

Open Access Original Article



Crescent Journal of Medical and Biological Sciences

Vol. 1, No. 2, Spring 2014, 49-53

eISSN: 2148-9696

Effects of Artemisia lanata Extract on Reproductive Parameters of Female Rats

Nava Ainehchi^{1*}, Afshin Zahedi²

Abstract

Objective: Until date, there is no report on safety of Artemisia lanata. This study aimed to determine the possible undesirable effects of A. lanata on reproduction of female rats.

Materials and Methods: The pregnant rats were treated (i.p.) with vehicle or 200 and 400 mg/kg of A. lanata hydroalcoholic extract from the 2-8 day of pregnancy. Then, number and weight of neonates, duration of pregnancy, and percent of dead fetuses were determined. Furthermore, cytotoxicity of this plant was tested using fibroblast (L929) and Chinese hamster ovary (Cho) cell lines.

Results: The A. lanata had no significant effect on duration of pregnancy, average number of neonates, and weight of neonates. However, administration of 200 and 400 mg/kg of the extract led to 30 and 44% abortion in animals, respectively. The extract at concentrations \geq 200 µg/ml significantly (P < 0.001) inhibited the proliferation of L929 fibroblast cells. Regarding the Cho cells, the extract induced toxicity only at concentration of 800 µg/ml (P < 0.010).

Conclusion: Our results showed that continuous consumption of A. lanata in pregnancy may increase the risk of abortion and also may have toxic effect on some cells.

Keywords: Artemisia lanata, Female Rats, Reproductive

Introduction

Until date, there is no report on safety of Artemisia kopetdaghensis. This study aimed to determine the possible undesirable effects of A. kopetdaghensis on reproduction of female rats. The pregnant rats were treated (i.p.) with vehicle or 200 and 400 mg/kg of A. kopetdaghensis hydroalcoholic extract from the 2-8 day of pregnancy. Then, number and weight of neonates, duration of pregnancy, and percent of dead fetuses were determined. Furthermore, cytotoxicity of this plant was tested using fibroblast (L929) and Chinese hamster ovary (Cho) cell lines. The A. kopetdaghensis had no significant effect on duration of pregnancy, average number of neonates, and weight of neonates. However, administration of 200 and 400 mg/kg of the extract led to 30 and 44% abortion in animals, respectively. The extract at concentrations \geq 200 µg/ml significantly (P < 0.001)

inhibited the proliferation of L929 fibroblast cells. Regarding the Cho cells, the extract induced toxicity only at concentration of 800 μ g/ml (P < 0.010). Our results showed that continuous consumption of A. kopetdaghensis in pregnancy may increase the risk of abortion and also may have toxic effect on some cells.

Today, medicinal plants are widely used around the world as an alternative to pharmaceutical drugs. Although herbal products are considered to have fewer adverse effects compared with synthetic drugs, they are not completely free from side effects or toxicity (1). Adverse effects of medicinal plants may result from contamination of herbs with toxic metals, adulteration with active synthetic compounds, improperly prepared herbal products, misidentification of herbal ingredients, and inherent toxicity of certain herbs (2). Therefore, the potential side effects of any medicinal plant need to be

Received: 16 Dec 2013, Revised: 24 Jan 2014, Accepted: 23 Feb 2014, Available online: 15 Apr 2014

¹ Women's Reproductive Health Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

² Department of Pathology, College of Vet Medicine, Rasht Branch, Islamic Azad University, Rasht, Iran

^{*}Corresponding Author: Nava Ainehchi, Women's Reproductive Health Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

determined before its clinical applications. Special care should be taken when an herbal product is used by pregnant women, children, and geriatrics. Unfortunately, unlike those synthetic drugs not recommended for use in pregnancy because of known unwanted effects, there are insufficient data about undesirable maternal and perinatal consequences of use of herbal agents.

Artemisia lanata, aromatic shrubs belonging to the Asteraceae family, is traditionally used worldwide for its anti-inflammatory, antimicrobial, antifungal, and sedative activities (3,4). However, to date, there is no report on safety or toxicity of this plant. Only Ebrahimi et al. reported that methanolic extract and essential oil of A. lanata exhibited tumor growth induction at some concentrations and cytotoxicity at other concentrations (5). The aim of the present study was to determine the possible undesirable effects of A. lanata on reproduction of female rats. Also, the possible cytotoxicity of this plant was assessed using fibroblast and ovary cells in vitro.

