

Original Article

Modeling of Environmental Factors Affecting the Prevalence of Zoonotic and Anthroponotic Cutaneous, and Zoonotic Visceral Leishmaniasis in Foci of Iran: A Remote Sensing and GIS Based Study

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Abstract

Background: Leishmaniasis is a re-emerging serious international public health problem, and both visceral and cutaneous types of leishmaniasis became important endemic diseases in Iran. In this study, the relationships between environmental factors (vegetation and elevation) and the prevalence of diseases have been investigated.

Methods: All international and national online databases were searched by terms such as leishmaniasis, incidence, prevalence and other related words attributed to Iran and published until first quarter of 2015. The developed database in Excel, later imported to the ArcMap for spatial analyst and mapping. Afterwards, the software was used for modeling the relationship between the prevalence/incidence and environmental variables (vegetation and elevation) by both linear and nonlinear regression.

Results: After mapping the prevalence data from 144 studies, considering non-parametric ANOVA, the tendency of zoonotic visceral leishmaniasis to presence in high elevation and high vegetation was more than Anthroponotic and zoonotic cutaneous leishmaniasis. While linear regression showed weaker results for modeling, however, additive nonparametric regression analysis suggested that 10km buffers for elevation, and 10 as well as 50km buffers for vegetation could contribute in better fitness in modeling of these variables.

Conclusion: The detailed maps for distribution of disease concluded. The nonlinear regression is a reliable predictor of the relationship between environmental factors and disease incidence, although more and wide researchers are needed to confirm it.

Keywords: Leishmaniasis, GIS, Remote sensing, Environmental variables, Nonlinear regression

Introduction

Leishmaniasis –a re-emerging serious international public health problem- is considered as one of the most neglected tropical diseases. The disease is distributed in new foci due to influence of many risk factors, including environmental factors (1-4). Almost 200000 to 400000 new cases of visceral leishmaniasis (VL), 0.7 million to 1.3 million new cases of cu-

taneous leishmaniasis (CL) and 20000 to 30000 deaths annually" are the estimations of WHO last report about importance of the disease (2).

Two dominant types of leishmaniasis there in Iran are: zoonotic visceral or kala-azar (ZVL), the most serious and fatal type of the disease, and both types of cutaneous leishmaniasis namely urban form or anthroponotic cutaneous

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leishmaniasis (ACL) and rural form or zoonotic cutaneous leishmaniasis (ZCL). *Leishmania donovani* complex are the causes of ZVL protozoan disease which can lead to patient mortality (5). Evidence indicates an increase in the incidence of disease in the Old and New Worlds at the early years of the present century. In recent decades, ZVL became an important endemic disease in the Northwest (6), South and Southwest (7), and Northeast (8) of Iran, especially in nomads. The majority of ZVL cases (92.8%) in country were found among children with age under 12yr old (9). The most recently, direct agglutination test (DAT) as a simple, affordable and Practical test, has found its way as a special quick and accurate test. Thus it has been recommended and applied widely for clinical diagnosis and seroprevalence studies of ZVL in zoonosis and anthroponotic reservoirs in endemic and nonendemic areas of Iran (10). Due to ecologically dependence of the disease vectors and reservoirs to environment, outbreaks mostly are common in lowlands rural or suburban areas (less than 600M elevation), with a heavy annual rainfall, a moderate temperature (15–38 °C), and almost dense vegetation (2).

Countries of Mediterranean coasts (Algeria and Syrian Arab Republic), South Americas (Brazil and Colombia), and Middle East (Islamic Republic of Iran and Afghanistan) and Central Asia countries are the most reported foci for CL and the major contribution of CL occur in this area (2). The CL cases in Iran during the period of 2001 to 2008 dramatically has been increased more than two-fold and also over half of 31 provinces of country have CL endemic foci (11). As reported by Center for Disease Control, the annual incidence of leishmaniasis in the country due to referring to health centers has reached about 20000 cases. While the actual cases of disease may be far more than these reports that may even reach up to five times (12).

As for the first time carried by John Snow in 1854 due to diarrheal disease detection in

London, spatial modeling as a useful tool in epidemiological studies could be used. Today, Geographic Information System (GIS) in relation to the processed images from remote sensing (RS) has been used to map the geographical distributions of disease prevalence, factors affecting the transmission and control of disease, and the spatial modeling of environmental factors that have meaningful impacts on disease occurrences (9, 13). Vector control programs are associated with logistic problems, mainly due to poor understanding of vector ecology (14). Expanding inexpensive and effective models of integrated management for better control of CL is a critical target which need a thorough will, alongside epidemiology studies and understanding of the ecology of the disease (11) and the role of social, ecological and specially environmental variables for risk factors such as the vegetation and elevation (two most widespread and accessible environmental variables). The relationship between epidemiological components of leishmaniasis (incidence, prevalence, and seroprevalence) and environmental variables (vegetation, elevation, and/or other factors) can be used in GIS modeling and may be useful for secondary uses including prediction. Therefore, contributing to better planning for control activities and the establishment of early warning systems, incorporating remotely sensed information in epidemiological studies can help public health policy makers in having a better notion and comprehensive understanding to find the relationships between disease and its environmental risk factors (15).

The remotely sensed Normalized Difference Vegetation Index (NDVI) –the predominantly index which used for vegetation coverage– has found its widespread applications due to fluctuations it has along with meteorological and environmental components (14). This index together with the other remotely sensed data (11, 16) like the Digital Elevation Model (DEM), have been used extensively in monitoring some vector-borne diseases in-

cluding leishmaniasis throughout the world. While many studies have surveyed the associations between the mean of NDVI and DEM with incidence or prevalence rate of a type of leishmaniasis locally, however, they do not study all variables in a vast area.

In this study, we systematically reviewed all studies regarding the incidence and/or prevalence rate of all categories of leishmaniasis investigated in Iran from 1976 to 2015. After mapping the results, using Spatial Analysis and Regression techniques, statistical association between variables have been modeled in detail.

Materials and Methods

To identify the incidence and prevalence of leishmaniasis in endemic and nonendemic area of ZVL, ZCL, and ACL of Iran, we searched and reviewed the literature based on international database including MEDLINE, ISI, SCOPUS, and Google Scholar, and national databases such as SID, IRANDOC, MagIran, and ISC. Search protocols have been based on using the terms related to all types of leishmaniasis, fauna, vectors, reservoirs, incidence, prevalence, seroprevalence, Direct Agglutination Test (DAT), and their Persian equivalents with the geographic term "Iran".

Overall, 503 articles published from 1976 through Jun 2015 in English, French, and Persian were founded in first search from databases. Besides, by referring to CV of famous professors, abstract seminars, thesis, non-digital papers, and the reference/citations of articles, 121 additional sources were found. After the elimination of duplicate and inappropriate articles by two separate teams, we consequently included 144 studies that clearly addressed confirmed clinical cases for human seroprevalence ($\text{DAT} \geq 1$: 3200)/ incidence of ZVL and active lesion prevalence/total prevalence (active lesion + scar)/incidence of ACL and ZCL in Iran. Disease data were extracted by researchers directly from body text of arti-

cles to database tables (Dbase format using the MS Excel software) containing all cities (1246 point) and villages with a population more than 500 people (12034 points), disease information, and descriptive data. Depending on the raw data presented in each article, one or more than one category of epidemiologic data was included in the main table. These numeric values (all in percent) for ACL and ZCL including active lesion prevalence, total prevalence (active lesion + scar), and incidence and for VCL including seroprevalence and incidence were directly or indirectly extracted using study population. The developed database later imported to the ArcMap[®] software version 9.3 (ESRI, Redlands, USA) software platform for Spatial Analyst and mapping all types of leishmaniasis. R software version 3.2.2 (17) was used for analyzing the relationship between the prevalence/incidence and continues spatial variables.

Shapefile maps for political subdivisions of provinces, counties, rural districts, and points of cities, towns, and villages with their attribute tables were obtained from National Cartographic Center, Iran. The Moderate Resolution Imaging Spectroradiometer (MODIS) onboard the Terra satellite, is one of most applied NDVI images source in literature. NDVI calculated from surface reflectance in the near-infrared ρNIR and red ρR portions of the electromagnetic spectrum, using the equation 1 (18):

$$1) \text{NDVI} = (\rho\text{NIR} - \rho\text{R}) / (\rho\text{NIR} + \rho\text{R})$$

The 16d, 250m NDVI raster maps (MOD 13Q1 series, version 5) projected in Geographic Coordinate System WGS_1984, derived from MODIS Reprojection Tool (MRT) (19) for the years 2000 to 2014. Sixteen days maps unified to annual average NDVI in Spatial Analysis, then reclassified to 1–20. Altitude was based on DEM, obtained from the 100m ASTER Global Digital Elevation Model (ASTER GDEM) data center Version 2 (20). These maps then reprojected to Lambert Conformal Conic in ArcMap, and additional Raster Analysis such as reclassification, annual and 15yr average

and zonal statistics were performed on them. Overall, 5km, 10km, 20km, and 50km buffer zones centered on each city/village point reported values for leishmaniasis, and their zonal Min, Max, Minority, Majority, Median and Mean statistics were calculated and exported in Dbase tables. Finally, massive joined tables were called in R, and statistical calculations were completed for each.

Data analysis

CSV tables for any purposes derived from the main disease data table and called in R console. We chose to test the normality of data with One-sample Kolmogorov-Smirnov and Shapiro-Wilk normality test. Next, we modeled the relationships between prevalence/incidence values and the continuous independent variables. Two regression scenarios were considered. First, using simple linear regression models, a stepwise backward elimination procedure was used with the variables, finally resulting in the model contains merely the variables which associated with the outcome. Secondly, using generalized additive models -which use spline or local regression- a stepwise backward elimination procedure, was used again to reach the final model. The additive nonparametric regression model is presented in equation 2 (21):

$$2) y = \beta_0 + m_1(x_1) + m_2(x_2) + \dots + m_k(x_k) + \varepsilon$$

Where the partial-regression functions m_j are fit using a simple-regression smoother, such as local polynomial regression or smoothing splines. In nonparametric regression, in contrast, the object is to estimate the regression function directly without specifying its form explicitly. In nonparametric comparing with simple regression, the ultimate goal is to obtain the regression function directly from internal computation without any open aspect (22). The level of 5% significance for all tests was considered.

Results

We found 144 studies that directly informed the active lesion/scar prevalence and/or incidence of leishmaniasis in Iran from 1976–2015 (Table 1). These data considered as a base for construction of 89 ACL points including 15, 19, and 71 urban/rural points for active lesion, total prevalence (active lesion + scar), and incidence respectively. The same way, data for 420 ZCL points included 67, 68, and 362 points. Cases for 355 ZVL points for the seroprevalence and incidence were 283 and 82 respectively. Figures 1, 2, and 3 show the maps of three types of leishmaniasis in Iran separately. The DEM map and 15yr average of NDVI map also attached on VL and ACL maps respectively. Rating in the categories are High, Middle and Low for prevalence data and obtained based on expert consensus.

Scatter diagrams and normality plot of each independent variable and the disease incidence/prevalence were created and visually inspected before running regression. Some of these plots traced in Fig. 4. Normal probability of data in our variables has been questioned, then, the normality of data statistically tested. Therefore, using both One-sample Kolmogorov-Smirnov and Shapiro-Wilk normality test, the data are without normality. However, both linear and nonparametric regression fitted on data.

