



## Screening of Novel Bacteriophage Infection in *Pseudomonas putida* Isolated From Potato Disease

Abbas Soleimani Delfan<sup>1</sup>, Zahra Etemadifar<sup>1\*</sup>, Majid Bouzari<sup>1</sup>, Giti Emtiazi<sup>1</sup>

<sup>1</sup> Department of Biology, Faculty of Sciences, University of Isfahan, Isfahan, IR Iran

### ARTICLE INFO

Article type:  
Original Article

Article history:  
Received: 11 Dec 2011  
Revised: 29 Feb 2012  
Accepted: 03 Apr 2012

Keywords:  
*Pseudomonas putida*  
Bacteriophages  
Cystoviridae

### ABSTRACT

**Background:** In recent decades bacteriophages have been used as treating agents against some pathogens. Also, bacteriophages could be considered as alternatives of antimicrobial agents in plant protection.

**Objectives:** This study aimed to isolate a pathogen from potato (*Solanum tuberosum*) tubers (*Pseudomonas putida*), and of its specific bacteriophage from soil and wastewater.

**Materials and Methods:** Infected samples of soft rot potato tubers were collected from Flavarjan farms (Isfahan province, Iran) to isolate and identify *P. putida* by biochemical and molecular methods. Soil and wastewater samples were obtained locally to isolate the bacteriophage attributed to *P. putida*. Soil suspension was centrifuged and then filtrated by 0.2 micrometer Millipore filter. The wastewater was directly filtrated by 0.2 micrometer filter after centrifugation. After incubation of isolated bacteria together with phage contained solution, plaques were detected in nutrient agar. Subsequently, clearance of *P. putida* liquid culture incubated by added filtrated bacteriophage was observed. The structure and morphological characteristics of *P. putida* related bacteriophage was remarked by transmission electron microscopy (TEM).

**Results:** Isolated strain sda2 was identified as *Pseudomonas putida* with related accession number HQ423667. The result showed that isolated bacteriophage belonged to *Cystoviridae* family.

**Conclusions:** We isolated a new strain of *P. putida* as well as a novel bacteriophage through which potato disease caused by the bacterium could be treated.

Published by Kowsar Corp, 2012. cc 3.0.

### ► Implication for health policy/practice/research/medical education:

This study was performed to isolate bacteriophage against plant pathogen and suggest a biocontrol method for plant diseases.

### ► Please cite this paper as:

Soleimani Delfan A, Etemadifar Z, Bouzari M, Emtiazi G. Screening of Novel Bacteriophage Infection in *Pseudomonas putida* Isolated From Potato Disease. *Jundishapur J Microbiol.* 2012;5(4):550-4. DOI: 10.5812/jjm.3786

## 1. Background

*Pseudomonas putida* as a symbion bacteria is found in plant rhizosphere that is known as an antagonist against plants pathogens bacteria (1). Some bacteria such as *Ralstonia solanacearum* and *Erwinia* sp. cause soft rot disease

in potato and *Solanaceae* family (2). Nevertheless, there are some data that demonstrate oxidase positive *Pseudomonas* such as *P. putida* may cause soft rot disease in potato, too (3). Bacteriophages are among the most effective inhibitor agents to control bacterial population and change bacterial genome during transduction (4-7). In

\* Corresponding author: Zahra Etemadifar, Department of Biology, Faculty of Sciences, University of Isfahan, Azadi Sq, Daneshgah St., P.O. BOX: 81746-73441, Isfahan, IR Iran. Tel: +98-3117932367, Fax: +98-3117932456, E-mail: z.etemadifar@sci.ui.ac.ir

DOI: 10.5812/jjm.3786

© 2012 Ahvaz Jundishapur University of Medical Sciences; Published by Kowsar Corp.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

recent decades bacteriophages have been used as treatment agents against some human and animal pathogens such as *E.coli* and gastrointestinal infections (8-10). Also, the use of bacteriophage in the plants may protect them via reduction in plant bacterial disorders (11).

## 2. Objectives

This study aimed to isolate an infecting bacterium from infected potato (*Solanum tuberosum*), and of its bacteriophage from wastewater and farm soils for possible use in treatment of infected potato or biocontrol of the disease.

## 3. Materials and Methods

Chemical materials were purchased from Merck and Sigma Companies. DNA extraction kit was provided from Cinnagen (Cat No: DN8115C, Iran). Two primers for 16S rRNA gene manipulation were prepared by TAG Copenhagen A/S, Denmark. Millipore filter was defined as 0.22 µm acetate cellulose filters (Orange scientific, Spanish). Lyophilization instrument (HETOSICC, Denmark) was applied to concentrate phage solution. Sonication of samples was performed by a sonicator (Transonic 660/H, Germany). Transmission electron microscopy (TEM) (Phillips CM10) was employed for fine detection of phage particles.

