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# **Original Article**

# Phytochemical Analysis, GCMS Identification, and Estimation of Antioxidant Activity of Iraqi Vitex negundo L.

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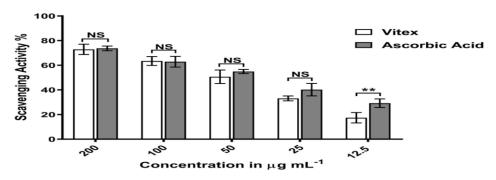
# KEYWORDS

Vitex negundo Linn GC/MS analysis Pinene Sabinene

#### ABSTRACT

Vitex negundo Linn, known as chaste tree, belongs to Verbenaceae family. It is a woody medicinal tree; an aromatic and flowering-ornamental plant has different ethnobotanical and pharmacological uses. The aims of this study included phytochemical analysis, detection of two flavonoids by TLC, GC/MS analysis of hexane extract, and the antioxidant estimation in leaves of Iraqi Vitex negundo activity. The results showed the important phytochemicals in hydroalcoholic and hexane extracts. In addition, the main constituents obtained by GC MS were Sabinene 3.23% and Pinene 0.87% as monoterpenes, alpha-Farnesene 0.5%, Pregnan-3,11-diol-20-one 1.55%, and alpha-Tocopherol 1.55%. Those Iraqi Vitex negundo L. were considered as an important source for chemical constituents with a significant antioxidant activity at low concentration with IC50 26.7, as compared with ascorbic acid.

## GRAPHICALABSTRACT



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#### Introduction

Vitex negundo Linn, known as chaste tree, belongs to Verbenaceae family. It is a woody medicinal tree, native to the Mediterranean regions, while now it is cultivated all over the world as an aromatic and flowering-ornamental plant. Vitex negundo is considered as one of the common species among the genus (vitex), which consists 250 species such as Vitex negundo, Vitex agnuscastu, Vitex trifolia, Vitex polygama, Vitex leucoxylon Linn, Vitex mollis, and Vitex altissima Linn [1]. Vitex tree is characterized as the beautiful little deciduous a medium sized about 3-4 m in high with leaves have approximately 5 leaflets in a hand shape (palmate) arrangement and rounded fruit, black when ripe, with 3-4 mm, while the flower characterized with the aromatic brilliant color varies from violet-deep purple [2]. Furthermore, an ethnobotanical uses of vitex was mainly to reduce breast tenderness, menstrual cycle pain, and amenorrhea symptoms. Likewise, it is regulating the menstrual cycle, regulate hormones related to fertility, and diuretic effects [3]. Most pharmacological effects of this species were as anti-inflammatory, reduce the level of prolactin hyperprolactinemia serum mastodyni, anti-oxidant, **GIT** disturbance, analgesic effects, antimicrobial, and anti-tumor activity [4]. Vitex negundo contains various potentially bioactive phytochemicals such as flavonoids, flavonoids glycosides, phenolic compounds, terpenes, and phyto steroids. Terpenoids are considered as one of the most abundant class of the secondary metabolites in vitex plants. Chemically, they consist of five carbon isoprene units to form the building blocks of hemiterpenes (C5), monoterpenes (ten carbon atoms), sesquiterpenes (fifteen carbon atoms), diterpenes (twenty carbon atoms), triterpenes (thirty carbon atoms) [5]. Moreover, previous literatures revealed that V. negundo contains a large number of terpenes in leaves oil such as caryophyllene epoxide, δ-guaiene, and ethyl-hexadecenoate, while in flowers oil contain  $\alpha$ -selinene, (*E*)-nerolidol, carryophyllene epoxide, and germacren-4-ol [6, 7].

#### **Materials and Methods**

#### Plant's collection

Vitex *negundo* plant was collected in February 2021 from AL-Zawraa Gardens in Iraq. The authentication of plant was done by Prof. Dr. Ibrahim Salih Abass, in Pharmacognocy Department, College of Pharmacy, Mustansiriyah University, Baghdad, Iraq. The leaves's part was dried in shade and ground to a fine powder by mechanical grinder.