To compare of the means of main independent variables (DEM and NDVI) versus types of leishmaniasis, homogeneity of variances rejected by Fligner-Killeen test, but due to nonparametric Kruskal-Wallis test, we result in significant difference between means of variables. Regard to boxplots for comparison of means in Fig. 5, and nonparametric analysis of variances in Table 2, tendency of ZVL to presence in high elevation and high vegetation is more than ACL and ZCL. Similarly, we compare the means of types of leishmaniasis occurrences and epidemiological variables of disease in Table 3.

The regression models were evaluated using R software packages. Tables 4 and 5 generally describe data structure and assessment of the effect of, or relationship between, explanatory variables on the response by using multiple linear regression models.

To determine the best final fitted models for each group of data, based on P-value, adjusted R-squared, and AIC (Akaike's 'An Information Criterion'), using stepwise backward simple linear regression, following model produced: (although second order or interaction models had a little more R-squared, due to the complexity were excluded).

- 3) Total prevalence= $12.14 - 0.014 (\text{MEAN\$DEM}_{20}) + 0.005 (\text{MEAN\$DEM}_{50}) - 1.64 (\text{MIN\$NDVI}_{20}) + 1.8 (\text{MEAN\$NDVI}_{20})$
 Multiple R-squared: 0.2111, Adjusted R-squared: 0.198
 F-statistic: 16.06 on 4 and 240 DF, P: $1.151e^{-11}$

Important plots of main model presented in Fig. 6. Regarding weak results of linear regression (R-squared located near the zero), eventually, Additive Nonparametric Regression analysis, using GAM (Generalized Additive Model) command in library "mgcv", ap

plied to find best parameters and buffers for modeling variables of this study. Here two models were fitted: first with the highest percentage of predictability by equation 4, second with the convenience practical application (same parameter mean of variables in unique buffer 10km) which came in equation 5.

- 4) Total prevalence= $7.69 + s (\text{MEAN\$DEM}_{10}) + s (\text{MAX\$NDVI}_{50})$
 R-sq. (adj)= 0.524 Deviance explained= 54.5%
- 5) Total prevalence= $7.42 + s (\text{MEAN\$DEM}_{10}) + s (\text{MEAN\$NDVI}_{10})$
 R-sq. (adj)= 0.371 Deviance explained= 39.6%

The "s" function, used in specifying the model formula, indicates that each term is to be fitted with a smoothing spline. The surface that fitted on the model and smoothing splines have been shown in Fig. 7 and 8. In equation 5, the 5.43 parameters are used for the MEAN\$NDVI₁₀ (mean of 10km buffer for NDVI) term, and 7.98 for the MEAN\$DEM₁₀ term, the degrees of freedom for the model is the sum of these plus 1 for the regression constant.

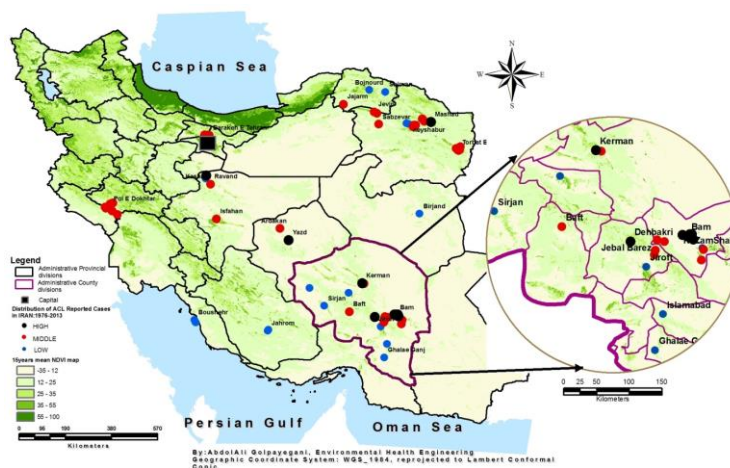


Fig. 1. Distribution of anthroponotic cutaneous leishmaniasis city points in Iranian studies from 1976–2013 on 15yr annual average Normalized Difference Vegetation Index map. High: active lesion $\geq 2\%$ or active lesion+scar $\geq 20\%$ or incidence $\geq 2\%$; Middle: $2\% > \text{active lesion} \geq 0.5\%$ or $20\% > \text{active lesion+scar} \geq 5\%$ or $2 > \text{incidence} \geq 0.5\%$, Low: $0.5\% > \text{active lesion}$ or $5\% > \text{active lesion+scar}$ or $0.5 > \text{incidence}$

Table 1. Literatures published for prevalence of leishmaniasis, divided by leishmaniasis types and provinces of Iran

Province	ACL references	ZCL references	ZVL references
Alborz			Fagih-Nayini et al. 2002(23)
Ardabil			Mazloumi Gavgani et al. 2002(24), Soleimanzadeh et al. 1993(25), Mohebali et al. 2010(26), Edrissian 1996(27), Mohebali et al. 2011(28), Mahami et al. 2006(29), Mohammadi-Kheyrabadi et al. 2004(30), Nadim et al. 1998(31), Arshi et al. 2002(32), Mazloumi Gavgani et al. 2011(33)
Azerbaijan, East			Mazloumi Gavgani et al. 2002(24), Mohebali et al. 2011(28), Mirsamadi et al. 2003(34), Mazloumi Gavgani et al. 2011(33)
Azerbaijan, West			
Bushehr	Yaghoobi-Ershadi et al. 2013(35)	Yaghoobi-Ershadi et al. 2013(35), Hamzavi et al. 2000(36)	Mohebali et al. 2001(37)
Chahar Mahaal and Bakhtiari			
Fars	Ghatee et al. 2013(38)	Razmjou et al. 2009(39), Ghatee et al. 2013(38), Rassi et al. 2004(40), Fakoorziba et al. 2011(41), Davami et al. 2010(42), Sharafi et al. 2013(43), Noorpisheh et al. 2013(44), Entezari and Eskandari 2014(45)	Fakhar et al. 2008(46), Edrissian et al. 1993(47), Sahabi et al. 1992(48), Fakhar et al. 2006(49)
Gilan			
Golestan		Mollalo et al. 2015(50), Rahbarian et al. 2009(51), Sofizadeh et al. 2012(52), Baghaei et al. 2012(53), Mesgarian et al. 2010(54)	(Fakhar et al. 2014)
Hamadan		Hanafi Bojd et al. 2006(55), Nazari 2012(56)	
Hormozgān		Azizi et al. 2012(57)	
Ilam		Asgari Nezhad et al. 2012(58), Shazad et al. 2005(59), Bahrami et al. 2011(60), Yazdanpanah and Rostamianpur 2013(61), Roghani et al. 2012(62)	
Isfahan	Shiee et al. 2012(63), Zahraei-Ramazani et al. 2007(64), Doroodgar et al. 2012(65)	Talari et al. 2006(66), Yaghoobi-Ershadi et al. 2001(67), Mohammadian et al. 1999(68), Yaghoobi-Ershadi and Javadian 1995(69), Yaghoobi-Ershadi et al. 2006(70), Nilfroushzadeh et al.	

Table 1. Continued ...

		2002(71), Doroodgar et al. 2009(72), Ahmadi et al. 2013(73), Ebadi and Hejazi 2003(74), Yaghoobi-Ershadi et al. 1999(75), Doroudgar et al. 1996(76)	
Kerman	Nadim and Aflatoonian 1995(77), Sharifi et al. 2011(78), Sharifi et al. 2015(3), Yaghoobi-Ershadi 1977(79), Mirzaei et al. 2012(80), Aflatoonian et al. 2014(81), Sharifi et al. 1998(82), Sharifi et al. 2011(83), Ghatee et al. 2013(38), Aflatoonian et al. 2013(84), Nadim et al. 1995(85), Sharifi et al. 2015(86), Aflatoonian and Sharifi 2011(87), Aflatoonian and Sharifi 2010(88), Mirzadeh et al. 2008(89), Aflatoonian et al. 2014(90), Aflatoonian and Sharifi 2007(91)	Sharifi et al. 2015(3), Akhavan et al. 2007(92), Aghaei-Afshar et al. 2013(93), Khosravi et al. 2013(94), Sharifi et al. 2008(95)	Mahmoudvand et al. 2011(96), Barati et al. 2008(97)
Kermanshah		Hamzavi and Khademi 2015(98), Nazari et al. 2012(99), Hamzavi and Khademi 2013(100)	Hamzavi et al. 2012(101)
Khorasan, North	Alavinia et al. 2009(102)	Alavinia et al. 2009(102)	Torabi et al. 2007(103)
Khorasan, Razavi	Sattar Pagheh et al. 2013(104), Moosa-Kazemi et al. 2007(105), Saadabadi et al. 2013(106), Hassanpour et al. 2014(107), Hoseini Farash et al. 2011(108), Khajedaluae et al. 2014(109), Mohajery et al. 2008(110)	Yaghoobi-Ershadi et al. 2003(111), Hassanpour et al. 2014(107), Khajedaluae et al. 2014(109), Karimi Zarchi et al. 2004(112), Sahabi et al. 1999(113), Akbari et al. 2014(114)	
Khorasan, South		Karamian et al. 2013(115)	
Khuzestan		Kassiri et al. 2012(116), Kassiri et al. 2012(117), Kassiri et al. 2014(118), Kassiri et al. 2014(119), Vazirianzadeh et al. 2014(120), Kassiri et al. 2013(121), Spotin et al. 2014(122), Ghasemian et al. 2011(123), Kassiri et al. 2013(124), Nejati et al. 2014(125), Behbahani et al. 2012(126), Kassiri et al. 2011(127)	
Kohgiluyeh and Boyer-Ahmad			Sarkari et al. 2010(128), Sarkari et al. 2007(129)
Kurdistan			

Table 1. Continued ...

Lorestan	Kheirandish et al. 2013(130)	Amraee et al. 2013(131), Ahmadi et al. 2013(132), Kheirandish et al. 2013(130), Chegeni-Sharafi et al. 2011(133)	Chegeni-Sharafi et al. 2005(134)
Markazi			
Mazandaran			
Qazvin			
Qom		Rostami et al. 2013(135), Rassi et al. 2013(12), Akhavan et al. 2003(136), Saghafipour et al. 2013(137), Saghafipour et al. 2012(138)	(Rakhshanpour et al. 2014), (Fakhar et al. 2004)
Semnan		Mohammadi Azni et al. 2010(139), Mohammadi Azni et al. 2011(140), Mohammadi Azni et al. 2010(141), Rafati et al. 2007(142)	
Sistan and Baluchestan		Fazaeli et al. 2008(143), Fazaeli et al. 2009(144), Fouladi et al. 2007(145)	
Tehran	Salehzadeh and Seyedi Rashti 1996(146), Hossein Abadi 2004(147)		
Yazd	Yaghoobi-Ershadi et al. 2002(148)	Yaghoobi-Ershadi et al. 2004(149), Jaafary et al. 2007(150), Mozafari and Bakhshi Zadeh 2011(151), Hanafi Bojd et al. 2003(152), Yaghoobi-Ershadi et al. 2008(153)	
Zanjan			
Multy-provinces			Mohebbali et al. 2006(6)

Table 2. Nonparametric analysis of variances between types of zoonotic, anthroponotic cutaneous, and zoonotic visceral leishmaniasis and environmental variables in Iran (P< 0.05), buffer = 5km

	DEM_MEAN	DEM_VARIANCE	NDVI_MEAN*	NDVI_VARIANCE
ACL	1216	213046	8.57	3.93
ZCL	666	385752	10.12	12.16
ZVL	1296	332153	11.93	6.28

