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Effect of defoliation on leaf physiology of sugar beet cultivars subjected to water stress and re-watering

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Abstract

Water stress causes defoliation, which can reduce yield and root quality of sugar beets (Beta vulgaris L.) through altered gas exchange characteristics of the leaves. In a two-year experiment, three sugar beet cultivars (Europa, Rival and Corsica) were subjected to three defoliation levels (control-C, moderate-MD, severe-SD) and re-watering after their exposition to drought for a month. Leaf physiological traits including net photosynthesis (A), transpiration rate (E), stomatal conductance (g_s) , intracellular CO_2 (C_i) , water use efficiency $(WUE_1 - A/E \text{ and } WUE_i - A/g_s)$, leaf N concentration, petiole NO₃-N concentration, specific leaf area (SLA), leaf water potential (WP) and leaf water content (LWC), were determined before defoliation and 15, 30 and 40 days after defoliation (DAD). On contrary to previous reports, water-stressed cultivars differed significantly in their leaf physiology; the late-season cultivar Corsica had the lowest E and g_s values without any significant reduction in A. Thus, Corsica was the most water-conservative cultivar. Re-watering rapidly restored leaf physiology but a gradual decline, with the progress of DAD, was evident for A, E, g_s and C_i. After re-growth, cultivars differed only in WP and LWC with Europa, the early-harvested cultivar, to have the highest values. Thus, the better response (higher yield increase and lower root quality degradation) of Corsica to re-watering and the subsequent re-growth, as reported by Tsialtas et al. (2009), could be ascribed to its better LAI maintenance compared to Europa and Rival rather than to a better leaf physiological response. The SD plants showed the highest leaf physiological activity since they had the highest A, E, g_s and leaf N concentration. The high leaf activity and the low respiration rate are the means by which sugar beets tolerate foliage losses up to 75% and compensate yield losses after defoliation.

Keywords: Beta vulgaris L.; Drought; Leaf physiology; Re-growth.

Abbreviations

A: net photosynthesis; C: control; C_i : intracellular CO_2 concentration; DAD: days after defoliation; dw: dry weight; E: transpiration rate; g_s : stomatal conductance; LA: leaf area; LAI: leaf area index; LWC: leaf water content; MD: moderate defoliation; SD: severe defoliation; RGR: relative growth rate; SLA: specific leaf area; WP: leaf water potential; WUE_L : A/E; WUE_i : A/g_s .

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Introduction

Productivity of sugar beets grown under semi-arid, Mediterranean conditions is mainly restricted by water availability (Morillo-Velarde and Ober, 2006). However, water scarcity is also a limiting factor in central and northern Europe and its negative effect on sugar beet productivity could be intensified in the near future due to climatic change (Jones et al., 2003).

In Mediterranean basin, rainfall during July and August, when water demands are maximized, is negligible and thus, supplemental irrigation is applied almost to the entire acreage cropped with sugar beets. In central Greece, irrigation need during the growing season is estimated to ~550 mm, a quantity not easily available (Analogides, 1993). Sugar beet, a drought tolerant species (Francois and Maas, 1994), responds to drought stress by losing part of the foliage (defoliation) to minimize its water demand (Vesk and Westoby, 2003). This is the case during the summer in central Greece where sugar beets are grown on clayey soils under water shortage conditions. In fact, the situation is more complicated since under the semi-arid, irrigated conditions, sugar beets face a combination of drought, temperature, light and salinity stresses, which can not easily distinguished (Chaves et al., 2002; Munns, 2002). Erratic rainfalls after mid-August or restored irrigation supply force sugar beets to re-grow. Re-watering after a prolonged drought period affects sugar beet physiology and yield (Owen and Watson, 1956).

Sugar beets are considered to be tolerant to defoliation since foliage losses up to 75% can be compensated and yield is not significantly affected (Afanasiev, 1964; Tsialtas et al., 2009). The selection of a suitable cultivar can also restrict root quality degradation (Tsialtas et al., 2009). It is assumed that the mechanism responsible for this compensation is that the newly expanded leaves after defoliation photosynthesize more and respire less than the older, intact leaves (French and Humphries, 1977; Carter et al., 1978). However, evidence of such mechanism in field conditions is lacking. Recently, several works were conducted to identify physiological traits for the selection of sugar beet genotypes suitable for water stress conditions (Tsialtas and Karadimos, 2003; Ober et al., 2005; Pidgeon et al., 2006). However, we are unaware of any study on sugar beet cultivar response to re-growth under Mediterranean conditions. The aim of this work was to study the leaf physiological response of three sugar beet cultivars subjected to defoliation and re-watering after drought.

