Vascular Endothelial Growth Factors and Their Inhibitors in Ocular Neovascular Disorders

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The introduction of pharmacotherapy has been one of the most important advances in the management of retinal disorders. Age-related macular degeneration (AMD) was once considered as an irrepressible disease leading to permanent macular damage, however anti-vascular endothelial growth factor (anti-VEGF) agents are now believed to halt the progression of the condition and improve vision in a considerable proportion of patients. Anti-VEGF agents are among the most commonly used drugs in ophthalmology but questions and uncertainties still surround their indications, efficacy and complications. This paper reviews the role of VEGF under physiologic and pathologic conditions in the eye and available anti-VEGF agents in current ophthalmic practice.

Key words: Vascular Endothelial Growth Factor; Macular Degeneration

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INTRODUCTION

More than 60 years ago, Michaelson proposed the concept of stimulation of neovascular retinopathies by an unknown factor, the so-called "X factor". Nowadays the role of vascular endothelial growth factors (VEGFs) has been increasingly clarified in the pathogenesis of neovascularization processes and other mediators are expected to be recognized in future.¹ A large body of evidence suggests VEGF-A to be the main regulator of pathologic angiogenesis.² Therefore, targeted inhibition of this mediator seems to be biologically plausible for treatment of these conditions.

Pathologic angiogenesis has a central role in age-related macular degeneration (AMD) which is the leading causes of blindness in elderly subjects in developed countries.¹ Recently, anti-VEGF agents have received FDA approval for treatment of AMD and have greatly changed the outcomes of treatment in these patients.^{3,4} Off-label administration of anti-VEGF agents is also commonly practiced for treatment of proliferative diabetic retinopathy (PDR) and diabetic macular edema (DME) and less commonly for retinal vein occlusion (RVO), other retinal vasoproliferative disorders and neovascular glaucoma (NVG).⁵⁻¹⁰

This review outlines current information on the mechanisms of action of VEGF under physiologic and pathologic conditions in the eye and overviews the clinical utility of available anti-VEGF agents.

VASCULAR ENDOTHELIAL GROWTH FACTOR (VEGF)

The VEGF family includes placental growth factor (PLGF), VEGF-A, VEGF-B, VEGF-C, VEGF-D and VEGF-E. These agents are the most important regulators of angiogenesis and vascular permeability; VEGF-A in particular, plays a pivotal role in pathologic ocular angiogenesis.¹¹ The VEGF-A gene has been localized to chromosome 6p12.3 and comprises of 8 ex-

ons and 8 intermediate introns. VEGF-A has 9 isoforms including VEGF₁₂₁, VEGF₁₄₅, VEGF₁₄₈, VEGF₁₆₂, VEGF₁₆₅, VEGF₁₆₅, VEGF₁₈₃, VEGF₁₈₉ and VEGF₂₀₆. These isoforms differ from each other by the number of amino acids and heparin-binding affinity.^{12,13} Heparin-binding affinity is nil for VEGF₁₂₁, maximum for VEGF₁₈₉ and VEGF₂₀₆, and intermediate for VEGF₁₆₅. This property determines the affinity of the molecule for binding to heparin-sulfate proteoglycans in cell membranes.¹⁴

VEGF appears to be essential for development and maintenance of functionally efficient retinal vasculature as well as for integrity of the retinal pigment epithelium (RPE), Bruch's membrane and choroidal endothelial cells. VEGF release mainly occurs on the basal surface of the RPE. VEGF-A production is induced by factors including hypoxia, nitric oxide, and other biologic growth factors including hypoxia-induced factor 1. The latter comprises of α and β subunits, the production of the α subunit acts as a VEGF-A gene stimulant and has been shown to be increased under hypoxic conditions.¹⁵

VEGF-A induces angiogenesis, increases vascular permeability, may affect female reproductive function, hair growth and wound healing in adults. It also seems to inhibit endothelial cell apoptosis and is therefore considered a survival factor.¹⁶

VEGF acts via two tyrosine kinase receptors including VEGF receptor-1 (VEGFR-1) and VEGFR-2 which are primarily present on vascular endothelial cell surfaces. Although the affinity of VEGFR-2 for VEGF is lower than VEGFR-1 it plays a more significant role in neovascularization processes.¹⁷ Another receptor named neuropilin-1 has also been recognized for VEGF₁₆₅ and is present on neurons and tumor cells in addition to endothelial cells.

PLGF binds to VEGFR-1 resulting in production of VEGF-A. PLGF is a weak mitogen.¹⁸ VEGF-B binds to both VEGFR-1 and neuropilin-1, causes destruction of extracellular matrix and has a role in cell migration. VEGF-C and VEGF-D are distinguished from other members of the family by possessing C and N terminal structures and affect endothelial cell mitosis and migration. Both are preproproteins which require plasmin for activation. VEGF-C has a major role in lymphangiogenesis in general while VEGF-D mediates lymphangiogenesis only in lymphatic tumors. VEGF-E and VEGF-F are non-human homologues of VEGF-A. The former acts via VEGFR-2 and strongly induces endothelial cell proliferation but the role of the latter remains unknown.¹⁹⁻²¹

RETINAL NEOVASCULARIZATION

Hypoxia is the major trigger of VEGF-associated neovascularization. Induction of hypoxia in vitro increases production of VEGF-A in different cell lines in the retina. Similarly, under hypoxic conditions in vivo, increased levels of VEGF-A were observed in the retina, vitreous and aqueous humor.^{22,23} Miller et al²⁴ showed that VEGF-A levels in ocular tissues change proportionately with increase and decrease in the amount of new vessels.

