Journal Homepage: vrf.iranjournals.ir

Optimization of incubation temperature in embryonated chicken eggs inoculated with H9N2 vaccinal subtype of avian influenza virus

Iraj Khalili*, Rahim Ghadimipour, Ali Ameghi, Saeed Sedigh-Eteghad

Department of Research and Development, Razi Vaccine and Serum Research Institute, Marand, Iran.

Article Info	Abstract
Article history:	There are little information about growth properties of low pathogenic (LP) avian influenza
Ai ticle history.	virus (AIV) in embryonated chicken eggs (ECEs) at different incubation temperatures.
Received: 18 August 2012	Knowledge of this information increases the quantity and quality of antigen in vaccine
Accepted: 01 December 2012	production process. For this purpose, 10-5 dilution of AIV (A/Chicken/Iran/99/H9N2) was
Available online: 15 September 2013	inoculated (Intra-allantoic) into 400, 11-day old specific pathogen free (SPF) ECEs in the 0.1 mL
	per ECE rate and incubated in 32, 33, 34, 35, 36, 37.5, 38, 39 °C for 72 hr in 65% humidity. Early
Key words:	death embryos in first 24 hr were removed. Amnio-allantoic fluid was withdrawn into the
	measuring cylinder, and tested for hemagglutination (HA) activity and egg infective dose 50
Inactivated influenza vaccine	(EID ₅₀). The utilizable ECEs and amnio-allantoic fluid volume was significantly increased in
Incubation temperature	35 °C, ($p < 0.05$). Significant difference in HA and EID ₅₀ titers, were seen only in 39 °C group.
Optimization	Therefore, 35°C is an optimum temperature for incubation of inoculated ECEs.
1	
	© 2013 Urmia University. All rights reserved.

بهینه سازی دمای انکوباسیون تخم مرغ های جنین دار تلقیح شده با ویروس آنفلوانزای طیور سویه واکسینال (H9N2)

چکیدہ

اطلاعات محدودی در ارتباط با روند رشد ویروس های آنفلوانزای پرندگان با پاتوژنسیته پایین در تخم مرغ های چنین دار و در دماهای انکوباسیون مختلف وجود دارد. اطلاعات در این زمینه می تواند باعث افزایش کمیت و کیفیت در روند تولید واکسن این نوع ویروس گردد. بدین منظور، رقت ^۵-۱۰ از ویروس آنفلوانزای پرندگان (A/Chicken/Iran/99/H9N2) در ۴۰۰ عدد تخم مرغ جنین دار عاری از بیماری خاص ۱۱ روزه، به مقدار ۰/۱ میلی لیتر تلقیح گردید. تخم مرغ های تلقیح شده به تعداد مساوی در دماهای انکوباسیون ۳۳ ۳۴، ۳۵ ۲۵ ۲۸ و ۳۹ درجه سانتیگراد به مدت ۲۷ ساعت و رطوبت ۶۵ درصد قرار گرفتند. تخم مرغ های تلقیح شده به تعداد مساوی در دماهای انکوباسیون ۳۳ ۳۴، ۳۵ ۲۵ ۲۸ ۲۰ ۲۵ و ۳۹ درجه سانتیگراد به مدت ۷۲ ساعت و رطوبت ۶۵ درصد قرار گرفتند. تخم مرغ های تلف شده در ۲۴ ساعت اول حذف گردیدند. سپس مایع آمنیوآلانتوئیک تخم مرغ ها استخراج شده و پس از اندازه گیری حجم، تست های HA و موات ۶۵ درصد قرار گرفتند. تخم مرغ های قابل برداشت و مقدار مایع برداشتی در دمای ۳۵ ۳۵ ۲۵ داری نسبت به دیگر گروه های تحت مطالعه تست های HA و موات 90 دروی آنها انجام گردید. تحم مرغ های قابل برداشت و مقدار مایع برداشتی در دمای ۳۵ درجه سانتیگراد به میزان معنی داری نسبت به دیگر گروه های تحت مطالعه افزایش نشان داد (۰/۵). میانگین تست هماگلوتینین و و EID50 به غیر از دمای ۳۹ درجه سانتیگراد در دیگر گروه ها تغییر معنی داری نداشت. بنابراین بر خلاف تصورات قبلی، ۳۵ درجه سانتیگراد در می داری نداشت. بنابراین بر خلاف تصورات قبلی، ۳۵ درجه سانتیگراد می تواند دمای مناسی برای انکوباسیون تخم مرغ های تلقیح از دمای ۳۵ در جه سانتیگراد در دیگر گروه ها تغییر معنی داری نداشت بر خلاف تصورات قبلی، ۳۵ درجه سانتیگراد