*(The range is 1–20 for NDVI)

Table 3. Nonparametric Analysis of variances between types of zoonotic, anthroponotic cutaneous, and zoonotic visceral leishmaniasis and epidemiological variables of disease in Iran (P< 0.05)

	MEAN			VARIANCE		
	ACL (%)	ZCL (%)	ZVL (%)	ACL	ZCL	ZVL
Active lesion prevalence	1.48	4.42	Na*	3.47	37.97	Na
Total prevalence	11.88	25.02	2.69	125.38	622.39	3.73
Incidence	0.76	1.04	1.01	2.62	8.47	1.78

*Na: Not available

Table 4. Multiple linear regression models for types of zoonotic, anthroponotic cutaneous, and zoonotic visceral leishmaniasis and Digital Elevation Model subcategory (buffer distance around the cities or villages)

	DEM 5km_total	DEM 5km_ACL	DEM 5km_ZVL	DEM 5km_ZCL	Buffer types	Buffer types_ACL	Buffer types_ZVL	Buffer types_ZCL
Active lesion prevalence	~MEAN+MAX AR [§] : 0.1004	~MEAN+MIN AR: 0.284	Na***	~MEAN AR: 0.077	~MEAN20km AR: 0.035	ns	ns	~MEAN5km AR: 0.077
Total prevalence	~MAX AR: 0.1028	~MEAN AR:0.1649	~MAX AR:0.0484	~MEAN+MAX AR: 0.234	~MEAN20km AR: 0.1178	ns	~MEAN50km AR: 0.029	~MEAN20km AR: 0.25
Incidence	~MEAN+MAX AR: 0.029	Ns**	~MIN AR:0.2954	~MAX+MAJORITY AR: 0.077	~MEAN5km AR: 0.011	ns	~MEAN20km AR: 0.21	~MEAN5 AR: 0.031

*AR: Adjusted R-squared **Ns: Not significant ***Na: Not available

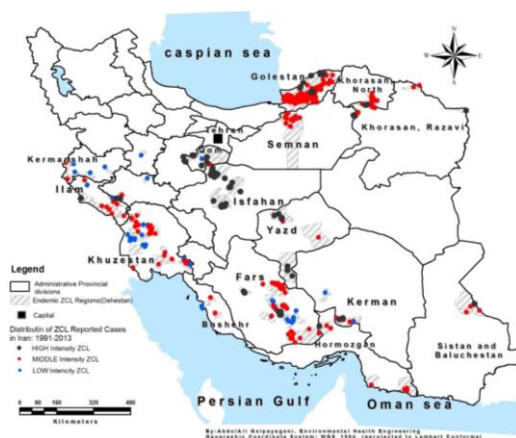


Fig. 2. Distribution of zoonotic cutaneous leishmaniasis rural region points in Iranian studies from 1991–2013. High: active lesion $\geq 2\%$ or active lesion+scar $\geq 20\%$ or incidence $\geq 2\%$, Middle: $2\% >$ active lesion $\geq 0.5\%$ or $20\% >$ active lesion+scar $\geq 5\%$ or $2 >$ incidence $\geq 0.5\%$, Low: $0.5\% >$ active lesion or $5\% >$ active lesion+scar or $0.5 >$ incidence

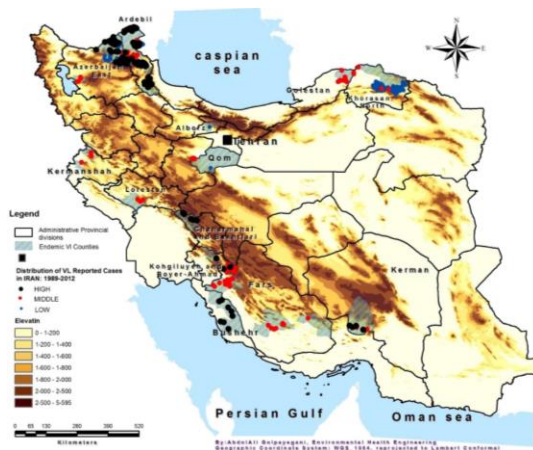


Fig. 3. Distribution of zoonotic visceral leishmaniasis county region points in Iranian studies from 1989–2012 on Digital Elevation Model map. High: DAT $\geq 2\%$ or incidence $\geq 1\%$, Middle: $2\% >$ DAT $\geq 0.5\%$ or $1 >$ incidence $\geq 0.1\%$, Low: $0.5\% >$ DAT or $0.1 >$ incidence

Table 4. Multiple linear regression models for types of zoonotic, anthroponotic cutaneous, and zoonotic visceral leishmaniasis and Digital Elevation Model subcategory (buffer distance around the cities or villages)

	DEM 5km_total	DEM 5km_ACL	DEM 5km_ZVL	DEM 5km_ZCL	Buffer types	Buffer types_ACL	Buffer types_ZVL	Buffer types_ZCL
Active lesion prevalence Total prevalence Incidence	~ MEAN+MAX AR*: 0.1004 ~ MAX AR: 0.1028 ~MEAN+MAX AR: 0.029	~ MEAN+MIN AR: 0.284 ~ MEAN AR:0.1649 Ns**	Na*** ~ MAX AR:0.0484 ~ MIN AR:0.2954	~ MEAN AR: 0.077 ~ MEAN+MAX AR: 0.234 ~ MAX+MAJORITY AR: 0.077	~ MEAN20km AR: 0.035 ~MEAN20km AR: 0.1178 ~ MEAN5km AR: 0.011	ns ns ns	ns ~MEAN50km AR: 0.029 ~ MEAN20km AR: 0.21	~ MEAN5km AR: 0.077 ~MEAN20km AR: 0.25 ~ MEAN5 AR: 0.031

*AR: Adjusted R-squared **Ns: Not significant ***Na: Not available

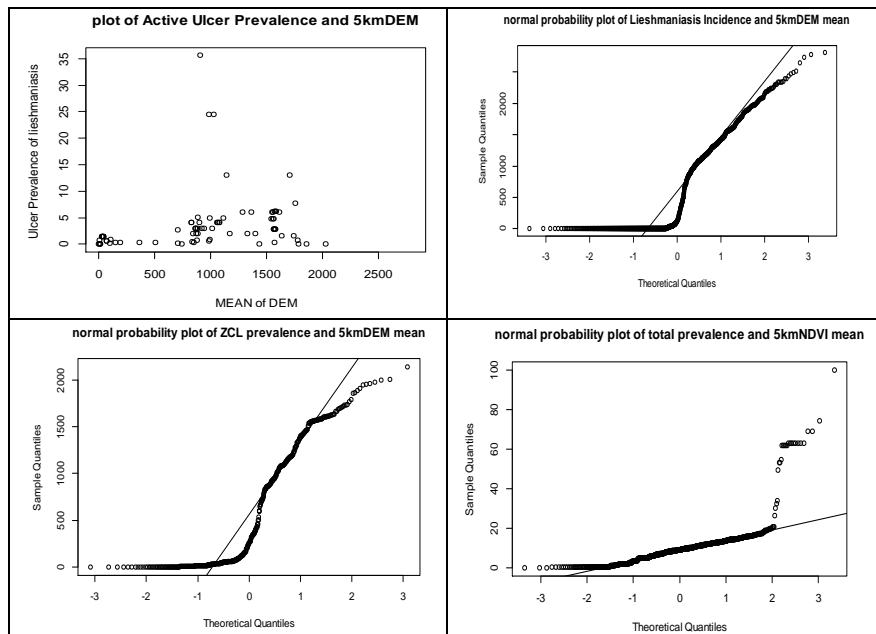


Fig. 4. Typical plots and normality plots of independent and dependent variables

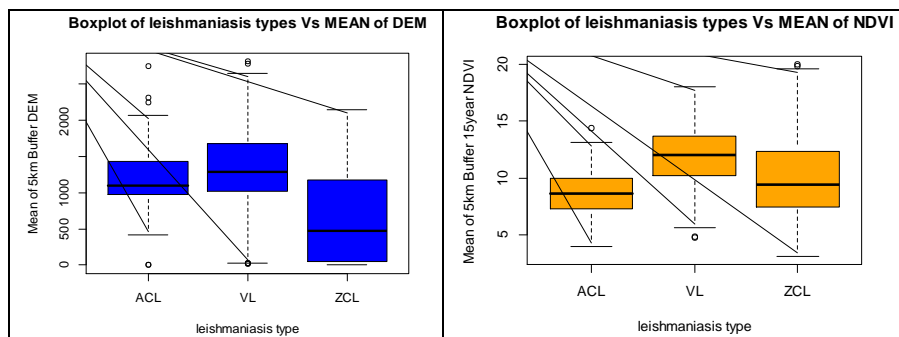


Fig. 5. Boxplots for compares of means of 5km-buffer Digital Elevation Model and Normalized Difference Vegetation Index ((in the range of 1–20)) for anthroponotic cutaneous leishmaniasis, zoonotic cutaneous leishmaniasis, and zoonotic visceral leishmaniasis in Iran for study period (1976–2013)

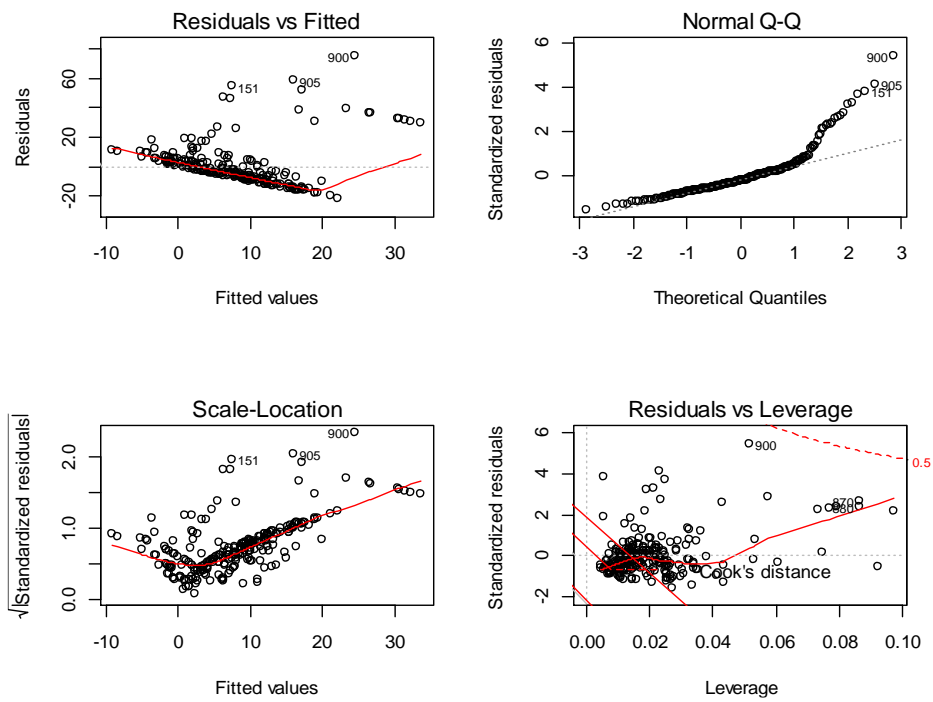


Fig. 6. Plots of final linear regression model for leishmaniasis and environmental variables (Digital Elevation Model and Normalized Difference Vegetation Index)