Materials and Methods

Three sugar beet cultivars (*Europa*, *Rival*-SESVANDERHAVE NV/SA, Tienen, Belgium and *Corsica*-Maribo Seed International ApS, Holeby, Denmark) were grown on a clay (pH 8.3, total N 1.23 g kg⁻¹, NO₃-N 7.7 mg kg⁻¹, P-Olsen 6.1 mg kg⁻¹, exchangeable K 292 mg kg⁻¹, exchangeable Na 174 mg kg⁻¹) in eastern Thessaly Plain, central Greece (39° 43' N, 22° 28' E, 76 m asl) in 2003 and 2004 growing seasons. *Europa* is an early-harvested cultivar, *Rival* is mid-season and *Corsica* is a late-season one. Figure 1 presents the mean monthly temperature and monthly precipitation pattern during the growing seasons. More details on soil and climatic conditions are given by Tsialtas et al. (2009).

Seeds were drilled (Hege 80, Wintersteiger AG, Ried, Austria) in rows 8 m long, 45 cm apart and 9.1 cm spacing in the row. Each plot was consisted of 12 rows. Winter rainfalls delayed sowing in 2003 (17 April). In 2004, sowing was conducted one month earlier (18 March). Adequate fertilization was provided as basal and top-dressing (150 kg N ha⁻¹ and 90 kg P ha⁻¹). No K fertilization was added due to the high soil K concentration. At the 2-leaf stage, plants were thinned by hand to achieve a population of ~100 000 plants ha⁻¹. Full protection against weeds, insects and fungi (cercospora and powdery mildew) was taken by sprayings.

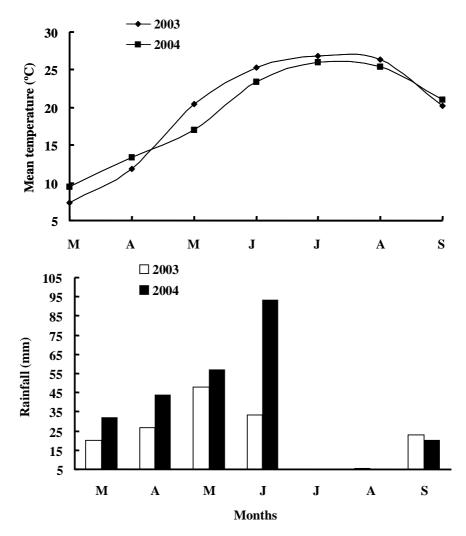


Figure 1. Seasonal patterns of mean monthly temperature and monthly precipitation during growing season.

Supplemental irrigation was given till the beginning of July (a total of 250 mm), and then sugar beets left without irrigation for a month. At early August, leaf removal treatments were applied. The three levels were control (C), moderate (MD) and severe (SD) defoliation. MD plants were defoliated to leave only the newly expanded leaves (estimated $LAI \sim 25\%$ of the C) and SD plants to leave only the meristematic leaves (LAI) almost zero). The experimental design was a split-plot with four replications where cultivars were in the main plots and defoliation levels in the sub-plots. After defoliation, irrigation $(\sim 70 \text{ mm})$ was applied to enhance re-growth.

The determination of leaf physiological parameters was conducted four times; the first on 10^{th} August (before defoliation) only in C plots and the others took place 15, 30 and 40 days after defoliation (DAD). In each plot, gas exchange (net photosynthesis-A, transpiration rate-E, stomatal conductance- g_s , intracellular CO_2 concentration- C_i) were measured in five upper, full-expanded, intact, and full sun-light leaves using LC_i portable photosynthesis system (ADC BioScientific Ltd, Hoddesdon, UK). Instantaneous water-use efficiency (WUE_L) and photosynthetic water-use efficiency (WUE_i) were estimated as A/E and A/g_s ratios, respectively.

Six leaves, similar to those used for gas exchange measurements, were collected per plot, put on an ice-chest and transferred to the Crop Physiology Lab of Larissa factory, Hellenic Sugar Industry SA. Three leaves were used for leaf area (*LA*) determinations using *WinDias* image analysis system (Delta-T Devices Ltd, Cambridge, UK) and then dried in oven at 75 °C for 48 h. Specific Leaf Area (*SLA*) was calculated as the ratio of *LA* to leaf dry weight. Leaf water content (*LWC*) was estimated by comparing fresh and dry leaf weights. Dried leaves were ground to fine powder using a microhammer cutter mill (Glen Creston Ltd, Stanmore, Middlesex, UK). Total leaf *N* concentration was measured on a *Kjeltec 2300* analyzer unit (Foss Tecator, Höganäs, Sweden). The other three leaves from each plot were used for leaf water potential (*WP*) determinations using *WP 4* system (Decagon Devices, Pullman, USA). From each plot, 10-12 petioles of upper, full-expanded leaves were collected and dried at 75 °C for 48 h. Petiole *NO*₃-*N* analysis, according to Johnson and Ulrich (1959), was conducted on an Orion Meter (Model 920A) using an Orion Nitrate electrode 9307 ionplus (Thermo Scientific, Nijkerk, The Netherlands).

