

FULL PAPER

The effect of secretome injection on tendon healing of rotator cuff tear on new zealand white rabbit through the expression of tgf- β , scleraxis, collagen type iii, and histological evaluation

Baskoro Kusumo Riswanto  | Cery Tarise Hajali  | Teddy Heri Wardhana* 

Department of Orthopedic and Traumatology,
Faculty of Medicine, Universitas Airlangga, Dr.
Soetomo General Academic Hospital, Surabaya,
Indonesia

Rotator cuff tear is the major cause of morbidity which affects over 40% of patients aged 60 years and older. Surgical repair is often unsuccessful and has high complications. Tissue engineering therapies, such as applying Mesenchymal stem cells (MSCs) secretomes, have shown beneficial effects on tendon repair. This study aims to assess the effect of secretome application on rotator cuff tendon tears by examining changes in transforming growth factor beta (TGF- β), Scleraxis, and Collagen type III levels. A total of 20 post-injured supraspinatus tendon New Zealand white rabbits were divided into 2 groups: injected with of frozen-dried tenogenic secretome (treatment group) and no treatment (control group). Histologic evaluation on the repaired site was conducted on the second and the fourth weeks; analysing immunohistochemistry (IHC), TGF- β , Scleraxis, collagen type III levels, modified tendon maturing score, blood vessel, fibroblast, and fibrotic tissue. Immunoreactive Score (IRS) result for TGF- β , Scleraxis, and collagen type III on treatment group were moderate. Meanwhile, IRS result for control group was a mix of mild and moderate. Compared to the control group, a significant difference was found ($p < 0.05$). Histologically, Modified Tendon Maturing Score, amount of blood vessel, and fibroblast were found to be significantly higher ($p < 0.05$) in treatment group. Meanwhile, fibrotic tissues were significantly lower ($p < 0.05$). Secretomes promote the healing of specific tissues such as tendons in vitro and in vivo. TGF- β is crucial for collagen synthesis, while scleraxis facilitates tendon-bone attachment. Collagen type III initiates tendon repair, requiring increased blood vessel and fibroblast growth and lower fibrotic tissue.

***Corresponding Author:**

Teddy Heri Wardhana

Email: teddy-heri-w@fk.unair.ac.id

Tel.: + 62 817-0336-2793

KEYWORDSMSCs secretomes; TGF- β ; Scleraxis; collagen type III.**Introduction**

Rotator cuff tendon tears are a significant cause of pain, reduced shoulder function, and weakness, affecting over 40% of patients over 60 years old. In the United States, approximately 30,000 to 75,000 tendon repairs are performed annually [1]. Tendons

are difficult to heal due to their hypo-vascular nature, leading to scar tissue and weaker ectopic bone [2]. Despite the development of surgical techniques for tendon repair, failure, and complications rates remain high even after therapy [3].

Tissue engineering strategies have been developed to create functional replacements

or accelerate natural healing of tendon defects. Applying stem cells directly to the injured site provides new precursor cells that proliferate and differentiate into new cells - but this has limitations, including low concentration of cells in the injured site and the long process of making the composite [4]. Recent studies are prioritizing cell therapy using mesenchymal stem cell (MSCs) secretomes instead of providing the stem cell itself to the injured site. Secretome MSCs significantly reduce the number of cells needed for transplantation and increase the ratio of production quantities through dynamic control of laboratory conditions (bioreactor). This allows for phenotypic adjustment and therapeutic potential [5,6]. Several bioactive molecules play a crucial role in coordinating cellular responses during tendon healing. Growth factors like insulin-like growth factor-I (IGF-I), Transforming growth factor beta (TGF- β), basic fibroblast growth factor (bFGF), platelet-derived growth factor (PDGF), vascular endothelial growth factor (VEGF), Bone morphogenetic proteins (BMP), Scleraxis (Scx), collagens, and connective tissue growth factor (CTGF) contribute to different stages of healing process. Other bioactive molecules, such as growth regulators and extracellular matrix (ECM) degradation enzymes like matrix metalloproteinases (MMPs) and their inhibitors- tissue inhibitor metalloproteinases (TIMPs), are also essential for tendon repair [7].