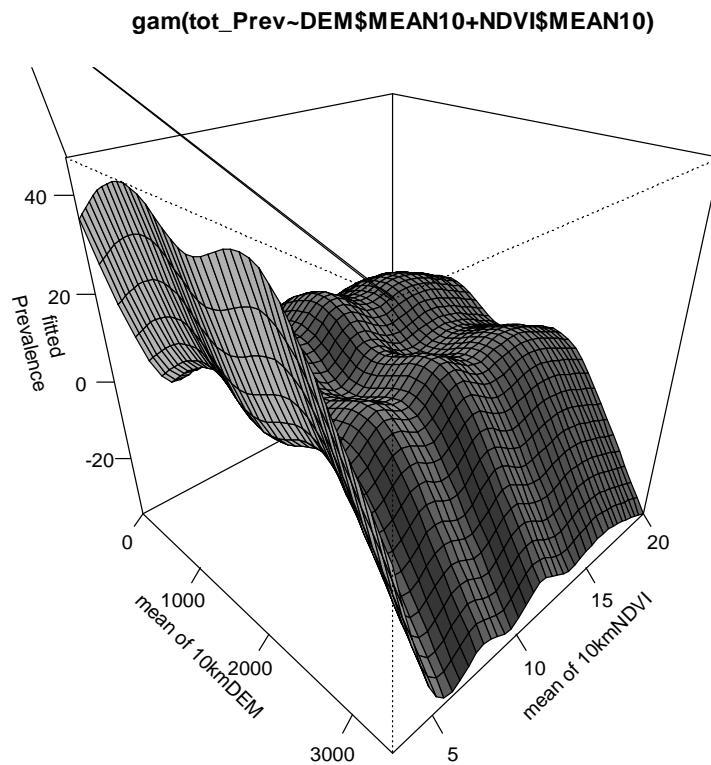


Fig. 7. Fitted surface for the additive nonparametric regression of total prevalence of leishmaniasis on mean of 10km buffer for Digital Elevation Model and Normalized Difference Vegetation Index

Table 5. Multiple linear regression models for disease mode and Normalized Difference Vegetation Index subcategory (buffer distance around the cities or villages)

	NDVI 5km_total	NDVI 5km_ACL	NDVI 5km_ZVL	NDVI 5km_ZCL	Buffer types
Active lesion prevalence	~MEAN+MAJORITY AR*: 0.0338	**Ns	***Na	Ns	~MAX50km AR: 0.093
Total prevalence	~MIN AR: 0.012	Ns	~MEAN AR:0.39	~ MIN + MI-NORITY AR: 0.3479	~ MIN5km AR: 0.012
Incidence	~MEAN AR: 0.0085	~ MINORITY AR: 0.042	Ns	~MEDIAN AR: 0.0259	~MEAN20km AR: 0.007

*AR: Adjusted R-squared **Ns: Not significant ***Na: Not available

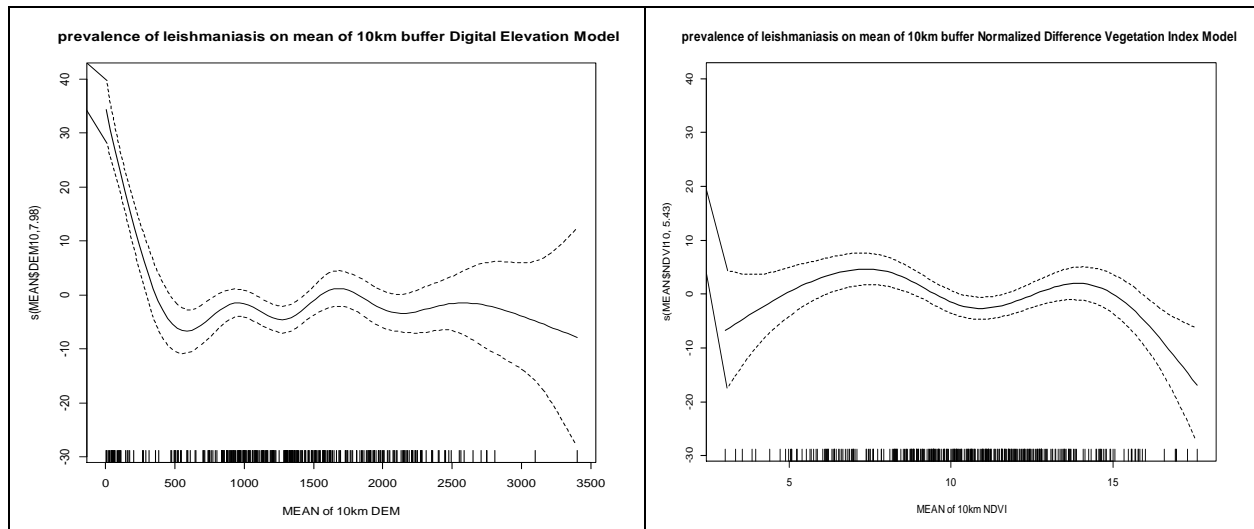


Fig. 8. Partial-regression functions for the additive regression of total prevalence of leishmaniasis on mean of 10km buffer Digital Elevation Model and Normalized Difference Vegetation Index. The broken lines give point-wise 95-percent confidence envelopes around the fit

Discussion

Maps based on the actual data for distribution of all leishmaniasis types in Iran, with identifying the detailed variables of environmental factors affecting the disease distribution, are the most important findings of this study. Due to the limited access to official leishmaniasis data of Center for Disease Control, integration all data from both CL types and rural and urban foci in the heart of the county official reports (lack of separation species), tens of kilometers

distance between some foci and center of counties often identified as disease points (lack of spatial resolution), unknown types of CL in most of official data, and passive screening in Public Health Centers, we decided to use the real data available in the literatures with a detailed focus on disease points. Many of scientific epidemiological studies predominantly focus on foci of diseases and have very valuable and verified information about all aspects of

the disease, such as prevalence, incidence, vectors, reservoirs, and control of disease data. Maps that contain all of the scientific information investigate by researchers of a country over several decades could well represent the disease situation in that country. Although the nature of the leishmaniasis itself was not the aim of our study, however, distribution of disease to the breakdown mapping can present more accurate view of disease for researchers.

Though vegetation statistically has a significant relationship with the prevalence of vector-borne diseases, this study (especially for the simple linear regression models), with a weak confirmation, implicitly point to a complex and indirect relationship. Therefore, more detailed studies and comprehensive investigations would be needed to prove the theorem. A study on effect of minimum, maximum and mean NDVI on the distribution of the VL vector in district of Bihar, India showed that max and mean variables with an R^2 about 0.6 strongly associated with the disease (14). Werneck et al. with a mention to the NDVI, emphasized on the impact of multilevel variable modeling on the development of leishmaniasis in Brazil (154). Study of environmental factors influencing the distribution of vectors involved in transmission of disease in north-eastern Italy revealed that areas with high winter NDVI may be related to the survival of the larvae in moist soil (155). According to the results of relationship between CL and NDVI in north-east province of Iran, Golestan, arid and semi-arid regions with low vegetation are the most involved area for CL occurrence (11). In our study, as expected due to their rural and nomadic natures of ZCL and ZVL in Iran, the values for NDVI index increase for ACL, ZCL, and ZVL respectively (Table 2). There is not much variation among them, and all three types are almost inclined to intermediate vegetation (neither too sparse nor too dense). The distinction in the result of numerous studies would be aroused from differences between climate factors, socio-economic variables, in-

strumental confounders and differences in the interpretation of the results.

Incidence of various types of leishmaniasis in different heights may result from the diversity in the reservoirs and vectors of the disease. Whereas ACL and ZVL had an interest for presence in high altitude areas, ZCL have been distributed in lower altitude (Table 2). This result for ZVL has obtained also in Ilam Province spatial modeling, and elevation had a negative impact on CL prevalence (156). Multivariate analysis in a VL study in eastern Sudan found the elevation as important variables, while the primary analysis did not show any correlation with disease incidence (157). However, while DEM can use as a good variable for topographic purposes, it may do not has most appropriate function as elevation in all regions (155).

The role of environmental factors in the development of disease vectors is obvious. While the CL is the main vector-borne disease in Iran, about 80% of cases are zoonotic and spread in 17 out of 31 provinces (158). Some studies have been modeled the environmental factors for vectors and reservoirs of leishmaniasis. Altitude and land cover was found to be from most important factors for suitability of niches for sand flies. Areas with 990m were found and 1235m average altitude were related to probability of presence of ZCL and ACL vectors respectively (159). Niche modeling of main reservoir hosts in Iran showed that among topography variables, slope has the most contribution in forecasting risk of disease (160). A predictive degree-day model was used for development time, population dynamics and activities of VL vector sandflies in field in northwest Iran (161). Among climate factors, minimum of temperature, mean of humidity, and rainfall had the most impact on ZVL distribution in Golestan, north-east of Iran (162). None of these studies include NDVI as a variable.

The vegetation coverage has been discussed as a risk factor for VL and CL types

in previous studies (163, 164). The NDVI has not considered for intrinsic specifications itself but also it covers some other important environmental and ecological variables such as soil types and moisture, humidity, slope degree and even elevation of its ambient land (16). The majority of studies, evaluated the presence/absence of vegetation or have used a cut-off value point for NDVI, but less attention has been paid to the sheer numerical modeling. Linear regression results in this study show that none of subcategories of independent variables have a favorable associated with DEM and NDVI and do not fit on the linear model very well (low R-squared). Only some of models when focused on leishmaniasis types, the R-squared reach to 0.2–0.4. This fact could be predicted from plots associated with variables (Fig. 4). Despite the lack of goodness of fitness of simple linear models, Additive Nonparametric Regression analysis presents a better prediction model for data of this study. These models do not output real intercepts for variables, but using their internal formulas, they are able to provide predictive models for input data. As a nonparametric regression attributes, no obvious estimates were seen for parameters (21), and then to figure out regression results, regards to necessitate of using fitted regression graphics, we used this method as shown in Fig. 7. Data frame used to find predicted values on the regression surface, fitted to data of model to drawn this figure.

For more clarification, functions which explained previously that present in inherent content of the additive regression, have been shown in partial regression of our model in Fig. 8.

Conclusion

Although the results of modeling and predictive power of the models in this study was not great but was somewhat satisfied. Modeling environmental factors which affecting ecological disease, need attention to the role

of all these factors together. Moreover, integration the scenarios such as finding hotspots in GIS, using statistical logistic regression, more specific factors of diseases such as vector and reservoir species, can be used in modeling the relationships, more meaningful and more clear. However, a good model needs to consider all the factors involved the prevalence and incidence of a disease.

