

# Study of the Association of Mutant *HBsAg* Gene and Hodgkin and Non-Hodgkin Lymphoma

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## Abstract

**Background:** Hepatitis B Virus (HBV) is responsible for chronic, acute, and fulminant hepatitis, which are prevalent worldwide. Chronic HBV may lead to cirrhosis and hepatocellular carcinoma. Several epidemiological studies have indicated that hepatitis B virus is involved in B-cell Hodgkin and Non-Hodgkin Lymphoma (NHL).

**Objectives:** The aim of this study was to evaluate the association between hepatitis B infection and Hodgkin and non-Hodgkin Lymphoma.

**Materials and Methods:** Paraffin embedded of 41 block samples including 12 (29.26%) Hodgkin and 29 (70.73%) non-Hodgkin patients were collected. Next, DNA extraction was carried out for all the samples followed by HBV DNA detection by the nested polymerase chain reaction (PCR). The positive HBV DNA samples were sequenced, and HBV genotypes and HBV subtypes were determined.

**Results:** Three out of 12 (25%) Hodgkin samples and seven out of 29 (24.13%) non-Hodgkin showed positive HBV DNA results. The results of sequencing revealed that the D genotype was predominant among the positive HBV patients. Interestingly an unpredictable amino acid proline was detected in position 88 of the *HBs* gene, which indicates a new mutation in the "S" region of HBV DNA in patients with Hodgkin and non-Hodgkin lymphoma.

**Conclusions:** A high rate of 25% and 24.13% of HBV DNA was detected among patients with Hodgkin and non-Hodgkin lymphoma, respectively.

**Keywords:** Hepatitis B Virus, Hodgkin Disease, Polymerase Chain Reaction

## 1. Background

The association of Hepatitis B Virus (HBV) and hematologic malignancies, including Hodgkin lymphoma and Non-Hodgkin lymphoma (NHL) has been previously studied (1). Hepatitis B Virus causes a wide range of liver diseases, including subclinical infection, acute, chronic and fulminant hepatitis. Chronic HBV may lead to cirrhosis or Hepatocellular Carcinoma (HCC) (2). Hepatitis B Virus is a DNA virus, which belongs to the Hepadnaviridae family (3). The genome of HBV is a partially double stranded circular DNA with molecular weight of 3.2 Kbp. The viral genome encodes four open reading frames (ORFs: S, C, P and X). The ORF S encodes *HBsAg*, which consists of pre-S1, pre-S2 and S regions. The ORF core/pre-core encodes *HBcAg*, and soluble *HBeAg*. The ORF X encodes the X protein (HBX) and is a transactivator that influences the transcription of HBV genes by regulating the activity of transcriptional promoters and ORF P, which encodes DNA

polymerase and reverse transcriptase (4).

The HBV is classified into eight genotypes, A to H (5). Genotypes A and D are prevalent in Europe, United States, and Africa, whereas genotypes B and C are prevalent in Asia. Genotype D is predominant in the Mediterranean, Middle East and India. Genotype E is the dominant genotype in West Africa whereas F and H have been isolated in the America (5). Based on amino acid substitutions in positions 122 and 160 of the S region of HBV DNA, four major subtypes: *adw*, *adr*, *ayw* and *ayr*, and nine minor subtypes have been described (6). Hepatitis B Virus is a hepatotropic virus but recently HBV DNA has been detected in Peripheral Blood Mononuclear Cell (PBMC) of patients with acute and chronic infection (7-9). Some studies have suggested that the HBV DNA could replicate in PBMC and proved this suggestion by detection of HBV mRNAs and expression of *HBsAg* and *HBeAg* in PBMCs (10-15).

These observations show that the lymphoid system could be an important reservoir for Hepatitis B Virus (16). The HBV DNA integration in PBMCs has been demonstrated previously (7, 17-19). Hodgkin lymphoma and non-Hodgkin lymphoma are lymphoid malignant tumors, and are prevalent worldwide. Recent studies have revealed the association between HBV infection and Hodgkin and non-Hodgkin lymphoma (20-22).

