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### Original Article

## Molecular and Seroepidemiological Survey of Visceral Leishmaniasis among Humans and Domestic Dogs in Mazandaran Province, North of Iran

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### ABSTRACT

**Background:** New cases of visceral leishmaniasis (VL) have been reported recently in some parts of Mazandaran Province, north of Iran where the first human case of VL was reported in 1949. This study aimed to determine the present status of *Leishmania infantum* infection among humans and domestic dogs using serological and molecular methods in central parts of Mazandaran Province.

**Methods:** In this cross-sectional study, blood samples were randomly collected from 402 humans and forty-nine domestic dogs throughout 2009 and 2010 in the central part of Mazandaran Province including Semeskadeh and Kiakola districts where recent cases of human visceral leishmaniasis had been reported there. All the collected samples were tested by direct agglutination test (DAT) for the detection of anti-*Leishmania infantum* antibodies as well as convenience PCR assay on whole blood samples for detection of leishmanial infection and identification of *Leishmania* species.

**Results:** None of 402 collected human (402) and dog (49) blood samples showed anti *Leishmania infantum* antibodies at titers 1:3200 and 1:320 as cut-off values of DAT, respectively but only 2 of domestic dogs (4.1 %) were found PCR-positive corresponding to *L. infantum*.

**Conclusion:** This study confirms the circulation of *L. infantum* at least among domestic dogs and highlights the sporadic pattern of VL in the studied areas. Further investigations regarding to sand flies fauna and wild canines as reservoir hosts of the disease, are recommended.

**Keywords:** Visceral leishmaniasis, *Leishmania infantum*, Seroprevalence, Direct agglutination test, convenience PCR, Iran

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## Introduction

Visceral leishmaniasis (VL), so-called Kala-azar, is a systemic disease caused by *Leishmania donovani* complex intracellular parasites, which are transmitted by different species of sand flies. The annual occurrence of human visceral leishmaniasis (HVL) cases worldwide is estimated to be 500,000 and accounts for 75,000 deaths (1-2). Nevertheless, these diseases are still considered as neglected diseases (3). Leishmaniasis still constitutes a major public health problem and burden is increasing (4). In addition *Leishmania*-HIV co-infections in the adult population are being reported with increasing frequency (1). The clinical signs of VL in humans include prolonged fever, hepatosplenomegaly, substantial weight loss, progressive anemia, and death (it is fatal in left treated cases) (5). *Leishmania infantum* is responsible for Mediterranean visceral leishmaniasis (MVL) in children and infants in the Mediterranean basin countries including Iran and domestic dogs are considered a major reservoir hosts for MVL (6). In Iran, the main endemic foci for VL are Fars and Bushehr provinces, in the south-west, the districts of Meshkin-shahr and Kaleybar in the north-west and Qom Province in the center of Iran (7-12). "Other parts of Iran are considered as sporadic areas for VL. Visceral leishmaniasis is common (over 98%) among children under 12 years old in different endemic foci in Iran and adult cases frequently present with subclinical and asymptomatic forms in endemic regions" (9-10).

The first case of human visceral leishmaniasis (HVL) in Iran has been reported by Pouya (13) from rural areas of Tonekabon, in western zone of Mazandaran Province, North of Iran. Moreover, at the same time, he reported the first case of

canine visceral leishmaniasis in this area. Afterward, a few infantile cases of HVL were reported in different parts of this province (14-16). At the present time, it is known as an endemic disease in some parts of five provinces of Iran and other parts of country are considered as sporadic areas of VL. Over the last decade, the incidence of VL has increased in many districts of the province of Mazandaran, in northern Iran (14-16).

This study aimed to determine prevalence of human and canine visceral leishmaniasis for the first time in the province. Since there are some reports correspond to exist possibly the *Leishmania* visceral infection among rodents in Semeskandeh district (17) along with one report of infected dogs and jackals in around of the area by Hamidi et al. (18) as well as new report of human VL case (identified as *L. infantum*) in Kiakola district from the Central zone of the province (15). Therefore, we designed a preliminary molecular and seroepidemiological investigation in these suspicious districts from the Central zone of this province.

## Materials and Methods

### Study area

The study was conducted throughout 2009-2010 in two suspicious districts of the Central zone of Mazandaran Province including Semeskandeh district (as a mountainous area including 7 villages) where five kilometer far from Sari city, capital of Mazandaran, and Kiakola district (as a coastal plain including 3 villages) where it's located at the littoral of Caspian Sea, Mazandaran Province is located in the north of Iran (53°6' E, 36°23' N). From the geographical point of view, Mazandaran Province is divided into two parts i.e. coastal

plain and the mountainous area. The central zone of the province has a humid weather and also has an annual mean rainfall of 977 mm (19).

