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**Research** Article

# The First Macrolide-Resistant Bordetella pertussis Strains Isolated From Iranian Patients

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Background: Whooping cough was considered as one of the major causes of childhood morbidity and mortality worldwide. Resistant isolates of Bordetella pertussis to macrolides in some countries have been recently reported.

Objectives: Recent reports on macrolide-resistant B. pertussis isolates and lack of evidence for such resistance in clinical isolates of the Iranian patients led the authors of the current study to study antibiotic susceptibility of the collected isolates in the country. Susceptibility of the B. pertussis isolates to three antibiotics was studied. Relatedness of the strains recovered in this research was also examined.

Materials and Methods: The antibacterial activities of erythromycin, azithromycin, and clarithromycin antibiotics against the recovered isolates of 779 nasopharyngeal swabs were examined using MIC (Minimum Inhibitory Concentration) method. Relationship of the strains was characterized by Pulsed-field Gel Electrophoresis (PFGE).

Results: Among the specimens, 11 cases (1.4%) were culture-positive. Among these isolates, only two isolates had high MIC values for erythromycin and clarithromycin. Pulsed-field gel electrophoresis analysis of the isolates revealed 6 PFGE profiles (A-F) among which three and two isolates had the same patterns in profiles A and B, respectively.

Conclusions: Azithromycin can be a good drug of choice to treat patients infected by B. pertussis in Iran. Clonal relationship of the isolates showed that the same *B. pertussis* strains were isolated from different patients in Iran.

Keywords:Bordetella pertussis, Macrolides; PFGE; Antibiotic Resistance

## 1. Background

Bordetella pertussis causes an important respiratory tract disease known as whooping cough. This illness was considered as one of the major causes of childhood morbidity and mortality worldwide. After introduction of the whole-cell vaccines in 1940s, the rate of pertussis decreased (1). However, the incidence of this contagious disease has been gradually increasing since early 1980s. It seems that there are some compelling explanations for this increase during vaccination programs in the world. It has been suggested that waning immunity along with pathogen adaptation are the main causes for the resurgence of pertussis (2-4).

Antibiotic treatment is a critical step to prevent the spread of diseases such as pertussis. As pertussis is a highly contagious respiratory disease, monitoring the emergence of *B. pertussis* specially the isolates which showed resistance to antibiotics is very important. Overuse and inappropriate use of antibiotics provide an opportunity for easy spread of these resistant and contagious bacteria in the community.

Among macrolide antibiotics, erythromycin is the first choice of treatment for pertussis infections (5, 6). Since resistance to antimicrobial agents in *B. pertussis* was rare, testing susceptibility to antibiotics in clinical isolates of this bacterium was therefore not deemed necessary (7). Prior to 1994, resistance to erythromycin in *B. pertussis* had not been observed. Since then few reports of resistant isolates of B. pertussis to macrolides in some countries such as the United States and Taiwan have been published (8-12). In addition, resistance to guinolones as an alternative drug has also emerged in *B. pertussis* isolates in Japan (13). Mechanisms of resistance in *B. pertussis* are being understood. Recent studies suggest that mutation in the genes related to erythromycin could result in resistance to this macrolide antibiotic (8,9).

Among various microbial strain-typing methods,

Implication for health policy/practice/research/medical education:

Screening for antimicrobial resistance of Bordetella pertussis as a respiratory tract infection agent that has re-emerged recently, and the clonal relationship of the strains would be essential in order to control the spread of these bacteria. Clonal spread of such strains that are transferred from one person to another through the respiratory tract may cause some important outbreak of the disease and should be considered.

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Pulsed-field Gel Electrophoresis (PFGE) has been used to establish genetic relationships between the *B. pertussis* strains (14). Some studies have also revealed significant changes in the *B. pertussis* population over time by this useful method (15, 16). Here PFGE was used to study clonality of the isolated strains from Iranian patients.

## 2. Objectives

Previous studies in Iran showed increase of pertussis incidence in the community (17, 18). Recent reports on macrolide-resistant *B. pertussis* isolates and lack of evidence of such resistance in clinical isolates of Iranian patients led the authors to determine antibiotic susceptibility of the collected isolates in the country. Susceptibility of the isolates to erythromycin, azithromycin, and clarithromycin as the drugs of choice to treatment of whooping cough was studied. Finally, relatedness of the isolated clinical strains recovered in this research was examined by PFGE.

## 3. Methods and Materials

## 3.1. Bacterial Isolates

A total of 779 nasopharyngeal swabs were collected from pertussis suspected patients with the age ranging from less than one month to more than ninety years and delivered to the Pertussis Reference Laboratory at the Pasteur Institute of Iran from May 2009 to December 2010. Specimen swabs (Dacron-tipped swabs) were transported from different provinces of Iran to the laboratory on Regan-Lowe transport medium. Specimens were cultured and streaked on fresh Regan-Lowe, and Bordet Gengou mediums (Difco, USA) containing 10% defibrinated horse blood with and without cephalexin (40 µg/mL) (Sigma Chemical Co., USA). After incubation, suspected colonies were confirmed as *B. pertussis* using conventional biochemical tests and specific agglutination reaction with *B. pertussis* antiserum (Difco, USA) (17-20).