Despite the secretomes potential for tendon healing, their specific role remains unelicited. This research aims to assess the effect of secretome application on rotator tendon cuff tear by examining changes in TGF- β , scleraxis, collagen type III Levels, and histological evaluation.

Materials and methods

This study was a true experimental with a randomized post-test only with a control

group design. In this study we divided the sample into two groups, namely the control group (not injected with freeze-dried tenogenic secretome) and treatment groups (injected with freeze-dried tenogenic secretome). Grouping is decided by randomization using lottery. Each rabbit was given a number, and then a draw was carried out for each group with a total of two groups. All animal procedures were in accordance with The Animal Care and use Committee of Veterinary Faculty Medicine of Airlangga University (Ethical Clearance No. 2.KEH.151.09.2023). The study subjects were the injured supraspinatus tendon of adult New Zealand white rabbits. To fit the sample, the rabbit must meet the inclusion criteria of male sex, adult aged (6-9 months), have an average weight of 2000 ± 500 grams, no body defect, and is healthy yet active. Meanwhile, rabbits which are aged less than 6 months or more than 9 months and rabbits which suffered from illness or other infections were excluded from this study. Briefly speaking, tendon tear was created on the right shoulder surgically on week 0. After that, we repaired the torn supraspinatus tendon using size no. 8-0 end-to-end, and then the freeze dried tenogenic secretomes were injected into the tendons for the treatment group and repaired the tendons. Four weeks after injection, the rabbits were terminated and the supraspinatus tendons were taken to make histological preparations. The minimum number of samples was calculated using statistical formula and it was found that a minimum of 10 samples was required for each group. Histologic evaluation on the repaired site was conducted on the second and fourth week. Modified Tendon Maturing Score, amount of fibroblast, amount of blood vessel, and fibrotic tissue were noted. The scleraxis level, TGF- β , and collagen type III would also be analysed from each sample four weeks after treatment. The collected data will be analysed statistically using the SPSS 26 program. Normality is firstly tested

using Saphiro-Wilk test. If normal, independent t-test will be used to test for significance. Mann-Whitney is used as alternative non-parametric test. $p < 0.05$ will be accepted as significant. In this study, bivariate ordinal data were obtained so that a test would be carried out using the Mann-Whitney.

Rotator cuff tear preparation

- Rabbits were imported from breeders who understood animal health. Rabbits with a body weight of 2000-2500 grams were used. They were placed in cages measuring 50 x 70 cm in a special room for rabbit placement. The rabbits were acclimated to the experimental animal laboratory's ambient conditions for a period of 7-10 days. They were provided with unlimited access to water using the ad libitum method, and their daily food intake consisted of 300 grammes of pellets. The rabbit excrement is cleaned each morning. Infirm rabbits were precluded from being placed in the enclosure. Nevertheless, in the case of a slight sickness, the rabbit will receive either vitamins or antiparasitic medication.

- The rabbits were administered ketamine (40 mg/kg) and xylazine (10 mg/kg) intramuscularly in the supraspinatus area for anaesthesia induction. Subsequently, a maintenance dose of 10 mg/kg was administered intramuscularly if the rabbits exhibited a reaction. Subsequently, the rabbits underwent a period of fasting lasting up to 6 hours after the surgery. The rabbit's forelimbs were depilated using a razor, leaving a 5 cm space from the operation field. The treated region was disinfected with a 10% solution of povidone iodine, followed by the use of a sterile serviette to restrict the operative field.

- The area to be treated for the injury model is at the base of the hind limbs of the rabbit, which is 1 cm proximal to the muscle-tendon junction of the supraspinatus muscle.

An incision is made in the skin and deepened layer by layer until it reaches the muscle layer. Tendon excision was carried out with a length of 1cm, a width of 0.5 cm, and a depth of 0.5 cm.

- As a marker, a knot is given using a nonabsorbable monofilament thread measuring 3.0 - 5 mm proximal and distal from the incision site to facilitate the identification of the incision site when observing later. An incision was made in the tendon of the right forelimb to be given the secretome injection treatment, and then the rabbits were taken care at the Faculty of Veterinary Medicine Airlangga University. The freeze-dried tenogenic secretome was isolated and made at the cell tissue bank of Dr. Soetomo General Academic Hospital.