## 2. Objectives

The aim of this study was to determine the prevalence of HBV DNA and S region mutations among block samples of patients with Hodgkin and non-Hodgkin lymphoma in Ahvaz city, Iran.

## 3. Materials and Methods

Paraffin embedded tissues of 41 block samples including 12 (29/26%) Hodgkin and 29 (70.73%) non-Hodgkin patients were collected from Imam Khomeini hospital located in Ahvaz, Iran, during 2002 to 2011. The following steps were then carried out for the detection of HBV DNA in the patient samples.

### 3.1. Statistical Analysis

Statistical analysis was done with the Chi square test using the SPSS software, version 20.

### 3.2. DNA Extraction

Initially all the samples were deparaffinized with xylene. The DNA extraction of all the tissue samples was carried out using the high pure PCR template kit (Roche, Germany), according to the manufacturer's instructions.

### 3.3. Nested Polymerase Chain Reaction

The nested PCR for detection of HBV DNA was performed for all the tissue samples. The primers for partial

sequencing of the "S" region are presented in Table 1 (23). For the first round, 5 µL of the extracted DNA from each sample was added to a PCR reaction mixture containing 0.5 µL dNTP (10 Mm), 2.5 µL PCR buffer (10x), 0.15 µL 5U Taq DNA polymerase (Roch, Germany), 50 pmol/µL of the FHBS1 and RHBS1 primers, 0.5 µL dNTP (10 Mm) and 15.85 µL distilled water. The samples were Placed in the thermal cycler (Techne Company, UK) and the first round amplification was carried out with initial denaturation at 94°C for five minutes, followed by denaturation at 94°C for 30 seconds, annealing at 56°C for 30 seconds and extension at 72°C for 30 seconds, for a total of 30 cycles. For the second round, 5 µL of the PCR product was added to 25 µL of reaction mixture containing the same components mentioned in the first run including dNTP, PCR buffer and Taq DNA polymerase with 50 pmol/µL of each of FHBS2 and RHBS2 primers. Amplification was carried out in the thermal cycler with the same program as the first round. Next, 8 µL of the nested PCR product (417 bp) was analyzed by 2% agarose gel electrophoresis.

### 3.4. Sequencing Analysis

Amongst the ten samples positive for HBV DNA, six samples were sequenced (Applied BI Benier Company, South Korea). The sequences of all the six samples were submitted to NCBI and registered in GEN Bank (Table 2 and Figure 1). Hepatitis B Virus sequences were aligned with the HBV reference sequence from Gene Bank using the HBV data-base and online Blast. A phylogenetic tree was constructed using the Mega 5 software. To evaluate the HBs subtypes and S region mutations of the positive samples, (Figure 1) the amino acid substitutions in positions 122 and 160 of the S region of HBV DNA were compared with the HBV reference sequence. Interestingly, the presence of an unpredictable amino acid substitution (L88P) was observed, which indicated a new mutation in the "S" region of HBV DNA in patients with Hodgkin and non-Hodgkin lymphoma, (Figure 2).

**Table 1.** Sequences of Primer Pairs Used For Hepatitis B Virus Polymerase Chain Reaction Detection

Primer Name	Nucleotide Sequence	Nucleotide Region
FHBS1	5'-GAG TCT AGA CTC GTG GTG GAC TTC-3'	244 - 267
RHBS1	5'-CGT GGT GGA CTT CTC TCA ATT TTC-3'	668 - 691
FHBS2	5'-AAA TKG CAC TAG TAA ACT GAG CCA-3'	255 - 278
RHBS2	5'-GCC ARG AGA AAC GGR CTG AGG CCC-3'	648 - 671

**Table 2.** The Profiles of Patients With Hepatitis B Virus Genotype Along With the Accession Numbers

Isolate	Gender	Age	Hodgkin	Non-Hodgkin	Genotype	Blast Identity
KJ398939	F	25	+	--	D1	98%
KJ398940	M	29	+	--	D1	98%
KJ398941	M	54	--	+	D1	98%
KJ398942	M	65	--	+	D1	98%
KJ398943	F	46	--	+	D1	98%
KJ398944	M	53	--	+	D1	98%