# Chemicals

Ethyl acetate, n-hexane, acetic acid, and chloroform (95-97%) were purchased from Merck (Darmstadt, Germany), while the standard kampferol ( $\geq$ 98%), and apigenin ( $\geq$ 97%) were purchased from Hyperchem Company (China) and DPPH (China).

# Extracts preparation

50 g of *V.negundo* leaves was defatted with hexane by Soxhlet for 12 hours, and then the residue was dried and extracted with 90% methanol. Both extracts were concentrated under the reduced pressure and stored in glass containers for further analysis.

# Preliminary phytochemical screening

The hydroalcoholic leaves extract was screened by different chemical tests to investigate the major secondary metabolites in this plant [8-10].

# Screening of Flavonoids

Alkali test depends on using ethanolic KOH (2 mL) mixed with 3 ml of alcoholic extract. The yellow color is indicated the flavonoids presence.

# Screening of tannins

Few drops of crude extract were mixed with 1% aqueous ferric chloride change of dark green to the blue color can predict the tannin presence.

# Screening of saponin

Froth assay was used to screen saponin by shaking distilled water vigorously with 5 mL of crude extract for approximately fifteen minutes.

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A persistent froth (1 cm) can be indicator for the saponins presence.

# Screening of terpenoids

Salkowski test were used as screening of terpenoids confirmed by using 4 ml of the hexane and alcoholic extracts were treated with 4 mL extract was added to 2 mL of chloroform and 2 mL of concentrated sulphuric acid, the positive result gives a reddish- brown color.

# Screening of sterols and steroids

Dried alcoholic and hexane extract were treated with 1.5 mL of the equal quantity of acetic anhydride and chloroform according to Liebermann reaction, and then the drops of concentrated  $H_2SO_4$  was added gradually. The appearance of the green color indicates the sterol presence.

# Screening of reducing sugar

Fehling test (solution I and II) was done by the addition of 20 mL of diluted sulphuric acid ( $H_2SO_4$ ) to 4 mL of the dried extract in a test tube and boil for 10 min, and then it was cooled and 4 mL of Fehling solution and 10% potassium hydroxide were added, the red to blue precipitate was considered as the positive results.

## *Screening of alkaloids*

Approximately few drops of the crude extract were dissolved individually in few volumes of 1% hydrochloric acid, and then it was filtered. After that, the filtrate was reacted with Mayer's reagent; a positive result for alkaloids was indicated by the presence of white precipitate.

# Analytical thin layer chromatography (TLC)

Approximately 2 mg of hydroalcoholic extract were suspended in about 1 mL of absolute methanol, applied on plate of analytical TLC was precoated with a silica gel GF254, and then developed with chloroform-ethyl acetate-acetic acid (5:5:0.1) for detecting the constituents, as compared with the standard.

# GC-MS analysis condition

The GC-MS identification was carried out to the hexane extract. Chromatograph analysis was done on Agelint (7820A) USA Gas Chromatography/ Mass Spectrometer (GC-MS) at the Ministry of Industry and Minerals- Ibn Al-Baitar research center-Baghdad. The conditions were as follow: carrier gas was helium column, the split ratio was 2.0, the column was used Agelint HP-5ms Ultra Inleit (30 m  $\times$  250  $\mu m$   $\times$  0.25  $\mu m$ ), 250-300 °C, column temperature, rate 10 °C/min, and the injection volume 1  $\mu l$ .