- Sampling of muscle tissue in the treatment area was carried out at the end of the fourth week according to the division of groups that had been determined sterilely. Network retrieval is carried out between the two marker nodes that have been previously given. The collected tissue is put into a container that has been labelled and filled with formalin buffer.

Analysis of scleraxis, TGF- β , and collagen type III through immunohistochemical analysis

The glass objects that had been deparaffinized were then stained using IHC reagents for respective parameter: scleraxis, TGF- β or collagen type III. The glass objects for each parameter were stained using antibodies and then rinsed using Phosphate-buffered saline (PBS) for 3x5 minutes. The object glass was counterstained using 4',6-diamidino-2-phenylindole (DAPI), and then incubated for 10 minutes and then rinsed again with PBS for 3x5 minutes. The amount of scleraxis, TGF- β or collagen type III was evaluated under the microscope by estimating the intensity and averaging the scleraxis percent of, TGF- β or collagen type III clusters that were visible in the six evaluated

segments. Result is classified based on immunoreactivity score (IRS) classification into mild, moderate, and strongly positive.

Analysis of modified tendon maturing score, amount of fibroblast, amount of blood vessel, and fibrotic tissue

Analysis of Modified Tendon Maturing Score was done based on previous scoring method [7]. Meanwhile, amount of fibroblast, blood vessel, and fibrotic tissues were done by manual calculation counting on the detectable fibroblast, blood vessel, and fibrotic tissue on

histological preparation stained with haematoxylin eosin. Calculation was based on the average of countable fibroblast, blood vessel, or fibrotic tissue on at least five viewing fields.

Results and discussion

Evaluation result of collagen type III, TGF- β , and scleraxis for the control and treatment groups are elaborated. At glance, it appears that tissue exposed to tenogenic secretomes shows increased expression in all three parameters (Figure 1).