Data of the physiological traits determined before re-growth in *C* plots, were analyzed as a Randomized Complete Block design combined over years with cultivars as the main factor. The data after re-growth were subjected to Analysis of Variance (*ANOVA*) and means were compared using Least Significant Difference (*LSD*) test at 0.05 level. The statistical analysis was conducted by *MSTAT-C* (version 1.41, Crop and Soil Sciences Department, Michigan State University).

Results

Physiological traits in C plots before re-growth

Year did not affect leaf gas exchange parameters $(A, E, g_s, C_i, WUE_L, WUE_i)$ but affected significantly leaf N, SLA, WP and LWC (Table 1). Leaf N, SLA and LWC were higher in 2003 compared to 2004 but the adverse was evident for leaf WP (Table 1).

Cultivars did not differ significantly for *A* (*Europa*: 13.04 µmol m⁻² s⁻¹, *Rival*: 12.19 µmol m⁻² s⁻¹, *Corsica*: 11.94 µmol m⁻² s⁻¹). *Corsica* had the lowest *E*, *g*_s and *C*_i and the highest *WUE*_i (Table 1). No significant difference was found for *WUE*_L (*Europa*: 1.92 µmol mmol⁻¹, *Rival*: 1.81 µmol mmol⁻¹, *Corsica*: 2.03 µmol mmol⁻¹). *Corsica* and *Europa* had the highest leaf *N* concentration (40.6 and 39.2 mg g⁻¹ *dw*, respectively) differing significantly from *Rival* (37.3 mg g⁻¹ *dw*). *Europa* and *Corsica* had higher but not significant different petiole *NO*₃-*N* concentration compared to *Rival* (2662, 2056, 1605 mg kg⁻¹ *dw*, respectively). *Europa* had higher *SLA* and *LWC* compared to *Rival* and *Corsica* (Table 1). *Corsica* showed the lowest *SLA* and *LWC* while no significant difference was found for *WP* (*Europa*: -3.79 MPa, *Rival*: -3.81 MPa, *Corsica*: -4.24 MPa).

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able 1. real	as not significant	ance 1. rear and cuttivar effect on the lear physiological traits before defoliation. Means labeled with the same fetter did not differ significantly at r<0.05. The cuttivaryeat teraction was not significant for any trait.	ıysıological tral	its belore deloi	iation. Means iat	Seled with the St	ame letter on	ı not diller sış	gnillcantiy at	r<0.03. 1 ne	cumvar×year
	A		őű				LeafN	NO ₃ -N	SLA	WP	LWC
	µmol m ⁻² s ⁻¹	mmol m ⁻² s ⁻¹	mol m ⁻² s ⁻¹	umol mol-1	umol mmol-1	umol mol-1	mg g-	mg kg-	cm^2 g ⁻¹	MPa	- - - - - -
Year				ı	ı	ı					
2003	12.12a	6.36a	0.20a	210.4a	1.90a	62.6a	42.0a	1694a	196.4a	-4.43b	0.896a
2004	12.62a	6.71a	0.26a	203.8a	1.94a	62.7a	36.0b	2521a	99.2b	-3.47a	0.871b
Cultivar											
Europa	13.04a	7.00a	0.26a	206.9b	1.92a	58.4b	39.2a	2662a	166.4a	-3.79a	0.894a
Rival	12.19a	6.76a	0.23a	215.6a	1.81a	57.4b	37.3b	1605a	139.5b	-3.81a	0.879b
Comercia	11 045	5 954	0.195	100 75	2.035	77 72	40.60	20560	1275h	0101	0.0701

Physiological traits after re-growth

Year and sampling time affected significantly all the traits except of A, which showed marginally insignificant (P=0.06) differences between years (Table 2). With the exception of C_i and WUE_L , defoliation level affected significantly the physiological traits (A, E, g_s , WUE_i , leaf N, petiole NO_3 -N, SLA, WP, LWC). Defoliation levels interacted significantly with years (A, g_s , leaf N, WP) and samplings (E, g_s). Cultivars did not differ significantly as regarded the gas exchange parameters but they showed significant differences for the other physiological traits (leaf N, petiole NO_3 -N, SLA, WP, LWC) (Table 2).

Table 2. ANOVA of the physiological traits determined after re-growth.