## 4. Results

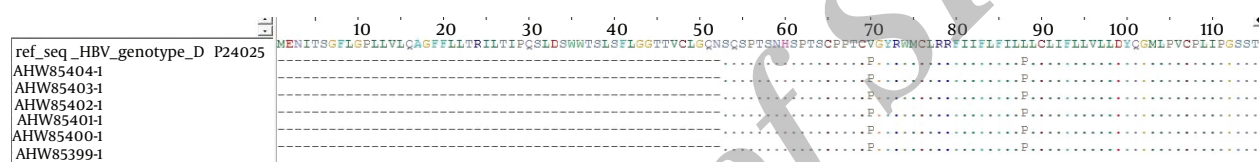
Out of 12 Hodgkin samples, seven (58.33%) and five (41.66%) were males and females, respectively. The age of the patients with Hodgkin lymphoma was between 5 and 34 with a mean age of  $20 \pm 7$  years. Of the 29 Non-Hodgkin samples, 17 (58.62%) and 12 (41.37%) were males and females, respectively, Table 3. Out of the 41 samples, three out of twelve (25%) Hodgkin samples and seven of twenty-nine (24.13%) non-Hodgkin were positive for HBV DNA, Table 4. All the positive HBV patients were within the normal range for Alanine aminotransferase (20 - 32 IU/DL) and Aspartate aminotransferase (18- 27 IU/DL). The youngest Hodgkin patient was a female with 17 years of age and the oldest was a male with 29 years of age. The youngest non-Hodgkin patient with HBV infection was a

male with two years of age and the oldest patient was a female with 75 years of age.

### 4.1. Results of Sequencing

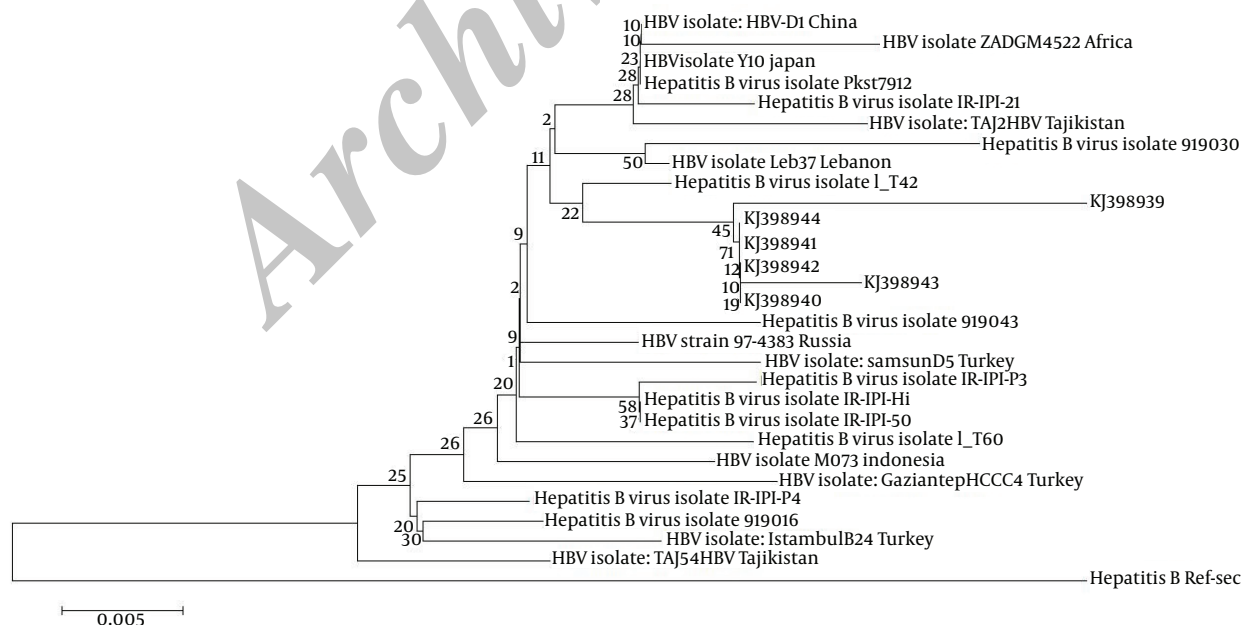
The sequences of six positive samples (two Hodgkin and four non-Hodgkin lymphoma) were registered in Gen-Bank with the following accession numbers, KJ398939.1 to KJ398944.1. The results of the alignment showed that the D genotype was the predominant genotype among the patients with Hepatitis B infection. Mutation analysis indicated amino acid substitutions in positions 88, 122 and 160 of the "S" region of HBV DNA. All of the six sequences were recognized as the "awy" subtype because of substitution in positions 122 and 160.