Materials and Methods

High glucose Dulbecco's modified eagles medium (DMEM) and fetal bovine serum (FBS) were purchased from gibco. Penicillin, streptomycin, and 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetraz olium bromide (MTT) were obtained from sigma. Dimethyl sulfoxide (DMSO) was purchased from Fluka. Tween 80 was purchased from Merck.

The fresh A. lanata was collected from native stores and identified by the Pharmacognosia Department of Tabriz Research Institute, Iran. The aerial parts of plant were cleaned and grounded to fine powder with a blender. Then, macerated extract was prepared as described previously (6,7), briefly by suspension of 200 g of the powder in 500 mL of 50% ethanol and incubation for 72 h at 37 °C. The hydroalcoholic extract was then dried on a water bath and the yield dissolved in distilled water containing 1% Tween 80.

Male and female Wistar rats (200-250 g) and female mice (26-32 g) were obtained from Laboratory Animals Research Center and housed in cages with controlled lighting (12 h dark, 12 h light) and temperature (22 \pm 2 °C). The animals were given standard pellets diet and water ad libitum. All animal procedures were in accordance with Ethical Guidelines approved by the Animal Care Use Committee of Tabriz Research Institute, Iran.

Prior to the mating, the female rats were isolated for 30 days to rule out preexisting pregnancy. Then, they were caged overnight with a male rat of proven fertility in the ratio of 1:1. Rats exhibiting vaginal plug on the following morning were separated, and that day was considered as the 1^{st} day of pregnancy. The pregnant rats were randomized into three groups: (1) control group receiving 1% Tween 80 as vehicle (n = 8), (2) experimental rats treated with 200 mg/kg of A. lanata extract (n = 10), and (3)

experimental rats receiving 400 mg/kg of the plant extract (n = 9). The extract was injected intraperitoneally from the 2^{nd} to the 8^{th} day of pregnancy (early period of organogenesis). The animals were kept individually in cages until parturition. Then, number and weight of neonates, duration of pregnancy, and percent of dead fetuses were determined.

Acute toxicity of A. lanata extract was evaluated by the method of Akhila et al. (8). Five groups of two mice received vehicle (1% Tween 80) or 400, 800, 1600, and 3200 mg/kg of the plant extract intraperitoneally. The treated animals were monitored for 24 h and also 1 week for mortality. The lowest dose which led to death of animals and the highest dose which did not kill any mice were recorded.

The L929 (mouse fibroblast) and Cho cells were seeded in 96-well plates and cultured for 24 h in DMEM supplemented with 10% FBS, penicillin (100 units/ml), and streptomycin (100 μ g/ml) at 37 °C and 5% CO₂. Then, the medium was changed to fresh one containing vehicle (1% DMSO) or 50-800 μ g/ml of A. lanata extract. The cells were further incubated for 24 h at 37 °C and 5% CO₂. At the end of the treatment, the cell proliferation was measured using MTT assay as previously described (9-11). The assay was carried out using 2 culture plates, 4 wells for each concentration (n = 8).

The values were compared using the one-way analysis of variance, followed by Tukey's post-hoc test for multiple comparisons. The P < 0.050 were considered to be statistically significant. All results are presented as mean \pm standard error of mean.

Results

Effect of A. lanata on reproduction

As shown in table 1, the A. lanata extract at concentrations of 200 and 400 mg/kg had no significant effect on duration of pregnancy. However, administration of 200 and 400 mg/kg of the extract led to 30 and 44% abortion in animals, respectively. The percent of dead neonates was 6% in control group and 7% and 0% in experimental groups treated with 200 and 400 mg/kg, respectively. The average number of the neonates in the animals receiving vehicle during pregnancy was 8.13 ± 1.5 (Figure 1). None of the A. lanata doses could cause a significant change in the neonate number. Likewise, the extract had virtually no significant effect on weight of neonates (Figure 2).

Acute toxicity of A. lanata

Different groups of mice (n = 2) were treated with 400, 800, 1600, and 3200 mg/kg of A. lanata hydroalcoholic extract. After 24 h, it was found that 1600 and 3200 mg/kg are the highest dose which did not kill any mice and the lowest dose which led to death of both mice, respectively. The treated animals were further monitored until 1 week and no mortality or any sign of toxicity was observed at doses \leq 1600 mg/kg.