Main sources of VEGF-A in the retina and vitreous are RPE, Muller and ganglion cells. Injection of VEGF-A in non-human primates resulted in clinical features similar to diabetic retinopathy (DR) and NVG.^{25,26} In humans, VEGF-A has been implicated in the pathogenesis of retinal and choroidal neovascularization in conditions such as DR,^{22,23} RVO,^{23,27} iris neovascularization,²⁷ retinopathy of prematurity (ROP)²⁸ and AMD.²⁹⁻³¹

VEGF-A may also play a role in intraocular inflammation through increased vasopermeability^{32,33} which is probably mediated via the following mechanisms: leukocyte-mediated endothelial cell damage, destruction of tight junctions, fenestrations in vessels, and transcellular bulk flow. The concentration of VEGF and contact time for induction of vasopermeability and break-down of the blood-retinal barrier may be less than that required for angiogenesis.²⁶ The mechanism of action of corticosteroids in improving macular edema may be decreased leukocyte-mediated damage induced by VEGF-A.³⁴

AGE-RELATED MACULAR DEGENERATION

AMD is the most prevalent disease associated with choroidal neovascularization (CNV) and

the most common subject for anti-VEGF studies in ophthalmology. Progressive macular degeneration results in drusen formation and pigmentary changes in the RPE in dry AMD leading to geographic atrophy of the macula and severe visual loss. Neovascular (wet) AMD is associated with formation of choroidal neovascular membranes. Although wet type AMD comprises only 10% of cases of AMD, it is responsible for 90% of severe visual loss due to the condition.³⁵ Development of CNV is associated with increased levels of VEGF.²⁹⁻³¹

Oxidant agents seem to precipitate in Bruch's membrane resulting in stimulation of complement and inflammatory pathways leading to up-regulation of VEGF secretion from RPE cells eventually resulting in CNV.³⁶ Patients with moderate to severe types of dry type AMD profit from antioxidants.³⁷

Many therapeutic modalities have been employed for neovascular AMD. Macular laser photocoagulation provides relative success in a small percentage of patients with extrafoveal CNV but may cause severe scotoma and immediate loss of vision. Photodynamic therapy (PDT) with verteporfin acts via activating a photosensitizing dye within the pathologic vessels by infrared laser leading to occlusion of choroidal new vessels with minimal damage to the retina. PDT can delay loss of vision in certain cases of CNV especially in small-sized CNVs but does not improve vision.³⁸

Anti-VEGF agents have changed the paradigm of treatment for neovascular AMD with dramatic outcomes. Pegaptanib is the first FDA-approved anti-VEGF agent for intravitreal injection in the treatment of neovascular AMD.³ The VISION study³⁹ showed that intravitreal injection of 0.3 mg pegaptanib every 6 weeks reduced the rate of visual loss of more than 30 letters to 10% as compared to 22% in controls; after 2 years visual acuity remained unchanged but no visual improvement was seen.

The advent of ranibizumab raised the expectations of treatment for neovascular AMD.⁴⁰ The MARINA study^{40,41} evaluated the effect of ranibizumab injection in patients with minimally classic or occult CNV and the ANCHOR study⁴²⁻⁴⁴ assessed ranibizumab injection in patients with predominantly classic CNV; both

studies used monthly injections and reported that 90% of treated patients maintained vision or experienced only mild visual loss. After 2 years of follow-up, 40% of treated patients had visual acuity of 20/40 or better in both studies. The PIER study⁴⁵ utilized 3 injections during a 2-month period followed by repeat injections every 3-4 months, but the outcomes were not as favorable as those achieved in the MARINA and ANCHOR studies.

Intravitreal injection of bevacizumab has been shown to be effective in the management of neovascular AMD and has been widely used since 2005.46,47 Ahmadieh et al48 evaluated the effect of initial triple therapy using PDT combined with intravitreal bevacizumab and triamcinolone followed by additional intravitreal injections of bevacizumab when active disease persisted thereafter and concluded that this method is beneficial in terms of retinal thickness reduction and visual acuity improvement after mean follow-up of 50±15 weeks. The Comparison of AMD Treatments Trial Study compares bevacizumab with ranibizumab using monthly injections versus 3 consecutive injections followed by additional injections as needed (www.clinicaltrials.gov).

Other therapeutic methods include a single intravenous injection of VEGF-Trap, which is under phase I trial. Intravenous VEGF-Trap seems to result in adverse effects with doses higher than 1 mg/kg.⁴⁹ Intravitreal injection of VEGF-Trap has also been used.⁵⁰ Another treatment also under phase I trial is a single injection of siRNA that resulted in an average increase of 15 letters of visual acuity in 23% of treated patients.⁵¹ More than 40 studies are currently under execution in the USA under supervision by the National Health Institute of USA evaluating different anti-VEGF therapies in AMD which are available at the clinical trials website (www.clinicaltrials.gov).