# Anti-oxidant analysis

In this work, we used 1,1-Diphenyl-2 picrylhydrazyl; (DPPH assay), using a colorimetric technique defined by Anna Floe gel and coworkers by using a methanolic dilution of DPPH. About 2.95 mL of DPPH solution (approximately 1 Mm DPPH in eighty percent of methanol) was assorted with 0.05 mL of sample or standard (different concentrations ranging from 12.5 to 200 mcg per 1 mL methanol). The assay was completed in triplicate, and the preparing mixture was incubated in the dark condition for about half an hour with aluminum foil paper over it. The absorbance decrease was observed at 517 nm [11, 12].

#### **Result and Discussion**

The phytochemical analysis revealed a positive result for the presence flavonoids, tannins, reducing sugar, terpenoids, steroids, and saponin, while negative result were found for alkaloids.

#### TLC

TLC was applied to detect apigenin and Kaempferol in hydroalcoholic extract, as compared with the standard at 0.5 and 0.7, respectively (Figure 1).

# Hexane extract identification by GC/MS

Analysis and determination of bioactive chemical components identified in leaves of Iraqi Vitex *negundo* hexane extract shows the different components and groups of terpenes and steroids, as displayed in Figure 2. Table 1 presents the major components identified in hexane extract of *Vitex negundo* leaves. The results reveal the

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percentage of the main constituents were Sabinene 3.23% and Pinene 0.87% as monoterpenes, alpha-Farnesene 0.5% as a sesquiterpene, in addition to the other important

terpenes Pregnan-3,11-diol-20-one 1.55% and alpha-Tocopherol 1.55%.

The GC-Mass spectra of the main known chemical constituents are shown in Figure 3.

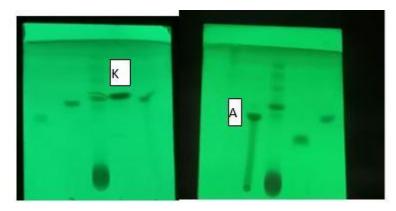


Figure 1: TLC analysis for crude extract compared with apigenin (A) and kaempferol (K) reference standard

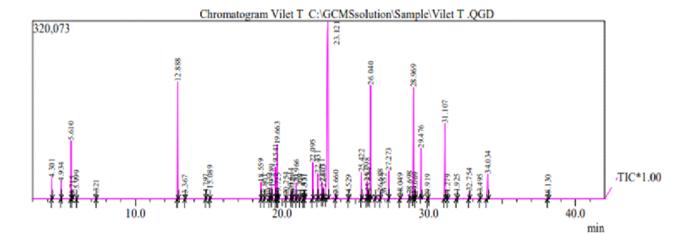


Figure 2: GC/MS chromatogramo of Vitex negundo leaves

| Peak No. | Name                                    | Retention time | Area (%) |
|----------|---|----------------|----------|
| 2        | Pinene                                  | 4.934          | 0.87     |
| 3        | Thujene, Sabinen                        | 5.608          | 3.23     |
| 11       | P-Mentha-[1(7),8]-diene 2-hydroperoxide | 18.558         | 0.07     |
| 14       | (Z,E)-alpha-Farnesene                   | 19.292         | 0.50     |
| 19       | Coprostan-16 betaol                     | 20.617         | 0.41     |
| 25       | Alpha-Cholestan-16-ol                   | 22.092         | 0.09     |
| 32       | Pregnan-3,11-diol-20-one                | 25.425         | 1.55     |
| 33       | 2-Bromocholestanone                     | 25.800         | 1.12     |
| 43       | Alpha-Tocopherol                        | 29.475         | 3.89     |
| 48       | Beta-Amyrene                            | 32.750         | 0.77     |