Source	df	A	Е	$g_{\rm s}$	$C_{\rm i}$	WUE_{L}	WUE _i	Leaf N	NO ₃ -N	SLA	WP	LWC
Year (Y)	1	P=0.06	***	***	***	***	***	**	***	***	*	***
Defoliation (D)	2	***	***	***	ns	ns	**	***	***	*	***	***
$Y \times D$	2	*	ns	*	ns	ns	ns	*	ns	ns	*	ns
Cultivar (C)	2	P=0.07	ns	ns	ns	ns	ns	***	***	**	***	***
$Y\times C$	2	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns
$D \times C$	4	ns	ns	ns	ns	ns	ns	ns	ns	ns	*	ns
$Y\times D\times C$	4	ns	ns	ns	ns	ns	ns	ns	**	ns	ns	ns
Sampling (S)	2	***	***	***	***	***	***	***	*	***	***	***
$Y\times S$	2	**	***	***	***	***	***	***	ns	***	***	***
$D\times S$	4	ns	*	**	ns	ns	ns	ns	ns	ns	ns	ns
$Y\times D\times S$	4	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns
S×C	4	ns	ns	ns	ns	ns	ns	*	ns	ns	ns	ns
$Y\times C\times S$	4	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns
$D \times C \times S$	8	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns
$Y\times D\times C\times S$	8	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns	ns
CV (%)		20.95	15.20	34.65	13.35	16.62	23.60	9.16	48.59	26.33	16.70	1.76

ns, not significant, *P<0.05, **P<0.01, ***P<0.001.

Defoliation levels increased significantly E and g_s but the differences among the levels were bridged with the progress of DAD (Figure 2). The respective trend for g_s was more pronounced in 2003 and the differences among defoliation levels were gradually eliminated with DAD. SD plants showed the highest A values in both years but significant differences were found only in 2003 (Figure 3). Although marginally insignificant (P=0.07), Europa had the highest A values (15.79 µmol m⁻² s⁻¹) followed by Rival (15.44 µmol m⁻² s⁻¹) and Corsica (14.58 μ mol m⁻² s⁻¹) (Table 3). Undefoliated plants (C) showed the highest WUE_i (73.32 µmol mol⁻¹), SD plants had the lowest values (63.88 µmol mol⁻¹) and MD sugar beets had moderate values (68.32 µmol mol⁻¹). Defoliation levels and cultivars affected petiole NO₃-N, SLA, WP and LWC (Table 3). The SD treatment showed the highest petiole NO₃-N, SLA, WP and LWC. Significant differences between C and MD plants were found only for WP. Europa and Corsica had the highest NO₃-N concentration differing significantly from Rival. Also, Europa showed the highest SLA, WP and LWC while the lowest values for these parameters were found in Corsica. Leaf N was highest in SD plants in 2004 but no significant difference between SD and MD plants was found in 2003 (Figure 4). Corsica had the highest leaf N concentration 15 and 30 DAD. In 40 DAD, Rival and Corsica had similar leaf N concentrations, which were significantly higher than that of Europa (Figure 4).

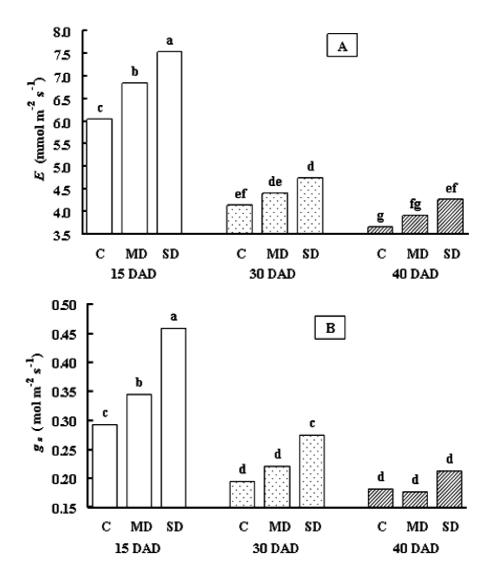


Figure 2. Transpiration rate-E, (A) and stomatal conductance-g, (B) for the defoliation level×sampling interaction. Columns labeled with the same letter did not differ significantly at P<0.05. DAD: days after defoliation; C: control; MD: moderate defoliation; SD: severe defoliation.



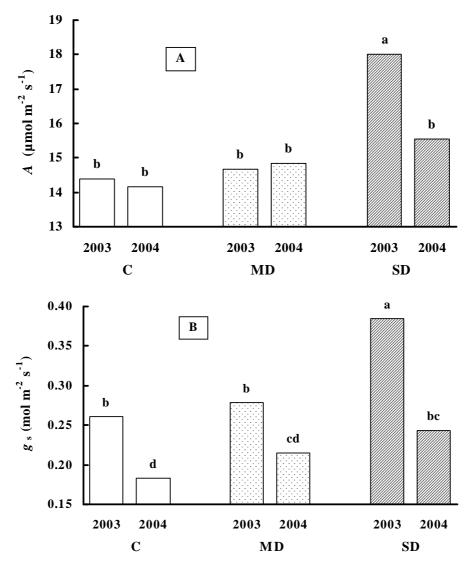


Figure 3. Net photosynthesis-A (A) and stomatal conductance- g_s (B) for the year×defoliation level interaction. Columns labeled with the same letter did not differ significantly at P< 0.05. C: control; MD: moderate defoliation; SD: severe defoliation.