واژه های کلیدی: بهینه سازی، دمای انکوباسیون، واکسن غیر فعال آنفلوانزا

*Correspondence:

Iraj Khalili. DVM, PhD

Department of Research and Development, Razi Vaccine and Serum Research Institute, Marand, Iran. **E-mail:** Iraj_DV4953@yahoo.com

Introduction

Among the avian influenza virus (AIV) subtypes, H9N2 virus has the potential to cause an influenza pandemic because of its wide prevalence in avian species and the ability to infect humans.¹ An outbreak of H9N2 infection in poultry farms was first reported in 1998 in a layer farm in Tehran-Iran.^{2,3} At the same time the similar respiratory disease outbreaks were observed in broiler and breeder flocks located in Tehran and neighbor provinces.²

This disease is now endemic and vaccination against this subtype is practiced, routinely.⁴ This virus cause mild disease among terrestrial birds. However, it can also cause severe outbreaks in poultry depending on the circumstances.⁵

Nowadays, two types of influenza vaccines are approved for clinical use, and both of them are manufactured in embryonated chicken eggs (ECEs): live attenuated influenza vaccines and inactivated influenza vaccines.^{6,7}

Inactivated influenza vaccine subtype H9N2 production was started in Iran in 1998 and nowadays this product is one of best inactived vaccines from quality and quantity point view.⁸

The ECE methods are widely used in inactivated H9N2 whole-virus vaccines manufacturing⁹ and this kinds of vaccines were shown to bear better immunogenic responses than other vaccine types.⁶

For the production of vaccines the growth characteristics of the vaccine strains and the yields of the viral antigens (quantity and quality) are important issues.¹⁰ The incubation temperature of inoculated ECEs is closely associated with quantity and quality of antigen in vaccine production process. Therefore, the number of utilizable ECEs and total volume of yielded antigen are very important factor from economical point view.

The effects of different incubation temperatures on some subtypes of AIV replication has been well studied in various infection systems, including cultured cells and animal models. But there are little information about the growth properties of low pathogenic (LP) H9N2 AIVs in ECEs at different incubation temperatures.¹¹

Therefore, optimization of ECEs incubation temperatures, in H9N2 vaccine production process seems necessary for reduction of extra production costs and increase amount of vaccine output.

Material and Methods

Virus preparations. Standard vaccine strain AIV (A/Chicken/Iran/99/H9N2) was used to inoculate 11-day old specific pathogen free ECEs. The eggs were observed for 24-72 hr post inoculation. Allantoic fluid of the inoculated eggs were collected and centrifuged at 1200 rpm for 30 min and supernatant were assayed.

Egg infective dose 50 (EID $_{50}$) of the samples was calculated by the method of Reed and Muench in SPF eggs with 11-day-old embryos.¹²

Hemagglutination assay (HA) was performed in Vbottom 96-well plates with 1% chicken red blood cell, as described with Burleson *et al.*¹³

Study design. Prepared 10^{-5} diluted working seed of AIV (A/Chicken/Iran/99/H9N2) solution (EID₅₀ = 9.8 log₁₀ and HA titer = 10 log₂) was inoculated into 400 SPF ECEs at the rate of 0.1 mL per ECE, via intra-allantoic way. All eggs were sealed with wax. Inoculated eggs were randomly divided into 8 groups (n = 50) and incubated in various temperatures. The experimental temperatures were 32, 33, 34, 35, 36, 37.5 (standard temperature), 38 and 39 °C. Inoculated ECEs were incubated for 72 hr with 65% humidity. Eggs were candled daily and early death embryos in first 24 hr were removed.

In the next step, the eggs were chilled at 4 °C for 24 hr to kill the embryo and to reduce the contamination of the allantoic fluid with blood during harvesting.

In harvesting stage, after removing the shell and shell membranes at the blunt end of the eggs, the amnioallantoic fluid samples were withdrawn into the small measuring cylinders by pressing the suction bulb until the total volume of each egg allantoic liquid was obtained. Allantoic fluid from surviving embryos was tested for HA activity and EID₅₀. Assays were performed in three replicates.

Statistical analysis. Data were analyzed using SPSS (version 17 for Windows, SPSS Inc., Chicago, IL, USA), and comparisons were made using the descriptive statistics and one way ANOVA tests.

Results

The utilizable ECEs and amnio-allantoic fluid volume was noticeably influenced by some temperatures. Amount of utilizable ECEs in 33 and 35 °C incubation temperature groups were significantly higher than standard incubation temperature (37.5 °C) and the greatest total antigen volume of 609.00 \pm 70.37 mL occurred under 35 °C incubation temperature, that was greater than other experimental groups (*p* < 0.05).

Mean of HA titers and EID_{50} tests in the study groups are shown in Table 1. No significant differences were seen for mentioned tests between groups. Only in 39 °C incubation temperature, HA titer and EID_{50} test results were significantly decreased.