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References

1. Cerbino Neto J, Werneck GL, Costa CHN (2009) Factors associated with the incidence of urban visceral leishmaniasis: an ecological study in Teresina, Piauí State, Brazil. *Cadernos de Saúde Pública*. 25(7): 1543–1551.
2. WHO (2016) Leishmaniasis fact sheet no. 375. World Health Organization, Geneva, Switzerland. Available at: <http://www.who.int/mediacentre/factsheets/fs375/en/>.
3. Sharifi I, Aflatoonian MR, Fekri AR, Hakimi Parizi M, Aghaei Afshar A,

- Khosravi A, Sharifi F, Aflatoonian B, Khamesipour A, Dowlati Y, Modabber F, Nadim A (2015) A comprehensive review of cutaneous leishmaniasis in Kerman Province, southeastern Iran—narrative review article. *Iran J Public Health*. 44(3): 299–307.
4. Kassiri H, Sharifinia N, Jalilian M, Shemshad K (2012) Epidemiological aspects of cutaneous leishmaniasis in Ilam Province, west of Iran (2000–2007). *Asian Pac J Trop Med*. 2: 382–386.
 5. Mohebbali M, Edrissian GH, Shirzadi MR, Akhoundi B, Hajjaran H, Zarei Z, Molaei S, Sharifi I, Mamishi S, Mahmoudvand H, Torabi V, Moshfe A, Malmasi A, Motazedian MH, Fakhar M (2011) An observational study on the current distribution of visceral leishmaniasis in different geographical zones of Iran and implication to health policy. *Travel Med Infect Dis*. 9(2): 67–74.
 6. Mohebbali M, Edrissian GH, Nadim A, Hajjaran H, Akhoundi B, Hooshmand B, Zarei Z, Arshi S, Mirsamadi N, Naeini Manouchehri K (2006) Application of direct agglutination test (DAT) for the diagnosis and seroepidemiological studies of visceral leishmaniasis in Iran. *Iran J Parasitol*. 1(1): 15–25.
 7. Ghatee MA, Sharifi I, Haghdoost AA, Kannejad Z, Taabody Z, Hatam G, Abdollahpanah A (2013) Spatial correlations of population and ecological factors with distribution of visceral leishmaniasis cases in southwestern Iran. *J Vector Borne Dis*. 50(3): 179–187.
 8. Mohebbali M (2012) Epidemiological status of visceral leishmaniasis in Iran: experiences and review of literature. *Int J Clin Exp Pathol*. S3: 003.
 9. Masimalai P (2014) Remote sensing and Geographic Information Systems (GIS) as the applied public health and environmental epidemiology. *Int J Med Sci Public Health* 3(12): 1430–1438.
 10. Ready PD (2010) Leishmaniasis emergence in Europe. *Euro Surveill*. 15(10): 19505–19516.
 11. Mollalo A, Alimohammadi A, Shahrivand M, Shirzadi MR, Malek MR (2014) Spatial and statistical analyses of the relations between vegetation cover and incidence of cutaneous leishmaniasis in an endemic province, northeast of Iran. *Asian Pac J Trop Dis*. 4(3): 176–180.
 12. Rassi Y, Saghaipour A, Abai MR, Oshaghi MA, Mohebbali M, Mostafavi R (2013) Determination of *Leishmania* Parasite Species of Cutaneous Leishmaniasis Using PCR Method in Central County, Qom Province. *Zahedan J Res Med Sci*. 15(12): 13–16.
 13. Palaniyandi M (2014) GIS for disease surveillance and health information management in India. *Geospatial Today*. 13(5): 44–46.
 14. Bhunia GS, Kesari S, Chatterjee N, Mandal R, Kumar V, Das P (2012) Seasonal relationship between normalized difference vegetation index and abundance of the *Phlebotomus kalarazar* vector in an endemic focus in Bihar, India. *Geospatial Health*. 7(1): 51–62.
 15. Tsegaw T, Gadisa E, Seid A, Abera A, Teshome A, Mulugeta A, Herrero M, Argaw D, Jorge A, Aseffa A (2013) Identification of environmental parameters and risk mapping of visceral leishmaniasis in Ethiopia by using geographical information systems and a statistical approach. *Geospatial Health*. 7(2): 299–308.
 16. Palaniyandi M, Anand PH, Maniyosai R (2014) Climate, landscape and the environments of visceral leishmaniasis

- transmission in India, using remote sensing and GIS. *J Geophys Remote Sens.* 3(3): 1–6.
17. R Core Team (2015) R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing. Vienna, Austria.
 18. Hereher ME (2015) Environmental monitoring and change assessment of Toshka lakes in southern Egypt using remote sensing. *Environ Earth Sci.* 73(7): 3623–3632.
 19. Land Processes DAAC NASA (2015) USGS Earth Resources Observation and Science (EROS) Center, MODIS Reprojection Tool (MRT). Sioux Falls, South Dakota. Available at: <https://mrtweb.cr.usgs.gov/>.
 20. NASA and Japan ASTER Program (2015) ASTER Global Digital Elevation Model (ASTER GDEM) data center. Version 2. Available at: <http://gdem.ersdac.jspacesystems.or.jp/search.jsp>.
 21. Fox J, Weisberg S (2010) Nonlinear regression and nonlinear least squares in R: An appendix to an R companion to applied regression. Retrieved from McMaster University: <http://socserv.mcmaster.ca/jfox/Books/Companion/appendix/Appendix-Nonlinear-Regression.pdf>.
 22. Fox J, Weisberg S (2011) An {R} Companion to Applied Regression. Second ed. Sage, Thousand Oaks, CA.
 23. Fagih-Nayini F, Mohebbali M, Javadian E (2002) Epidemiology of Visceral leishmaniasis in Kordan-savojbalagh. Pajoohandeh J. 7(2): 9–15.
 24. Mazloumi Gavvani AS, Hodjati MH, Mohite H, Davies CR (2002) Effect of insecticide-impregnated dog collars on incidence of zoonotic visceral leishmaniasis in Iranian children: a matched cluster randomised trial. *The Lancet.* 360(9330): 374–379.
 25. Soleimanzadeh G, Edrissian GH, Movahhed-Danesh AM, Nadim A (1993) Epidemiological aspects of kala-azar in Meshkin-Shahr, Iran: human infection. *Bull World Health Organ.* 71(6): 759–762.
 26. Mohebbali M, Edrissian GH, Shirzadi MR, Hosseingholizadeh G, Pashaei MH, Ganji A, Zarei Z, Kousha A, Akhondi B, Hajjaran H, Malekafzali H (2010) Integrated visceral leishmaniasis surveillance system in primary care for children in Meshkin-Shahr district, north-western Islamic Republic of Iran. *Eastern Mediterranean health journal = La revue de sante de la Mediterranee orientale = al-Majallah al-sihhiyah li-sharq al-mutawasit.* 16(10): 1050–1054.
 27. Edrissian GH (1996) Visceral leishmaniasis in Iran and the role of serological tests in diagnosis and epidemiological studies. *Parasitology for 21st Century ICOPA VIII, Izmir, Turkey: CAB International*, pp. 63–78.
 28. Mohebbali M, Edrissian GH, Shirzadi MR, Hosseingholizadeh Y, Pashaei MH, Ganji A, Zarei Z, Kousha A, Akhondi B, Hajjaran H (2011) Establishment and integration of a visceral leishmaniasis surveillance system in the primary health care system and its evaluation, Meshkin-Shahr District, Iran. *J Sch Public Health Inst Public Health Res.* 9(2): 1–8.
 29. Mahami M, Moheb Ali M, Keshavarz H, Hajarani H, Akhondi B, Zarei Z, Charedar S (2006) A seroepidemiological survey of visceral leishmaniasis (KALA-AZAR) in Germe District, Ardabil Province. *J Sch Public Health Inst Public Health Res.* 4(1): 45–55.
 30. Mohammadi-Kheyraadi K, Mohebbali M, Mamishi S, Arshi S (2004) Epidemiological characteristics of Kala-Azar

- in hospitalized patients in Ardebil Province. *J Sch Public Health Inst Public Health Res.* 2(2): 11–24.
31. Nadim A, Tahvildar Birooni GH, Mohebbali M (1998) A report of cutaneous and post kala-azar dermal leishmaniasis in Moghan, Ardabil Province *Hakim Res J.* 1(1): 57–60.
 32. Arshi S, Mohebbali M, Akhouni B, Sadeghi-Bazargani H, Sepehram V, Zarei Z, Hajikhani S, Sezavar SH (2002) Identification of a new endemic focus of kala-azar and seroepidemiological study of visceral *Leishmania* infection in Ardabil Province. *J Sch Public Health Inst Public Health Res.* 1: 9–18.
 33. Mazloumi Gavvani AS, Malaki Ravasan N, Mazloumi Gavvani F (2011) Comparison of Nomadic and non-Nomadic lifestyles in transmission of Visceral Leishmaniasis. *J Gorgan Univ Med Sci.* 13(1): 94–100.
 34. Mirsamadi N, Mohebbali M, Atari MR, Edrisian GH (2003) Serological survey of Visceral leishmaniasis (Kala-Azar) in Azarshahr, Azarbaijan Province, northwest of Iran. *Hakim Res J.* 6(1): 17–22.
 35. Yaghoobi-Ershadi MR, Shahbazi F, Darvishi M, Akhavan AA, Jafari R, Khajeian M, Rassi Y, Soleimani H, Shirzadi MR, Hanafi-Bojd AA, Darabi H, Arandian MH, Sanei-Dehkordi A, Heidari M (2013) Molecular epidemiological study of cutaneous leishmaniasis in the focus of bushehr City, southwestern Iran. *J Arthropod Borne Dis.* 7(2): 113–121.
 36. Hamzavi V, Mohebbali M, Edrissian G, Fourozani A (2000) Epidemiological study of cutaneous leishmaniasis in Dashti and Dashtestani district of Bushehr Province. *Iran J Public Health.* 29: 177–191.
 37. Mohebbali M, Hamzavi Y, Edrissian GH, Forouzani A (2001) Seroepidemiological study of visceral leishmaniasis among humans and animal reservoirs in Bushehr Province, Islamic Republic of Iran. *Eastern Mediterranean health journal = La revue de sante de la Mediterranee orientale = al-Majallah al-sihhiyah li-sharq al-mutawassit.* 7(6): 912–917.
 38. Ghatee M, Sharifi I, Mirhendi H, Kanannejad Z, Hatam G (2013) Investigation of Double-Band Electrophoretic Pattern of ITS-rDNA Region in Iranian Isolates of *Leishmania tropica*. *Iran J Parasitol.* 8(2): 264–272.
 39. Razmjou S, Hejazy H, Motazedian MH, Baghaei M, Emamy M, Kalantary M (2009) A new focus of zoonotic cutaneous leishmaniasis in Shiraz, Iran. *Trans R Soc Trop Med Hyg.* 103(7): 727–730.
 40. Rassi Y, Javadian E, Jalali M, Motazedian MH, Vatndoost H (2004) Investigation on Zoonotic Cutaneous Leishmaniasis, Southern Iran. *Iran J Public Health.* 33(1): 31–35.
 41. Fakoorziba MR, Baseri A, Eghbal F, Rezaee S, Azizi K, Moemenbellah-Fard MD (2011) Post-earthquake outbreak of cutaneous leishmaniasis in a rural region of southern Iran. *Ann Trop Med Parasitol.* 105(3): 217–224.
 42. Davami MH, Motazedian MH, Sarkari B (2010) The changing profile of cutaneous leishmaniasis in a focus of the disease in Jahrom District, southern Iran. *Ann Trop Med Parasitol.* 104(5): 377–382.
 43. Sharafi M, Pezeshki B, Reisi A, Kalantari M, Naghizadeh MM, Dast Manesh S (2013) Detection of Cutaneous Leishmaniasis by PCR in Fasa District in 2012. *J Fasa Univ Med Sci.* 3(3): 266–270.
 44. Noorpisheh S, Naghizadeh MM, Nikrouz L (2013) A study on the life quality