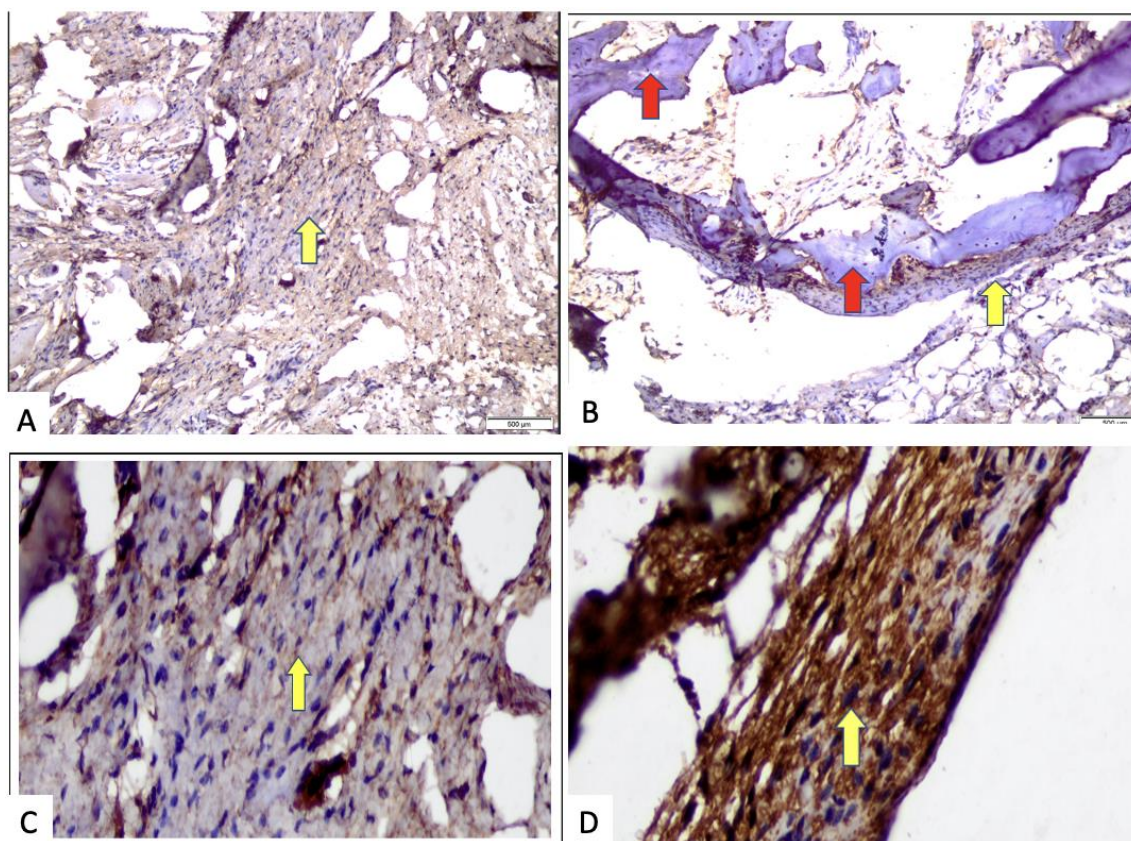


FIGURE 1 Image of IHC staining results. Tendon fibroblast cells (yellow arrow), bone trabeculae (red arrow) are seen on the slide, **Image A** shows the slide with 100x magnification of the Control Group which has light intensity in the cytoplasm of tendon fibroblast cells, **Image B** demonstrates a slide with 100x magnification of the Treatment Group which has moderate intensity in the cytoplasm of tendon fibroblast cells, **Image C** shows the slide results with 400x magnification of the Control Group which has a light intensity in the cytoplasm of tendon fibroblast cells, and **Image D** shows the slide with 400x magnification of the Treatment Group which has moderate intensity in the cytoplasm of tendon fibroblast cells

Based on the IRS, staining is interpreted as negative for IRS 0-1, positive with weak expression for IRS 2-3, positive with moderate expression for IRS 4-8, and positive with strong expression for IRS 9-12. In addition, the IRS classification provides further insight, where IRS 0 is considered negative, IRS 1 indicates positive with weak expression, IRS 2 indicates positive with moderate expression, and IRS 3 indicates positive with strong expression. Overall, this

analysis provides an in-depth understanding of the presence and intensity of the intended marker in the cells or tissues examined through immunohistochemical staining.

Both in terms of percentage of cell expression and the intensity of the expression of the parameters in question. As a result, the results of the IRS classification also showed better results than the control group (Tables 1 and 2).

TABLE 1 IRS result of collagen type III, TGF-β, and Scleraxis on Control Group

Sample No.	Evaluation Result of Control Group		
	Collagen type III	TGF-β	Scleraxis
1	Moderate	Moderate	Moderate
2	Mild	Moderate	Mild
3	Moderate	Moderate	Mild
4	Mild	Moderate	Mild
5	Mild	Moderate	Mild
6	Mild	Moderate	Mild
7	Moderate	Moderate	Mild
8	Mild	Mild	Moderate
9	Moderate	Moderate	Mild
10	Moderate	Mild	Moderate

Note: IRS 0 is considered negative, IRS 1 indicates positive with weak expression, IRS 2 indicates positive with moderate expression, and IRS 3 indicates positive with strong expression.

TABLE 2 IRS Result of Collagen type III, TGF-β, and Scleraxis on Treatment Group

Sample No.	Evaluation Result of Treatment Group		
	Collagen type III	TGF-β	Scleraxis
1	Moderate	Moderate	Moderate
2	Moderate	Moderate	Moderate
3	Moderate	Moderate	Moderate
4	Moderate	Moderate	Moderate
5	Moderate	Moderate	Moderate
7	Moderate	Moderate	Moderate
8	Moderate	Moderate	Moderate
9	Moderate	Moderate	Moderate
10	Moderate	Moderate	Moderate

Statistical tests were carried out for three variables to test whether there were significant differences between the control group and the treatment group. It was found that there were significant differences in the three parameters between the two groups (p<0.05) (Table 3).