Discussion

All avian eggs necessarily lose water because of porous eggshell.¹⁴ About 10-11% of the water in an embryo is lost during the incubation period.¹⁵ Some studies show that, the change in volume and composition of amnio-allantoic fluids are closely associated with water metabolism of avian egg.¹⁶ Reportedly, the phase change from liquid water to water vapor requires heat and every extra calorie/hr increase the amount of evaporation and water lost.¹⁷

Incubation temperature (°C)	Early death embryos (no.)	Utilizable eggs (no.)	Total antigen volume (mL)	Antigen volume (mL) / Utilizable eggs	Antigen volume (mL) / All eggs	HA Log ₂	EID50 Log ₁₀
32	7.33 ± 2.51	42.66 ± 2.51	470.00 ± 67.26	10.98 ± 0.97	9.40 ± 1.34	9.02 ± 0.21	9.46 ± 0.06
33	6.33 ± 2.08*	43.66 ± 2.08*	536.66 ± 31.75*	12.29 ± 0.82	10.73 ± 0.63*	8.77 ± 0.19	9.66 ± 0.16
34	7.33 ± 2.08	42.66 ± 2.08	560.00 ± 28.61*	13.12 ± 0.13	$11.18 \pm 0.58^*$	9.22 ± 0.25	9.54 ± 0.52
35	4.66 ± 2.08*	45.33 ± 2.08*	609.00 ± 70.37*	13.45 ± 1.69	$12.18 \pm 1.40^*$	9.44 ± 0.95	9.55 ± 0.09
36	11.33 ± 3.21	38.66 ± 3.21	480.00 ± 61.44	12.45 ± 1.64	9.60 ± 1.22	9.30 ± 0.67	9.49 ± 0.18
37.5	13.33 ± 1.15	36.66 ± 1.15	385.66 ± 65.54	10.50 ± 1.66	7.71 ± 1.31	9.13 ± 0.55	9.06 ± 0.35
38	17.00 ± 2.00	33.00 ± 2.00	299.66 ± 57.70	9.02 ± 1.19	5.99 ± 1.15	8.91 ± 1.04	8.25 ± 0.64
39	$22.00 \pm 2.64^*$	$28.00 \pm 2.64^*$	248.66 ± 5.13	8.91 ± 0.65	4.97 ± 0.10	5.50 ± 0.25*	7.13 ± 0.27*

Table 1. Interaction of incubation temperature with antigen volume and quality factors (Three replicates mean values ± SD).

* indicates significant differences compared to 37.5 °C (Standard temperature) at p < 0.05.

Romanoff and Hayward showed that extreme temperatures (39.5 and 34.5 °C) significantly reduced the amnio-allantoic fluids¹⁶ but in the present study, the best result in antigen volume (mL) per utilizable ECEs in vaccine manufacturing process was achieved in 35 °C incubation temperature.

On the other hand, the mortality of inoculated ECEs 24 hr post-inoculation is generally considered non-specific¹⁸ and according to the Office International des Epizooties (OIE) manual should be discarded.¹⁹ Therefore, increasing of first 24 hr mortality can affect the quality and economic factors in vaccine industry.

Our results showed that temperature variation was the effective factor in this rate; utilizable eggs in the 35 °C incubation temperature were significantly higher than standard incubation temperature.

The rapid replication of vaccine virus with highly active HA and neuraminidase may be a burden to embryos, and may cause early embryonic mortality. High embryonic virulence may result in a decrease of the amount of allantoic fluid and an increase in the number of discarded ECEs.²⁰ Lang *et al.* showed that avian influenza virus can be isolated from ECE within 48 hr by incubation at 37 °C or 39 °C, whereas 72 hr is required if ECE are incubated at 35 °C.¹¹ Also, Hahon *et al.* study results showed that an incubation temperature of 35 °C is favored for the multi-plication of virus on the chorio-allantoic membrane of ECEs, while higher temperatures are found to be less optimal.²¹

In the present paper the HA titer and EID_{50} test were evaluated in different incubation temperatures for prepared AIV and there were not any significant differences between treatment groups (except 39 °C group). Also in Lang *et al.* study there are no statistically significant differences in the mean HA titers and EID_{50} of low pathogen avian influenza viruses (LPAIV) at different incubation temperatures.¹¹ However, in the Blumenthal *et al.* work, the HA titers are the highest and best sustained in ECEs incubated at 34 °C,²² that may be related with the virus strain.

Therefore, 35 °C is an optimum temperature for incubation of inoculated ECEs, in inactivated AI (A/Chicken/Iran/99/H9N2) vaccine production process and this optimization help reduce extra production costs and increase amount of vaccine output.