- of patients suffering from leishmaniasis. J Fasa Univ Med Sci. 3(2): 155–162.
45. Entezari M, Eskandari F (2014) Relationship between Climatic Factors and the Prevalence of Cutaneous Leishmaniasis in Larestan City J Mil Med. 16 (2): 99–104.
 46. Fakhari M, Motazedian MH, Hatam GR, Asgari Q, Kalantari M, Mohebbali M (2008) Asymptomatic human carriers of *Leishmania infantum*: possible reservoirs for Mediterranean visceral leishmaniasis in southern Iran. Ann Trop Med Parasitol. 102(7): 577–583.
 47. Edrissian GH, Ahanchin AR, Gharachahi AM, Ghorbani M, Nadim A, Ardehali S, Hafizi A (1993) Seroepidemiological studies of visceral leishmaniasis and search for animal reservoirs in Fars province, southern Iran. Iran J Med Sci. 18(3–4): 99–105.
 48. Sahabi Z, Seyedi Rashti MA, Nadim A, Javadian E, Kazemeini M, Abai MR (1992) A Preliminary Report on the Natural Leptomonad Infection of *Phlebotomus Majorin* an Endemic Focus of Visceral Leishmaniasis (VI) in Fars Province, South of Iran. Iran J Public Health. 21(1–4): 87–93.
 49. Fakhari M, Motazedian M, Asgari G, Mohebbali M, Mehrbani D (2006) A new endemic focus of Visceral Leishmaniasis in southern Iran. Armaghane-Danesh. 11(2): 103–113.
 50. Mollalo A, Alimohammadi A, Shirzadi MR, Malek MR (2015) Geographic information system-based analysis of the spatial and spatio-temporal distribution of zoonotic cutaneous leishmaniasis in Golestan Province, north-east of Iran. Zoonoses Public Health. 62(1): 18–28.
 51. Rahbarian N, Mesgarian A, Mahmoudi Rad M, Hajaran H, Shahbazi F, Mesgarian Z, Taghipour N (2009) Identification of *Leishmania* species isolated from human cutaneous leishmaniasis using PCR method. J Res Health Sci. 9(2): 48–51.
 52. Sofizadeh A, Cherabin M, Mehravaran A (2012) Cutaneous leishmaniasis in Gonbad Kavos, North of Iran (2009–11): an epidemiological study. J Gorgan Univ Med Sci. 14(4): 100–106.
 53. Baghaei A, Parvizi P, Amirkhani A, Honarvar MR, Badiei F (2012) Identification of *Leishmania* using microscopic and molecular methods in suspected patients of Cutaneous Leishmaniasis by targeting ITS-rDNA gene, Golestan Province, Iran (2009–10). J Gorgan Univ Med Sci. 14(3): 72–81.
 54. Mesgarian F, Rahbarian N, Hajaran H, Shahbaz F, Mesgarian Z, Taghipour N (2010) Identification of *Leishmania* species isolated from human cutaneous Leishmaniasis in Gonbad-e-Qabus city using a PCR method during 2006–2007. J Tehran Univ Med Sci. 68(4): 250–256.
 55. Hanafi Bojd AA, Yaghoobi-Ershadi MR, Zamani GH, Barzekar A, Jafari R, Pour Abazari G (2006) Epidemiologic aspect of Cutaneous Leishmaniasis in Hajiabad, Hormozgan, Iran (2003). Medical J Hormozgan Univ Med sci. 10(1): 63–70.
 56. Nazari M (2012) Cutaneous leishmaniasis in Hamadan, Iran (2004–2010). Zahedan J Res Med Sci. 13(9): 39–42.
 57. Azizi K, Soltani A, Alipour H (2012) Molecular detection of *Leishmania* isolated from cutaneous leishmaniasis patients in Jask County, Hormozgan Province, Southern Iran, 2008. Asian Pac J Trop Med. 5(7): 514–517.
 58. Asgari Nezhad H, Borhani M, Norouzi M, Merzaie M (2012) Cutaneous Leishmaniasis in school children in a border area at south-west of Iran. Sci Parasitol. 13(4): 153–158.

59. Shazad B, Abbaszadeh B, Khamesipour A (2005) Comparison of topical paromomycin sulfate (twice/day) with intralesional meglumine antimoniate for the treatment of cutaneous leishmaniasis caused by *L. major*. *Eur J Dermatol*. 15(2): 85–87.
60. Bahrami AM, Delpisheh A, Zamandousti A (2011) Epidemiologic aspects of cutaneous Leishmaniasis and therapeutic effect of traditional ointment containing onion juice compared with Amphotricin B. *J Kermanshah Univ Med Sci*. 15(4): 251–259.
61. Yazdanpanah HA, Rostamianpur M (2013) Analysis of Spatial Distribution of Leishmaniasis and its Relationship with Climatic Parameters (Case Study: Ilam Province). *Bull Env Pharmacol Life Sci*. 2(12): 80–86.
62. Roghani A, Yasemi M, Jalilian M, Abdi J, Rezayee K (2012) Epidemiology of cutaneous leishmaniasis in Ilam. *Pajohesh dar Pezeshki*. 36(5): 50–53.
63. Shiee MR, Mohebbali M, Doroodgar A, Teimouri A, Afzali H, Shirzadi MR (2012) A molecular and parasitological survey on cutaneous leishmaniasis patients from historical city of Kashan in Isfahan Province, center of Iran. *Asian Pac J Trop Dis*. 2(6): 421–425.
64. Zahraei-Ramazani AR, Yaghoobi-Ershadi MR, Mokhtari AR, Akhavan AA, Abdoli H, Arandian MH (2007) Anthroponotic Cutaneous Leishmaniasis in Nonendemic Quarters of a Central City in Iran. *Iran J Public Health*. 36(2): 7–11.
65. Doroodgar A, Sayyah M, Doroodgar M, Mahbobi S, Nemetian M, Rafizadeh S, Rassi Y (2012) Progressive increasing of cutaneous leishmaniasis in Kashan district, central of Iran. *Asian Pac J Trop Med*. 2(4): 260–263.
66. Talari S, Rezvan Talaei R, Shajari G (2006) A clinical and epidemiology survey of cutaneous leishmaniasis in childhood. *J Med Sci*. 6(4): 693–697.
67. Yaghoobi-Ershadi M, Hanafi-Bojd A, Akhavan A, Zahraei-Ramazani A, Mohebbali M (2001) Epidemiological study in a new focus of cutaneous leishmaniasis due to *Leishmania major* in Ardestan town, central Iran. *Acta Trop*. 79(2): 115–121.
68. Mohammadian A, Almasi M, Kheybari H, Karimi F, Tehrani S (1999) Identification of species and characteristics of an outbreak of cutaneous leishmaniasis in a new focus of Iran. *Biomedical J Iran*. 3(1): 31–39.
69. Yaghoobi-Ershadi M, Javadian E (1995) leishmaniose cutanée zoonotique au nod d'Ispahan: le point sur l'infection humaine en 1991. *Bull Soc Pathol Exot*. 88(1): 42–45.
70. Yaghoobi-Ershadi M, Moosa-Kazemi S, Zahraei-Ramazani A, Jalai-Zand A, Akhavan A, Arandian M, Abdoli H, Houshmand B, Nadim A, Hosseini M (2006) Evaluation of deltamethrin-impregnated bed nets and curtains for control of zoonotic cutaneous leishmaniasis in. *Bull Soc Pathol Exot*. 99(1): 43–48.
71. Nilforoushzadeh MA, Tahmores J, Ataei B, Jaafary F (2002) Evaluation of integrated pest management method on controlling zoonotic Cutaneous Leishmaniasis in Emamzadeh Agha Ali Abbas, Natanz County, Isfahan Province, 1996–1998. *Iran J Dermatol*. 6(2): 78–83.
72. Doroodgar A, Mahbobi S, Nemetian M, Sayyah M, Doroodgar M (2009) An epidemiological study of cutaneous leishmaniasis in Kashan (2007–2008). *Koomesh*. 10(3): 177–184.
73. Ahmadi NA, Ghafarzadeh M, Jalali Galoosang A, Gholami Parizad E (2013) An epidemiological study of Cutaneous Leishmaniasis with emphasis on incidence rate in Kashan, Isfahan Prov-

- ince. Scientific J Ilam Univ Med Sci. 21(2): 1–9 (In Persian).
74. Ebadi M, Hejazi SH (2003) The epidemiological study of cutaneous Leishmaniasis situation in the students of primary school in Isfahan Borkhar region. *J Kerman Univ Med Sci.* 10(2): 92–98.
 75. Yaghoobi-Ershadi M, Hanafi A, Akhavan A, Zahraei Ramazani A, Mohebbali M (1999) Cutaneous leishmaniasis in Ardestan Town. *Hakim Res J.* 1(3): 206–215.
 76. Doroudgar A, Dehghani R, Hooshyar H, Sayyah M (1996) Study of the prevalence of the cutaneous Leishmaniasis in south-east part of Kashan. *J Kerman Univ Med Sci.* 3(2): 80–86.
 77. Nadim A, Aflatoonian MR (1995) Anthroponotic cutaneous leishmaniasis in the city of Bam, southeast Iran. *Iran J Public Health.* 24(1–2): 15–24.
 78. Sharifi I, Nakhaei N, Aflatoonian M, Parizi MH, Fekri A, Safizadeh H, Shirzadi M, Gooya M, Khamesipour A, Nadim A (2011) Cutaneous leishmaniasis in bam: a comparative evaluation of pre- and post-earthquake years (1999–2008). *Iran J Public Health.* 40(2): 49–56.
 79. Yaghoobi-Ershadi MR (1977) Cutaneous leishmaniasis in Kerman. [MSPH theses]. School of Public Health, Tehran University, Iran.
 80. Mirzaei M, Sharifi I, Poursmaelian S (2012) A new focus of anthroponotic cutaneous leishmaniasis and identification of parasite species by nested PCR in Jiroft, Iran. *Comp Clin Path.* 21(5): 1071–1075.
 81. Aflatoonian MR, Sharifi I, Hakimi Parizi M, Fekri AR, Aflatoonian B, Sharifi M, Khosravi A, Khamesipour A, Sharifi H (2014) A prospective cohort study of cutaneous leishmaniasis risk and opium addiction in south eastern Iran. *PLoS One.* 9(2): e89043.
 82. Sharifi I, Fekri AR, Aflatoonian MR, Nadim A, Nikian Y, Kamesipour A (1998) Cutaneous leishmaniasis in primary school children in the south-eastern Iranian city of Bam, 1994–95. *Bull World Health Organ.* 76(3): 289–293.
 83. Sharifi I, Poursmaelian S, Aflatoonian MR, Ardakani RF, Mirzaei M, Fekri AR, Khamesipour A, Parizi MH, Harandi MF (2011) Emergence of a new focus of anthroponotic cutaneous leishmaniasis due to *Leishmania tropica* in rural communities of Bam District after the earthquake, Iran. *Trop Med Int Health.* 16(4): 510–513.
 84. Aflatoonian MR, Sharifi I, Poursmaelian S, Hakimi-Parizi M, Ziaali N (2013) The Emergence of Anthroponotic Cutaneous Leishmaniasis Following the Earthquake in Southern Villages of Bam District, southeastern Iran, 2010. *J Arthropod Borne Dis.* 7(1): 8–14.
 85. Nadim A, Motabar M, Houshmand B, Keyghobadi K, Aflatoonian MR (1995) Evaluation of pyrethroid impregnated bednets for control of anthroponotic cutaneous leishmaniasis in Bam (Islamic Republic of Iran), WHO/LEISH/95. 37: Geneva. p. 21.
 86. Sharifi I, Aflatoonian MR, Aflatoonian B, Kermanizadeh A (2015) The severity of cutaneous leishmaniasis before and after the earthquake in Bam, southeastern Iran. *J Parasit Dis.* 39(4): 741–744.
 87. Aflatoonian MR, Sharifi I (2011) The Epidemiology of cutaneous leishmaniasis in the city and suburb of Bam in 2010: Active case-finding, treatment and health education of the school children. *Iran J Epidemiol.* 7(3): 52–57.
 88. Aflatoonian MR, Sharifi I (2010) Prevalence rate of cutaneous leishmaniasis in Bam District during 20 Years (1988–2007). *J Kerman Univ Med Sci.* 17(4): 297–306.