NON-AMD ASSOCIATED CNV

CNV may be seen in conditions other than AMD such as high myopia, angioid streaks, ocular histoplasmosis, and choroidal osteoma. Anti-VEGF therapy in these conditions is almost limited to off-label use of bevacizumab and there are a few small-sized reports in the literature in this regard.⁵²⁻⁵⁶

DIABETIC RETINOPATHY

Prolonged hyperglycemia activates the β isoform of protein kinase-C (PKC- β) leading to VEGF synthesis. Elevated levels of VEGF lead to PDR, vascular leakage and fluid collection manifesting as DME. Early vascular changes in diabetic retinopathy demonstrate leukostasis (adhesion of leukocytes to the vascular bed), platelet aggregation, abnormal blood flow, degeneration of pericytes, and thickening of basement membranes.^{22,23}

Retinal capillary occlusion increases synthesis of angiogenesis stimulant factors such as VEGF via regional hypoxia. These factors loosen junctions between vascular endothelial cells increasing vasopermeability.^{22,23} VEGF is an important survival factor for endothelial cells, however proliferated endothelial cells degenerate and result in capillary occlusion, generation of acellular capillaries and capillary nonperfusion.^{22,23}

Based on these findings, inhibition of VEGF and PKC pathways are pivotal in the treatment of diabetic retinopathy. Pegaptanib is under phase II trials for treatment of DME; it has improved visual acuity and decreased macular edema in comparison to placebo. A retrospective evaluation of these patients also revealed regression of retinal neovascularization.⁵ Several studies have reported rapid regression of neovascularization in the iris (NVI), optic nerve (NVD) and retina (NVE) following intravitreal injection of bevacizumab in patients with PDR.8-10 Some authors have used bevacizumab as an adjunct to vitrectomy in patients with vitreous hemorrhage due to PDR.57-59

Ruboxistaurin has been evaluated for diabetic retinopathy in 2 large scale multicenter studies as follows: in the PKC-DRS study,⁶⁰ oral ruboxistaurin (32 mg) had no effect on progression of PDR but decreased the severity of visual loss probably by decreasing macular edema. In the PKC-DRS2 study,⁶¹ less visual loss, visual improvement and reduction in macular edema were reported. One may conclude that in patients with PDR or DME, standard treatments such as panretinal photocoagulation (PRP), macular photocoagulation (MPC) and vitrectomy with established indications and outcomes may be advisable. However, in cases with severe and persistent macular edema unresponsive to standard therapy, anti-VEGF agents may be used as adjunctive treatment. Additionally, in patients with vitreous hemorrhage associated with advanced PDR, preoperative injection of anti-VEGF agents (usually one week before vitrectomy) seems to be advantageous.⁵⁷⁻⁵⁹

CENTRAL AND BRANCH RETINAL VEIN OCCLUSION

In retinal vein occlusions, VEGF levels are increased proportionate to the degree of retinal ischemia and severity of macular edema. Primary studies have reported favorable results using ranibizumab and bevacizumab.⁶²⁻⁶⁴ Anti-VEGF drugs are potent agents for reduction of edema; they reduce vasopermeability but cannot eliminate the pathologic process completely. The duration of action is not clear but their effect seems to decrease after a few weeks and re-injections are required after 4 weeks.^{65,66}

NEOVASCULAR GLAUCOMA (NVG)

NVG develops due to iris and angle neovascularization in response to retinal ischemia. This process may complicate diabetic retinopathy, vascular occlusions, sickle cell anemia, old retinal detachments and the ocular ischemic syndrome. Standard treatment for NVG consists of relieving retinal ischemia by using PRP along with medical or surgical control of elevated intraocular pressure (IOP). When a large area of the anterior chamber angle is involved by neovascular tissue, shunting procedures are required. Elevated levels of VEGF in the aqueous humor and vitreous have been shown in a few studies.² Several studies have reported rapid regression of NVI following intravitreal injection of bevacizumab.67-69 The combination of PRP and IVB seems an appropriate treatment for NVG; IVB prevents angle closure in the time PRP needs time to take effect.⁷⁰

RETINOPATHY OF PREMATURITY (ROP)

ROP is characterized by incomplete vascularization of the peripheral retina in a premature neonate leading to retinal neovascularization. Risk factors for ROP include premature birth, low birth weight and oxygen therapy. Experimental studies have shown that high concentrations of oxygen impair vascular formation which in turn causes hypoxia and elevates VEGF levels.71,72 Increased levels of VEGF in the vitreous have been reported in patients with ROP.73 It is notable that VEGF has a physiologic role in the formation of normal retinal vasculature. Another factor implicated in the pathogenesis of ROP is insulin-like growth factor-1 (IGF-1) which stimulates angiogenesis in a pathway independent of VEGF.⁷⁴ Ongoing studies⁵¹ are evaluating the use of anti-IGF-1 and anti-VEGF agents for treatment of ROP, the results of which are not published yet.

ANTI-ANGIOGENESIS AGENTS

Pegaptanib sodium

Pegaptanib sodium (Macugen, Evetech Pharmaceuticals, New York, USA) is a modified 28base pegylated RNA aptamer which binds to VEGF₁₆₅ and other larger isoforms of VEGF-A.³ It was the first VEGF inhibitor to receive FDA approval for treatment of CNV due to AMD in December 2004. In its phase III trial, three different doses of intravitreal pegaptanib (0.3, 1 and 3 mg) were used every 6 weeks for 2 years in comparison with placebo in three types of AMD (with minimally classic, classic and occult CNV). After 12 months, 30% of the 0.3 mg group and 45% of controls experienced loss of vision more than 15 letters equal to 3 lines (P=0.001). On the other hand, 6% of the 0.3 mg group and 2% of the placebo group achieved 3 lines or more visual improvement (P=0.04). After 2 years, pegaptanib significantly decelerated loss of vision but did not improve it.39