### 3.1. Isolation of *Pseudomonas putida*

Infected potato tubers with soft rot symptom were collected from Flavarjan, Isfahan province, Iran. The tissue surface was sterilized by 70% ethanol and rinsed 4-5 times by sterile distilled water; then tubers were cut into pieces with 5mm dimensions and suspended in test tubes containing 5ml sterile double distilled water. The test tubes were incubated at 37°C for 60 minutes; this bacterial suspension (i.e. suspended potato tubers in water) was placed in an agar medium supplemented by 15 gr/l Agar, 5 gr/l Peptone, 5 gr/l NaCl, 2 gr/l Yeast extract, and 1 gr/l beef extract for 24-48 hours.

### 3.2. Biochemical Tests For Identification of Isolated *P. putida*

To identify biochemical characteristics of *P. putida*, various tests were applied including oxidase, catalase, oxidative/fermentative (O/F), motility, utilization of glucose and trehalose, and lecithin tests, as well as growth occurrence of the bacterium in 41°C and 4°C.

### 3.3. 16S rRNA Gene Sequencing.

Bacterial genomic DNA obtained from pure culture was extracted by DNA extraction kit (Cinnagen Cat No: DN8115C). This extract was examined by electrophoresis in 1% agarose gel. For 16S rRNA gene amplification, universal primers including RW01 as forward primer (5' AACTGGAG-GAAGGTGGGGAT 3') and DG74 as reverse primer (5' AGGAG-GTGATCCAACCGCA 3') were used (12). PCR conditions were set as follows: initial denaturation at 95°C for 5 minutes (1

cycle), denaturation at 94°C for 45 seconds, annealing at 54°C for 30 seconds, extension by synthesized pieces of 16S rRNA genes at 72°C for 45 seconds, in 30 cycles, and final extension at 72°C for 5 minutes.

### 3.4. Isolation of Bacteriophage

To isolate bacteriophage, wastewater and farm soils were collected from Flavarjan, Isfahan province. In the first stage, the wastewater was centrifuged at 12000 rpm for 10 minutes followed by supernatant filtration by 0.22 µm acetate cellulose filters (Orange scientific). In the second stage, in order to isolate bacteriophage from the soil, 30 - 50 ml of 1% potassium citrate supplemented by 10 gr/l  $K_3C_6H_5O_7 \cdot H_2O$ , 44.1 gr/l  $Na_2HPO_4 \cdot 7H_2O$ , 24 gr/l  $KH_2PO_4$ , pH 7 (13) was mixed with 10 grams of soil sample and sited on ice. Samples were sonicated (transonic 660/H) in three one-minute intervals with one minute rest on ice. This suspension, then, was placed at 4°C for 24 hours and centrifuged at 12000 rpm in 10 minute. The supernatant was filtrated by 0.22 µm acetate cellulose filter and stored at 4°C in refrigerator.

### 3.5. Bacteriophage Enrichment

To enrich isolated bacteriophage, 1% of filtrated bacteriophage suspension was included to 50 ml overnight prepared culture of *P. putida*, followed by rotation of this combination in a shaker (50 rpm) at 28°C for 24 hours. The medium was filtrated by 0.22 µm acetate cellulose filters (Orange scientific) and stored at 4°C on 1% of glycine as well as on several drops of chloroforms (13).

### 3.6. Bacteriophages Plaque Assay

The pure culture of *P. putida* was prepared on nutrient agar (or Muller Hinton agar). After 2 hours, 10 µL of filtrated bacteriophage was added to culture plate and incubated at 28°C for 24 hours.

### 3.7. Bacteriophage Purification and Concentration

Bacteriophage purification and concentration are usually performed by the use of two methods including CsCl density-gradient for sample ultracentrifugation and PEG 8000 for purification (14, 15). In this study, lyophilization method was used for bacteriophage concentration and purification, instead. Thirty ml of bacteriophage solution in test flask was incubated in lyophilization instrument (HETOSICC, DENMARK), that was reduced finally to 5 ml. The concentrated solution was then passed through 0.22 µm cellulose acetate filters and prepared for electron microscopy staining.

### 3.8. Electron Microscopy

For bacteriophage staining, phage solution was deposited on carbon coated copper grid for two minutes and stained by 2% (w/v) uranyl acetate, pH 4 - 4.5. The solution was drained through filter paper and phage particles were observed through transmission electron micros-

copy (TEM) (Phillips CM10).