TABLE 3 Test statistic result on three parameters

Item	Difference (p)
Collagen type III	<0.05
TGF- β	<0.05
Scleraxis	<0.05

Detailed result of histological analysis was detailed on Table 4. From histological analysis, evaluation of tendon healing with Modified Tendon Maturing Score has significant difference in the 2-week activity comparing the control and treatment groups

($p < 0.05$). Histological analysis also found significant difference in number of fibroblasts, blood vessel, and fibrotic tissue ($p < 0.05$) indicating a tangible increase in necessary tissue for tendon healing (Table 4).

TABLE 4 Histological analysis result

Item	Control Group	Treatment Group	Difference (p)
Modified Tendon Maturing Score	8.9 \pm 2.23	17.3 \pm 1.57	<0.05
Blood Vessel amount	6.08 \pm 2.79	14.61 \pm 1.25	<0.05
Fibroblast amount	9.27 \pm 0.29	42.64 \pm 2.75	<0.05
Fibrotic tissue	3.41 \pm 0.14	1.38 \pm 0.14	<0.05

The progress in tissue engineering technologies enables the production and analysis of a secretome, which has the ability to stimulate the development of particular tissues like tendons both in laboratory settings (*in vitro*) and in living organisms (*in vivo*) [9]. This Secretome is referred to as the tenogenic secretome. Research has demonstrated that the secretome derived from tendons can promote the growth and multiplication of tendon stem cells. Furthermore, it was discovered that the secretome obtained from tendons has a greater ability to sustain its tenogenic phenotype compared to tissue culture plastic alone and matrices derived from bone or skin [10]. Studies have demonstrated that electrospun aligned Poly L-Lactic Acid (PLLA) fibres enhance the development of spindle-shaped cells, significantly increase the expression of tendon-specific genes (such as collagen type I, collagen type III, collagen type XIV, scleraxis, and elastin), and decrease the expression of osteogenic genes (such as osteocalcin,

alkaline phosphatase (ALP), RUNX2), even when exposed to osteogenic media[11].

Several studies have evaluated the regeneration capabilities of the tenogenic secretome, either by itself or in conjunction with a carrier system, in preclinical models including small animals [9-11]. The secretome of mesenchymal stem cells (MSCs) contains various cytokines, chemokines, growth factors, anti-inflammatory factors, and proteins that have potential applications in various diseases. MSCs release TGF- β , a key role in immunomodulation, and produce growth factors like VEGF, HGF-1, and LIF. The galectin network mediates the immunomodulatory effects of MSCs. Extracellular Vesicles (EVs) are a complex communication system that affects recipient cells' functions. They contain 20S proteasome members and regulate cell stability and translation of proteins. MSC-EVs have immunoregulatory and regenerative properties, with miRNAs playing a role in cell cycle regulation and apoptosis [12].

Nevertheless, there is currently a lack of study about preclinical models involving large animals and clinical trial trials. Furthermore, the reparative capacity of pre-treatment tenogenic secretomes has only been evaluated in two trials. The application of Connective Tissue Growth Factor (CTGF) and ascorbic acid to the tenogenic secretome of the rat patellar tendon induces the development of tendon-like tissue in a mouse patella tendon defect model. This is supported by the observed upregulation of tenomodulin, collagen type I, and collagen type III, as well as the downregulation of collagen type II and osteocalcin [13]. Human tenogenic secretomes, when exposed to 5% oxygen tension and applied to Matrigel™, led to the development of adipose, cartilage-like, and bone-like tissues. However, when placed into a matrix produced from tendons, they specifically formed tendon-like tissues in living organisms [14].