### **Sampling and testing**

Blood samples were collected in EDTA-coated tubes from a total 20 % of less than 12 years old children and 10% of their parents, as well 20% of owner dogs from 11 villages. All samples were collected by cluster sampling methods. Totally, 402 human blood samples and 49 domestic dogs' blood samples were collected. All the samples centrifuged at  $1000\times g$  for 5 min and then plasma and buffy coat were collected individual micro tubes in order to DAT and PCR examination and were stored at  $-20^{\circ}\text{C}$  until examined. All plasma samples were tested by DAT and buffy coat were examined by PCR. Department of Parasitology, School of Public Health in Tehran University of Medical Sciences, supplied the DAT antigen and stored at  $4^{\circ}\text{C}$  until used. Plasma samples were tested by DAT according to the methods described by Harith et al. (20). First dilutions were prepared from 1:10 to 1:80 for dog samples and 1:10 to 1:800 for human samples. Samples with titers 1:80 for dogs, 1:800 for humans were diluted further to give final titers. Known negative and positive controls were tested in each plate. In this investigation, we considered anti-*Leishmania* antibodies titers at equal and above of 1:3200 and 1:320 (cut off point) as *Leishmania* infection for the human and dogs respectively. Total DNA was extracted from blood buffy coat as described by Motazedian et al. (21). Briefly, 200  $\mu\text{l}$  of buffy coat was homogenized with 200  $\mu\text{l}$  lyses buffer [50 mM Tris-HCl (pH=7.6), 1 mM EDTA and 1% Tween 20] and 10  $\mu\text{l}$  of proteinase K solution (containing 19 mg of the enzyme/ml), in a 1.5 ml micro centrifuge tube. The homogenate was then incubated at  $37^{\circ}\text{C}$  overnight

before 200  $\mu\text{l}$  of a phenol: chloroform: isoamyl alcohol mixture was added. After being shaken vigorously, the tube holding the mix was centrifuged ( $10,000\times g$  for 10 min) and then the DNA in the supernatant solution was precipitated with 400  $\mu\text{l}$  cold, pure ethanol, re-suspended in 50  $\mu\text{l}$  double-distilled water and then stored at  $4^{\circ}\text{C}$  until it could be tested. It was re-suspended in 100  $\mu\text{l}$  sterile distilled water and stored at  $4^{\circ}\text{C}$  until it could be tested in a modified genus-specific PCR for a sequence from the kinetoplast DNA (k DNA) of *Leishmania*. The primers used, RV<sub>1</sub> (5'-CTT TTC TGG TCC CGC GGG TAG G-3') and RV<sub>2</sub> (5'-CCA CCT GCG CTA TTT TAC ACC A-3') amplify a 145-bp sequence from the LT1 fragment of the parasites' kDNA minicircles, according to the methods as described by Fakhar et al. (22).

The PCR products were separated by electrophoresis in a 2% agarose gel, stained with ethidium bromide, visualized under ultra-violet trans-illumination, and sized by comparison with a 100bp ladder. Each sample found PCR-positive for Leishmanial DNA was then investigated using the PCR described by Fakhar et al. (22), which is based on the species-specific primers LINR4 and LIN17 to confirm that the DNA detected was that of *L. infantum*. Reference strain of *L. infantum* (MCAN/IR/96/Lon49) was used as standard.

### **Data analysis**

Chi-squared ( $\chi^2$ ), Mac Nemar and Fisher exact tests were used to compare seroprevalence values relative to gender, age groups and two studied areas. Analyses were performed with SPSS (version 13.5; SPSS Inc, Chicago, IL, USA) and Epi-Info software, with a probability ( $P$ ) value of  $<0.05$  were considered as statistically significant.