Acknowledgments

This study was financially supported by the Razi Vaccine and Serum Research Institute and Education and Research Deputy of Jihad-e-Agriculture Ministry with the grant No. 2-81-18-89039.

References

- Fedorko DP, Nelson NA, McAuliffe JM, et al. Performance of rapid tests for detection of avian influenza a virus types H5N1 and H9N2. J Clin Microbiol 2006; 44(4): 1596-1597.
- Vasfi-Marandi M, BozorgmehriFard MH. Isolation of H9N2 subtype of avian Influenza viruses during an outbreak in chickens in Iran. Iran Biomed J 2002;6(1):13-17.
- 3. Alizadeh E, Hosseini SM, Kheiri MT, et al. Avian Influenza (H9N2) among poultry workers in Iran. Iran J Microbiol 2009; 1: 3-6.
- 4. Vasfi-Marandi M, Bozorgmehri-Fard MH, Hashemzadeh M. Efficacy of inactivated H9N2 avian influenza vaccine against non-highly pathogenic A/Chicken/Iran/ZMT-173/1999 infection. Arch Razi Ins 2002; 53: 23-26.
- 5. Park KJ, Kwon HI, Song MS, et al. Rapid evolution of low-pathogenic H9N2 avian influenza viruses following poultry vaccination programmes. J Gen Virol 2011; 92: 36-50.
- 6. Singh N, Pandey A, Mittal SK. Avian influenza pandemic preparedness: developing prepandemic and pandemic vaccines against a moving target. Exp Rev Mol Med 2011: 1-29.
- 7. Takada A, Matsushita S, Ninomiya A, et al. Intranasal immunization with formalin-inactivated virus vaccine induces a broad spectrum of heterosubtypic immunity against influenza A virus infection in mice. Vaccine 2003; 21: 3212-3218.
- 8. Moghadam-Pour M, Momayez R, Akhavizadegan MA. The efficacy of inactivated oil emulsion H9N2 avian influenza vaccine. Iran J Vet Res 2006; 15: 85-87.
- Hickling J, D'Hondt E. A review of production technologies for influenza virus vaccines, and their suitability for deployment in developing countries for influenza pandemic preparedness. Available at: http://www.who. int/ vaccine_search/diseases/influenza/Flu_vacc_manuf_

tech_report.pdf. Accessed: May 21, 2012.

- 10. Rimmelzwaan GF, Osterhaus AD. Influenza vaccines: New developments.Curr Opin Pharmacol 2001; 1:491-496.
- 11. Lang V, Marjuki H, Krauss SL, et al. Different incubation temperatures affect viral polymerase activity and yields of low-pathogenic avian influenza viruses in embryonated chicken eggs. Arch Virol 2011; 156: 987-994.
- 12. Reed LJ, Muench H. A simple method of estimating fifty percent endpoints. Am J Hyg 1938; 27: 493-497.
- Burleson FG, Chambers TM, Wiedbrauk DL. Virology: A laboratory manual. London, UK: Academic press 1992; 132-136.
- 14. Pearson JT, Seymour RS, Baudinette RV, et al. Respiration and energetics of embryonic development in a large altricial bird, the Australian pelican (*Pelecanus conspicillatus*). J Exp Biol 2002; 205: 2925-2933.
- 15. Mao KM, Murakami A, Iwasawa A, et al. The asymmetry of avian egg-shape: An adaptation for reproduction on dry land. J Anatomy 2007; 210: 741-748.
- 16. Romanoff AL, Hayward FW. Changes in volume and physical properties of allantoic and amniotic fluids under

normal and extreme temperatures. Bio Bull 1943; 84: 141-147.

- 17. French NA. Modeling incubation temperature: The effects of incubator design, embryonic development, and egg size. Poult Sci 1997; 76: 124-133.
- Terregino C, Capua I. Avian influenza and Newcastle disease: A field and laboratory manual. Milan, Italy: Springer; 2009; 312-320.
- 19. OIE. Manual of diagnostic tests and vaccines for terrestrial animals. Avian influenza. 5th ed. Paris, France; Office international des epizooties 2004; 7-8.
- 20. Kwon HJ, Cho SH, Ahn YJ, et al. Characterization of a chicken embryo-adapted H9N2 subtype avian influenza virus. Vet Sci J 2009; 3: 9-16.
- 21. Hahon N, Ratner M, Kozikowski E. Factors influencing variola virus growth on the chorioallantoic membrane of emberyonated eggs. J Bacteriol 1958 75: 707-712.
- 22. Blumenthal HT, Greiff D, Pinkerton H, et al. The hemagglutination and infectivity titre curves of PR8 Influenza virus cultivated in embryonated eggs at different temperatures. J Exp Med 1950; 91: 321-329.