TGF- β is an important regulator in tendon healing [7,15-16]. TGF- β affects cell function and gene expression primarily through activation of the Smad signalling pathway. In the context of tendon healing, TGF- β has an important role in collagen synthesis [17]. During the tendon healing phase after an injury, the more elastic collagen type III is initially expressed, and then slowly remodelled into type 1 [18]. The absence of TGF- β interferes with this remodelling process and thus of course collagen type III. As a result, tendons formed under conditions of TGF- β deficiency will have lower tensile strength. Scx has been found to promote attachment between tendon and bone through its important role in tendogenesis which is also important for enthesis repair [19,20-21]. Deficiencies in Scx expression alone can interfere with enthesis development. Specifically, Scx plays a role in inhibiting the formation of osteoclasts by secreting miR-6924-5p-rich exosomes. These exosomes will eventually strengthen the end result of tendon healing [22,23]. Scx as a

transcription factor most likely prevents osteoclast formation by regulating the expression of target genes by binding to genetic loci [24]. However, the remaining details of how Scx can affect the inhibition of osteoclast formation are unknown [25]. In addition, Scx is able to affect protein expression from mesenchymal cells and also exosomes which in turn prevents osteoclastogenesis. Collagen type III is essential for fibrillogenesis in tendon tissues, with high levels observed in basic medium and culture medium supplemented with TGF- β . Collagen type III expression decreased by 14 days and increased again by 21 days. Scleraxis, a transcription factor, was upregulated by 7 days in basic and EGF media, especially in TGF- β , and increased in EGF supplemented medium by 14 and 21 days [26]. TGF- β 2 treatment significantly upregulated collagen production, cytokines, and growth factors which are relevant to tendon healing [27].

Modified Tendon Maturing Score (MTMS) has an important role in evaluating the regeneration of tendon from histological perspective. This instrument measures the number of cells, the amount of fibrosis, vascularity, parallel cells, and insertions. Healing of tendons is characterized by the formation of scar tissue which can be functional or non-functional. If it is non-functional, the scar tissue can cause tendon injury again at a later time, even causing instability and changes in joint function. Scar tissue deposition usually interferes with the tendon from gliding and moving. This is most visible in tendons covered by a synovial layer, such as in zone II of the hand flexor tendon system. Therefore, we ideally seek for decrease in fibrotic tissues but increase in blood vessel and fibroblast [28].

In this study, it was found that there was an increase in TGF- β , collagen type III, and Scleraxis compared to the control group. Moreover, histologically there is evidence of increase in blood vessel and fibroblast along

with decrease in fibrotic tissue. This shows that the administration of tenogenic secretomes is indeed able to induce tendons regeneration indicated by the increased factors that support the regeneration of the tendons. Collagen type III as the progenitor of more mature collagen was also found to increase, indicating that the administration of tenogenic secretomes succeeded in encouraging the collagen formation in damaged tissue in the sample.

Conclusion

The secretome of mesenchymal stem cell (MSCs) contains various cytokines, chemokines, growth factor, anti-inflammatory factors, and protein that have potential as an adjuvant therapy in various diseases. The ability of human tenogenic secretomes to form specific tissues under different condition suggests the potential for future tailored tissue engineering and regenerative medicine approaches for tendon injuries and related conditions.

Acknowledgements

We would like to express our gratitude to the Orthopedic and Traumatology Department of Dr. Soetomo General Academic Hospital (Surabaya, Indonesia) for supporting this study and Cell and Tissue Bank Dr Soetomo General Hospital staff for their kind and dedicated assistance during the preparation of the secretome.

Conflict of Interest

This research has no conflict of interest.

Orcid:

Baskoro Kusumo Riswanto:

<https://orcid.org/0009-0003-2422-9854>

Cery Tarise Hajali:

<https://orcid.org/0000-0001-5109-2716>

Teddy Heri Wardhana*:

