



Relative Frequency of Norovirus Infection in Children Suffering From Gastroenteritis and Referred to Aboozar Hospital, Ahvaz, Iran

Shahram Jalilian¹, Ali Reza Samarbaf-Zadeh^{1*}, Seyed Hamid Reza Mozhgani¹, Manoochehr Makvandi¹, Mehdi Parsa-nahad¹, Roya Pirmoradi¹, Ahmad Shamsi-Zadeh²

¹ Department of Medical Microbiology, School of Medicine and Infectious and Tropical Disease Research Center, Ahvaz Jundishapur University of Medical Science, Ahvaz, IR Iran

² Departments of Pediatrics, Ahvaz Aboozar Hospital, Ahvaz, IR Iran

ARTICLE INFO

Article type:
Original Article

Article history:
Received: 01 Sep 2010
Revised: 10 Dec 2010
Accepted: 01 Jun 2011

Keywords:
Human Norovirus
Gastroenteritis
Nested-RT-PCR

ABSTRACT

Background: Noroviruses belong to the *Norovirus* genus in the *Caliciviridae* family. Noroviruses are the most common causes of gastroenteritis and have a great impact on public health. They have been identified as a common cause of acute gastroenteritis in children. **Objectives:** The aim of this study was to determine the prevalence of Norovirus in children suffering from gastroenteritis.

Patients and Methods: Fecal samples (n = 143) were collected from children under 5 years of age who were suffering from gastroenteritis. All the children were referred to Ahvaz Aboozar Hospital, located in southwestern Iran. Norovirus RNA was extracted by Trizol, and RNA was detected using nested reverse transcriptase polymerase chain reaction (nested-RT-PCR).

Results: Norovirus infection was detected in 9 of the 143 collected samples (6.3%). All positive samples belonged to genogroup II. Five positive samples were obtained from male patients and 4 were obtained from female patients. Most of the positive cases were from patients between 3 and 5 years of age (n = 5, 56%). There was no relationship between gender and virus prevalence. The rate of infection peaked in winter (n = 6, 66.9%), and we did not detect any positive cases in summer.

Conclusions: The prevalence of this virus in Ahvaz is similar to that reported by other researchers. Because this virus is transmitted by contaminated food or water, we recommend adult education and improved personal hygiene to reduce the incidence of Norovirus infection in children. This study improves our epidemiological knowledge of the prevalence of this virus in Ahvaz and Iran.

©2012, AJUMS. Published by Kowsar M.P.Co. All rights reserved.

► Implication for health policy/practice/research/medical education:

This study improves our knowledge about the significance of noroviral gastroenteritis in clinics.

► Please cite this paper as:

Jalilian S, Samarbaf-Zadeh AR, Mozhgani SHR, Makvandi M, Parsa-nahad M, Pirmoradi R, et al. Relative Frequency of Norovirus Infection in Children Suffering From Gastroenteritis and Referred to Aboozar Hospital, Ahvaz, Iran. *Jundishapur J Microbiol.* 2012;5(1): 355-8. DOI: 10.5812/kowsar.20083645.2370

* Corresponding author: Ali Reza Samarbaf-Zadeh, Department of Medical Microbiology, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, IR Iran. Tel: +98-6113354389, Fax: +98-6113361544. E-mail: Alirezasarbaf_78@hotmail.com

1. Background

Noroviruses belong to the *Norovirus* genus in the *Caliciviridae* family (1). The virion contains a single positive-strand RNA genome with icosahedral symmetry. Based on the genetic sequence of viral RNA-dependent RNA polymerase and the capsid proteins, noroviruses are divided into 5 genogroups GI-GV (2). Traditionally, noroviruses were detected using electron microscopy. The

limitation of this method was the low load of viruses, because EM visualization requires 10^6 viral particles per gram of fecal sample (3).

Noroviruses are the most common causes of gastroenteritis, and they have a great impact on public health (4). They have been identified as a common cause of acute gastroenteritis in children (5). Patients infected with Norwalk-like viruses (NV) present with the following signs and symptoms: nausea (79%), vomiting (69%), diarrhea (66%), abdominal cramps (30%), headache (22%), fever (subjective; 37%), chills (32%), myalgia (26%), and sore throat (18%). Bloody stools have not been reported until recently (6). Attempts to develop an inclusive serological test for detection of all noroviruses have not been successful because of the antigenic diversity of these viruses (7).

