14(1):19-27.

46. Winters JL. Apheresis in the treatment of idiopathic dilated cardiomyopathy. *J Clin Apher* 2012; 27(6):312-9.

47. McLeod BC, Price TH, Owen H, Ciavarella D, Sniecinski I, Randels MJ, *et al.* Frequency of immediate adverse effects associated with apheresis donation. *Transfusion* 1999; 39: 282-8.

48. Owens MR, Sweeney JD, Tahhan RH, Fortkolt P. Influence of type of exchange fluid on survival in therapeutic apheresis for TTP. *J Clin Apheresis* 1995; 10: 178-82.

49. Weinstein R. Prevention of citrate reactions during therapeutic plasma exchange by constant infusion of calcium gluconate with the return fluid. *J Clin Apheresis* 1996; 11: 204-10.

50. Pierce LR, Gaines A, Finlayson JS, Varricchio F, Epstein JS. Hemolysis & acute renal failure due to the administrations of albumin diluted in sterile water [letter]. *Transfusion* 1999; 39: 110-1.

51. Hattersley JG, Chappell MJ, Zehnder D, Higgins RM, Evans ND. Describing the effectiveness of immunosuppression drugs and apheresis in the treatment of transplant patients. *Comput Methods Programs Biomed* 2013; 109(2):126-33.

52. Stigelman WH, Henry DH, Talbert RL, Townsend RJ. Remove of prednisone & prednisolone by plasma exchange. *Clin Pharmacy* 1984;3: 402-7.

53. Ibrahim RB, Liu CY, Cronin SM, Murphy BC, Cha R, Swerdlow P, *et al.* Influence of plasma exchange on the disposition of the fourth generation cephalosporin cefepime. *J Oncol Pharm Pract* 2009; 15(4):217-22.

54. Wood GJ, Hall GM. Plasmapheresis and plasma cholinesterase. *Br J Anaesth* 1978; 50(9):945-9.

55. Perseghin P, Capra M, Baldini V, Sciorelli G. Bradykinin production during donor plasmapheresis

procedures. *Vox Sang* 2001; 81(1):24-8.

56. Owen HG, Brecher ME. Partial colloid replacement for therapeutic plasma exchange. *J Clin Apheresis* 1997; 12: 87-92.

57. Sultan Y, Bussel A, Maisonneuve P, Poupene M, Sitty X, Gajdos P. Potential danger of thrombosis after plasma exchange in the treatment of patients with immune disease. *Transfusion* 1979; 19: 558-93.

58. Domen RE, Kennedy MS, Jones LL, Senhauser DA. Hemostatic imbalances produced by plasma exchange. *Transfusion* 1984; 24: 336-8.

59. Flaum MA, Cuneo RA, Appelbaum FR, Deisseroth AB, Engel WK, Gralnick HR. The hemostatic imbalance of plasma exchange transfusion. *Blood* 1979;54(3):694-702.

60. Karussis D. Immunotherapy of multiple sclerosis: the state of the art. *BioDrugs*. 2013; 27(2):113-48.

61. Kamel H, Tomasulo P, Bravo M, Wiltbank T, Cusick R, James RC, *et al.* Delayed adverse reactions to blood donation. *Transfusion* 2010; 50(3):556-65.

62. Salam A, Hosain GM, Narvios A, Sazama K, Lichtiger B. Immediate adverse reactions to platelet transfusions: whole blood derived versus apheresis platelets. *Mymensingh Med J* 2013; 22(1):143-7.

63. McClellan SD, Whitaker CH, Friedberg RC. Removal of vancomycin during plasmapheresis. *Ann Pharmacother* 1997; 31(10):1132-6.

64. Strauss RG. Mechanisms of adverse effects during hemapheresis. *J Clin Apher* 1996; 11(3):160-4.

65. Silberstein LE, Naryshkin S, Haddad JJ, Strauss Jf. Calcium hemostasis during therapeutic plasma exchange. *Transfusion* 1986; 26(2):151-5.

66. Crookston KP, Simon TL. Physiology of apheresis. In: Mcleod BC, Price MJ, *et al.* Apheresis: principles & practice. 2th ed. Bethesda., MD: AABB press. 2003,

67. Szymanski IO. Ionized calcium during platelet-pheresis. *Transfusion* 1978; 18(6):701-8.

68. Karagiorgou LZ, Pantazopoulou MN, Mainas NC, Beloukas AI, Kriebardis AG. Knowledge about umbilical cord blood banking among Greek citizens. *Blood Transfus* 2013 ; 3:1-7.

69. Hester JP, Ayyar R. Anticoagulant & electrolytes. *J*

Clin Apheresis 1984; 2: 41-51.

70. Bolan CD, Cecco SA, Wesley RA, Horne M, Yau YY, Remaley AT, *et al.* Controlled study of citrate effects and response to IV calcium administration during allogenic peripheral blood progenitor cell donation. *Transfusion* 2002; 42: 935-46.

71. Korach JM, Berger P, Giraud C. Role of replacement fluids in the immediate complications of plasma exchange. *Intensive Care Med* 1998; 24: 452-8.

72. Dutcher JP, Aisner J, Hogge DE, Schiffer CA. Donor reaction to hydroxyethyl starch during granulocytapheresis. *Transfusion* 1984; 24(1):66-7.

73. Ring J, Messmer K. Incidence & severity of anaphylactoid reactions to colloid volume substitutes. *Lancet* 1977 26; 1(8009):466-9.

74. Kannan S, Milligan KR. Moderately severe anaphylactoid reaction to pentastarch(200/0.5) in a patient with acute severe asthma. *Intensive Care Med* 1999; 25(2):220-2.

75. Martínez Álvarez JC. Antibodies, human leukocyte antigens, and biomodulators in transfusion-related acute adverse effects. *Gac Med Mex* 2013; 149(1):81-8.

76. Nollet KE, Ohto H, Yasuda H, Hasegawa A. The great East Japan earthquake of March 11, 2011, from the vantage point of blood banking and transfusion

medicine. *Transfus Med Rev* 2013; 27(1):29-35.

77. Leitman SF, Boltansky H, Alter HJ, Pearson FC, Kaliner MA. Allergic reactions in healthy plateletpheresis donor caused by sensitization to ethylene oxide gas. *N Eng J Med* 1986; 315: 1192-6.

78. Owen HG, Brecher ME. Atypical reactions associated with use of ACEI & apheresis. *Transfusion* 1994; 34(10):891-4.

79. Aghishi T. Anion-blood contact reaction (ABC reaction) in patients treated by LDL apheresis with dextran sulfate-cellulose column while receiving ACE inhibitors. *JAMA* 1994; 271(3):195-6.

80. Olbricht CJ, Schaumann D, Fischer D. Anaphylactoid reactions, LDL apheresis with dextran sulfate & ACE inhibitors. *Lancet* 1993; 341: 60-1.

81. Jaime-Perez JC, Monreal-Robles R, Colunga-Pedraza J, Mancías-Guerra C, Rodríguez-Romo L, Gómez-Almaguer D. Cord blood banking activities at a university hospital in northeast Mexico: an 8-year experience. *Transfusion* 2012; 52(12):2606-13.

82. Newman BH. Donor reactions & injuries from whole blood donation. *Transfus Med Rev* 1997; 11(1):64-75.

83. Montacer-kuhsari J, Voller H, Keller F. Pulmonary air embolism. *Intensive Care Med* 1994;20(2):166-7.