<https://orcid.org/0000-0002-6936-8730>

References

- [1] F. Familiari, O. Galasso, F. Massazza, M. Mercurio, H. Fox, U. Srikumaran, G. Gasparini, Artificial intelligence in the management of rotator cuff tears, *International Journal of Environmental Research and Public Health*, **2022**, *19*, 16779. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [2] C. Loiacono, S. Palermi, B. Massa, I. Belviso, V. Romano, A. Di Gregorio, F. Sirico, A.M. Sacco, Tendinopathy: pathophysiology, therapeutic options, and role of nutraceuticals. A narrative literature review, *Medicina*, **2019**, *55*, 447. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [3] S.W. Chung, J.Y. Kim, M.H. Kim, S.H. Kim, J.H. Oh, Arthroscopic repair of massive rotator cuff tears: outcome and analysis of factors associated with healing failure or poor postoperative function, *The American Journal of Sports Medicine*, **2013**, *41*, 1674-1683. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [4] S. Wong, A. Ning, C. Lee, B.T. Feeley, Return to sport after muscle injury, *Current Reviews in Musculoskeletal Medicine*, **2015**, *8*, 168-175. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [5] J. Phelps, A. Sanati-Nezhad, M. Ungrin, N.A. Duncan, A. Sen, Bioprocessing of mesenchymal stem cells and their derivatives: toward cell-free therapeutics, *Stem Cells International*, **2018**, 2018. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [6] J.R. Ferreira, G.Q. Teixeira, S.G. Santos, M.A. Barbosa, G. Almeida-Porada, R.M. Gonçalves, Mesenchymal stromal cell secretome: influencing therapeutic potential by cellular pre-conditioning, *Frontiers in Immunology*, **2018**, *9*, 2837. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [7] T. Molloy, Y. Wang, G.A. Murrell, The roles of growth factors in tendon and ligament healing, *Sports Medicine*, **2003**, *33*, 381-394. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [8] H. Yurie, R. Ikeguchi, T. Aoyama, M. Tanaka, H. Oda, H. Takeuchi, S. Mitsuzawa, M.

- Ando, K. Yoshimoto, T. Noguchi, S. Akieda, Bio 3D conduits derived from bone marrow stromal cells promote peripheral nerve regeneration, *Cell Transplantation*, **2020**, *29*, 963689720951551. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [9] J. Zhang, B. Li, J.H. Wang, The role of engineered tendon matrix in the stemness of tendon stem cells in vitro and the promotion of tendon-like tissue formation in vivo, *Biomaterials*, **2011**, *32*, 6972-6981. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [10] Z. Yin, X. Chen, J.L. Chen, W.L. Shen, T.M.H. Nguyen, L. Gao, H.W. Ouyang, The regulation of tendon stem cell differentiation by the alignment of nanofibers, *Biomaterials*, **2010**, *31*, 2163-2175. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [11] J. Zhang, J.H. Wang, Characterization of differential properties of rabbit tendon stem cells and tenocytes. *BMC musculoskeletal disorders*, **2010**, *11*, 1-11. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [12] S. Eleuteri, A. Fierabracci, Insights into the secretome of mesenchymal stem cells and its potential applications, *International Journal of Molecular Sciences*, **2019**, *20*, 4597. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [13] M. Ni, Y.F. Rui, Q. Tan, Y. Liu, L.L. Xu, K.M. Chan, Y. Wang, G. Li, Engineered scaffold-free tendon tissue produced by tendon-derived stem cells, *Biomaterials*, **2013**, *34*, 2024-2037. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [14] J. Zhang, J.H.C. Wang, Human tendon stem cells better maintain their stemness in hypoxic culture conditions, *PloS one*, **2013**, *8*, 61424. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [15] M. Ngo, H. Pham, M.T. Longaker, J. Chang, Differential expression of transforming growth factor- β receptors in a rabbit zone II flexor tendon wound healing model, *Plastic and Reconstructive Surgery*, **2001**, *108*, 1260-1267. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [16] J. Chang, D. Most, E. Stelnicki, J.W. Siebert, M.T. Longaker, K. Hui, W.C. Lineaweaver, Gene expression of transforming growth factor beta-1 in rabbit zone II flexor tendon wound healing: evidence for dual mechanisms of repair, *Plastic and Reconstructive Surgery*, **1997**, *100*, 937-944. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [17] P. Bornstein, H. Sage, Regulation of collagen gene expression, *Progress in Nucleic Acid Research and Molecular Biology*, **1989**, *37*, 67-106. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [18] T.W. Lin, L. Cardenas, L.J. Soslowky, Biomechanics of tendon injury and repair, *Journal of Biomechanics*, **2004**, *37*, 865-877. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [19] R. Schweitzer, J.H. Chyung, L.C. Murtaugh, A.E. Brent, V. Rosen, E.N. Olson, A. Lassar, C.J. Tabin, Analysis of the tendon cell fate using Scleraxis, a specific marker for tendons and ligaments, **2001**. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [20] E. Blitz, A. Sharir, H. Akiyama, E. Zelzer, formed modularly by a distinct pool of Scx- and Sox9-positive progenitors, *Development*, **2013**, *140*, 2680-2690. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [21] K. Ideo, T. Tokunaga, C. Shukunami, A. Takimoto, Y. Yoshimoto, R. Yonemitsu, T. Karasugi, H. Mizuta, Y. Hiraki, T. Miyamoto, Role of Scx+/Sox9+ cells as potential progenitor cells for postnatal supraspinatus enthesis formation and healing after injury in mice, *PloS One*, **2020**, *15*, 242286. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [22] Y. Sugimoto, A. Takimoto, H. Akiyama, R. Kist, G. Scherer, T. Nakamura, Y. Hiraki, C. Shukunami, Scx+/Sox9+ progenitors contribute to the establishment of the junction between cartilage and tendon/ligament, *Development*, **2013**, *140*, 2280-2288. [[Crossref](#)], [[Google Scholar](#)], [[Publisher](#)]
- [23] S. Agarwal, S.J. Loder, D. Cholak, J. Peterson, J. Li, C. Breuler, R. Cameron Brownley, H. Hsin Sung, M.T. Chung, N. Kamiya, S. Li, Scleraxis-lineage cells