2. Objectives

Since the total genome of noroviruses has been sequenced, detection of these viruses can be performed by sensitive molecular methods such as nested RT-PCR. This method is between 10 and 1000 times more sensitive than conventional RT-PCR (8). Therefore, we used nested RT-PCR to detect noroviruses in fecal specimens collected from children less than 5 years of age who were referred to Ahvaz Aboozar Hospital in Iran. This information will improve our epidemiological knowledge about the prevalence of this virus in Ahvaz and Iran.

3. Patients and Methods

Fecal specimens were collected from children under 5 years of age who had gastroenteritis and were referred to Aboozar Hospital in Ahvaz between 2008 and 2009. Samples from 143 patients were collected and stored at -80°C . Samples from patients with bacterial infections or parasite infestations were excluded from the study. We tested all the samples for noroviral infection by using nested RT-PCR.

3.1. RT-PCR

The RNA of viruses was extracted by Trizol (Roche Germany). The extracted RNA was denatured by incubation at 60°C for 5 min and then incubated on ice for 2 min. We used a Fermentas RT-PCR kit (Lithuania) to construct cDNA from 9 μl of extracted RNA, according to the manufacturer's instructions. Five microliters of the cDNA was used as a template for PCR with the following primers: NV 35 (5'-CTTGTTGGTTGAGGCCATAT-3') and NV 36 (5'-ATAAAGTTGGCATGAACA-3') (9). These primers amplify 470 bp from open reading frame (ORF) 1a that encodes RNA polymerase. We used the following PCR conditions: 35 cycles of 1 min at 94°C , 1 min at 49°C , and 1 min at 72°C , followed by a final extension of 6 min at 72°C . The products of the first round of PCR were subjected to electrophoresis in 3% agarose gel, stained by ethidium bromide, and visualized under a UV Transilluminator (Vilber

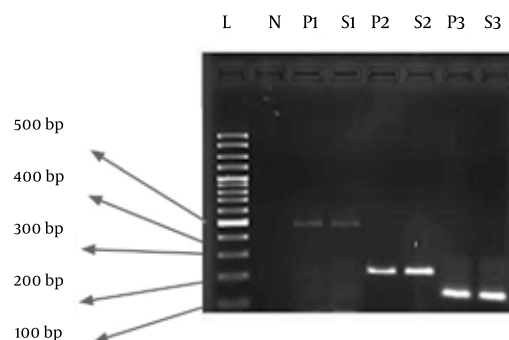
Lourmat, France). We took a 1- μl subsample of each PCR product of the first round of PCR, including those that were both positive and negative for Norovirus. Using NV 51 (5'-GTTGACACAATCTCATCATC-3') and NV 3 (5'-GCAC-CATCTGAGATGGATGT-3') primers, we amplified 206 bp under the following PCR conditions: 30 cycles of 1 min at 94°C , 1 min at 42°C , and 1 min at 72°C , followed by a final extension of 6 min at 72°C . The PCR products were loaded on a 3% agarose gel, stained, and visualized under a UV Transilluminator (10).

To determine whether samples contained Norovirus from genogroup I or genogroup II, 2 pairs of specific genogroup primers were used in a PCR reaction. For group 1, these were CAP A (5'-GGC (A/T) GTT CCC ACA GGC TT-3') and CAP B (5'-TAT GTT GAC CCT GAT AC-3'). For group 2, the specific primers were NVP 110 (5'-AC(A/T/G) AT(C/T)TCATCATCACCATA-3') and SR 46 (5'-TGGAATTC-CATCGCCCACTGG-3') (11). These primers amplify 178 bp from ORF 2 that encodes a capsid protein in genogroup I and 119 bp from ORF 1a that encodes RNA polymerase in genogroup II. The PCR conditions were as follows: 30 cycles of 30 sec at 94°C , 30 sec at 50°C , and 45 sec at 72°C followed by a final extension of 5 min at 72°C (Figure 1).

4. Results

After the second round of PCR, Norovirus was detected in 9 of 143 (6.3%) fecal samples. All the positive samples belonged to genogroup II and there were no positive Norovirus genogroup I samples. Five positive samples were from male patients and 4 were from female patients. There was no relationship between gender and the prevalence of the virus. The peak infection rate was observed in winter (66.9% of positive cases), and we did not detect any positive cases during the summer.