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	A	E	ρδ	Ü	$WUE_{\scriptscriptstyle m L}$	WUE_{i}	LeafN	NO_{3} - N	SLA	WP	LWC
	umol m ⁻² s ⁻¹	mmol m ⁻² s ⁻¹	mol m ⁻² s ⁻¹	umol mol ⁻¹	umol mmol ⁻¹	umol mol ⁻¹	mg g-	mg kg-1	$\mathrm{cm}^2\mathrm{g}^{-1}$	MPa	- 50 50
Defoliation											
C	14.28b	4.60c	0.22b	194.8a	3.33a	73.3a	34.8b	1557b	158.0b	-2.91c	0.894b
MD	14.76b	5.04b	0.25b	195.1a	3.14a	68.3ab	36.4a	1504b	163.1ab	-2.66b	0.899b
SD	16.77a	5.50a	0.31a	197.5a	3.26a	63.9b	37.4a	2028a	176.9a	-2.22a	0.915a
Cultivar											
Europa	15.79a	5.07a	0.27a	196.4a	3.49a	68.1a	34.9b	1835a	177.4a	-2.45a	0.912a
Rival	15.44a	5.14a	0.26a	195.9a	3.54a	68.2a	35.4b	1348b	166.4ab	-2.58b	0.904b
Corsica	14.58a	4.94a	0.25a	195.1a	3.82a	69.2a	38.2a	1907a	154.3b	-2.76c	0.893b



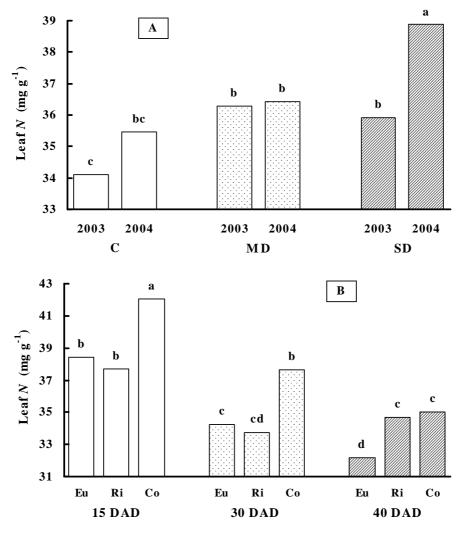


Figure 4. Leaf N concentration for the year×defoliation level (A) and sampling×cultivar (B) interactions. Columns labeled with the same letter did not differ significantly at P<0.05. DAD: days after defoliation; C: control; MD: moderate defoliation; SD: severe defoliation; Eu: Europa; Ri: Rival; Co: Corsica.

Discussion

Under the conditions of central Greece, the water shortages during July and August defoliate sugar beets; the intensity of the phenomenon depends on the severity of the abiotic conditions. However, sugar beet is considered as a well-adapted crop to water deficits in mid-or late season without any significant sugar yield reduction when irrigation is limited

to 70% of the normal (Carter et al., 1980; Winter, 1980). In central Greece, re-watering (irrigation and/or rainfall) from the beginning of August and onwards causes the rapid re-growth of sugar beets with negative effects on yield and quality. In accordance to Afanasiev (1964), Tsialtas et al. (2009) found that sugar beets can be subjected to extended defoliation (up to 75%) without significant negative effects on yield. Also, the selection of a suitable cultivar minimizes the degradation of root quality.

The compensation of yield losses in defoliated sugar beets is ascribed to that newly expanded leaves of re-grown plants are more photosynthetically active and respire less compared to intact sugar beets (French and Humphries, 1977; Carter et al., 1978).

