

Molecular Characterization of Community-Associated Methicillin-Resistant *Staphylococcus aureus* in Iranian Burn Patients

Samira Tajik¹, Shahin Najar-Peerayeh^{1*} , Bita Bakhshi¹ , Reza Golmohammadi² 

1. Department of Bacteriology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran

2. Molecular Biology Research Center, Systems Biology and Poisonings Institute, Baqiyatallah University of Medical Sciences, Tehran, Iran

KEYWORDS

Burn
Patients
MRSA,
SCCmec

Scan to discover online



Main Subjects:
Microbiology

Received 22 Nov 2018;

Accepted 12 Jul 2019;

Published Online 22 Sep 2019;

 10.30699/ijp.2019.94189.1917

PMCID:

PMID:

ABSTRACT

Background & Objective: Methicillin-resistant *Staphylococcus aureus* (MRSA) is reported as one of the important bacterial causes of burn wound infections. This study was carried out to investigate molecular characterization of community-associated MRSA (CA-MRSA) isolated from Iranian burn patients.

Methods: A total of 31 isolates of *S. aureus* were collected from the Motahari Burns Hospital (Tehran, Iran) in 2016. All isolates were collected from outpatients and inpatients within 48 hours of admission. The *mecA*, *pvl*, *tsst-I*, *hla-a*, and *psma* genes detecting, SCCmec, *agr* and PFGE typing were done.

Results: A total of 13 (41.9%) isolates were cefoxitin-resistant and *mecA*-positive, which were considered as MRSA. The SCCmec typing MRSA strains revealed type II in 1 (7.7%), type III in 9 (69.2%), and other types in 3 isolates (23.7%) cases. The *agr* typing of all 31 isolates showed that 14 (45.2%), 1 (3.2%), 6 (19.4%), and 10 (32.3%) strains belonged to *agr* groups 1, 3, 4, and unknown type, respectively. The *pvl*, *tsst-I*, *hla-a*, and *psma* genes were positive in 3 (9.7%), 4 (12.9%), 21 (67.7%), and 31 (100%) isolates, respectively. Considering the cut-off values of $\geq 50\%$, 3 groups of related isolates (cluster A1, B1, and C1) in PFGE study were observed.

Conclusion: The MRSA strains of this study were initially isolated as Community-associated *S. aureus* (CA-MRSA); however molecular characterization showed that a significant proportion of them had hospital-associated MRSA (HA-MRSA) features. Therefore, it is likely that the HA-MRSA strains are spread among the community.

Corresponding Information: Shahin Najar-Peerayeh, Department of Bacteriology, Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran Email: najarp_s@modares.ac.ir

Copyright © 2019. This is an open-access article distributed under the terms of the Creative Commons Attribution- 4.0 International License which permits Share, copy and redistribution of the material in any medium or format or adapt, remix, transform, and build upon the material for any purpose, even commercially.

Introduction

Due to the skin damaged and weakened innate immunity, burn patients become susceptible to various infections (1, 2), among which bacterial infections are the main problems in hospitalized burn patients (3, 4).

Staphylococcus aureus is one of the most common bacteria isolated from burn infections (5). Methicillin-resistant *S. aureus* (MRSA) including hospital-associated MRSA (HA-MRSA) (3, 6) and community-associated MRSA (CA-MRSA) (7) are reported as the important bacterial causes of burn wound infections. The centers for disease control (CDC) defined a CA-MRSA infection as any MRSA infection diagnosed for an outpatient or within 48 hours of hospitalization if the patient lacks such healthcare-associated MRSA risk factors as dialysis or use of invasive medical devices, surgery, and residence in a long-term care facility or hospitalization during the previous year; all other MRSA infections were considered to be HA-MRSA (8-10).

Cefoxitin is a potential inducer of the *mecA* regulatory system and is used for screening MRSA strains (11). Staphylococcal cassette chromosome *mec* (SCCmec) element of MRSA strains carry *mecA* gene that mediates resistance to methicillin (10, 12). HA-MRSA strains carry large SCCmec types II and III, which are responsible for resistance to other antibiotics. However, SCCmec types IV and V elements are found in CA-MRSA strains (10, 13).