contribute to ectopic bone formation in muscle and tendon, *Stem Cells*, **2017**, *35*, 705-710. [Crossref], [Google Scholar], [Publisher]

[24] V. Leéjard, G. Brideau, F. Blais, R. Salingcarnboriboon, G. Wagner, M.H. Roehrl, M. Noda, D. Duprez, P. Houillier, J. Rossert, Scleraxis and NFATc regulate the expression of the pro- $\alpha 1$ (I) collagen gene in tendon fibroblasts, *Journal of Biological Chemistry*, **2007**, *282*, 17665-17675. [Crossref], [Google Scholar], [Publisher]

[25] W. Feng, Q. Jin, Y. Ming-Yu, H. Yang, T. Xu, S. You-Xing, B. Xu-Ting, C. Wan, W. Yun-Jiao, W. Huan, Y. Ai-Ning, MiR-6924-5p-rich exosomes derived from genetically modified Scleraxis-overexpressing PDGFR α (+) BMMSCs as novel nanotherapeutics for treating osteolysis during tendon-bone healing and improving healing strength, *Biomaterials*, **2021**, *279*, 121242. [Crossref], [Google Scholar], [Publisher]

[26] A.I. Goncalves, M.T. Rodrigues, S.J. Lee, A. Atala, J.J. Yoo, R.L. Reis, M.E. Gomes, Understanding the role of growth factors in modulating stem cell tenogenesis, *PLoS*

one, **2013**, *8*, 83734. [Crossref], [Google Scholar], [Publisher]

[27] D.W. Koch, L.V. Schnabel, I.M. Ellis, R.E. Bates, A.K. Berglund, TGF- $\beta 2$ enhances expression of equine bone marrow-derived mesenchymal stem cell paracrine factors with known associations to tendon healing, *Stem Cell Research & Therapy*, **2022**, *13*, 477. [Crossref], [Google Scholar], [Publisher]

[28] N. Sato, T. Taniguchi, Y. Goda, H. Kosaka, K. Higashino, T. Sakai, S. Katoh, N. Yasui, K. Sairyu, H. Taniguchi, Proteomic analysis of human tendon and ligament: solubilization and analysis of insoluble extracellular matrix in connective tissues, *Journal of Proteome Research*, **2016**, *15*, 4709-4721.

[Crossref], [Google Scholar], [Publisher]

How to cite this article: Baskoro Kusumo Riswanto, Cery Tarise Hajali, Teddy Heri Wardhana*, The effect of secretome injection on tendon healing of rotator cuff tear on new zealand white rabbit through the expression of tgf- β , scleraxis, collagen type iii, and histological evaluation. *Journal of Medicinal and Pharmaceutical Chemistry Research*, 2024, 6(4), 401-410. **Link:** http://jmpcr.samipubco.com/article_185692.html