Before defoliation

On contrary to previous reports (Bloch et al., 2006), sugar beet cultivars subjected to water stress before defoliation showed significant differences in their gas exchange physiology. Although no significant genotypic differences in long-term WUE (assessed by carbon isotope discrimination) have been reported previously (Tsialtas and Karadimos, 2003; Rytter, 2005), the late-season cultivar Corsica showed the most conservative water use (highest WUE_L) as a result of its lowest g_s . However, Corsica was not severely stressed since g_s was not lower than 0.10-0.15 mol m⁻² s⁻¹, which is the value indicating severe water stress in C₃ species (Flexas et al., 2004). Although g_s regulates A in sugar beets (Kosobryukov et al., 2004; Monti et al., 2006), cultivars did not differ significantly for this parameter but they showed significant differences in their transpiration rate (E), which affects cultivar water economy (Mudrik et al., 2003). Although Corsica had the highest leaf N concentration, it seems that N was not invested in the photosynthetic machinery but in secondary compounds since it had the lowest SLA (Onoda et al., 2004; Tsialtas et al., 2004). Low SLA is considered as an adaptation of plants to water deficits (Jensen et al., 2000; Nautiyal et al., 2002). Cultivars had LWC close to 90%, an indication of succulence reported for Chenopodiaceae species under drought (Vendramini et al., 2002; Tsialtas et al., 2010). LWC is related with SLA and thus, with cultivar photosynthesis and water economy (Leidi et al., 1999).

After re-growth

Cultivars differed significantly only for WP and LWC; Europa had the highest values. These differences were not translated to differences in gas exchange physiology $(A, E, g_s, C_i, WUE_L, WUE_i)$ among the three cultivars. The better yield and quality performance of Corsica compared to Europa and Rival (Tsialtas et al., 2009) should be ascribed to its better LAI maintenance than to the more photosynthetically active leaves of this cultivar. Based on field observations and growers' experience, Europa is an early-harvested cultivar since it shows high growth rates early in the growing season. This could be, probably, ascribed to its elongated tap-root, which accesses water at deep soil layers and contributes to high WP and LWC values as found here.

Sugar beets show a rapid recovery of their gas exchange physiology after re-watering (Vomáčka and Pospíšilova, 2003) but with the progress of *DAD*, a gradual decline of *A*, *E*,

 g_s , and C_i was found. Adversely, water use efficiency (WUE_L , WUE_i) increased with the progress of DAD. The SD plants showed the highest A, E, g_s and leaf N values and MD plants followed them. However, the differences tended to be bridged with the course of DAD. Thus, the newly expanded leaves of re-grown sugar beets were more physiologically active and thus, increased sucrose concentration in source leaves, as a result of the higher photosynthesis. This has a positive effect on phloem loading and finally on sucrose storage in tap-roots (Winzer et al., 1996). These findings confirmed previous assumptions that higher photosynthetic rate along with lower respiration contributed to the compensation of yield losses in the defoliated sugar beets (French and Humphries, 1977; Carter et al., 1978).

The SD sugar beets showed a suite of traits that were indicative of their higher physiological activity. First of all, the higher photosynthetic rate (A) was related with the higher leaf N concentration (Reich et al., 1998; Tsialtas and Maslaris, 2008), which was confirmed by our findings. Sugar beet re-growth is based on both current N up-take and remobilization of N stored in roots as α -amino N (Pocock et al., 1990). A reliable indication of N nutritional status of sugar beets is the petiole NO_3 -N (Ulrich and Hills, 1990; Oliveira et al., 1993), which was highest in SD plants supporting the highest leaf N concentration and A values found. Also, petiole NO_3 -N in sugar beets is used as an osmoticum since metabolic pool is rapidly saturated (Izmailov, 2004) and thus, NO_3 -N contributed to the highest WP found in SD plants. Confirming previous reports (Marron et al., 2003; Tsialtas et al., 2010), high WP was related with high LWC and SLA. Higher SLA is indicative of thinner leaves, shorter leaf life span and higher RGR (Castro-Díez et al., 2000; Wright and Westoby, 2001; Vendramini et al., 2002).

Conclusions

Before defoliation when cultivars were subjected to water stress, the late-season cultivar *Corsica* showed the most conservative water use (highest WUE_i) due to its lowest E and g_s . After the re-watering and the subsequent re-growth, cultivars differed only for WP and LWC with the early-harvested cultivar Europa to have the highest values. Thus, the good yield and quality performance of Corsica after re-growth, as reported by Tsialtas et al. (2009), should be ascribed to its better LAI maintenance than to its leaf physiological superiority compared to Europa and Rival. The SD plants were the most physiologically active showing the highest A, E, g_s and leaf N concentration. This finding accompanied by the assumption that the newly expanded leaves respire less could be the mechanism by which defoliated sugar beets compensate for yield and quality losses (Tsialtas et al., 2009).