It should be noted that there are several problems in terms of using CA-MRSA and HA-MRSA. First, both strains can be circulated in the community; second, it is possible that CA-MRSA strains are gradually converted to nosocomial pathogens (10, 14).

The pathogenicity of *S. aureus* is associated with various virulence factors genes including accessory gene regulator (*agr*) system, panton-valentine leukocidin (*pvl*), toxic shock syndrome toxin (*tsst*), α -hemolysin (*hla*), and alpha-type phenol soluble modulins (*psma*).

These virulence factors can make extensive infection and severe damage (7, 12, 15, 16). The *agr* regulatory system has a crucial role in the development of skin infection cases by most CA-MRSA USA300 strains (17). The PVL toxin can be found in both MSSA and MRSA strains (18) but typically is associated with CA-MRSA strains (7). TSST-1 is a superantigen that is absorbed from cutaneous infections and it localizes the kidneys (19). It is reported that the pore-forming α -hemolysin of *S. aureus* plays a key role in soft tissue infection (20). The *psma* may play a role in biofilms development and lysis of human cell types such as leukocytes and erythrocytes; also, it stimulates inflammation response (21).

Phenotypic and genotypic characterization of CA-MRSA and HA-MRSA can be helpful in the treatment, infection control, and prevention of the spread of antibiotic-resistant bacterial strains (1, 22, 23). This study was carried out to investigate molecular characterization of CA-MRSA isolated from Iranian burn outpatients and hospitalized patients within 48 hours of admission.

Materials and Methods

Bacterial Isolates

A total of 31 isolates of *S. aureus* were collected from Shahid Motahari Burns Hospital (Tehran, Iran) in the time period of May to October 2016. All isolates were collected from outpatients and inpatients within 48 hours of admission. Bacterial morphology study and biochemical tests including catalase, tube and slide coagulase, DNase, and mannitol-fermenting were done

to identify *S. aureus*. After identification, all strains were preserved in trypticase soy broth medium containing 15% glycerol at -70°C.

Antimicrobial Susceptibility Testing

Antimicrobial susceptibility testing was done by disk diffusion method according to the clinical and laboratory standards institute (CLSI) 2015 guidelines. Antibiotic discs containing ceftiofloxacin (30 µg), gentamicin (10 µg), erythromycin (15 µg), tetracycline (30 µg), rifampin (5 µg), clindamycin (2 µg), ciprofloxacin (5 µg), trimethoprim-sulfamethoxazole (1.25/23.75 µg), and linezolid (30 µg) (MAST UK) were used. *S. aureus* ATCC 25923 was used as a control strain.

Molecular Detection and Typing

DNA Extraction

Four to five fresh colonies of bacteria were suspended in 400 µL of lysis solution buffer containing lysostaphin (200µg/mL) (Sigma-Aldrich Co, Germany). Then, DNA was extracted by phenol/chloroform/isoamyl alcohol (Merck Co).

PCR Amplification

The *mecA* was used as target gene for the detection of MRSA strains (24). The primers described by Zhang *et al.* (24) and Shopsis *et al.* (25) were used for SCC*mec* typing and *agr* typing, respectively. RN6390, RN6607, RN8465, and RN4550 strains were used as positive control for *agr* typing. The *pvl*, *tsst-1*, *hla-a* (26), and *psma* (27) genes were used as target gene for virulotyping. The list of primers sequences of this study, target genes, and the length of PCR products are presented in Table 1.

Table 1. The list of primers sequences for *mecA*, *pvl*, *tsst-1*, *hla-a*, and *psma* genes detection, SCC*mec* and *agr* typing and PCR products length