Ranibizumab

Ranibizumab (Lucentis, Genentech, San Fran-

cisco, USA) received FDA approval for treatment of neovascular AMD after pegaptanib in 2006. It is a humanized Fab fragment of a murine monoclonal anti-VEGF-A antibody which binds to all VEGF-A isoforms and their active cleavage by-products.⁴

The efficacy and safety of ranibizumab have been demonstrated in all types of AMDrelated CNV based on the results of two phase III clinical trials.⁴⁰⁻⁴⁴ In the MARINA study^{40,41} patients with minimally classic or occult disease underwent monthly injections of 0.3 or 0.5 mg ranibizumab for 2 years. Ultimately, visual acuity increased by 7 letters in treated groups but decreased by 10.5 letters in controls (P<0.01). Another phase III clinical trial compared ranibizumab (0.3 and 0.5 mg) with PDT in the treatment of classic CNV. After 2 years, monthly injections of ranibizumab were superior to PDT.42,43 Ranibizumab was the first anti-VEGF agent capable of increasing visual acuity in patients with neovascular AMD in addition to preventing visual loss.⁴⁰⁻⁴⁴ The rate of ocular complications related to ranibizumab was less than 1.7% in both of the abovementioned studies.

Bevacizumab

Bevacizumab (Avastin, Genentech, San Francisco, USA) is a full-length humanized monoclonal antibody similar to ranibizumab that binds all isoforms of VEGF-A and their active by-products. It was approved by the FDA exclusively for intravenous administration in the treatment of colorectal malignancies.⁷⁵ It has also been used as an off-label drug for ocular neovascular diseases such as AMD and has shown to be safe and effective with shortterm follow-up.⁷⁶⁻⁷⁷ The long-term safety is yet to be established; this demands large scale clinical trials with longer follow-up. Such studies have been initiated and their results are anticipated.

VEGF-Trap

VEGF-Trap (Regeneron, Pharmaceuticals, New York, USA) is a protein molecule which binds to VEGFR-1 and VEGFR-2, inhibiting the effect

of all isoforms of VEGF and PLGF.⁷⁸ Intravenous administration for treatment of AMD is under phase I/II clinical trials.⁴⁹

Sirna

Sirna (Sirna Therapeutics, Boulder, Colorado) is a short ribonucleic acid (RNA) that interferes with VEGFR-1 production at the gene level. This agent is under phase II clinical trial for treatment of AMD. Cand-5 (Acuity Pharmaceuticals, Philadelphia, USA) acts in a fashion similar to Sirna.⁵¹

Ruboxistaurin mesylate

Ruboxistaurin mesylate (Arxxant, Lilly, Indianapolis, USA) acts as a selective systemic inhibitor of PKC- β . It has been shown to be safe and effective in maintaining visual acuity in diabetic patients.^{60,61,79}

Multireceptor Tyrosine Kinase Inhibitors

Simultaneous inhibition of several tyrosine kinase receptors including VEGFR-1, VEGFR-2 and receptors for stem cell factor, fetal liver TK₃ and PDGF is theoretically advantageous to inhibition of VEGF alone. Vatalanib (Novartis, Pharma AG, Basel, Switzerland, PTK 787/ 2K222584) is an inhibitor of tyrosine kinase receptors and currently under phase I/II clinical trials in combination with PDT for treatment of AMD.⁸⁰ The following agents from this group have also been used in animal models: AG013764/AG013711 (Pfizer Inc, New York, USA), Sutent (Sunitinib, Pfizer Inc, New York, USA), Compound A (Merck and Co., Rome, Italy), and TG 100801 (Targeger, San Diego, USA).81-84

REFERENCES

- Friedman DS, O'Colmain BJ, Muñoz B, Tomany SC, McCarty C, de Jong PT, et al. Prevalence of agerelated macular degeneration in the United States. *Arch Ophthalmol* 2004;122:564-572.
- 2. Tripathi RC, Li J, Tripathi BJ, Chalam KV, Adamis AP. Increased level of vascular endothelial growth factor in aqueous humor of patients with neovascular glaucoma. *Ophthalmology* 1998;105:232-237.

Potential Hazards of Anti-VEGF Therapy

The most important adverse local effects related to anti-VEGF agents include uveitis, retinal detachment and cataracts. Based on the results of the VISION study,³⁹ the incidence of these complications over one year was 0.08% retinal detachment, 0.16% for for endophthalmitis, 0.7% for traumatic cataracts and 1% for moderate uveitis. For prevention of infectious complications, skin preparation and drapes, irrigation of the fornices, using a lid speculum and adherence to aseptic principles are required.

No adverse systemic effect was seen in the VISION study³⁹ up to 2 years after injection of pegaptanib but in the MARINA^{40,41} and ANCHOR^{42,43} studies, an increased incidence of MI and stroke was seen in patients who received 0.5 mg ranibizumab as compared to controls (1.3% versus 2.9%). Since then, Genentech stated that intravitreal injection of 0.5 mg ranibizumab may increase the risk of CVA in patients with a positive previous history.

Due to lack of large scale clinical trials potential side effects of bevacizumab remain vague, however this agent also seems to entail local and systemic complications similar to ranibizumab or pegaptanib.

Major concerns with anti-VEGF therapy for ocular diseases include: (1) repeat intravitreal injections; (2) risk of CVA or cardiovascular complications; (3) possible retinal and neural toxicity due to cumulative dosing; (4) interference with physiologic functions of VEGF; and (5) economic and cost-effectiveness concerns.