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References

- Afanasiev, M.M., 1964. The effect of simulated hail injuries on yield and sugar content of beets. J. Am. Soc. Sugar Beet Technol. 13, 225-237.
- Analogides, D.A., 1993. Estimating sugar beet irrigation requirements on the basis of climatic parameters in Greece: results and conclusions from a 6-year experimental study (1980-1985). P. IIRB Congr. 56, 259-269.
- Bloch, D., Hoffmann, C.M., Märländer, B., 2006. Impact of water supply on growth, photosynthesis, water use and carbon isotope discrimination of sugar beet in relation to genotypic variability. Eur. J. Agron. 24, 218-225.
- Carter, J.N., Traveller, D.J., Bosma, S.M., 1978. Sugarbeet yield and seasonal growth characteristics as affected by hail damage and nitrogen level. J. Am. Soc. Sugar Beet Technol. 20, 73-83.
- Carter, J.N., Jensen, M.E., Traveller, D.J., 1980. Effect of mid- to late-season water stress on sugarbeet growth and yield. Agron. J. 72, 806-815.
- Castro-Díez, P., Puyravaud, J.P., Cornelissen, J.H.C., 2000. Leaf structure and anatomy as related to leaf mass per area variation in seedlings of a wide range of woody plant species and types. Oecologia 124, 476-486.
- Chaves, M.M., Pereira, J.S., Maroco, J., Rodrigues, M.L., Ricardo, C.P.P., Osório, M.L., Carvalho, I., Faria, T., Pinheiro, C., 2002. How plants cope with water stress in field: photosynthesis and growth. Ann. Bot. 89, 907-916.
- Flexas, J., Bota, J., Cifre, J., Escalona, J.M., Galmés, J., Gulías, J., Lefi, E.K., Martínez-Cañellas, S.F., Moreno, M.T., Ribas-Carbó, M., Riera, D., Sampol, B., Medrano, H., 2004. Understanding down-regulation of photosynthesis under water stress: future prospects and searching for physiological toots for irrigation management. Ann. Appl. Biol. 144, 273-283.
- Francois, L.E., Maas, E.V., 1994. Crop response and management on salt-affected soils. In: Pessarakli, M. (Ed.), Handbook of Plant and Crop Stress. Marcel Dekker Inc., New York, pp. 149-181.
- French, S.A.W., Humphries, E.C., 1977. The effect of partial defoliation on yield of sugar beet. Ann. Appl. Biol. 87, 201-212.
- Izmailov, S.F., 2004. Saturation and utilization of nitrate pools in pea and sugar beet leaves. Russ. J. Plant Physiol. 51, 189-193.
- Jensen, C.R., Jacobsen, S.E., Andersen, M.N., Núñez, N., Andersen, S.D., Rasmussen, L., Mogensen, V.O., 2000. Leaf gas exchange and water relation characteristics of field quinoa (*Chenopodium quinoa* Willd.) during soil drying. Eur. J. Agron. 13, 11-25.
- Johnson, C.M., Ulrich, A., 1959. Analytical Methods for Use in Plant Analysis. California Agricultural Experimental Station Bulletin, No. 767.
- Jones, P.D., Lister, D.H., Jaggard, K.W., Pidgeon, J.D., 2003. Future climate impact on the productivity of sugar beet (*Beta vulgaris* L.) in Europe. Climatic Change 58, 93-108.
- Kosobryukhov, A.A., Bil', K.Y., Nishio, J.N., 2004. Sugar beet photosynthesis under conditions of increasing water deficiency in soil and protective effects of a low-molecular-weight alcohol. Appl. Biochem. Microbiol. 40, 581-587
- Leidi, E.O., López, M., Gorham, J., Gutiérrez, J.C., 1999. Variation in carbon isotope discrimination and other traits related to drought tolerance in upland cotton cultivars under dryland conditions. Field Crops Res. 61, 109-123.
- Marron, N., Dreyer, E., Boudouresque, E., Delay, D., Petit, J.M., Delmotte, F.M., Brignolas, F., 2003. Impact of successive drought and re-watering cycles on growth and specific leaf area of two *Populus×canadensis* (Moench) clones, 'Dorskamp' and 'Luisa_Avanzo'. Tree Physiol. 23, 1225-1235.
- Monti, A., Brugnoli, E., Scartazza, A., Amaducci, M.T., 2006. The effect of transient and continuous drought on yield, photosynthesis and carbon isotope discrimination in sugar beet (*Beta vulgaris L.*). J. Exp. Bot. 57, 1253-1262.
- Morillo-Velarde, R., Ober, E.S., 2006. Water use and irrigation. In: Draycott, P.A. (Ed.), Sugar Beet. Blackwell Publishing Ltd, Oxford, pp. 221-255.
- Mudrik, V., Kosobrukhov, A., Knyazeva, I., Pigulevskaya, T., 2003. Changes in the photosynthetic characteristics of *Plantago major* plants caused by soil drought stress. Plant Growth Regul. 40, 1-6.
- Munns, R., 2002. Comparative physiology of salt and water stress. Plant Cell Environ. 25, 239-250.
- Nautiyal, P.C., Rachaputi, N.R., Joshi, Y.C., 2002. Moisture-deficit-induced changes in leaf-water content, leaf carbon exchange rate and biomass production in groundnut cultivars differing in specific leaf area. Field Crops Res. 74, 67-79.
- Ober, E.S., Le Bloa, M., Clark, C.J.A., Royal, A., Jaggard, K.W., Pidgeon, J.D., 2005. Evaluation of physiological traits as indirect selection criteria for drought tolerance in sugar beet. Field Crops Res. 91, 231-249.