	Primer	Sequence (5'→3')	Size (bp)	Ref
<i>mecA</i> detection	<i>mecA</i>	F: GTGAAGATATACCAAGTGATT R: ATGCGCTATAGATTGAAAGGA	146	(24)
	SCC <i>mec</i> I	F: GCTTTAAAGAGTGTCTGTTACAGG R: GTTCTCTCATAGTATGACGTCC	613	
SCC <i>mec</i> typing	SCC <i>mec</i> II	F: CGTTGAAGATGATGAAGCG R: CGAAATCAATGGTTAATGGACC	398	(24)
	SCC <i>mec</i> III	F: CCATATTGTGTACGATGCG R: CCTTAGTTGTCTGTAACAGATCG	280	
	SCC <i>mec</i> IVa	F: GCCTTATTCGAAGAAACCG R: CTACTCTTCTGAAAAGCGTCG	776	
	SCC <i>mec</i> IVb	F: TCTGGAATTAATTCAGCTGC R: AAACAATATTGCTCTCCCTC	493	
	SCC <i>mec</i> IVc	F: ACAATATTTGTATTATCGGAGAGC R: TTGGTATGAGGTATTGCTGG	200	
	SCC <i>mec</i> IVd	F: CTCAAATACGGACCCCAATACA R: TGCTCCAGTAATTGCTAAAG	881	
	SCC <i>mec</i> V	F: GAACATTGTTACTTAAATGAGCG R: TGAAAGTTGTACCCCTTGACACC	325	
	<i>agr</i> typing	panF	ATGCACATGGTGCACATGC	
<i>agr</i> I-R		GTCACAAGTACTATAAGCTGCGAT	440	
<i>agr</i> II-R		GTATTACTAATTGAAAAGTGCCATAGC	572	
<i>agr</i> III-R		CTGTTGAAAAAGTCAACTAAAAGCTC	406	
<i>agr</i> IV-R		CGATAATGCCGTAATAC CCG	588	
	<i>pvl</i>	F: GGAAACATTTATTCTGGCTATAC	502	(26)

	Primer	Sequence (5'→3')	Size (bp)	Ref
Virulence genes typing		R: CTGGATTGAAGTTACCTCTGG		
	<i>tsst-1</i>	F: TTATCGTAAGCCCTTTGTTG R: TAAAGGTAGTTCTATTGGAGTAGG	398	
	<i>hla-a</i>	F: CGGTACTACAGATATTGGAAGC R: TGGTAATCATCACGAACTCG	744	
	<i>psma</i>	F: TATCAAAAGCTTAATCGAACAATTC R: CCCCTTCAAATAAGATGTTTCATATC	176	(27)

The final volume of 25 mL PCR reaction mixture containing reverse and forward primer (10 pmol) (Pishgam Biotech Co, Iran), DNA taq polymerase enzyme (1.5 units), dNTPs (0.02 mM), MgCl₂ (0.1 mM), buffer (Fermentase Co, Lithuania), deionized distilled water, and DNA template were used for PCR amplification. The PCR amplification performed in thermo cycler (BIO RAD T100) by following annealing temperatures: *mecA*; 55°C, SCC*mec* typing; 60°C, *agr* typing; 50°C, and virulence genes typing; 50°C for *pvl* and *hla-a*, and 51°C for *tsst-1* genes.

Pulsed-field Gel Electrophoresis (PFGE)

PFGE analysis based on *Sma*I restriction enzyme (Takara) digestion was carried out using the PulseNet standard protocol on 13 SCC*mec*-positive *S. aureus* strains (PulseNet, www.cdc.gov/pulsenet). After optimizing conditions, PFGE was run on CHEF MAPPER^{XA} (BIO RAD) device as follows: initial switch 5 s, final switch 40 s, run time 21 h, voltage 200 (6v/cm). 1% agarose gel (intron biotechnology) in TBE 0.5 X was used for electrophoresis. According to the PulseNet protocol, *Salmonella enterica* serotype Braenderup (H9812) was used as a control. After gel staining with ethidium bromide (10 mg/mL), imaging

was performed by UV light gel documentation system (BioDocAnalyze). GelCompar II version 6.6.11 software was used for drawing dendrogram graph and interpretation of results. Cut-off values of ≥50% were considered to define clusters.

Results

Patients

A total of 31 non-duplicate *S. aureus* isolates were investigated in this study. All isolates were collected from burn patients within 48 hours of admission. 20 (64.5%) of these patients were inpatients and 11 (35.5%) outpatients.

Antimicrobial Susceptibility Testing

Antibiotic susceptibility test results revealed that a total of 13 (41.9%) isolates were resistant to cefoxitin, which were considered as MRSA. All isolates were susceptible to Linezolid. The resistance rates of other antibiotics were as follows: erythromycin 15 (48.4%), tetracycline 11 (35.5%), ciprofloxacin 11 (35.5%), gentamicin 10 (32.3%), clindamycin 9 (29.0%), rifampin 6 (19.4%), and trimethoprim-sulfamethoxazole 4 (12.9%) (Table 2).