- Gragoudas ES, Adamis AP, Cunningham ET Jr, Feinsod M, Guyer DR, VEGF Inhibition Study in Ocular Neovascularization Clinical Trial Group. Pegaptanib for neovascular age-related macular degeneration. N Engl J Med 2004;351:2805-2816.
- Rosenfeld PJ, Brown DM, Heier JS, Boyer DS, Kaiser PK, Chung CY, et al. Ranibizumab for neovascular age-related macular degeneration. *N Engl J Med* 2006; 355:1419-1431.
- 5. Cunningham ET Jr, Adamis AP, Altaweel M, Aiello LP, Bressler NM, D' Amico DJ, et al. A phase II

randomized double-masked trial of pegaptanib, an antivascular endothelial growth factor aptamer, for diabetic macular edema. *Ophthalmology* 2005;112:1747-1757.

- 6. Soheilian M, Ramezani A, Bijanzadeh B, Yaseri M, Ahmadieh H, Dehghan MH, et al. Intravitreal bevacizumab (avastin) injection alone or combined with triamcinolone versus macular photocoagulation as primary treatment of diabetic macular edema. *Retina* 2007;27:1187-1195.
- Ahmadieh H, Ramezani A, Shoeibi N, Bijanzadeh B, Tabatabaei A, Azarmina M, et al. Intravitreal bevacizumab with or without triamcinolone for refractory diabetic macular edema; a placebocontrolled, randomized clinical trial. *Graefes Arch Clin Exp Ophthalmol* 2008;246:483-489.
- 8. Moradian S, Ahmadieh H, Malihi M, Soheilian M, Dehghan MH, Azarmina M. Intravitreal bevacizumab in active progressive proliferative diabetic retinopathy. *Graefes Arch Clin Exp Ophthalmol* 2008;246:1699-1705.
- Stahl A, Agostini H, Hansen LL, Feltgen N. Bevacizumab in retinal vein occlusion, results of a prospective case series. *Graefes Arch Clin Exp Ophthalmol* 2007;245:1429-1436.
- Yazdani S, Hendi K, Pakravan M. Intravitreal bevacizumab (Avastin) injection for neovascular glaucoma. *J Glaucoma* 2007;16:437-439.
- Leung DW, Cachianes G, Kuang WJ, Goeddel DV, Ferrara N. Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science* 1989;246:1306-1309.
- 12. Takahashi H, Shibuya M. The vascular endothelial growth factor (VEGF)/VEGF receptor system and its role under physiological and pathological conditions. *Clin Sci (Lond)* 2005;109:227–241.
- Lange T, Guttmann-Raviv N, Baruch L, Machluf M, Neufeld G. VEGF162, a new heparin-binding vascular endothelial growth factor splice form that is expressed in transformed human cells. *J Biol Chem* 2003;278:17164-17169.
- 14. Houck KA, Leung DW, Rowland AM, Winer J, Ferrara N. Dual regulation of vascular endothelial growth factor bioavailability by genetic and proteolytic mechanisms. *J Biol Chem* 1992;267:26031-26037.
- Levy NS, Chung S, Furneaux H, Levy AP. Hypoxic stabilization of vascular endothelial growth factor mRNA by the RNA-binding protein HuR. *J Biol Chem* 1998;273:6417-6423.
- Ferrara N. Vascular endothelial growth factor: basic science and clinial progress. *Endocr Rev* 2004;25:581-611.
- Soker S, Takashima S, Miao HQ, Neufeld G, Klagsbrun M. Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor. *Cell* 1998;92:735-745.
- 18. Yonekura H, Sakurai S, Liu X, Migita H, Wang H,

Yamagishi S, et al. Placenta growth factor and vascular endothelial growth factor B and C expression in microvascular endothelial cells and pericytes: implication in autocrine and paracrine regulation of angiogenesis. *J Biol Chem* 1999;274:35172-351728.

- 19. Wise LM, Veikkola T, Mercer AA, Savory LJ, Fleming SB, Caesar C, et al. Vascular endothelia growth factor (VEGF)-like protein from orf virus NZ2 binds to VEGFR2 and neuropilin-1. *Proc Natl Acad Sci U S A* 1999;96:3071-3076.
- 20. Junqueira de Azevedo IL, Farsky SH, Oliveira ML, Ho PL. Molecular cloning and expression of a functional snake venom vascular endothelium growth factor (VEGF) from the Bothrops insularis pit viper. A new member of the VEGF family of proteins. *J Biol Chem* 2001;276:39836-39842.
- 21. Takahashi H, Hattori S, Iwamatsu A, Takizawa H, Shibuya M. A novel snake venom vascular endothelial growth factor (VEGF) predominantly induces vascular permeability through preferential signaling via VEGF receptor-1. *J Biol Chem* 2004;279:46304-46314.
- 22. Adamis AP, Miller JW, Bernal MT, D'Amico DJ, Folkman J, Yeo TK, et al. Increased vascular endothelial growth factor levels in the vitreous of eyes with proliferative diabetic retinopathy. *Am J Ophthalmol* 1994;118:445-450.
- 23. Aiello LP, Avery RL, Arrigg PG, Keyt BA, Jampel HD, Shah ST, et al. Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. *N Engl J Med* 1994;331:1480-1487.
- 24. Miller JW, Adamis AP, Shima DT, D'Amore PA, Moulton RS, O'Reilly MS, et al. Vascular endothelial growth factor/vascular permeability factor is temporally and spatially correlated with ocular angiogenesis in a primate model. *Am J Pathol* 1994;145:574-584.
- 25. Tolentino MJ, Miller JW, Gragoudas ES, Jakobiec FA, Flynn E, Chatzistefanou K, et al. Intravitreous injections of vascular endothelial growth factor produce retinal ischemia and microangiopathy in an adult primate. *Ophthalmology* 1996;103:1820-1828.
- 26. Tolentino MJ, Miller JW, Gragoudas ES, Chatzistefanou K, Ferrara N, Adamis AP. Vascular endothelial growth factor is sufficient to produce iris neovascularization and neovascular glaucoma in a nonhuman primate. *Arch Ophthalmol* 1996;114:964-970.
- Pe'er J, Shweiki D, Itin A, Hemo I, Gnessin H, Keshet E. Hypoxia-induced expression of vascular endothelial growth factor by retinal cells is a common factor in neovascularizing ocular diseases. *Lab Invest* 1995;72:638-645.
- 28. Lashkari K, Hirose T, Yazdany J, McMeel JW, Kazlauskas A, Rahimi N. Vascular endothelial growth factor and hepatocyte growth factor levels