**Figure 1.** A Phylogenic Tree Constructed With the Neighbor Joining Method and 500 Replication Bootstrap Test Using the Partial Nucleotide Sequences of the S Region of Hepatitis B Virus, Genotype D1



Reference sequences were retrieved from Gen Bank with their accession numbers and origin. Bootstrap values are indicated in the tree roots.

**Figure 2.** Comparison of the Predicted Amino Acid Sequences of the S Region With Proteins of Accession Number AHW85404.1 to AHW85399.1 Recorded in Gen Bank



The consensus amino acid sequence of the reference D genotype is shown on the top. The partial sequences of all the six HBs genes in position 88 compared with the consensus sequence amino acid indicated a proline substitution with leucine. So far this type of mutation has not been reported in any HBV D genotype. The dots show conserved amino acid and the amino acid code indicates differences with the consensus amino acid.

**Table 3.** The Frequency of Hodgkin and Non-Hodgkin Lymphoma

Age Group, y	Males Amongst Hodgkin Patients	Females Amongst Hodgkin Patients	Males Amongst Non-Hodgkin Patients	Females Amongst Non-Hodgkin Patients	Total
< 9	-	1	7	-	8
10 - 19	1	3	-	2	6
20 - 29	5	1	-	2	8
30 - 39	1	-	-	1	2
40 - 49	-	-	1	1	2
50 - 59	-	-	3	2	5
60 - 69	-	-	3	1	4
> 70	-	-	3	3	6
<b>Total</b>	7	5	17	12	41
	12		29		

**Table 4.** Distributions of Hepatitis B Virus Infection Amongst the Hodgkin and Non-Hodgkin Patients

Positive/Negative Cases	Males Amongst Hodgkin Patients	Females Amongst Hodgkin Patients	Males Amongst Non-Hodgkin Patients	Females Amongst Non-Hodgkin Patients	Total
<b>HBV positive<sup>a</sup></b>	2 (16.66)	1 (8.33)	5 (17.24)	2 (6.89)	10
<b>HBV negative<sup>a</sup></b>	6 (50)	3 (25)	12 (41.37)	10 (34.48)	31
<b>Pos/total</b>	3/12		7/29		10/41
<b>Neg/Total</b>	9/12		22/29		31/41

<sup>a</sup>Data are presented as No. (%).

## 5. Discussion

Several epidemiological studies have revealed the persistence of Hepatitis B Virus (HBV) infection amongst patients with Hodgkin (HL) and Non-Hodgkin Lymphoma (NHL). In our study, 25% of Hodgkin and 24.13% of non-Hodgkin patients had positive HBV DNA, which indicates the high rate of HBV infection among these patients. The youngest Hodgkin patient with HBV infection was a female with 17 years of age and the oldest was a male with 29 years of age. The youngest non-Hodgkin patient with HBV infection was a male with two years of age and the oldest was a female with 75 years of age. The frequency of HBV infection among the male and female patients was not significantly different ( $P > 0.05$ ). In our study all the HBV patients had the *awy* subtype.

Distribution of HBsAg subtype has been reported in different regions of the world. Fernández et al. in Spain reported the *ad* and *ay* HBs subtypes amongst 40 chronic HBV patients (24). De Souza et al. from Brazil reported HBsAg subtypes *adw2*, *adw4*, and *ayw* amongst patients with mental problems (25). In our study the protein sequence, of all the six "S" genes, showed mutation at position 88 (L88P). So far this type of mutation has not been reported in any HBV D genotype. In the study conducted by Okamoto et al. a point mutation was found in the S gene of hepatitis B virus for two blood donors carrying a surface antigen with subtype *adyr* or *adwr* (26). All of the HBV pa-

tients had normal SGPT and SGOT tests. The patient's history revealed that they were not tested for HBV and hepatitis C virus and it is not clear whether these patients had chronic HBV or occult HBV infection. However, there are some reports about the association of occult HBV infection and non-Hodgkin lymphoma (27, 28).