**Table 1:** The main chemical constituents of *Vitex negundo* leaves

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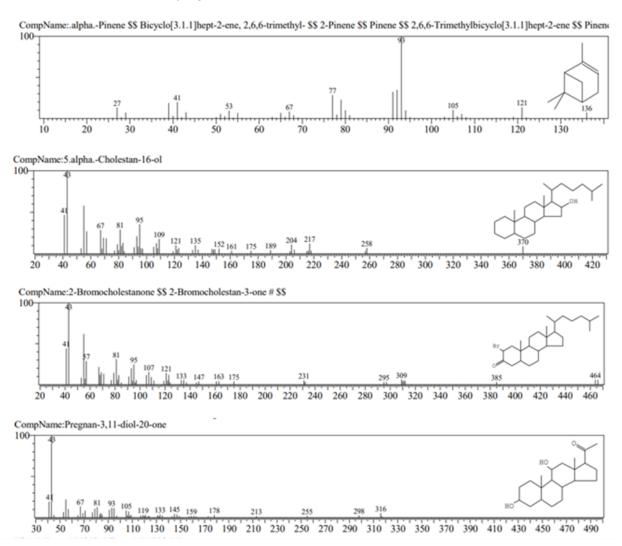


Figure 3: The GC-Mass spectra of the main known chemical constituents in hexane extract

Previous studies reported that major components of essential oils in different vitex species consist of sesquiterpene hydrocarbons mainly was caryophyllene which identified in four vitex species which are: V. limonifolia in Thailand [13], V. polygama, V. rufescens, and V. megapotamica in Brazil [14]. Besides caryophylleme, another presented sesquiterpene were  $\delta$ -cadinene, ciscalamenene, and 6,9-guaiadiene. In addition, the monoterpene hydrocarbons and oxygenated monoterpenes were identified in different regions. The main monoterpene was reported is B-pinene. α-pinene [15],and 1,8-cineole. Moreover, the other terpenoids reported in vitex genus such as abietane-type diterpene, labdanetype diterpenoids, and nor labdane-type

diterpenoids [16], triterpenoids oleanane-type, and Oleanolic acid [17].

#### **Antioxidant**

Vitex negundo leaves extract indicate a good antioxidant activity when compared with ascorbic acid as shown in Figure 4 and Table 2, 3 and 4. This activity may belong to the presence of a good concentration of various terpenoids or terpenes in Iraqi plant of vitex leaves. The reported previous studies reveled the antioxidant activities of the phytochemical components [18-20].

The results show a significant antioxidant activity at a low concentration with IC50 26.7 as compared with ascorbic acid with IC50=34.74.

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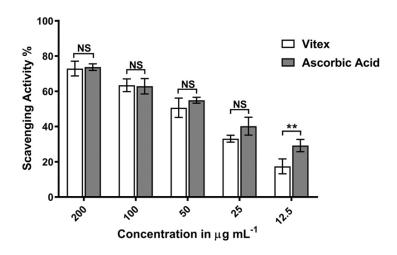


Figure 4: The scavenging activity of Vitex negundo leaves extract and ascorbic acid

**Table 2:** Antioxidant assay of ascorbic acid as a control

|                    | 200   | 100   | 50     | 25    | 12.5  |
|--------------------|-------|-------|--------|-------|-------|
| Number of values   | 3     | 3     | 3      | 3     | 3     |
| Mean               | 73.69 | 62.85 | 54.90  | 40.20 | 29.21 |
| Std. Deviation     | 1.882 | 4.357 | 1.675  | 5.091 | 3.506 |
| Std. Error of Mean | 1.086 | 2.515 | 0.9670 | 2.939 | 2.024 |

**Table 3:** The scavenging activity of *Vitex negundo* leave extract. Bauhinia leaves extract IC50= 26.70

|                    | 200   | 100   | 50    | 25    | 12.5  |
|--------------------|-------|-------|-------|-------|-------|
| Number of values   | 3     | 3     | 3     | 3     | 3     |
| Mean               | 72.88 | 63.43 | 50.62 | 33.10 | 17.40 |
| Std. Deviation     | 4.175 | 3.614 | 5.480 | 1.978 | 4.207 |
| Std. Error of Mean | 2.410 | 2.086 | 3.164 | 1.142 | 2.429 |