are differentially elevated in patients with advanced retinopathy of prematurity. *Am J Pathol* 2000;156:1337-1344.

- Kvanta A, Algvere PV, Berglin L, Seregard S. Subfoveal fibrovascular membranes in age-related macular degeneration express vascular endothelial growth factor. *Invest Ophthalmol Vis Sci* 1996;37:1929-1934.
- 30. Lopez PF, Sippy BD, Lambert HM, Thach AB, Hinton DR. Transdifferentiated retinal pigment epithelial cells are immunoreactive for vascular endothelial growth factor in surgically excised agerelated macular degeneration-related choroidal neovascular membranes. *Invest Ophthalmol Vis Sci* 1996;37:855-868.
- 31. Otani A, Takagi H, Oh H, Koyama S, Ogura Y, Matumura M, et al. Vascular endothelial growth factor family and receptor expression in human choroidal neovascular membranes. *Microvasc Res* 2002;64:162-169.
- 32. Qaum T, Xu Q, Joussen AM, Clemens MW, Qin W, Miyamoto K, et al. VEGF-initiated blood-retinal barrier breakdown in early diabetes. *Invest Ophthalmol Vis Sci* 2001;42:2408-2413.
- 33. Vinores SA, Youssri AI, Luna JD, Chen YS, Bhargave S, Vinores MA, et al. Upregulation of vascular endothelial growth factor in ischemic and non-ischemic human and experimental retinal disease. *Histol Histopathol* 1997;12:99-109.
- 34. Nauck M, Karakiulakis G, Perruchoud AP, Papakonstantinou E, Roth M. Corticosteroids inhibit the expression of the vascular endothelial growth factor gene in human vascular smooth muscle cells. *Eur J Pharmacol* 1998;341:309-315.
- 35. Ferris FL 3rd, Fine SL, Hyman L. Age-related macular degeneration and blindness due to neovascular maculopathy. *Arch Ophthalmol* 1984;102:1640-1642.
- 36. Eichler W, Reiche A, Yafai Y, Lange J, Wiedemann P. Growth-related effects of oxidant-induced stress on cultured RPE and choroidal endothelial cells. *Exp Eye Res* 2008;87:342-348.
- 37. Chucair AJ, Rotstein NP, Sangiovanni JP, During A, Chew EY, Politi LE. Lutein and zeaxanthin protect photoreceptors from apoptosis induced by oxidative stress: relation with docosahexaenoic acid. *Invest Ophthalmol Vis Sci* 2007;48:5168-5177.
- 38. Bressler NM; Treatment of Age-Related Macular Degeneration with Photodynamic Therapy (TAP) Study Group. Photodynamic therapy of subfoveal choroidal neovascularization in age-related macular degeneration with verteporfin: two-year results of 2 randomized clinical trials-tap report 2. Arch Ophthalmol 2001;119:198-207.
- 39. VEGF Inhibition Study in Ocular Neovascularization (VISION) Clinical Trial Group, D'Amico DJ, Masonson HN, Patel M, Adamis AP, Cunningham ET Jr, Guyer DR, et al. Pegaptanib sodium for neovascular age-related macular

degeneration: two-year safety results of the two prospective, multicenter, controlled clinical trials. *Ophthalmology* 2006;113:992-1001.