The presence of unpredictable amino acid proline at position 88 of HBs gene indicates a new mutation in the "S" region of HBV DNA in patients with Hodgkin and non-Hodgkin lymphoma. In our survey the D1 genotype was dominant in all the HBV patients, which is similar to the report of Marcucci et al. (29). In the present study the role of other viruses including hepatitis C virus in Hodgkin and non-Hodgkin lymphoma was not studied however this requires further investigations (30, 31). There have been reports about the reactivation of Hepatitis B and Hepatitis C virus (HCV) infections during NHL treatment; in these studies it was revealed that the patients who were under chemotherapy showed HBV and HCV reactivation, yet other patients who received lamivudine before chemotherapy didn't show any reactivation for HBV or HCV infection (32-35). In conclusion, a high prevalence of HBV infection (25% and 24.13%, respectively) was found in patients with Hodgkin and non-Hodgkin lymphoma, indicating the importance of HBV and HCV screening by sensitive tests such as PCR be-



fore chemotherapy to prevent HBV or HCV reactivation in patients with HL and NHL.

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## Footnotes

**Authors' Contribution:** Nastaran Ranjbari was responsible for the accuracy of the data and contributed to the design and performance of the study. Manoochehr Makvandi and Niloofar Neisi participated in the laboratory evaluation and performed the literature review. Manoochehr Makvandi drafted the manuscript and was the guarantor. All authors read and approved the final manuscript.