### 3.9. Chemicals

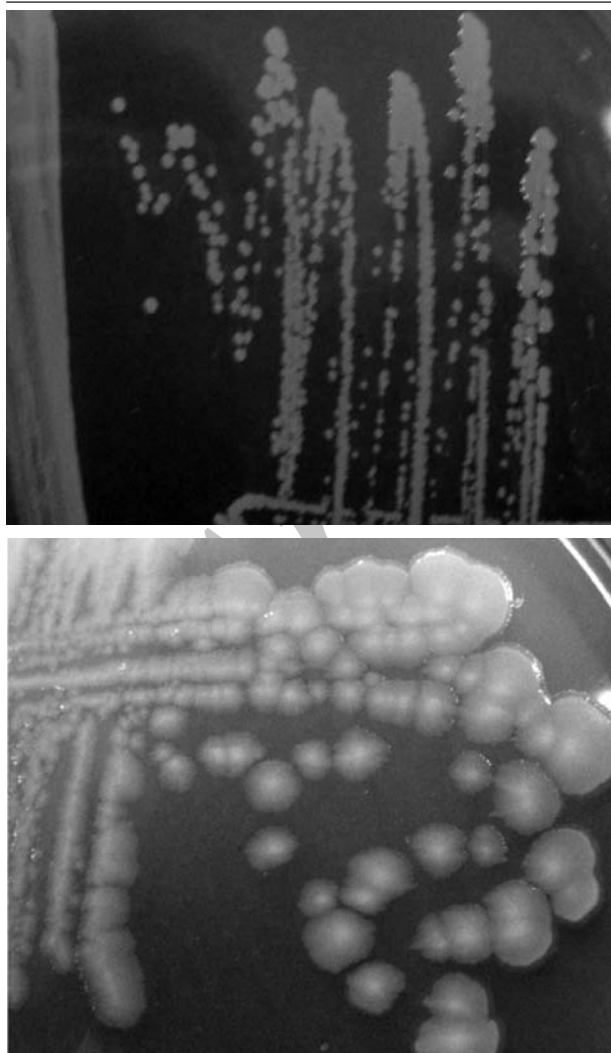
Other chemicals were purchased from Merck or Sigma representative companies in Iran.

## 4. Results

### 4.1. Biochemical Identification of Isolated *P. Putida*

The biochemical and morphological investigations showed that the isolated strain related to *P. putida*. Positives oxidase and catalase results as well as, positive glucose and negative trehalose utilization, negative growth at 41°C and 4°C, oxidative and non-fermentative (O<sup>+</sup>/F<sup>-</sup>) effect, and negative lecithin result were detected. Also, the morphology of isolated bacteria was different from that of other strains of this genus such as *P. putida* DSM 6125 (Figure 1).

**Figure 1.** Comparison between Colony Morphology of Standard *Pseudomonas putida* DSM 6125 (Above) and Isolated *P. putida* Strain sda2 Extracted From Infected Potato (*Solanum tuberosum*) in This Study (Below).



Bacteria Were Grown on Nutrient Agar Medium

### 4.2. 16S rRNA Sequencing

The sequence of 16S rRNA gene PCR product was blasted in NCBI database and mega BLAST program. Data showed that isolated bacterium was attributed to *P. putida*. Information of bacterial genome was registered in NCBI database as *Pseudomonas putida* strain sda2 with accession number HQ423667.

### 4.3. Bacteriophage of *P. Putida*

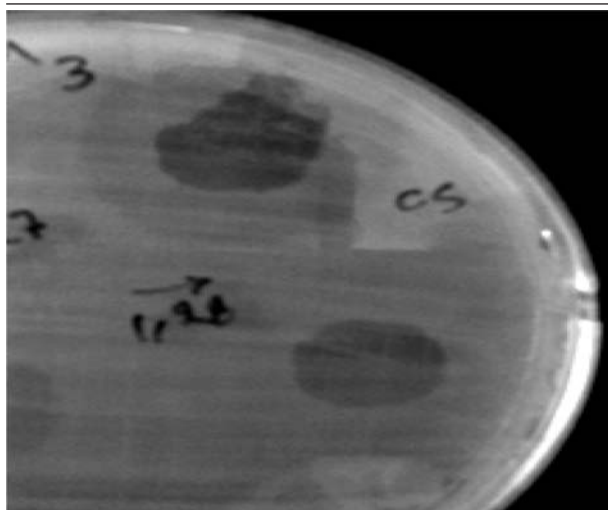
In this study, Broth enrichment method was employed for bacteriophage enrichment. After 10-24 hours, bacterial lysate sedimentation occurred in the tube indicating complete lysis of bacterial cells in the presence of bacteriophage (Figure 2). Small Drop method was used to observe bacteriophage plaques. After bacteriophage inoculation on bacterial culture and incubation for five hours, phage plaques appeared on MHA (Figure 3). Bacteriophage stained by 2% uranyl acetate in pH 4-4.5 was observed in transmission electron microscopy characterized by hexagonal nucleocapsid and about 74 nm across with no tail in its structure indicating that the phage was associated with *Cystoviridae* family, morphologically (Figure 4). A lipid envelope was detected around the head of bacteriophage.