89. Mirzadeh A, Hejazi Zadeh B, Mesgar pour B, Golezar A, Halakooiiy Nayini K (2008) The process of development of high-risk centers of cutaneous leishmaniasis in the city of Kerman in 2002–2005 years and evaluation of effective environmental factors by using GIS. *Iran J Epidemiol.* 4(3 and 4): 5–17.
90. Aflatoonian MR, Sharifi I, Aflatoonian B (2014) The Effects of Prevention Knowledge Compared to the Improvement of Housing Conditions in Control of Anthroponotic Cutaneous Leishmaniasis: a Natural Experience of Earthquake in Bam/Iran. *J Kerman Univ Med Sci.* 21(3): 247–258.
91. Aflatoonian MR, Sharifi I (2007) Prevalence of Cutaneous Leishmaniasis in School Children in Bam and Barawat. Iran in 2006. *J Kerman Univ Med Sci.* 14(2): 82–89.
92. Akhavan AA, Yaghoobi-Ershadi MR, Hasibi F, Jafari R, Abdoli H, Arandian MH, Soleimani H, Zahraei-Ramazani AR, Mohebbali M, Hajjaran H (2007) Emergence of cutaneous leishmaniasis due to *Leishmania major* in a new focus of southern Iran. *J Arthropod Borne Dis.* 1(1): 1–8.
93. Aghaei-Afshar A, Vatandoost H, Sharifi I, Rassi Y, Abai MR, Oshaghi MA, Yaghoobi-Ershadi MR, Rafizadeh S (2013) First determination of impact and outcome indicators following indoor residual spraying (IRS) with deltamethrin in a new focus of anthroponotic cutaneous leishmaniasis (ACL) in Iran. *Asian Pac J Trop Med.* 3(1): 5–9.
94. Khosravi A, Sharifi I, Dortaj E, Aghaei Afshar A, Mostafavi M (2013) The present status of cutaneous leishmaniasis in a recently emerged focus in South-west of Kerman Province, Iran. *Iran J Public Health.* 42(2): 182–187.
95. Sharifi I, Zemani F, Aflatoonian MR, Fekri AR (2008) A report of cutaneous leishmaniasis epidemic and its probable causative factors in Baft District, Kerman Province. *Iran J Epidemiol.* 4(1): 53–58.
96. Mahmoudvand H, Mohebbali M, Sharifi I, Keshavarz H, Hajjaran H, Akhoundi B, Jahanbakhsh S, Zarean M, Javadi A (2011) Epidemiological aspects of visceral leishmaniasis in Baft District, Kerman Province, Southeast of Iran. *Iran J Parasitol.* 6(1): 1–11.
97. Barati M, Parizi MHD, Sharifi I (2008) Epidemiological and Clinical aspects of kala-azar in hospitalized children of Kerman Province, during 1991–2006. *J Kerman Univ Med Sci.* 15(2): 148–155.
98. Hamzavi Y, Khademi N (2015) Trend of cutaneous leishmaniasis in Kermanshah Province, west of Iran from 1990 to 2012. *Iran J Parasitol.* 10(1): 78–86.
99. Nazari N, Faraji R, Vejdani M, Mekaeili A, Hamzavi Y (2012) Prevalence of Cutaneous leishmaniasis in Patients referred to Kermanshah hygienic centers (2006–2008) Zahedan. *J Res Med Sci.* 14(8): 77–79.
100. Hamzavi Y, Khademi N (2013) The Analytical study of Cutaneous Leishmaniasis in Kermanshah (2011–2012). *J Kermanshah Univ Med Sci.* 17(9): 582–589.
101. Hamzavi Y, Hamzeh B, Mohebbali M, Akhoundi B, Ajhang K, Khademi N, Ghadiri K, Bashiri H, Pajhouhan M (2012) Human visceral leishmaniasis in Kermanshah Province, Western Iran, during 2011–2012. *Iran J Parasitol.* 7(4): 49–56.
102. Alavinia S, Arzamani K, Reihani M, Jafari J (2009) Some epidemiological aspects of cutaneous leishmaniasis in Northern Khorasan Province, Iran. *J Arthropod Borne Dis.* 3(2): 50–54.
103. Torabi V, Mohebbali M, Edrissian GH, Keshavarz H, Mohajery M, Hajjaran

- H, Akhoondi B, Sanaati A, Zarei Z, Delshad A (2007) Seroepidemiology of visceral leishmaniasis by DAT in Bojnord-North Khorasan. *Iran J Epidemiol.* 4(3 and 4): 43–50.
104. Sattar Pagheh A, Fakhar M, Sharif M, Danesh V, Ahmadi Z (2013) Epidemiological Survey of Cutaneous Leishmaniasis due to *Leishmania tropica* in a New Focus in Khorasan Razavi Province. *J Mazandaran Univ Med Sci.* 23(103): 47–53.
105. Moosa-Kazemi SH, Yaghoobi-Ershadir MR, Akhavan AA, Abdoli H, Zahraei-Ramazani AR, Jafari R, Houshmand B, Nadim A, Hosseini M (2007) Deltamethrin-impregnated bed nets and curtains in an anthroponotic cutaneous leishmaniasis control program in northeastern Iran. *Ann Saudi Med.* 27(1): 6–12.
106. Saadabadi F, Mohajery M, Poostchi E, Shamsian SAA (2013) Identification of *Leishmania* species causing cutaneous leishmaniasis using Random Amplified Polymorphic DNA (RAPD-PCR) in Kharve, Iran. *Reports of Biochemistry and Molecular Biology.* 1 (2): 69–73.
107. Hassanpour K, Aghamollaei H, Golpich M, Amani J, Taheri A, Farnoosh G (2014) Molecular epidemiological study of cutaneous leishmaniasis in the east north of Iran. *Asian Pac J Trop Dis.* 4: 540–544.
108. Hoseini Farash BR, Mohajery M, Shamsian AA, Rezaie A (2011) Identification of *Leishmania* species causing cutaneous Leishmaniasis by PCR method in Torghabeh-Shandiz during 2009–2010. Forth Congress of Laboratory and Clinic, 2011 December 21–23, Tehran, Iran.
109. Khajedaluae M, Yazdanpanah M, Seyed Nozadi SM, Fata A, Juya MR, Masoudi MH, Najaf Najafi M (2014) Epidemiology of cutaneous leishmaniasis in Razavi Khorasan in 2011. *Medical J Mashhad Univ Med sci.* 57(4): 647–654.
110. Mohajery M, Hajaran H, Shamsian A, Tavakol Afshari J, Saadabadi F (2008) Identification of cutaneous leishmaniasis species by RAPD-PCR in Neyshabur. *Medical J Mashhad Univ Med sci.* 51(100): 79–86.
111. Yaghoobi-Ershadi MR, Akhavan AA, Zahraei-Ramazani AV, Abai MR, Ebrahimi B, Vafaei-Nezhad R, Hanafi-Bojd AA, Jafari R (2003) Epidemiological study in a new focus of cutaneous leishmaniasis in the Islamic Republic of Iran. *Eastern Mediterranean health journal = La revue de sante de la Mediterranee orientale = al-Majallah al-sihhiyah li-sharq al-mutawassit.* 9 (4): 816–826.
112. Karimi Zarchi AA, Mahmodzadeh A, Vatani H, Shirbazo S (2004) Epidemiological study of Cutaneous Leishmaniasis in Sarakhs District. *J Shahid Sedoghi Univ Med Sci.* 12(1): 30–35.
113. Sahabi Z, Mohajery M, Seyedi Rashti M (1999) Fauna of arthropods vectors and their medical importance in the free trade zone Sarakhs ferns, sand flies and leishmaniasis. *Hakim Res J.* 2(3): 177–182.
114. Akbari E, Mayvaneh E, Entezari A, Nazari M (2014) Survey of the Role of Bioclimatic Factors in the Outbreak of Cutaneous Leishmaniasis. *Iran J Epidemiol.* 10(3): 65–74.
115. Karamian M, Bojd F, Sedigh M, Hemmati M, Saadatjoo A, Barati DA (2013) Molecular identification of cutaneous leishmaniasis agents in Birjand, Iran. *J Birjand Univ Med Sci.* 20(2): 183–190.
116. Kassiri H, Shemshad K, Kassiri A, Shojaei S, Sharifinia N, Shemshad M (2012) Clinical laboratory and epidemiological research on cutaneous leish-

- maniasis in the south west of Iran. Archives of Clinical Infectious Diseases. 7(4): 128–131.
117. Kassiri H, Shojaei S, Kolahkaji K (2012) Epidemiological status of cutaneous leishmaniasis in Behbahan County, Khuzestan Province, Southwestern Iran. Int J Infect Dis. 16(Supplement 1): e348.
 118. Kassiri H, Lotfi M, Farajifard P, Kassiri E (2014) Laboratory diagnosis, clinical manifestations, epidemiological situation and public health importance of cutaneous leishmaniasis in Shushtar County, Southwestern Iran. J Acute Disease. 3(2): 93–98.
 119. Kassiri H, Feizhaddad MH, Abdehpanah M (2014) Morbidity, surveillance and epidemiology of scorpion sting, cutaneous leishmaniasis and pediculosis capitis in Bandar-Mahshahr County, Southwestern Iran. J Acute Disease. 3(3): 194–200.
 120. Vazirianzadeh B, Hoseini S, Pour Rezaee S, Gardani H, Amraee K (2014) Prevalence of Cutaneous Leishmaniasis in Ramshir, Iran, An Epidemiological Study. International Archives of Health Sciences. 1(1): 41–37.
 121. Kassiri H, Shemshad K, Lotfi M, Shemshad M (2013) Relationship Trend Analysis of of Cutaneous Leishmaniasis Prevalence and Climatological Variables in Shush County, South-West of Iran (2003–2007). Acad J Entomol. 6(2): 79–84.
 122. Spotin A, Rouhani S, Parvizi P (2014) The associations of *Leishmania major* and *Leishmania tropica* aspects by focusing their morphological and molecular features on clinical appearances in Khuzestan Province, Iran. Bio Med Res Int. 2014: 913510.
 123. Ghasemian M, Maraghi S, Samarbafzadeh AR, Jelowdar A, Kalantari M (2011) The PCR-based detection and identification of the parasites causing human cutaneous leishmaniasis in the Iranian city of Ahvaz. Ann Trop Med Parasitol. 105(3): 209–215.
 124. Kassiri H, Shemshad K, Shojaei S (2013) The incidence rate of cutaneous leishmaniasis in Behbahan County of Khuzestan Province, Southwest of Iran. Jundishapur J Microbiol. 6(5): e7045.
 125. Nejadi J, Mojadam M, Bojdi H, Ali A, Keyhani A, Habibi Nodeh F (2014) An epidemiological study of Cutaneous Leishmaniasis in Andimeshk (2005–2010). Scientific J Ilam Univ Med Sci. 21(7): 94–101.
 126. Behbahani A, Ahmadi S, Latifi SM, Sadeghi M (2012) Study of the frequency of Cutaneous Leishmaniasis in Omidieh District, Khouzestan Province, Southwest of Iran (2008–2010). Jundishapur J Health Sci. 4(4): 37–46.
 127. Kassiri H, Mortazavi SH, Kazemi S (2011) The epidemiological study of cutaneous leishmaniasis in Khorramshahr City, Khuzestan Province, Southwest of Iran. Jundishapur J Health Sci. 3(2): 11–20.
 128. Sarkari B, Pedram N, Mohebbali M, Moshfe AA, Zargar MA, Akhoundi B, Shirzadi MR (2010) Seroepidemiological study of visceral leishmaniasis in Booyerahmad district, southwest Islamic Republic of Iran. Eastern Mediterranean health journal = La revue de sante de la Mediterranee orientale = al-Majallah al-sihhiyah li-sharq al-mutawassit. 16(11): 1133–1136.
 129. Sarkari B, Moshfe AA, Pedram N, Zargar MA, Yazdanpanah B, Akhoundi B, Hasani Z, Mohebbali M (2007) Serological Study of Visceral Leishmaniasis in Boyer Ahmad Township in 2005. Armaghane-Danesh. 12(2): 69–77.
 130. Kheirandish F, Chegeni Sharafi A, Kazemi B, Mohebbali M, Sarlak A, Tarahi MJ, Holakouee K, Hajaran H (2013) Iden-