Table 2. Antibiotic resistance of MSSA and MRSA isolated from burn patient

Antibiotics	MSSA (n=18)	MRSA (n=13)	Total (n=31)
cefoxitin	0	13 (100%)	13 (41.9%)
erythromycin	6 (33.3%)	9 (69.2%)	15(48.4%)
tetracycline	3 (16.7%)	8 (61.5%)	11(35.5%)
ciprofloxacin	2 (11.1%)	9 (69.2%)	11(35.5%)
gentamicin	1 (5.5%)	9 (69.2%)	10(32.3%)
clindamycin	2 (11.1%)	7 (53.8%)	9(29.0%)
rifampin	1 (5.5%)	5 (38.5%)	6(19.4%)
trimethoprim-sulfamethoxazole	0	4 (30.8%)	4(12.9%)
Linezolid	0	0	0

Molecular Detection and Typing

Molecular studies showed that 13 (41.9%) isolates were *mecA*-positive (Table 3). The SCC*mec* typing MRSA strains revealed type II in 1 (7.7%), type III in 9 (69.2%), and other types in 3 isolates (23.1%) (Table 3).

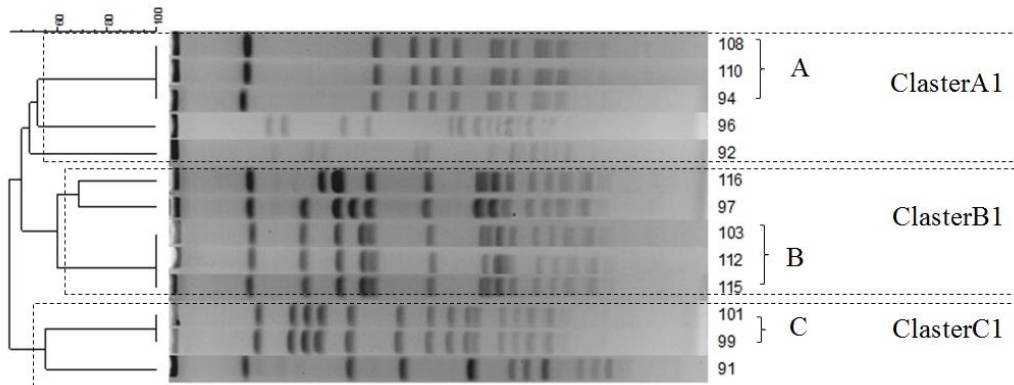
The *agr* typing of all 31 isolates showed that 14 (45.1%), 1 (3.2%), 6 (19.4%), and 10 (32.3%) strains belonged to *agr* groups 1, 3, 4, and unknown type, respectively (Table 3).

The *pvl*, *tsst-1*, *hla-a*, and *psma* genes were positive in 3 (9.7%), 4 (12.9%), 21 (67.7%), and 31 (100%), respectively.

All 13 MRSA isolates were studied by PFGE. Considering the cut-off values of ≥50%, 3 groups of isolates (cluster A1, B1, and C1) were observed in this study: cluster A1; 3 pulse types, cluster B1; 3 pulse types and cluster C1; 2 pulse types (Figure 1).

Table 3. The *mecA*, *pvl*, *tsst-1*, *hla-a*, and *psma* genes detection, and SCC*mec* and *agr* typing result (n=31)

gene	<i>mecA</i>	SCC <i>mec</i>	<i>agr</i>	<i>pvl</i>	<i>tsst-1</i>	<i>hla-a</i>	<i>psma</i>		
Positive	13(41.9%)	type3	9(69.2%)	type 1	14(45.1%)	3(9.7%)	4(12.9%)	21(67.7%)	31(100%)
		type2	1(7.7%)	type 3	1(3.2%)				
		Not typeable	3(23.1%)	type 4	6(19.4%)				
				Not typeable	10(32.3%)				
Negative	18(58.1%)	18	0(0%)	28(90.3)	27(87.1%)	10(32.3%)	0(0%)		

**Figure 1.** PFGE analysis of CA-MRSA in current study

Discussion

S. aureus is one of the common causes of burn wound infections (5). Drug-resistant strains of this organism such as MRSA can complicate the treatment process (3, 6, 7). In addition, the strains containing virulence genes such as *agr*, *pvl*, *tsst-1*, *hla-a*, and *psma* can make extensive infection and severe damage (7, 15, 16, 28, 29).