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## References

- Marcucci F, Mele A. Hepatitis viruses and non-Hodgkin lymphoma: epidemiology, mechanisms of tumorigenesis, and therapeutic opportunities. *Blood*. 2011;**117**(6):1792-8. doi: 10.1182/blood-2010-06-275818. [PubMed: 20959600]
- McMahon BJ. The natural history of chronic hepatitis B virus infection. *Hepatology*. 2009;**49**(5 Suppl):S45-55. doi: 10.1002/hep.22898. [PubMed: 19399792]
- Schafer DF, Sorrell MF. Hepatocellular carcinoma. *Lancet*. 1999;**353**(9160):1253-7. doi: 10.1016/S0140-6736(98)09148-X. [PubMed: 10217098]
- Rosner MT. Review: hepatitis B virus X-gene product: a promiscuous transcriptional activator. *J Med Virol*. 1992;**36**(2):101-17. [PubMed: 1583465]
- Khaled IAA, Mahmoud OM, Saleh AF, Baoumi EA. Prevalence of HBV genotypes in Egypt among hepatitis patients. *J Am Sci*. 2010;**6**(11):185-90.
- Couroucé-Pauty AM, Plançon A, Soulier JP. Distribution of HBsAg Subtypes in the World. *Vox Sanguinis*. 1983;**44**(4):197-211. doi: 10.1111/j.1423-0410.1983.tb01885.x. [PubMed: 6845678]
- Bouffard P, Lamelin JP, Zoulim F, Lepot D, Trepo C. Phytohemagglutinin and concanavalin A activate hepatitis B virus in peripheral blood mononuclear cells of patients with chronic hepatitis B virus infection. *J Med Virol*. 1992;**37**(4):255-62. [PubMed: 1402824]
- Calmus Y, Marcellin P, Beaurain G, Chatenoud L, Brechot C. Distribution of hepatitis B virus DNA sequences in different peripheral blood mononuclear cell subsets in HBs antigen-positive and negative patients. *Eur J Clin Invest*. 1994;**24**(8):548-52. [PubMed: 7982442]
- Malave Lara C, Gorrino MT, Campelo C, Lardelli P, Cisterna R. Detection of hepatitis B virus DNA and determination of surface antigen expression in peripheral blood mononuclear cells from patients with AIDS. *Eur J Clin Microbiol Infect Dis*. 1994;**13**(3):267-71. [PubMed: 8050444]
- Yoffe B, Noonan CA, Melnick JL, Hollinger FB. Hepatitis B virus DNA in mononuclear cells and analysis of cell subsets for the presence of replicative intermediates of viral DNA. *J Infect Dis*. 1986;**153**(3):471-7. [PubMed: 3005423]
- Baginski I, Chemin I, Bouffard P, Hantz O, Trepo C. Detection of polyadenylated RNA in hepatitis B virus-infected peripheral blood mononuclear cells by polymerase chain reaction. *J Infect Dis*. 1991;**163**(5):996-1000. [PubMed: 1708401]
- Roisman FR, Castello A, Fainboim H, Morelli A, Fainboim L. Hepatitis B virus antigens in peripheral blood mononuclear cells during the course of viral infection. *Clin Immunol Immunopathol*. 1994;**70**(2):99-103. [PubMed: 8299235]
- Stoll-Becker S, Repp R, Glebe D, Schaefer S, Kreuder J, Kann M, et al. Transcription of hepatitis B virus in peripheral blood mononuclear cells from persistently infected patients. *J Virol*. 1997;**71**(7):5399-407. [PubMed: 9188611]
- Feray C, Zignego AL, Samuel D, Bismuth A, Reynes M, Tiollais P, et al. Persistent hepatitis B virus infection of mononuclear blood cells without concomitant liver infection. The liver transplantation model. *Transplantation*. 1990;**49**(6):1155-8. [PubMed: 2360255]
- Mason A, Yoffe B, Noonan C, Mearns M, Campbell C, Kelley A, et al. Hepatitis B virus DNA in peripheral-blood mononuclear cells in chronic hepatitis B after HBsAg clearance. *Hepatology*. 1992;**16**(1):36-41. [PubMed: 1618481]
- Pontisso P, Vidalino L, Quarta S, Gatta A. Biological and clinical implications of HBV infection in peripheral blood mononuclear cells. *Autoimmun Rev*. 2008;**8**(1):13-7. doi: 10.1016/j.autrev.2008.07.016. [PubMed: 18706529]
- Murakami Y, Minami M, Daimon Y, Okanoue T. Hepatitis B virus DNA in liver, serum, and peripheral blood mononuclear cells after the clearance of serum hepatitis B virus surface antigen. *J Med Virol*. 2004;**72**(2):203-14. doi: 10.1002/jmv.10547. [PubMed: 14695661]
- Umeda M, Martusawa H, Seno H, Katsurada A, Nabeshima M, Egawa H, et al. Hepatitis B virus infection in lymphatic tissues in inactive hepatitis B carriers. *J Hepatol*. 2005;**42**(6):806-12. doi: 10.1016/j.jhep.2005.01.016. [PubMed: 15885350]
- Pontisso P, Poon MC, Tiollais P, Brechot C. Detection of hepatitis B virus DNA in mononuclear blood cells. *BMJ*. 1984;**288**(6430):1563-6. doi: 10.1136/bmj.288.6430.1563. [PubMed: 6426645]
- Kang J, Cho JH, Suh CW, Lee DH, Oh HB, Sohn YH, et al. High prevalence of hepatitis B and hepatitis C virus infections in Korean patients with hematopoietic malignancies. *Ann Hematol*. 2011;**90**(2):159-64. doi: 10.1007/s00277-010-1055-5. [PubMed: 20821327]
- Park SC, Jeong SH, Kim J, Han CJ, Kim YC, Choi KS, et al. High prevalence of hepatitis B virus infection in patients with B-cell non-Hodgkin's lymphoma in Korea. *J Med Virol*. 2008;**80**(6):960-6. doi: 10.1002/jmv.21168. [PubMed: 18428141]
- Nath A, Agarwal R, Malhotra P, Varma S. Prevalence of hepatitis B virus infection in non-Hodgkin lymphoma: a systematic review and meta-analysis. *Intern Med J*. 2010;**40**(9):633-41. doi: 10.1111/j.1445-5994.2009.02060.x. [PubMed: 19815561]
- Sitnik R, Pinho JR, Bertolini DA, Bernardini AP, Da Silva LC, Carilho FJ. Hepatitis B virus genotypes and precore and core mutants in Brazilian patients. *J Clin Microbiol*. 2004;**42**(6):2455-60. doi: 10.1128/JCM.42.6.2455-2460.2004. [PubMed: 15184419]
- Fernandez E, Rodrigo L, Garcia S, Riestra S, Blanco C. Hepatitis B surface antigen detection using pooled sera. A cost-benefit analysis. *Rev Esp Enferm Dig*. 2006;**98**(2):112-21. [PubMed: 16566643]
- de Souza MM, Barbosa MA, Borges AM, Daher RR, Martins RM, Cardoso D. [Seroprevalence of hepatitis B virus infection in patients with mental problems]. *Rev Bras Psiquiatr*. 2004;**26**(1):35-8. [PubMed: 15057838]
- Okamoto H, Imai M, Tsuda F, Tanaka T, Miyakawa Y, Mayumi M. Point mutation in the S gene of hepatitis B virus for a d/y or w/r subtypic change in two blood donors carrying a surface antigen of compound subtype ady or adw. *J Virol*. 1987;**61**(10):3030-4. [PubMed: 3041023]
- Hui CK, Sun J, Au WY, Lie AK, Yueng YH, Zhang HY, et al. Occult hepatitis B virus infection in hematopoietic stem cell donors in a hepatitis B virus endemic area. *J Hepatol*. 2005;**42**(6):813-9. doi: 10.1016/j.jhep.2005.01.018. [PubMed: 15885351]
- Chen MH, Hsiao LT, Chiou TJ, Liu JH, Gau JP, Teng HW, et al. High prevalence of occult hepatitis B virus infection in patients with B cell non-Hodgkin's lymphoma. *Ann Hematol*. 2008;**87**(6):475-80. doi: 10.1007/s00277-008-0469-9. [PubMed: 18327583]