- tification of *Leishmania* species using PCR assay on giemsa-stained slides prepared from cutaneous leishmaniasis patients. Iran J Parasitol. 8(3): 382–388.
131. Amraee K, Rastegar HA, Beiranvand E (2013) An epidemiological study of cutaneous leishmaniasis in Poledokhtar District, Lorestan Province, Southwestern of Iran, 2001–2011. Jundishapur J Health Sci. 5(1): 55–62.
132. Ahmadi NA, Modiri M, Mamdohi S (2013) First survey of cutaneous leishmaniasis in Borujerd county, western Islamic Republic of Iran. Eastern Mediterranean Health J = La revue de sante de la Mediterranee orientale = al-Majalah al-sihhiyah li-sharq al-mutawasit. 19(10): 847–853.
133. Chegeni-Sharafi A, Amani H, Kayedi MH, Yarahahmadi A, Saki M, Mehrdad M, Nasiri E (2011) Epidemiological survey of cutaneous leishmaniasis in Lorestan Province (Iran) and introduction of disease transmission in new local areas. J Ilam Univ Med Sci. 19(1):54–60. [In Persian].
134. Chegeni-Sharafi A, Ourmazdi H, Mohebbali M, Akhlaghi L, Sharafi MM, Akhondi B (2005) Seroepidemiological study of Visceral Leishmaniasis (human infection) in east Miankooch area in Lorestan Province by Direct Agglutination Test (DAT). Yafte J Med Sci. 7(3 and 4): 31–35.
135. Rostami MN, Saghafipour A, Vesali E (2013) A newly emerged cutaneous leishmaniasis focus in central Iran. Int J Infect Dis. 17(12): e1198–e1206.
136. Akhavan AA, Yaghoobi-Ershadi MR, Mehdipour D, Abdoli H, Farzinnia B, Mohebbali M, Hajjaran H (2003) Epidemic outbreak of cutaneous leishmaniasis due to *leishmania major* in Ghanavat Rural District, Qom Province, Central Iran. Iran J Public Health. 32(4): 35–41.
137. Saghafipour A, Rassi Y, Abai MR, Oshaghi MA, Farzinnia B, Mostafavi R, Karimian F (2013) Outbreak of Zoonotic Cutaneous Leishmaniasis: A Report. Arch Hyg Sci. 2(2): 45–54.
138. Saghafipour A, Amir A, Rassi Y, Mostafavi R (2012) Epidemiology of cutaneous leishmaniasis in Qom. J Qom Univ Med Sci. 6(1): 83–88.
139. Mohammadi Azni S, Nokandeh Z, Khorsandi AA, Sanei Dehkordi AR (2010) Epidemiology of cutaneous leishmaniasis in Damghan. Iran J Mil Med. 12(3): 131–135.
140. Mohammadi Azni s, Rassi Y, Oshaghi MA, Yaghoobi-Ershadi M, Mohebbali M, Abai MR, Mohtarami F, Nokandeh Z, Rafizadeh S, Khojani GM (2011) Determination of parasite species of cutaneous leishmaniasis using Nested PCR in Damghan-Iran, during 2008. J Gorgan Univ Med Sci. 13(1): 59–65.
141. Mohammadi Azni S, Nokandeh Z, Sanei Dehkordi AR (2010) Control of rural cutaneous leishmaniasis in Damghan in 2005–2006. Iran J Infect Dis. 15 (48): 29–32.
142. Rafati N, Shapouri Moghadam M, Ghorbani R (2007) Epidemiological study of cutaneous Leishmaniasis in Damghan (2000–2006). Scientific J Semnan Univ Med Sci. 8(4): 247–253.
143. Fazaeli A, Fouladi B, Hashemi-Shahri SM, Sharifi I (2008) Clinical features of cutaneous leishmaniasis and direct PCR-based identification of parasite species in a new focus in southeast of Iran. Iran J Public Health. 37(3): 44–51.
144. Fazaeli A, Fouladi B, Sharifi I (2009) Emergence of cutaneous leishmaniasis in a border area at south-east of Iran: an epidemiological survey. J Vector Borne Dis. 46(1): 36–42.
145. Fouladi B, Sharifi I, Hashemi SS, Morad GH, Sarabandi NA, Ebrahimzadeh A, Fazaeli A (2007) Evaluation of a

- direct PCR in comparison with routine microscopy and in vitro culture for diagnosis of cutaneous leishmaniasis. *Zahedan J Res Med Sci.* 9(3): 9–15.
146. Salehzadeh A, Seyedi Rashti M (1996) Cutaneous leishmaniasis in North of Tehran. *Scientific J Hamadan Univ Med Sci.* 4(1): 34–39.
147. Abadi M (2004) Survey the situation of anthroponotic Cutaneous leishmaniasis in North-West of Tehran. [PhD dissertation]. Tehran Islamic Azad university, Iran.
148. Yaghoobi-Ershadi MR, Hanafi-Bojd AA, Javadian E, Jafari R, Zahraei-Ramazani AR, Mohebbali M (2002) A new focus of cutaneous leishmaniasis caused by *Leishmania tropica*. *Saudi Med J.* 23(3): 291–294.
149. Yaghoobi-Ershadi MR, Jafari R, Hanafi-Bojd AA (2004) A new epidemic focus of zoonotic cutaneous leishmaniasis in central Iran. *Ann Saudi Med.* 24(2): 98–101.
150. Jaafary R, Mohebbali M, Dehghan Dehnavi AR, Soleimani H, Akhavan AA, Hajarani H, Dehghan shadkam A, Fatahi J (2007) Epidemiological status of cutaneous leishmaniasis in Bafgh City, Yazd Province 2005. *J Shahid Sadooghi Univ Med Sci.* 15(2): 76–83.
151. Mozafari GH, Bakhshi Zadeh F (2011) Analysis the Roll of bioclimatic Factors in Prevalence of Cutaneous Leishmaniasis in Yazd-Ardakan plain Geography and Development J. 23: 185–202.
152. Hanafi Bojd AA, Dehghani Tafti AA, Jafari R, Ehrampoosh MH (2003) Efficacy of control program of zoonotic cutaneous leishmaniasis in a new focus of the disease. *Medical J Mashhad Univ Med sci.* 46(80): 19–25.
153. Yaghoobi-Ershadi MR, Marvi Moghadam N, Jaafary R, Akhavan A, Soleimani H, Zahraei Ramazani AR, Arandian MH, Dehghan Dehnavi AR (2008) Study of certain epidemiological aspects of cutaneous Leishmaniasis in Khatam County, Yazd Province, Iran. *J Shahid Sedoghi Univ Med Sci.* 15(4): 47–52.
154. Werneck G, Costa C, Walker A, David J, Wand M, Maguire J (2007) Multi-level modelling of the incidence of visceral leishmaniasis in Teresina, Brazil. *Epidemiol Infect.* 135(2): 195–201.
155. Signorini M, Cassini R, Drigo M, di Regalbono AF, Pietrobelli M, Montarsi F, Stensgaard AS (2014) Ecological niche model of *Phlebotomus perniciosus*, the main vector of canine leishmaniasis in north-eastern Italy. *Geospat Health.* 9(1): 193–201.
156. Mokhtari M, Miri M, Nikoonahad A, Jalilian A, Naserifar R, Ghaffari HR, Kazembeigi F (2016) Cutaneous leishmaniasis prevalence and morbidity based on environmental factors in Ilam, Iran: Spatial analysis and land use regression models. *Acta Trop.* 163: 90–97.
157. Elnaiem DEA, Schorscher J, Bendall A, Obsomer V, Osman ME, Mekkawi AM, Connor SJ, Ashford RW, Thomson MC (2003) Risk mapping of visceral leishmaniasis: the role of local variation in rainfall and altitude on the presence and incidence of Kala-Azar in eastern Sudan. *Am J Trop Med Hyg.* 68(1): 10–17.
158. Karimi A, Hanafi-Bojd AA, Yaghoobi-Ershadi MR, Akhavan AA, Ghezelbash Z (2014) Spatial and temporal distributions of phlebotomine sand flies (Diptera: Psychodidae), vectors of leishmaniasis, in Iran. *Acta Trop.* 132: 131–139.
159. Hanafi-Bojd AA, Yaghoobi-Ershadi MR, Haghdoost AA, Akhavan AA, Rassi Y, Karimi A, Charrayh Z (2015) Modeling the distribution of cutaneous leishmaniasis vectors (Psychodidae: Phlebotom-

- inae) in Iran: a potential transmission in disease prone areas. *J Med Entomol.* 52(4): 557–565.
160. Gholamrezaei M, Mohebbali M, Hanafi-Bojd AA, Sedaghat MM, Shirzadi MR (2016) Ecological Niche Modeling of main reservoir hosts of zoonotic cutaneous leishmaniasis in Iran. *Acta Trop.* 160: 44–52.
161. Oshaghi M, Ravasan NM, Javadian E, Rassi Y, Sadraei J, Enayati A, Vatan-dooost H, Zare Z, Emami S (2009) Application of predictive degree day model for field development of sandfly vectors of visceral leishmaniasis in northwest of Iran. *J Vector Borne Dis.* 46(4): 247–255.
162. Shirzadi MR, Mollalo A, Yaghoobi-Ershadi MR (2015) Dynamic relations between incidence of zoonotic cutaneous leishmaniasis and climatic factors in Golestan Province, Iran. *J Arthropod Borne Dis.* 9(2): 148–160.
163. Sudhakar S, Srinivas T, Palit A, Kar SK, Battacharya SK (2006) Mapping of risk prone areas of Kala-Azar (Visceral leishmaniasis) in parts of Bihar State, India: an RS and GIS approach. *J Vector Borne Dis.* 43(3): 115–122.
164. Menezes JA, Ferreira EdC, Andrade-Filho JD, Sousa AMd, Morais MHG, Rocha AMS, Machado-Coelho GLL, Lima FP, Madureira AP, Garcia TC (2015) An Integrated Approach Using Spatial Analysis to Study the Risk Factors for Leishmaniasis in Area of Recent Transmission. *Bio Med Res Int.* 2015: 621854.