In current study 41.9% isolates were *mecA*-gene positive and cefoxitin-resistant, so were considered as MRSA. Based on the molecular detection of *mec* gene in different studies, MRSA strains have been reported as 86.4% (30), 88.6% (31), 81.2% (32), and 87.4% (18) in Iran and 55.3% in China (3).

According to the antibiotic susceptibility testing of all 31 strains, the highest resistance was related to erythromycin (48.4%) compared to 58.2% (2), 77.8% (30), and 90.6% (16) in Iran. All the 13 MRSA strains in this study were sensitive to linezolid, which was similar to the findings of the studies conducted by Montazeri (31, 32), and Goudarzi (16) from Iran and Murray (7) from the US.

The SCC*mec* typing of 13 MRSA strains showed that 9 (69.2%) strains belonged to SCC*mec* III, that's almost is consistent with some studies in Iran; 56.8% (2), 47.5% (33), 70.1% (15), and 71.7% (16) did not match the other studies in Iran; 95.7% (30), China; 95.4% (16), Brazil; 94.8% (34). 7.7% of the these 13 strains belonged to SCC*mec* II, compared with 10% (33) and 2.5% (15) studies in Iran, and (1%) in Brazil (34). Although 3 (23.1%) other types were not detected in this study, SCC*mec* I, IV, and V have been detected in other studies (2, 3, 15, 16, 34).

Based on the CDC definition of CA-MRSA and HA-MRSA strains (8-10) and according to other studies (10, 13), it seems that most of these SCC*mec* types belonged to HA-MRSA strains. Since sampling was done within 48 hours after admission, the strains were expected to be CA-MRSA. According to a hypothesis, this has been affected by various factors such as having the same source of infection or spread of infected patients with HA-MRSA strains among the community. Since checking the patients' referral records during the last year was not possible in this study, it is possible that most of these strains were HA-MRSA. Furthermore, having high relative resistance to selected antibiotics including clindamycin in most strains also confirms this probability. Eight (61.5%) of MRSA strains in this study and 75.5% in Goudarzi's study (16) were resistant to clindamycin, which indicates these strains had high resistance to other antibiotics. Also, 5 (38.5%) clindamycin-susceptible strains in this study had a lower resistance to other antibiotics. Therefore, it can be concluded that 8 (61.5%) of these strains correspond to HA-MRSA and 5 (38.5%) to CA-MRSA.

The PFGE cluster results with cut-off values of $\geq 50\%$ indicated that MRSA strains in this study, to some extent, had CA-MRSA characteristics. Also, the 3 strains in clusters A1, 3 in B1, and 2 in C1 have the same pattern. This similarity suggests that the source of infection has been the same in each cluster.

The PVL toxin typically is associated with CA-MRSA strains (7). In the present study, 3 (9.7%) MRSA strains were *pvl*-positive. In contrast to the study by

Goudarzi that reported the positive *pvl* strains belonged to SCCmec type IV (16), in this study 2 of 3 strains belonged to the SCCmec type III. Although all MSSA strains in this study were *pvl*-negative, some other studies in Iran reported that 7.23% (18), 15.1% (16), and 33.3% (18) of MSSA cases were positive.

The *agr* typing assay of all 31 isolates showed that the highest rate belonged to *agr* type 1 (45.1%) compared to 84.9% in Iran (16) and 96.5% in China (3); meanwhile, the lowest rate belonged to *agr* type 3 (3.2%) compared to 15.1% in Iran (16). In this study, 32.3% strains were not typeable, which might be related to the presence of different mutations, sensitivity, and specificity of primers.

The *tsst-1* gene detection showed that 12.9% of the isolates in this study belonged to MSSA strains, but this gene was not seen in MRSA strains. In contrast to this study, in a different study in Iran 18.9% of MRSA isolates were *tsst*-positive which were distributed among SCCmec type III (16).