Table 4: The relation of antioxidant activity results of Vitex negundo leave extract compared with ascorbic acid

| Sidak's multiple comparisons test | Significant | Summary | Adjusted P Value |
|-----------------------------------|-------------|---------|------------------|
| 200                               | No          | ns      | 0.9997           |
| 100                               | No          | ns      | >0.9999          |
| 50                                | No          | ns      | 0.6395           |
| 25                                | No          | ns      | 0.1582           |
| 12.5                              | Yes         | **      | 0.0058           |

#### **Conclusion**

Those Iraqi *Vitex negundo* L. were considered as an important source for chemical constituents with a significant antioxidant activity at low concentration with IC50 26.7, as compared with ascorbic acid.

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# **Authors' contributions**

All authors contributed to data analysis, drafting, and revising of the paper and agreed to be responsible for all the aspects of this work.

#### **Conflict of Interest**

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The author declared that they have no conflict of interest.

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#### References

- [1]. Borji M., Moradi M., Otaghi M., Tartjoman A., Relationship between Nutritional Status, Food Insecurity, and Causes of Hospitalization of Children with Infectious Diseases, *Journal of Comprehensive Pediatrics*, 2018, **9**:e63870 [Crossref], [Google Scholar], [Publisher]
- [2]. Tawfeeq T.A., Jasim G.A., Nasser A.A., Al-Sudani B.T., Isolation of Lupeol and Gallic acid with cytotoxic activity of two different extracts from the leaves of Iraqi Conocarpus erectus L., Research Journal of Pharmacy and Technology, 2021, 14:3495 [Crossref], [Google Scholar], [Publisher]
- [3]. Das N., Salgueiro A.C., Choudhury D.R., Mandal S.K., Logesh R., Hassan M.M., Devkota H.P., Traditional uses, phytochemistry, and pharmacology of genus Vitex (Lamiaceae), *Phytotherapy Research*, 2022, **36**:571 [Crossref], [Google Scholar], [Publisher]
- [4]. Ben Nouri A., Dhifi W., Bellili S., Ghazghazi H., Aouadhi C., Chérif A., Hammami M., Mnif W., Chemical composition, antioxidant potential, and antibacterial activity of essential oil cones of Tunisian Cupressus sempervirens, *Journal of Chemistry*, 2015, **2015**:538929 [Crossref], [Google Scholar], [Publisher]
- [5]. Tawfeeq A.A., Mahdi M.F., Abaas I.S., Alwan A.H., Isolation, quantification, and identification of rosmarinic acid, gas chromatography-mass spectrometry analysis of essential oil, cytotoxic effect, and antimicrobial investigation of rosmarinus officinalis leaves, *Asian Journal of Pharmaceutical and Clinical Research*, 2018, 11 [Crossref], [Google Scholar], [Publisher]
- [6]. Hajdú Z., Hohmann J., Forgo P., Martinek T., Dervarics M., Zupkó I., Falkay G., Cossuta D., Máthé I., Diterpenoids and flavonoids from the fruits of Vitex agnus-castus and antioxidant activity of the fruit extracts and their constituents, *Phytotherapy Research: An*