### 4.4. Storage of Bacteriophage

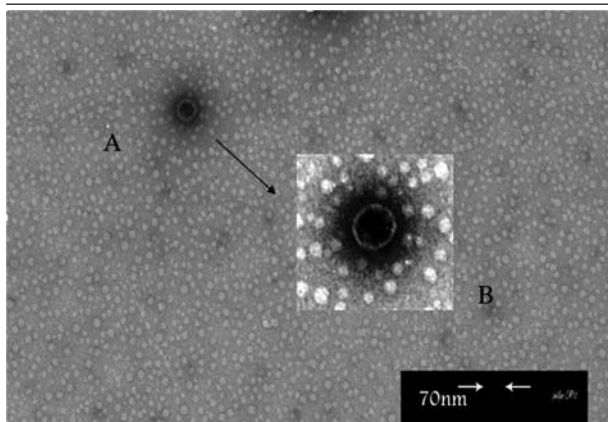
Stored bacteriophage samples treated by chloroform were damaged during storage period because of sensitivity to this buffer. However, population of the phage in the samples treated by 1% glycine remained intact.



**Figure 2.** Complete Lysis of Bacterial Cells (*P. putida* sda2) in the Presence (Right) and Absence (Left) of Respective Bacteriophage in Liquid Culture After 12 Hours.

**Figure 3.** Plaque Formation on *Pseudomonas putida* Strain Sda2 Culture.

Inoculated Muller Hinton Agar Medium by Propagated *P. putida* Sda2 Incubated with added Filtrated Bacteriophage at 28°C For 24 Hours.

**Figure 4.** Original Electron Micrograph of Bacteriophage Attributed to *Pseudomonas putida* Strain Sda2.

(A) Amplified Picture With High Resolution (B) An Envelope is Seen Around The Phage. This phage was propagated on *P. putida* Strain Sda2 by Overnight Incubation at 28°C. The Phage Suspension was concentrated by lyophilization, passed Through 0.22  $\mu\text{m}$  Cellulose Acetate Filter, and Prepared for Electron Microscopy by Uranyl Acetate Staining.

## 5. Discussion

In this survey *P. putida* sda2 was isolated from infected potato tubers. Biochemical tests and 16S rRNA gene homology indicated that isolated strain was attributed to *Pseudomonadales* family. Morphology of *P. putida* sda2 colony was different from which of standard strain *P. putida* DSM 6125. Janse and colleagues have reported that oxidase positive *Pseudomonas* such as *P. putida*, *P. aureofaciens*, *P. tolaassii* and *P. fluorescens* have soft rot activity (16). Usually, *P. putida* does not have any effect on plants and antagonizes *Fusarium* and *Ralstonia* effects (17). Shaburva and Krylov have obtained 5 bacteriophages: af, pf16,  $\phi$  2f,  $\phi$  15, and  $\phi$  27 from which  $\phi$  15,

$\phi$  27, and  $\phi$  2f were attributed to *Podoviridae* family, and phage pf16 to *Myoviridae* family (18). Lee and Yen studied the effect of bacteriophage gh-1 - isolated by Lee and Boezi in 1966 - on *P. putida* in oil enhanced recovery. This bacteriophage exhibited a regular hexagonal outline, about 50  $\mu\text{m}$  across, and a short, wedge-shaped tail attached to a corner corresponded to the morphology and size of bacteriophage head and non-contracted tail of *Podoviridae* family.

In this study, the bacteriophage observed under TEM was characterized by a hexagonal nucleocapsid, about 74 nm across, no visible tail, and being similar to *Cystoviridae* family in respect of morphology and size. In previous studies, two types of bacteriophages from *Podoviridae* and *Myoviridae* families were reported for *P. putida* but not from *Cystoviridae* family. Based on international committee on taxonomy of viruses (ICTV) only one bacteriophage (named Phage  $\phi$ 6) was reported to act against *Pseudomonas syringae*, but there was no report about *P. putida*.