In the present study, 67.7% strains were *hla*-positive compared to another study in Iran which reported 51.8% (15). Data analysis showed that 32.2% belonged to MRSA and 35.5% to MSSA.

Our results showed that 41.9% of isolates were MRSA, which indicates the presence of antibiotic resistance elements in a significant number of strains and necessity of performing antibiogram before starting treatment. Furthermore, the MRSA strains of this study were initially isolated as CA-MRSA, however molecular characterization showed that a significant proportion of them had HA-MRSA features. Therefore, it is likely that the HA-MRSA strains are spread among the community. Finally, the investigation of virulence factors in this study showed the presence of various toxin genes in isolates that can enhance their ecological niche ability and also make treatment more complicated, especially in burn patients.

Acknowledgements

The current study was supported by a grant from the Faculty of Medical Sciences, Tarbiat Modares University, Tehran, Iran.

Conflict of Interest

The authors declared no conflict of interest regarding the publication of this article.

References

- Schweizer M, Ward M, Cobb S, McDanel J, Leder L, Wibbenmeyer L, et al. The epidemiology of methicillin-resistant *Staphylococcus aureus* on a burn trauma unit. *Infection Control & Hospital Epidemiology*. 2012;33(11):1118-25. [DOI:10.1086/668032] [PMID] [PMCID]
- Namvar AE, Khodaei F, Bijari A, Lari AR. Detection of integrons and Staphylococcal Cassette Chromosome (SCCmec) types in *Staphylococcus aureus* isolated from burn and non-burn patients. *International journal of health sciences*. 2015;9(4):440. [DOI:10.12816/0031234] [PMID] [PMCID]
- Chen X, Yang H-h, Huangfu Y-c, Wang W-k, Liu Y, Ni Y-x, et al. Molecular epidemiologic analysis of *Staphylococcus aureus* isolated from four burn centers. *Burns*. 2012;38(5):738-42. [DOI:10.1016/j.burns.2011.12.023] [PMID]
- Choopani A, Golmohammadi R, Rafati H, Imani Fooladi AA. Prevalence of *Staphylococcus aureus* strains isolated from drug infection and drug sensitivity pattern, Tehran-Iran (2006-07). *Journal of Gorgan University of Medical Sciences*. 2012;14(3):135-40.
- Alebachew T, Yismaw G, Derabe A, Sisay Z. *Staphylococcus aureus* burn wound infection among patients attending Yekatit 12 hospital burn unit, Addis Ababa, Ethiopia. *Ethiopian journal of health sciences*. 2012;22(3).
- Kaiser ML, Thompson DJ, Malinoski D, Lane C, Cinat ME. Epidemiology and risk factors for hospital-acquired methicillin-resistant *Staphylococcus aureus* among burn patients. *Journal of Burn Care & Research*. 2011;32(3):429-34. [DOI:10.1097/BCR.0b013e318217f92d] [PMID]
- Murray CK, Holmes RL, Ellis MW, Mende K, Wolf SE, McDougal LK, et al. Twenty-five year epidemiology of invasive methicillin-resistant *Staphylococcus aureus* (MRSA) isolates recovered at a burn center. *Burns*. 2009;35(8):1112-7. [DOI:10.1016/j.burns.2009.02.013] [PMID]
- Horwitz RJ, Jernigan D, Powers J, Jernigan J. Strategies for clinical management of MRSA in the community: summary of an experts' meeting convened by the Centers for Disease Control and Prevention. 2006. 2009.
- David MZ, Daum RS. Community-associated methicillin-resistant *Staphylococcus aureus*: epidemiology and clinical consequences of an emerging epidemic. *Clinical microbiology reviews*. 2010;23(3):616-87. [DOI:10.1128/CMR.00081-09] [PMID] [PMCID]
- David MZ, Glikman D, Crawford SE, Peng J, King KJ, Hostetler MA, et al. What is community-associated methicillin-resistant *Staphylococcus aureus*? *Journal of Infectious Diseases*. 2008;197(9):1235-43. [DOI:10.1086/533502] [PMID]
- Anand K, Agrawal P, Kumar S, Kapila K. Comparison of cefoxitin disc diffusion test, oxacillin screen agar, and PCR for *mecA* gene for detection of MRSA. *Indian journal of medical microbiology*. 2009;27(1):27.
- Ghasemian A, Peerayeh SN, Bakhshi B, Mirzaee M. Detection of accessory gene regulator groups genes and cassette chromosome *mec* types among *Staphylococcus aureus* isolated from intensive care unit patients. *Asian Pacific journal of tropical disease*. 2015;5(2):153-7. [DOI:10.1016/S2222-1808(14)60643-5]
- Ito T, Ma XX, Takeuchi F, Okuma K, Yuzawa H, Hiramatsu K. Novel type V staphylococcal cassette chromosome *mec* driven by a novel cassette chromosome recombinase, *ccrC*. *Antimicrobial Agents and Chemotherapy*. 2004;48(7):2637-51. [DOI:10.1128/AAC.48.7.2637-2651.2004] [PMID] [PMCID]
- Huang H, Flynn NM, King JH, Monchard C, Morita M, Cohen SH. Comparisons of community-associated methicillin-resistant *Staphylococcus aureus* (MRSA) and hospital-associated MRSA infections in Sacramento, California. *Journal of Clinical Microbiology*. 2006;44(7):2423-7. [DOI:10.1128/JCM.00254-06] [PMID] [PMCID]
- Motallebi M, Jabalameli F, Asadollahi K, Taberikalani M, Emameini M. Spreading of genes encoding enterotoxins, haemolysins, adhesin and biofilm among methicillin resistant *Staphylococcus aureus* strains with staphylococcal cassette chromosome *mec* type IIIA isolated from burn patients.