## Results

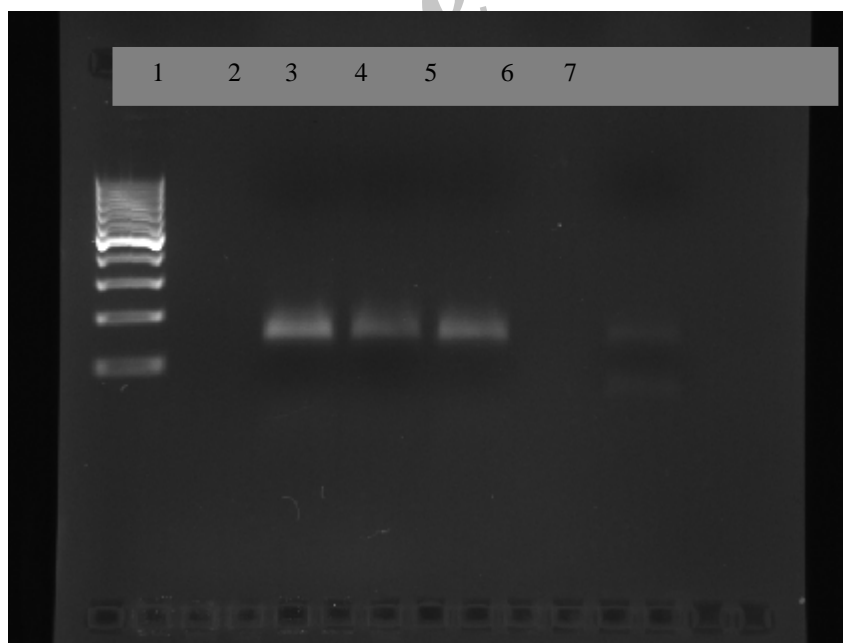
Of the 402 subjects (200 males and 202 females) investigated, 138 cases (31.9%) were showed *Leishmania* specific antibodies with titers 1:800, 8 cases (2%) with 1:1600 titers which those considered as suspicious cases. Also none of those were found DAT (at titers  $\geq 1:3200$ ) and PCR positive. Out of 49 dogs ( 38 males and 11 females) examined, 17 (34.6%) were showed *Leishmania* specific antibodies with titers 1:80 and only two dogs (4.1%), belonged to Kiakola district, were found PCR positive in the *Leishmania* genus -specific PCR based on the RV1/ RV2 primer set (Fig. 1) and none of those were found PCR positive from Semskande district.

In addition, none suspicious human cases ( $< 1:3200$  titers, as cut off) had clinical signs and only one of PCR positive dogs had clinical signs including skin lesions, alopecia

and weight loss along with anti *Leishmania* specific antibodies at 1:80 titer.

There was no significant difference between male and female at 1:800 and 1:1600 titers of anti-*Leishmania* specific antibodies in human population and in dog population as well. Although, the number of human cases with 1:800 and 1:1600 titers in females were higher than males. As well as, there was no significant difference in age groups of the subjects.

Moreover, there was no significant difference between two studied areas. Although all human suspicious cases ( $< 1:3200$  titers, as cut off) have now been followed-up for at least 11 months, none has yet shown any symptoms of VL.



**Fig. 1:** 2% agarose gel electrophoresis of PCR products from buffy coat DNA of dogs Lane 1: standard marker (100 bp), Lane 2: negative samples, Lane 3: Positive control, Lane 3: Standard *Leishmania infantum* (145 bp), lanes 4, 5: positive dogs, Lane 6: negative samples, Lane 7: positive sample

## Discussion

For the first time, our preliminary study was carried out in Mazandaran Province. Thus we used DAT for serological test on humans, and dogs' plasma samples as a practical and sensitive test in seroepidemiological studies (20, 23). Serological results by DAT showed in spite of 34.6% human populations in the studied areas had anti-*Leishmania* antibodies in titers of 1:800 and 2% had anti-*Leishmania* antibodies in titers 1:1600 but none of them were DAT positive at titers 1:3200 and higher as cut-off values of DAT on human sera. The direct agglutination test has been used in epidemiologic studies in several endemic areas and is used in large-scale screening of human VL as a simple, valid test with high sensitivity and specificity at cut-off titer 1:3200 (10, 23). The sensitivity and specificity of this method varies in different studies between 90–100% and 72–100% respectively (24).

As the type of sampling is noninvasive, we also used genus-specific PCR based on RV<sub>1</sub> and RV<sub>2</sub> specific primers for *L. infantum* on peripheral blood buffy coats of humans and dogs (24, 25). The results of PCR based on RV<sub>1</sub> and RV<sub>2</sub> primers for all human buffy coats were negative and all of them had no clinical signs.

Recently, the use of polymerase chain reaction with high sensitivity (70–100%) and specificity (100%) has become popular in different parts of the world (24, 25) and the sensitivity of 82.1% and specificity of 100% in our previous study in Fars Province, as known endemic region of VL, (22) in Iran.