## 3.2. Antibiotic Susceptibility Testing

The activities of erythromycin, azithromycin and clarithromycin (Shifa Pharmed Industrial Group, Iran) against *B. pertussis* isolates were examined using agar dilution by the direct suspension method in Muller-Hinton agar (Difco, USA) supplemented with 5% horse blood with incubation temperatures of 35-37°C for a period of 72 hours (21, 22). To determine MIC (Minimum Inhibitory Concentration), the concentrations of antimicrobial agents, range of 0.016 to 512  $\mu$ g/Ml were prepared. Antibiotic-free plates were inoculated to check for growth and purity. *B. pertussis ATCC* 9797 and *Staphylococcus aureus ATCC* 29213 served as the control strains. Strains with macrolide MICs less than or equal to 0.12  $\mu$ g/ml were considered susceptible (13, 22, 23).

# 3.3. Pulsed field Gel Electrophoresis

*B. pertussis* isolates were characterized by PFGE as described previously (14). The plug slices were incubated overnight at  $37^{\circ}$ C in 100 µL of buffer solution containing 30 U of XbaI (Fermentase, Vilnius, Lithuania). The pulsed-field program was modified (running times, 19 hours, and pulse times, 5 to 40 seconds, 6 V/cm). The cooling system was set to  $14^{\circ}$ C. Restricted fragments of chromosomal DNA of the isolates were separated in CHEFF DR-II apparatus (Bio-Rad, USA). *Salmonella braenderup* H9812 was used as molecular weight marker.

#### 4. Results

In the current study, out of 779 nasopharyngeal swabs moistened per individual, 11 specimens (1.4%) were culture-positive. Of the total samples examined in the current study, 657 (84.3%) specimens were collected from 10-years-old or younger patients. Ten out of 11 culture-positive strains were isolated from 10-years-old or younger patients. Nine cultures were collected from two-years-old or younger patients.

According to Table 1, MIC determination of the isolates revealed that only two isolates had high MIC values against erythromycin and clarithromycin (MIC  $\geq$  128 µg/mL). From the 11 strains whose MICs were examined, five and six strains showed MIC  $\geq$  2 µg/mL against erythromycin and clarithromycin, respectively. Except one isolate with MIC=0.12 µg/mL against azithromycin, all the isolates showed MIC  $\leq$  0.06 µg/mL against this antibiotic.

Pulsed-field gel electrophoresis analysis of 11 strains of *B. pertussis* in this research revealed six PFGE profiles (A-F) among which three (strains number 1 to 3) and two (strains number 6 and 7) isolates had the same patterns in profiles A1 and B, respectively (Figure 1). These isolates were collected from different provinces of Iran (Table 1). Five closely related isolates (stains number 1, 2, 3, 4 and 5) in profile A with similar susceptibility patterns (except for strain no. 5) were clustered (Table 1). Three of these strains were considered as profile A1 and the others as profiles A2 and A3 according to the minor difference in their banding patterns (Figure 1 and Table 1). In profile B, there were two strains with nearly similar susceptibility patterns isolated from different geographical locations at different dates.

Two strains (number 8 and 11) with high level of resistance to erythromycin and clarithromycin (MIC  $\geq$ 128) were isolated from two patients of different ages but living in the same province (Table 1). These two strains had distinct PFGE patterns, suggesting that they were not related. Hopefully, these strains were not among the isolates in profile A that were considered as a predominant profile (with 5 isolates) in this research.

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| Table 1. Antibiotic Susceptibility of Clinical B. pertussis Isolates <sup>a</sup> |                           |                         |              |              |                |              |
|---|---------------------------|-------------------------|--------------|--------------|----------------|--------------|
| Isolates  | Strains (Age of Patients) | Year (Province of Iran) | PFGE Pattern | МІС          |                |              |
| Number  |                           |                         |              | Erythromycin | Clarithromycin | Azithromycin |
| 1   | BpT1 (2 m)                | 2009 (Tehran)           | A1           | < 0.06       | < 0.06         | <0.06        |
| 2   | BpM1 (3 m)                | 2009 (Mazandaran)       | A1           | < 0.06       | < 0.06         | 0.125        |
| 3   | BpT2 (3 m)                | 2009 (Tehran)           | A1           | < 0.06       | < 0.06         | < 0.06       |
| 4   | BpE1 (2 y)                | 2009 (Esfahan)          | A2           | < 0.06       | < 0.06         | < 0.06       |
| 5   | BpQ1 (18 y)               | 2009 (Qom)              | A3           | < 0.06       | 2              | < 0.06       |
| 6   | BpM2 (3 y)                | 2009 (Mazandaran)       | В            | 4            | 8              | < 0.06       |
| 7   | BpK1 (5 m)                | 2010 (Khoozestan)       | В            | 4            | 32             | < 0.06       |
| 8   | ВрКО1 (4 у)               | 2009 (Khorasan)         | С            | 128          | >256           | < 0.06       |
| 9   | BpM3 (1 m)                | 2009 (Mazandaran)       | D            | < 0.06       | 0.5            | < 0.06       |
| 10  | BpK2 (1.5 m)              | 2009 (Khorasan)         | Е            | 2            | <0.06          | < 0.06       |
| 11  | BpK3 (26 d)               | 2009 (Khorasan)         | F            | 128          | > 256          | < 0.06       |

<sup>a</sup> Abbreviations: y, year; m, month; MIC, Minimum Inhibitory Concentration ; PFGE, Pulsed Field Gel Electrophoresis.