- Microbial pathogenesis. 2016;97:34-7. [DOI:10.1016/j.micpath.2016.05.017] [PMID]
16. Goudarzi M, Bahramian M, Tabrizi MS, Udo EE, Figueiredo AMS, Fazeli M, et al. Genetic diversity of methicillin resistant Staphylococcus aureus strains isolated from burn patients in Iran: ST239-SCCmec III/t037 emerges as the major clone. Microbial Pathogenesis. 2017;105:1-7. [DOI:10.1016/j.micpath.2017.02.004] [PMID]
 17. Cheung GY, Wang R, Khan BA, Sturdevant DE, Otto M. Role of the accessory gene regulator agr in community-associated methicillin-resistant Staphylococcus aureus pathogenesis. Infection and immunity. 2011;79(5):1927-35. [DOI:10.1128/IAI.00046-11] [PMID] [PMCID]
 18. Khosravi AD, Hoveizavi H, Farshadzadeh Z. The prevalence of genes encoding leukocidins in Staphylococcus aureus strains resistant and sensitive to methicillin isolated from burn patients in Taleghani Hospital, Ahvaz, Iran. Burns. 2012;38(2):247-51. [DOI:10.1016/j.burns.2011.08.002] [PMID]
 19. Mino MJ, Ortiz RT, Randad P, Moffatt LT, Jordan MH, Shupp JW. Localization of Superantigen Virulence Factors in Kidney Tissue of Animals With Staphylococcus aureus-Infected Burn Wounds. Journal of Burn Care & Research. 2013;34(1):142-50. [DOI:10.1097/BCR.0b013e31827778e3] [PMID]
 20. Adhikari RP, Thompson CD, Aman MJ, Lee JC. Protective efficacy of a novel alpha hemolysin subunit vaccine (AT62) against Staphylococcus aureus skin and soft tissue infections. Vaccine. 2016;34(50):6402-7. [DOI:10.1016/j.vaccine.2016.09.061] [PMID] [PMCID]
 21. Cheung GY, Joo H-S, Chatterjee SS, Otto M. Phenol-soluble modulins-critical determinants of staphylococcal virulence. FEMS microbiology reviews. 2014;38(4):698-719. [DOI:10.1111/1574-6976.12057] [PMID] [PMCID]
 22. Mirzaee M, Najar-Peerayeh S, Behmanesh M, Moghadam MF. Relationship between adhesin genes and biofilm formation in vancomycin-intermediate Staphylococcus aureus clinical isolates. Current microbiology. 2015;70(5):665-70. [DOI:10.1007/s00284-014-0771-9] [PMID]
 23. Najar-Peerayeh S, Mirzaee M, Behmanesh M. Molecular characterization of vancomycin-intermediate Staphylococcus aureus isolates from Tehran. Asian Pacific Journal of Tropical Disease. 2016;6(9):726-31. [DOI:10.1016/S2222-1808(16)61118-0]
 24. Zhang K, McClure J-A, Elsayed S, Louie T, Conly JM. Novel multiplex PCR assay for characterization and concomitant subtyping of staphylococcal cassette chromosome mec types I to V in methicillin-resistant Staphylococcus aureus. Journal of clinical microbiology. 2005;43(10):5026-33. [DOI:10.1128/JCM.43.10.5026-5033.2005] [PMID] [PMCID]