Figure 1. Representative Agarose Gel Electrophoresis of PCR Products From A Positive Norovirus Sample.



P lanes show results from a positive control and S lanes are the results from a field sample. The number 1 indicates results of first-round PCR, the number 2 indicates the results of nested PCR, and the number 3 indicates genogroup test results. A 100-bp DNA ladder was loaded into the L lane.

5. Discussion

Acute gastroenteritis is one of the most common diseases in the human population (12). Annually, up to 700

million cases of this disease occur in children under 5 years of age. Usually, gastroenteritis causes mild diarrhea. However, patients may show more severe symptoms ranging from relatively mild upper gastroenteritis symptoms, such as nausea and vomiting, to severe symptoms, such as profuse diarrhea that leads to dehydration or death (13). Human caliciviruses, especially noroviruses, are important causes of nonbacterial gastroenteritis. They are the most important pathogen transmitted through contaminated food (14).

According to studies from all over the world, the prevalence of this virus is between 6%-19% (15). For example, O'Neill et al found that the prevalence of this virus was 6.7% in Ireland (16). In Germany, Rohayem et al determined that the prevalence of Norovirus was 6.6% during one study interval and 9.5% during a second study interval (17). In our study, the prevalence of Norovirus was 6.3%, very similar to the prevalence reported by these authors. Children under 5 years of age show higher rates of infection with this virus than people in other age groups. Among these children, the peak infection rate occurs between the ages of 3 and 5 years. In a study performed in Bangladesh, researchers found Norovirus antibodies in 43% of the children between 2 and 49 months of age. The authors of this study found that only 7% of 2-7-month-old children harbored Norovirus antibodies, but 100% of 4-year-old children had them (18). We found that 6.3% of the fecal samples from children under 5 years of age contained noroviruses. Additionally, 55.6% of these samples were from children between 3 and 5 years of age. Children under 1 year of age had the lowest prevalence of Norovirus infections (22.2%). Our results are in concordance with studies undertaken in other areas of the world. Ike et al. reported that out of the 148 fecal samples they tested, 95% were positive for noroviruses belonging to genogroup II (19). This report matches our results and those of similar studies showing that genogroup II is the dominant group all over the world (20, 21). Human caliciviruses, especially Noroviruses, are important causes of nonbacterial gastroenteritis. The prevalence of this virus in Ahvaz is similar to that in other studies. Because this virus is transmitted through contaminated food or water, drinking-water sanitation greatly reduces the incidence of Norovirus gastroenteritis. In addition, this study improves our epidemiological knowledge about the prevalence of this virus in Ahvaz and Iran.

Acknowledgments

The authors wish to thank Prof. Pierre Pothiere from Centre National Reference des Virus Enteriques, Laboratoire de Virologie, Dijon, France for transferring a Norovirus-positive sample to them.

Financial Disclosure

All authors declare that they have no conflict of interest.

Funding/Support

Research Centre for Tropical and Infectious Diseases. Vice chancellor of Research and Technology, Ahvaz, Jundishapur University of Medical Sciences.