Figure 1. PFGE Pattern of the B. pertussis Strains Digested With XbaI

Lane M, genomic pattern of *S. braenderup* H9812 as a marker.

# 5. Discussion

Erythromycin used to be the drug of choice for treatment of pertussis however, because of resistance to this drug, azithromycin has been recently an alternative choice for children (24). In Japan, prophylactic administration of erythromycin caused adverse effects such as digestive organ symptoms, diarrhea, stomachache and abdominal distention in hospital staffs in 2009. It suggested that attention should be paid to erythromycin compliance during a pertussis outbreak (25).

In 2001 in Poland data showed that erythromycin remains the drug of choice for treatment of whooping cough and in case of resistant strains can be replaced by azithromycin. Azithromycin was found the most active antibiotic against *B. pertussis* infection (26). In the U.K in 2010, no evidence of resistance was found in the strains

tested against erythromycin, azithromycin and clarithromycin (27). In Taiwan all suspected isolates appeared to be susceptible to erythromycin, azithromycin, clarithromycin and co-trimoxazole (11). In Australia, no significant decrease was observed in the susceptibility to erythromycin among *B. pertussis* strains over 35 years (28). This is the first report on antimicrobial susceptibility of *B. pertussis* isolates in Iran. For evidence of such resistance in clinical isolates of *B. pertussis*, the susceptibility of isolates recovered from clinical specimens to three macrolides was examined.

As expected, the results showed that the rates of resistance to azithromycin were less than those of clarithromycin and erythromycin. Two isolates that indicated high resistance to the two latter antibiotics were susceptible to the former (Table 1). Therefore, similar to the other investigations, the current study data also demonstrated that azithromycin is the most efficient antibiotic against *B. pertussis* isolates (22, 24). Co-trimoxazole is a less active agent against *B. pertussis* strains. However, it can be used in children who cannot tolerate erythromycin or the ones infected by resistant isolates of *B. pertussis* (29).

Since the number of recovered isolates was low, the true rate of resistance could not be found in this research. The current study results suggested that macrolides, especially azithromycin can be a good choice to treat patients infected by *B. pertussis* in Iran. Different mechanisms may confer resistance to erythromycin in the isolates (30). Molecular methods can help to find the reason of resistance even in the absence of the isolates. Resistance to erythromycin is also likely due to a mutation of the erythromycin binding site in 23S rRNA gene (8, 9). In order to find the genetic relationship between the isolates, genomic patterns of the isolates were obtained by PFGE. Even though there are some new typing methods such as MLST (Multi-Locus Sequence Typing) and MLVA (Multiple-Locus Variable-Number Tandem Repeat Analysis) for epidemiologic study, PFGE has been used most widely for typing *B. pertussis* strains in view of its high discriminating power (14, 15, 31). By PFGE, it was shown that an international expansion of *B. pertussis* strains from a similar source resulted in high incidence of pertussis in 1997 in Europe and Taiwan (32).

Pulsed-field gel electrophoresis also found an outbreak and source of infection among *B. pertussis* recovered from Brazilian patients (33). Similarly, in Sweden, Advani. was able to successfully use PFGE as a tool to monitor the bacterial population in vaccine surveillance (14). PFGE also showed expansion of certain PFGE profiles within the *B. pertussis* population from five European countries (34). Based on the current study results, clonal relationship of the isolates showed that the same *B. pertussis* strains were isolated from different patients in Iran; Cluster A, including five strains, was found among the patients from different provinces of the country and may spread more easily than the other PFGE profiles.

Clonal spread of such strains, transferred from one person to another through the respiratory tract, may cause some important outbreak and should be considered. Fortunately, two highly resistant isolates in this research had unique patterns and had been isolated from different geographical locations. Since the rate of culture-positive strains was low, the obtained data in this research may not give the true incidence of pertussis in Iran. However, these data are the first information on the resistance and spread of *B. pertussis* isolates in different provinces of Iran.

In conclusion, screening for antimicrobial resistance of this respiratory tract infection agent may be warranted especially for strains that showed resistance to macrolides in order to control the spread of these bacteria. Moreover, molecular causes of resistance to antibiotics in the isolates need to be studied further.

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# **Authors' Contribution**

The presented work was carried out with collaboration of all the authors; Shahcheraghi defined the research theme; Lotfi, Parzadeh and Allahyar carried out the laboratory experiments including bacterial isolation and biochemical tests; Nikbin, Shooraj and Azizian performed antimicrobial testing; Shahcheraghi and Nikbin also interpreted the results and wrote the paper; Dr. Zahraei from Center for Communicable Diseases Control of Iran supported this research.

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