The *cystoviridae* family consists of icosahedral enveloped multilayer phages with segmented dsRNA. Our experiments demonstrated that the glycine buffer is a suitable medium to preserve bacteriophages in refrigerator without any effect on the population. In conclusion, we isolated a new strain of *P. putida* (accession number HQ423667), and a novel bacteriophage for *P. putida* from *Cystoviridae* family. In respect to treatment application of bacteriophages against some human and animal pathogens in recent decades, we suggest that phages isolated from natural habitat of plants against infectious agents might be employed to control plant diseases in the same area. Experiments to determine the effectiveness of phage against potato infections should be performed in order to develop a phage control treatment for the disease.

## Acknowledgements

We express our thanks to the University of Isfahan.

## Financial Disclosure

We thank the University of Isfahan for financial support of this study.

## Funding Support

This survey was financially supported by the University of Isfahan.

## References

1. Marugg JD, van Spanje M, Hoekstra WP, Schippers B, Weisbeek PJ. Isolation and analysis of genes involved in siderophore biosynthesis in plant-growth-stimulating *Pseudomonas putida* WCS358. *J Bacteriol.* 1985;164 (2):563-70.
2. Dickey RS. *Erwinia chrysanthemi*: a comparative study of phenotypic properties of strains from several hosts and other *Erwinia*

- species. *Phytopathology*. 1979;**69** (4):324.
3. Janse J, Derks J, Spit B, Van Der Tuin W. Classification of Fluorescent Soft Rot *Pseudomonas* Bacteria, Including *P. marginalis*-Strains, Using Whole Cell Fatty Acid Analysis. *Syst Appl Microbiol*. 1992;**15**(4):538-53.
  4. Fuhrman JA. Marine viruses and their biogeochemical and ecological effects. *Nature*. 1999;**399** (6736):541-8.
  5. Fuhrman JA, Schwalbach M. Viral influence on aquatic bacterial communities. *Biol Bull*. 2003;**204** (2):192-5.
  6. Miao EA, Miller SI. Bacteriophages in the evolution of pathogen-host interactions. *Proc Natl Acad Sci U S A*. 1999;**96** (17):9452-4.
  7. Weinbauer MG, Rassoulzadegan F. Are viruses driving microbial diversification and diversity? *Environ Microbiol*. 2004;**6** (1):1-11.
  8. Gorski A, Weber-Dabrowska B. The potential role of endogenous bacteriophages in controlling invading pathogens. *Cell Mol Life Sci*. 2005;**62** (5):511-9.
  9. Smith HW, Huggins MB, Shaw KM. The control of experimental *Escherichia coli* diarrhoea in calves by means of bacteriophages. *J Gen Microbiol*. 1987;**133** (5):1111-26.
  10. Verthe K, Possemiers S, Boon N, Vaneechoutte M, Verstraete W. Stability and activity of an *Enterobacter aerogenes*-specific bacteriophage under simulated gastro-intestinal conditions. *Appl Microbiol Biotechnol*. 2004;**65**(4):465-72.
  11. Jones JB, Jackson LE, Balogh B, Obradovic A, Iriarte FB, Momol MT. Bacteriophages for plant disease control. *Annu Rev Phytopathol*. 2007;**45**:245-62.
  12. Leong D, Greisen K. PCR detection of bacteria found in cerebrospinal fluid. In: Persing DH, editor. *Diagnostic Molecular Microbiology: Principles and Applications*. Washington: American Society for Microbiology; 1993.
  13. Clokie M, Kropinski A. Bacteriophages: methods and protocols, volume 1: isolation, characterization, and interactions. *Humana, New York*. 2009.
  14. Gnanamanickam SS. *Plant-Associated Bacteria*. Dordrecht: Springer; 2006.
  15. Sambrook J, Russell DW. *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor, N.Y: Cold Spring Harbor Laboratory Press; 2001.
  16. Shaburova OV, Krylov SV, Veiko VP, Pleteneva EA, Burkal'tseva MV, Miroshnikov KA, et al. [Search for destruction factors of bacterial biofilms: comparison of phage properties in a group of *Pseudomonas putida* bacteriophages and specificity of their halo-formation products]. *Genetika*. 2009;**45** (2):185-95.
  17. Chang PL, Yen TF. Interaction of *Pseudomonas putida* ATCC 12633 and Bacteriophage gh-1 in Berea Sandstone Rock. *Appl Environ Microbiol*. 1985;**50** (6):1545-7.
  18. Lee LF, Boezi JA. Characterization of bacteriophage gh-1 for *Pseudomonas putida*. *J Bacteriol*. 1966;**92** (6):1821-7.