References

1. Yan H, Yagyu F, Okitsu S, Nishio O, Ushijima H. Detection of norovirus (GI, GII), Sapovirus and astrovirus in fecal samples using reverse transcription single-round multiplex PCR. *J Virol Methods*. 2003;**114**(1):37-44.
2. Lew JF, Valdesuso J, Vesikari T, Kapikian AZ, Jiang X, Estes MK, et al. Detection of Norwalk virus or Norwalk-like virus infections in Finnish infants and young children. *J Infect Dis*. 1994;**169**(6):1364.
3. Doane F, Kapikian A. Electron microscopy for the detection of gastroenteritis inviral infections of the gastrointestinal tract. *Marcel Dekker Inc New York NY viruses*. 1994:101-30.
4. Foley B, O'Mahony J, Morgan SM, Hill C, Morgan JG. Detection of sporadic cases of Norwalk-like virus (NLV) and astrovirus infection in a single Irish hospital from 1996 to 1998. *J Clin Virol*. 2000;**17**(2):109-17.
5. Nordgren J, Bucardo F, Dienus O, Svensson L, Lindgren PE. Novel light-upon-extension real-time PCR assays for detection and quantification of genogroup I and II noroviruses in clinical specimens. *J Clin Microbiol*. 2008;**46**(1):164-70.
6. Kaplan JE, Gary GW, Baron RC, Singh N, Schonberger LB, Feldman R, et al. Epidemiology of Norwalk gastroenteritis and the role of Norwalk virus in outbreaks of acute nonbacterial gastroenteritis. *Ann Intern Med*. 1982;**96**(6 Pt 1):756-61.
7. Jiang X, Wilton N, Zhong W, Farkas T, Huang P, Barrett E, et al. Diagnosis of human caliciviruses by use of enzyme immunoassays. *J Infect Dis*. 2000;**181**(Supplement 2):S349.
8. Green J, Henshilwood K, Gallimore C, Brown D, Lees D. A nested reverse transcriptase PCR assay for detection of small round-structured viruses in environmentally contaminated mollusc shellfish. *Appl Environ Microbiol*. 1998;**64**(3):858.
9. Wang J, Jiang X, Madore HP, Gray J, Desselberger U, Ando T, et al. Sequence diversity of small, round-structured viruses in the Norwalk virus group. *J Virol*. 1994;**68**(9):5982-90.
10. Moe CL, Gentsch J, Ando T, Grohmann G, Monroe SS, Jiang X, et al. Application of PCR to detect Norwalk virus in fecal specimens from outbreaks of gastroenteritis. *J Clin Microbiol*. 1994;**32**(3):642-8.
11. Pang X, Lee B, Chui L, Preiksaitis JK, Monroe SS. Evaluation and validation of real-time reverse transcription-pcr assay using the LightCycler system for detection and quantitation of norovirus. *J Clin Microbiol*. 2004;**42**(10):4679-85.
12. Dolin R, Treanor JJ, Madore HP. Novel agents of viral enteritis in humans. *J Infect Dis*. 1987;**155**(3):365-76.
13. Snyder JD, Merson MH. The magnitude of the global problem of acute diarrhoeal disease: a review of active surveillance data. *Bull World Health Organ*. 1982;**60**(4):605-13.
14. Mead PS, Slutsker L, Dietz V, McCaig LF, Bresee JS, Shapiro C, et al. Food-related illness and death in the United States. *Emerg Infect Dis*. 1999;**5**(5):607-25.
15. Buesa J, Collado B, Lopez-Andujar P, Abu-Mallouh R, Rodriguez Diaz J, Garcia Diaz A, et al. Molecular epidemiology of caliciviruses causing outbreaks and sporadic cases of acute gastroenteritis in Spain. *J Clin Microbiol*. 2002;**40**(8):2854-9.
16. O'Neill HJ, McCaughey C, Coyle PV, Wyatt DE, Mitchell F. Clinical utility of nested multiplex RT-PCR for group F adenovirus, rotavirus and norwalk-like viruses in acute viral gastroenteritis in children and adults. *J Clin Virol*. 2002;**25**(3):335-43.
17. Rohayem J, Berger S, Juretzek T, Herchenroder O, Mogel M, Poppe M, et al. A simple and rapid single-step multiplex RT-PCR to detect Norovirus, Astrovirus and Adenovirus in clinical stool samples. *J Virol Methods*. 2004;**118**(1):49-59.
18. Black RE, Greenberg HB, Kapikian AZ, Brown KH, Becker S. Acquisition of serum antibody to Norwalk Virus and rotavirus and relation to diarrhea in a longitudinal study of young children in rural Bangladesh. *J Infect Dis*. 1982;**145**(4):483-9.

19. Ike AC, Brockmann SO, Hartelt K, Marschang RE, Contzen M, Oehme RM. Molecular epidemiology of norovirus in outbreaks of gastroenteritis in southwest Germany from 2001 to 2004. *J Clin Microbiol.* 2006;**44**(4):1262-7.
20. Fankhauser RL, Monroe SS, Noel JS, Humphrey CD, Bresee JS, Parashar UD, et al. Epidemiologic and molecular trends of "Norwalk-like viruses" associated with outbreaks of gastroenteritis in the United States. *J Infect Dis.* 2002;**186**(1):1-7.
21. Kirkwood CD, Bishop RF. Molecular detection of human calicivirus in young children hospitalized with acute gastroenteritis in Melbourne, Australia, during 1999. *J Clin Microbiol.* 2001;**39**(7):2722-4.

Archive of SID