- International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives, 2007, **21**:391 [Crossref], [Google Scholar], [Publisher]
- [7]. Yao J.L., Fang S.M., Liu R., Oppong M.B., Liu E.W., Fan G.W., Zhang H., A review on the terpenes from genus Vitex, *Molecules*, 2016, **21**:1179 [Crossref], [Google Scholar], [Publisher] [8]. Tawfeeq A.A., Mahdi M.F., Abaas I.S., Alwan A.H., Phytochemical and anti bacterial studies of leaves of Rosmarinus officinalis cultivated in Karbala, Iraq, *Al Mustansiriyah Journal of Pharmaceutical Sciences*, 2017, **17**:9 [Crossref], [Google Scholar], [Publisher]
- [9]. Izadi-Ajeerlo B., Bastaminejad S., Basati G., Upregulated expression of the growth arrest-specific-2 (gas2) gene in colorectal cancer, and its relation to cancer progression and prognosis, *Journal of Isfahan Medical School*, 2019, **37**:93 [Crossref], [Google Scholar], [Publisher]
- [10]. Mushtaq M., Peerzada S., Ishtiaq S., Pharmacognostic Features, Preliminary Phytochemical Screening and in Vitro Antioxidant Studies of Indigenous Plant of Lahore, International Journal of Advanced Biological and Biomedical Research, 2021, 9:204 [Crossref], [Google Scholar], [Publisher]
- [11]. Sreedevi A., Sangeetha S., Murali Mohan Achari K., Sruthi K., Vadlamudi Y., 'Phytochemical, in vitro and in silico screening of roots of Jasminum auriculatum for antioxidant activity', *Eurasian Chemical Communications*, 2022, **4**:768 [Crossref], [Google Scholar], [Publisher]
- [12]. Ukwubile C., Idriss U., Isah A., Phytochemical evaluation, in vitro-in vivo antioxidant and cytotoxicity activities of various layers of watermelon fruit Citrullus lanatus (Cucurbitaceae) Matsum. & Nakai. *Progress in Chemical and Biochemical Research*, 2022, 5:97 [Crossref], [Google Scholar], [Publisher]
- [13]. Suksamrarn A., Aphaijitt S., Brophy J.J., The volatile leaf oil of Vitex limonifolia Wall, *Flavour and fragrance journal*, 1990, **5**:53 [Crossref], [Google Scholar], [Publisher]
- [14]. Tarjoman A., Vasigh A., Safari S., Borji M., Pain management in neonatal intensive care units: A cross sectional study of neonatal nurses

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Tawfeeg T.A., et al. / J. Med. Chem. Sci. 2023, 6(4) 876-883

in Ilam City, *Journal of Neonatal Nursing*, 2019, **25**:136 [Crossref], [Google Scholar], [Publisher] [15]. Dai D.N., Thang T.D., Ogunwande I.A., Lawal O.A., Study on essential oils from the leaves of two Vietnamese plants: Jasminum subtriplinerve CL Blume and Vitex quinata (Lour) FN Williams, *Natural product research*, 2016, **30**:860

[16]. Ono M., Yanaka T., Yamamoto M., Ito Y., Nohara T., New diterpenes and norditerpenes from the fruits of Vitex rotundifolia, *Journal of Natural Products*, 2002, **65**:537 [Crossref], [Google Scholar], [Publisher]

[Crossref], [Google Scholar], [Publisher]

[17]. Adjei S., Amponsah I.K., Bekoe S.O., Harley B.K., Mensah K.B., Mensah A.Y., Baah M.K., Fosu-Mensah G., Fruits of Vitex doniana Sweet: Toxicity profile, anti-inflammatory and antioxidant activities, and quantification of one of its bioactive constituents oleanolic acid, *Heliyon*,

2021, **7**:e07910 [Crossref], [Google Scholar], [Publisher]

[18]. Eldalawy R., Naser N.M., Naqqash Z.A.A., Antimicrobial and antioxidant activity of Iraqi cupressuss empervirens cones, *AIP Conference Proceedings*, 2020, **2213**:020006 [Crossref], [Google Scholar], [Publisher]

[19]. Tian C., Liu X., Chang Y., Wang R., Lv T., Cui C., Liu M., Investigation of the anti-inflammatory and antioxidant activities of luteolin, kaempferol, apigenin and quercetin, *South African Journal of Botany*, 2021, **137**:257 [Crossref], [Google Scholar], [Publisher]

[20]. Ahmadi F.I., Fathollahi R., Dastan D., Phytochemical constituents and biological properties of Scutellaria condensata subsp. pycnotricha, *Journal of Applied Organometallic Chemistry*, 2022, **2**:119 [Crossref], [Google Scholar], [Publisher]

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