- 40. Boyer DS, Antoszyk AN, Awh CC, Bhisitkul RB, Shapiro H, Acharya NR; MARINA Study Group. Subgroup analysis of the MARINA study of ranibizumab in neovascular age-related macular degeneration. *Ophthalmology* 2007;114:246-252.
- 41. Chang TS, Bressler NM, Fine JT, Dolan CM, Ward J, Klesert TR; MARINA Study Group. Improved vision-related function after ranibizumab treatment of neovascular age-related macular degeneration: results of a randomized clinical trial. *Arch Ophthalmol* 2007;125:1460-1469.
- 42. Brown DM, Kaiser PK, Michels M, Soubrane G, Heier JS, Kim RY, et al; ANCHOR Study Group. Ranibizumab versus verteporfin for neovascular age-related macular degeneration. *N Engl J Med* 2006;355:1432-1444.
- 43. Kaiser PK, Brown DM, Zhang K, Hudson HL, Holz FG, Shapiro H, et al. Ranibizumab for predominantly classic neovascular age-related macular degeneration: subgroup analysis of firstyear ANCHOR results. *Am J Ophthalmol* 2007;144:850-857.
- 44. Brown DM, Michels M, Kaiser PK, Heier JS, Sy JP, Ianchulev T; ANCHOR Study Group. Ranibizumab versus verteporfin photodynamic therapy for neovascular age-related macular degeneration: Two-year results of the ANCHOR study. *Ophthalmology* 2009;116:57-65.e5.
- 45. Regillo CD, Brown DM, Abraham P, Yue H, Ianchulev T, Schneider S, et al. Randomized, double-masked, sham-controlled trial of ranibizumab for neovascular age-related macular degeneration: PIER Study year 1. *Am J Ophthalmol* 2008;145:239-248.
- Avery RL, Pieramici DJ, Rabena MD, Castellarin AA, Nasir MA, Giust MJ. Intravitreal bevacizumab (Avastin) for neovascular age-related macular degeneration. *Ophthalmology* 2006;113:363-372.e5.
- 47. Spaide RF, Laud K, Fine HF, Klancnik JM Jr, Meyerle CB, Yannuzzi LA, et al. Intravitreal bevacizumab treatment of choroidal neovascularization secondary to age-related macular degeneration. *Retina* 2006;26:383-390.
- 48. Ahmadieh H, Taei R, Soheilian M, Riazi-Esfahani M, Karkhaneh R, Lashay A, et al. Single-session photodynamic therapy combined with intravitreal bevacizumab and triamcinolone for neovascular age-related macular degeneration. *BMC Ophthalmol* 2007;7:10.
- 49. Nguyen QD, Shah SM, Hafiz G, Quinlan E, Sung J, Chu K, et al. A phase I trial of an IV-administered vascular endothelial growth factor trap for treatment in patients with choroidal neovascularization due to age-related macular degeneration. *Ophthalmology* 2006;113(9):1522.e1-1522.e14.

- 50. Stewart MW, Rosenfeld PJ. Predicted biological activity of intravitreal VEGF Trap. *Br J Ophthalmol* 2008;92:667-668.
- 51. Andreoli CM, Miller JW. Anti-vascular endothelial growth factor therapy for ocular neovascular disease. *Curr Opin Ophthalmol* 2007;18:502-508.
- 52. Arias L, Planas N, Prades S, Caminal JM, Rubio M, Pujol O, et al. Intravitreal bevacizumab (Avastin) for choroidal neovascularisation secondary to pathological myopia: 6-month results. *Br J Ophthalmol* 2008;92:1035-1039.
- 53. Teixeira A, Moraes N, Farah ME, Bonomo PP. Choroidal neovascularization treated with intravitreal injection of bevacizumab (Avastin) in angioid streaks. *Acta Ophthalmol Scand* 2006;84:835-836.
- 54. Schadlu R, Blinder KJ, Shah GK, Holekamp NM, Thomas MA, Grand MG, et al. Intravitreal bevacizumab for choroidal neovascularization in ocular histoplasmosis. *Am J Ophthalmol* 2008;145:875-878.
- 55. Chang LK, Spaide RF, Brue C, Freund KB, Klancnik JM Jr, Slakter JS. Bevacizumab treatment for subfoveal choroidal neovascularization from causes other than age-related macular degeneration. *Arch Ophthalmol* 2008;126:941-945.
- 56. Ahmadieh H, Vafi N. Dramatic response of choroidal neovascularization associated with choroidal osteoma to the intravitreal injection of bevacizumab (Avastin). Graefes Arch Clin Exp Ophthalmol 2007;245:1731-3.
- Chen E, Park CH. Use of intravitreal bevacizumab as a preoperative adjunct for tractional retinal detachment repair in severe proliferative diabetic retinopathy. *Retina* 2006;26:699-700.
- 58. Rizzo S, Genovesi-Ebert F, Di Bartolo E, Vento A, Miniaci S, Williams G. Injection of intravitreal bevacizumab (Avastin) as a preoperative adjunct before vitrectomy surgery in the treatment of severe proliferative diabetic retinopathy (PDR). *Graefes Arch Clin Exp Ophthalmol* 2008;246:837-842.
- 59. Arevalo JF, Maia M, Flynn HW Jr, Saravia M, Avery RL, Wu L, et al. Tractional retinal detachment following intravitreal bevacizumab (Avastin) in patients with severe proliferative diabetic retinopathy. *Br J Ophthalmol* 2008;92:213-216.
- 60. The PKC-DRS Study Group. The effect of ruboxistaurin on visual loss in patients with moderately severe to very severe nonproliferative diabetic retinopathy: initial results of the Protein Kinase C beta Inhibitor Diabetic Retinopathy Study (PKC-DRS) multicenter randomized clinical trial. *Diabetes* 2005;54:2188-2197.
- PKC-DRS2 Group, Aiello LP, Davis MD, Girach A, Kles KA, Milton RC, Sheetz MJ, et al. Effect of ruboxistaurin on visual loss in patients with diabetic retinopathy. *Ophthalmology* 2006;113:2221-2230.
- 62. Campochiaro PA, Hafiz G, Shah SM, Nguyen QD,

Ying H, Do DV, et al. Ranibizumab for macular edema due to retinal vein occlusions: implication of VEGF as a critical stimulator. *Mol Ther* 2008;16:791-799.