 25. Shopsis B, Mathema B, Alcabes P, Said-Salim B, Lina G, Matsuka A, et al. Prevalence of agr specificity groups among Staphylococcus aureus strains colonizing children and their guardians. Journal of clinical microbiology. 2003;41(1):456-9. [DOI:10.1128/JCM.41.1.456-459.2003] [PMID] [PMCID]
 26. Havaei SA, Azimian A, Fazeli H, Naderi M, Ghazvini K, Samiee SM, et al. Genetic characterization of methicillin resistant and sensitive, vancomycin intermediate Staphylococcus aureus strains isolated from different Iranian Hospitals. ISRN microbiology. 2012;2012. [DOI:10.5402/2012/215275] [PMID] [PMCID]
 27. Li M, Cheung GY, Hu J, Wang D, Joo H-S, DeLeo FR, et al. Comparative analysis of virulence and toxin expression of global community-associated methicillin-resistant Staphylococcus aureus strains. Journal of Infectious Diseases. 2010;202(12):1866-76. [DOI:10.1086/657419] [PMID] [PMCID]
 28. Kennedy AD, DeLeo FR. Epidemiology and virulence of community-associated MRSA. Clinical Microbiology Newsletter. 2009;31(20):153-60. [DOI:10.1016/j.clinmicnews.2009.09.004]
 29. Ghasemian A, Peerayeh SN, Bakhshi B, Mirzaee M. Accessory gene regulator specificity groups among Staphylococcus aureus isolated from hospitalized children. Archives of Pediatric Infectious Diseases. 2014;2(4). [DOI:10.5812/pedinfect.16096]
 30. Parhizgari N, Khoramrooz SS, Hosseini M, Asghar SA, Marashifard M, Yazdanpanah M, et al. High frequency of multidrug-resistant Staphylococcus aureus with SCCmec type III and Spa types t037 and t631 isolated from burn patients in southwest of Iran. Apmis. 2016;124(3):221-8. [DOI:10.1111/apm.12493] [PMID]
 31. Montazeri EA, Khosravi AD, Jolodar A, Ghaderpanah M, Azarpira S. Identification of methicillin-resistant Staphylococcus aureus (MRSA) strains isolated from burn patients by multiplex PCR. Burns. 2015;41(3):590-4. [DOI:10.1016/j.burns.2014.08.018] [PMID]
 32. Abbasi-Montazeri E, Khosravi AD, Feizabadi MM, Goodarzi H, Khoramrooz SS, Mirzaei M, et al. The prevalence of methicillin resistant Staphylococcus aureus (MRSA) isolates with high-level mupirocin resistance from patients and personnel in a burn center. Burns. 2013;39(4):650-4. [DOI:10.1016/j.burns.2013.02.005] [PMID]
 33. Namvar AE, Afshar M, Asghari B, Lari AR. Characterisation of SCCmec elements in methicillin-resistant Staphylococcus aureus isolated from burn patients. Burns. 2014;40(4):708-12. [DOI:10.1016/j.burns.2013.09.010] [PMID]
 34. Rodrigues MVP, Fortaleza CMCB, Riboli DFM, Rocha RS, Rocha C, de Souza MdLR. Molecular epidemiology of methicillin-resistant Staphylococcus aureus in a burn unit from Brazil. Burns. 2013;39(6):1242-9. [DOI:10.1016/j.burns.2013.02.006] [PMID]

How to Cite This Article