- Oliveira, M.D., Carranca, C.F., Oliveira, M.M., Gusmão, M.R., 1993. Diagnosing nutritional status of sugarbeet by soil and petiole analysis. In: Fragoso, M.A.C., Van Beusichem, M.L. (Ed.), Optimization of Plant Nutrition. Kluwer Academic Publishers, The Netherlands, pp. 147-151.
- Onoda, Y., Hikosaka, K., Hirose, T., 2004. Allocation of nitrogen to cell walls decreases photosynthetic nitrogenuse efficiency. Funct. Ecol. 18, 419-425.
- Owen, P.C., Watson, D.J., 1956. Effect on crop growth of rain after prolonged drought. Nature 177, 847.
- Pidgeon, J.D., Ober, E.S., Qi, A., Clark, C.J.A., Royal, A., Jaggard, K.W., 2006. Using multi-environment sugar beet variety trials to screen for drought tolerance. Field Crops Res. 95, 268-279.
- Pocock, T.O., Milford, G.F.J., Armstrong, M.J., 1990. Storage root quality in sugarbeet in relation to nitrogen uptake. J. Agr. Sci. 115, 355-362.
- Reich, P.B., Ellsworth, D.S., Walters, M.B., 1998. Leaf structure (specific leaf area) modulates photosynthesisnitrogen relations: evidence from within and across species and functional groups. Funct. Ecol. 12, 948-958.
- Rytter, R.M., 2005. Water use efficiency, carbon isotope discrimination and biomass production of two sugar beet varieties under well-watered and dry conditions. J. Agron. Crop Sci. 191, 426-438.
- Tsialtas, J.T., Karadimos, D.A., 2003. Leaf carbon isotope discrimination and its relation with qualitative root traits and harvest index in sugar beet (*Beta vulgaris* L.). J. Agron. Crop Sci. 189, 286-290.
- Tsialtas, J.T., Maslaris, N., 2008. Seasonal trends and relationships of light, temperature and leaf physiological traits of sugar beets (*Beta vulgaris* L.) grown under semi-arid, Mediterranean conditions. Int. J. Plant Prod. 2, 223-242.
- Tsialtas, J.T., Pritsa, T.S., Veresoglou, D.S., 2004. Leaf physiological traits and their importance for species success in a Mediterranean grassland. Photosynthetica 42, 371-376.
- Tsialtas, J.T., Soulioti, E., Maslaris, N., Papakosta, D., 2009. Genotypic response to re-growth of defoliated sugar beets after re-watering in a water-limited environment: effects on yield and quality. Int. J. Plant Prod. 3, 1-18.
- Tsialtas, J.T., Matsi, T., Maslaris, N., 2010. Plasticity of leaf anatomy, chemistry and water economy of irrigated sugar beets grown under Mediterranean conditions. Int. J. Plant Prod. 4, 99-114.
- Ulrich, A., Hills, F.J., 1990. Plant analysis as an aid in fertilizing sugarbeet. In: Westerman, R.L. (Ed.), Soil Testing and Plant Analysis, 3rd edition. Soil Science Society of America, Madison, pp. 429-447.
- Vendramini, F., Díaz, S., Gurvich, D.E., Wilson, P.J., Thompson, K., Hodgson, J.G., 2002. Leaf traits as indicators of resource-use strategy in floras with succulent species. New Phytol. 154, 147-157.
- Vesk, P.A., Westoby, M., 2003. Drought damage and recovery-a conceptual model. New Phytol. 160, 7-14.
- Vomáčka, L., Pospíšilová, J., 2003. Rehydration of sugar beet plants after water stress: effect of cytokinins. Biol. Plantarum 46, 57-62.
- Winter, S.R., 1980. Suitability of sugarbeets for limited irrigation in a semi-arid climate. Agron. J. 72, 118-123.
- Winzer, T., Lohaus, G., Heldt, H.W., 1996. Influence of phloem transport, N-fertilization and ion accumulation on sucrose storage in the taproots of fodder beet and sugar beet. J. Exp. Bot. 47, 863-870.
- Wright, I.J., Westoby, M., 2001. Understanding seedling growth relationships through specific leaf area and leaf nitrogen concentration: generalizations across growth forms and growth irradiance. Oecologia 127, 21-29.