- 63. Pieramici DJ, Rabena M, Castellarin AA, Nasir M, See R, Norton T, et al. Ranibizumab for the treatment of macular edema associated with perfused central retinal vein occlusions. *Ophthalmology* 2008;115:e47-54.
- 64. Hsu J, Kaiser RS, Sivalingam A, Abraham P, Fineman MS, Samuel MA, et al. Intravitreal bevacizumab (avastin) in central retinal vein occlusion. *Retina* 2007;27:1013-1019.
- 65. Spaide RF, Chang LK, Klancnik JM, Yannuzzi LA, Sorenson J, Slakter JS, et al. Prospective study of intravitreal ranibizumab as a treatment for decreased visual acuity secondary to central retinal vein occlusion. *Am J Ophthalmol* 2009;147:298-306.
- 66. Prager F, Michels S, Kriechbaum K, Georgopoulos M, Funk M, Geitzenauer W, et al. Intravitreal bevacizumab (Avastin) for macular oedema secondary to retinal vein occlusion: 12-month results of a prospective clinical trial. *Br J Ophthalmol* 2009;93:452-456.
- 67. Davidorf FH, Mouser JG, Derick RJ. Rapid improvement of rubeosis iridis from a single bevacizumab (Avastin) injection. *Retina* 2006;26:354-356.
- 68. Costagliola C, Cipollone U, Rinaldi M, della Corte M, Semeraro F, Romano MR. Intravitreal bevacizumab (Avastin) injection for neovascular glaucoma: a survey on 23 cases throughout 12month follow-up. *Br J Clin Pharmacol* 2008;66:667-673.
- 69. Duch S, Buchacra O, Milla E, Andreu D, Tellez J. Intracameral bevacizumab (Avastin) for neovascular glaucoma: a pilot study in 6 patients. *J Glaucoma* 2009;18:140-143.
- 70. Ehlers JP, Spirn MJ, Lam A, Sivalingam A, Samuel MA, Tasman W. Combination intravitreal bevacizumab/panretinal photocoagulation versus panretinal photocoagulation alone in the treatment of neovascular glaucoma. *Retina* 2008;28:696-702.
- 71. Smith LE, Wesolowski E, McLellan A, Kostyk SK, D'Amato R, Sullivan R, et al. Oxygen-induced retinopathy in the mouse. *Invest Ophthalmol Vis Sci* 1994;35:101-111.
- 72. Pierce EA, Avery RL, Foley ED, Aiello LP, Smith LE. Vascular endothelial growth factor/vascular permeability factor expression in a mouse model of retinal neovascularization. *Proc Natl Acad Sci US A* 1995;92:905-909.
- 73. Young TL, Anthony DC, Pierce E, Foley E, Smith LE. Histopathology and vascular endothelial growth factor in untreated and diode laser-treated retinopathy of prematurity. *J AAPOS* 1997;1:105-110.
- 74. Hellstrom A, Perruzzi C, Ju M, Engstrom E, Hard AL, Liu JL, et al. Low IGF-I suppresses VEGF-

survival signaling in retinal endothelial cells: direct correlation with clinical retinopathy of prematurity. *Proc Natl Acad Sci USA* 2001;98(10):5804-5808.

- 75. Hurwitz H, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. N Engl J Med 2004;350:2335-2342.
- 76. Schouten JS, La Heij EC, Webers CA, Lundqvist IJ, Hendrikse F. A systematic review on the effect of bevacizumab in exudative age-related macular degeneration. *Graefes Arch Clin Exp Ophthalmol* 2009;247:1-11.
- 77. Riazi-Esfahani M, Ahmadieh H, Faghihi H, Piri N, Taei R, Karkhaneh R, et al. Intravitreal Bevacizumab versus Combined Bevacizumab and Triamcinolone Acetonide for Neovascular Age-Related Macular Degeneration. J Ophthalmic Vis Res 2008;3:95-101.
- Konner J, Dupont J. Use of soluble recombinant decoy receptor vascular endothelial growth factor trap (VEGF Trap) to inhibit vascular endothelial growth factor activity. *Clin Colorectal Cancer* 2004;4 Suppl 2:S81-85.
- 79. Strom C, Sander B, Klemp K, Aiello LP, Lund-Andersen H, Larsen M. Effect of ruboxistaurin on blood-retinal barrier permeability in relation to

severity of leakage in diabetic macular edema. *Invest Ophthalmol Vis Sci* 2005;46:3855-3858.

- Chappelow AV, Kaiser PK. Neovascular age-related macular degeneration: potential therapies. *Drugs* 2008;68:1029-1036.
- 81. Kinose F, Roscilli G, Lamartina S, Anderson KD, Bonelli F, Spence SG, et al. Inhibition of retinal and choroidal neovascularization by a novel KDR kinase inhibitor. *Mol Vis* 2005;11:366-373.
- 82. Wang FE, Shi G, Niesman MR, Rewolinski DA, Miller SS. Receptor tyrosine kinase inhibitors AG013764 and AG013711 reduce choroidal neovascularization in rat eye. *Exp Eye Res* 2007;84:922-933.
- Takahashi H, Obata R, Tamaki Y. A novel vascular endothelial growth factor receptor 2 inhibitor, SU11248, suppresses choroidal neovascularization in vivo. J Ocul Pharmacol Ther 2006;22:213-218.
- 84. Palanki MS, Akiyama H, Campochiaro P, Cao J, Chow CP, Dellamary L, et al. Development of prodrug 4-chloro-3-(5-methyl-3-{[4-(2-pyrrolidin-1ylethoxy)phenyl]amino}-1,2,4-benzotriazin-7yl)phenyl benzoate (TG100801): a topically administered therapeutic candidate in clinical trials for the treatment of age-related macular degeneration. *J Med Chem* 2008;51:1546-1559.