The PCR technique has several advantages including the ability to work with small amounts of target material, fast detection of *Leishmania* in symptomatic patients and asymptomatic carriers as well as *Leishmania*/HIV co-infected patients and the

follow up of treatment as well as the assessment of the successful cure of visceral leishmaniasis (26-28).

Six primer pairs were compared for detecting *L. infantum* DNA by Gao et al. (29). The primer pairs RV<sub>1</sub> - RV<sub>2</sub> (0.1 parasite/ml blood) were most sensitive and suitable in detecting the asymptomatic infection of *L. infantum* and the prevalence of the asymptomatic infection is high in human population in the endemic area. In our study, we applied RV<sub>1</sub> - RV<sub>2</sub> primers set as described above. The PCR on peripheral blood sample can be used for treatment response evaluation and it is also very efficient for early detection of the disease especially in patients with cryptic infection, small children and immunocompromised patients (30) and also subclinical and asymptomatic infections in endemic areas (22).

Serological results by DAT showed 34.7% dog population had anti-*Leishmania* antibodies at titers 1:80 but none of them showed anti-*Leishmania infantum* antibodies as cut-off titer 1:320. The results of PCR based with RV<sub>1</sub>, RV<sub>2</sub> primer set revealed two dogs were found positive. Both of them were from the Kiakola district, one of them exactly was from the village where new human VL case had previously reported. Only one of PCR positive dogs had clinical signs. None of those was seropositive and had anti-*Leishmania* antibodies in titers 1:80. It is noticeable that the symptomatic dog lives in the same place where new human case of VL has been reported by Rahmati et al. (15).

After one year, in following up the first dog (asymptomatic dog) had not developed in clinical signs but another one was died. So it

seems the evidences of circulation *L. infantum* in the sporadic area.

Recent epidemiological reports in endemic regions of CVL, such as Iran, indicate that asymptomatic dogs' infections with *L. infantum* occur in more than 50%-70% of the seropositive dogs in the field investigations (9, 12, 31). The canidae families are main reservoir hosts because the parasites are multiplied in skin macrophages and readily transferred by feeding sand flies (32). In addition, the wild carnivores such as jackals and foxes are considered reservoir hosts in sylvatic cycle of MVL, principally in sporadic areas of Iran (9).

The diagnosis of VL is complex because commonly occurring diseases such as malaria, typhoid, and tuberculosis have clinical features similar to VL. Some VL cases have been misdiagnosed as autoimmune hepatitis, acute lymphoblastic leukemia, and malignant lymphoma (33-34). Most of these misdiagnosed patients are reported from non endemic regions where physicians do not expect the occurrence of the disease. Moreover, atypical cells and different blast may be observed in bone marrow aspiration of VL patients (34). Consequently, it seems that in the Mazandaran Province, some of VL cases (particularly, atypical and subclinical or asymptomatic) due to lack of specific and sensitive diagnostic tests such as DAT and PCR as well as lack of physician's awareness of existing the VL in the region (as a non endemic region) are most likely misdiagnosed. On the other hand some patients had referred to Tehran (as capital of Iran) hospitals in order to more management, early and accurate diagnosis.

As a whole, according to aforementioned and our investigations look like VL occur as cryptic forms in some parts of the province and it may be a potential risk for emerging or reemerging a new focus in future. Though, recently one autochthonous case of VL with

no history of traveling to endemic areas has been reported from the region. However, it is difficult to explain that how she acquired infection. We can not rule out the possibility that this *L. infantum* strain is less virulent and might be circulating in some animal reservoirs. It is likely that the disease has been re-introduced in the region and is spread by some local species of sandfly.

This preliminary study confirms the circulation of *L. infantum* at least among dogs population and highlights the sporadic pattern of VL in the studied areas. Moreover, several evidences such as neglecting MVL in the province, lack of physician's awareness, discontinue or non-effective spray insecticides against malaria vectors, environmental changes and climate conditions, nomadic movements and increasing incidence of VL (35) recommend the reemergence of MVL in this non-endemic area, where the first human case of visceral leishmaniasis had been reported in Iran.

Since all human and the majority of dog population were asymptomatic as well, it showed role of asymptomatic dogs as a possible reservoir host for VL. As a whole, further investigations regarding sandflies fauna and animal reservoirs and human populations are required in this province.

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