29. Marcucci F, Spada E, Mele A, Caserta CA, Pulsoni A. The association of hepatitis B virus infection with B-cell non-Hodgkin lymphoma - a review. *Am J Blood Res.* 2012;**2**(1):18-28. [PubMed: 22432084]
30. Eftekhari Y, Kazemi Arababadi M, Hakimi H, Rezazadeh Zarandi E. Common HBV genotype in southeastern Iranian patients. *Arch Iran Med.* 2010;**13**(2):147-9. [PubMed: 20187670]
31. Anderson LA, Engels EA. Hepatitis C virus infection and non-Hodgkin lymphoma: interesting association or causal relationship? *Int J Cancer.* 2008;**122**(8):x-xii. doi: 10.1002/ijc.23462. [PubMed: 18271007]
32. Ozguroglu M, Bilici A, Turna H, Serdengeci S. Reactivation of Hepatitis B Virus Infection with Cytotoxic Therapy in Non-Hodgkin's Lymphoma. *Med Oncol.* 2004;**21**(1):67-72. doi: 10.1385/mo:21:1:67. [PubMed: 15034216]
33. Kim JS, Hahn JS, Park SY, Kim Y, Park IH, Lee CK, et al. Long-term outcome after prophylactic lamivudine treatment on hepatitis B virus reactivation in non-Hodgkin's lymphoma. *Yonsei Med J.* 2007;**48**(1):78-89. [PubMed: 17326249]
34. Tsutsumi Y, Tanaka J, Kawamura T, Miura T, Kanamori H, Obara S, et al. Possible efficacy of lamivudine treatment to prevent hepatitis B virus reactivation due to rituximab therapy in a patient with non-Hodgkin's lymphoma. *Ann Hematol.* 2004;**83**(1):58-60. doi: 10.1007/s00277-003-0748-4. [PubMed: 14513286]
35. Jadali Z. Hepatitis C virus cryoglobulinemia and non-hodgkin lymphoma. *Hepat Mon.* 2012;**12**(2):85-91. doi: 10.5812/hepatmon.818. [PubMed: 22509184]

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