Table 1. Effect of A. lanata on reproduction of female rats. The pregnant rats were treated (i.p.) with vehicle or A. lanata hydroalcoholic extract from the $2-8^{th}$ day of pregnancy. Values are mean \pm SEM (n = 8-10)

Groups	Duration of pregnancy (day)	Abortion (%)	Percent of dead fetuses (%)
Control	22 ± 0.3	0	6
A. lanata (200 mg/kg)	23 ± 0.3	30	7
A. lanata (400 mg/kg)	22 ± 0.4	44	0

A. lanata: Artemisia lanata: SEM: Standard error of mean

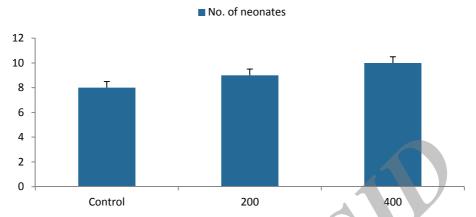


Figure 1. Effect of Artemisia lanata hydroalcoholic extract on number of neonates The pregnant rats were treated (i.p.) with vehicle or 200 and 400 mg/kg of A. lanata hydroalcoholic extract from the 2-8th day of pregnancy. Values are mean ± standard error of mean (n = 8-10)

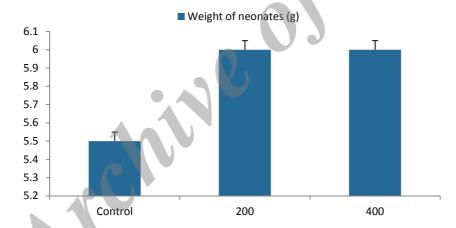


Figure 2. Effect of Artemisia lanata hydroalcoholic extract on weight of neonates The pregnant rats were treated (i.p.) with vehicle or 200 and 400 mg/kg of A. lanata hydroalcoholic extract from the 2-8th day of pregnancy. Values are mean ± standard error of mean (n = 8-10)

Cytotoxicity of A. lanata

Figures 3 and 4 show the effect of A. lanata hydroalcoholic extract on proliferation of L929 and Cho cells. Following incubation of L929 fibroblast cells with 50, 100, 200, 400, and 800 μ g/ml of the extract, approximately 15, 17, 45, 72, and 77 percent inhibition in cell growth, was observed, respectively, as compared with untreated cells. The cytotoxic effect of A. lanata was statistically significant at concentrations \geq 200 μ g/ml (P < 0.001). On the other hand, the extract induced toxicity on Cho cells only at concentration of 800 μ g/ml (P < 0.010). In the presence of 50, 100, 200, 400 and 800 μ g/ml of A. lanata, surviving of Cho cells was 103 \pm 1.3,

 100 ± 3.2 , 102 ± 2 , 102 ± 2 and $85 \pm 2\%$, respectively, as compared to untreated cells (100 ± 5).

Discussion

Many medicinal plants are used by pregnant women for their therapeutic effects. For example, it has been shown that about 36% of pregnant women in Norway use herbs (8,12). However, these plants are consumed mostly based on personal experience or traditional knowledge and in most cases it is unclear how safe the use of them is during pregnancy. Previous studies highlighted that some of plants have different antifertility activities (13). The present study was aimed to examine the possible toxic effects of A. lanata

reproduction of female rats. Our demonstrated that the plant extract has no effect on duration of pregnancy and number or weight of neonates. However, it can induce abortifacient effect when consumed at early period of pregnancy. This antifertility effect of A. lanata has been also reported for some other plants of the Asteraceae family such as Achillea millefolium and Aspilia africana which showed antispermatogenic and antiovulatory activities, respectively (14,15). On the other hand, we observed that administration of A. lanata extract (400 mg/kg) did not lead to stillbirth. The exact cause of this discrepancy should be explored in the future experiments. However, it may be attributed to high rate of abortion in animals receiving high concentration of the extract.

According to the previously published work, amphene, camphor, davanone, eucalyptol, eugenol, and geranial are of major components of A. lanata. Camphor accounted for about 1.5 g/100 g of this plant (16). Rabl et al. have reported that camphor crosses the placenta and may lead to abortion (17). In another study, Linjawi reported that camphor induces significant structural changes on uterus of pregnant rats (18). Therefore, it is rational to assume that camphor is involved in the abortifacient effect of A. lanata.

Cytotoxicity evaluation of A. lanata showed that its hydroalcoholic extract decreases proliferation of fibroblast cells. This finding may describe the camphor induced degeneration of luminal epithelium and decrease of endometrium thickness in uterus of pregnant animals (18).

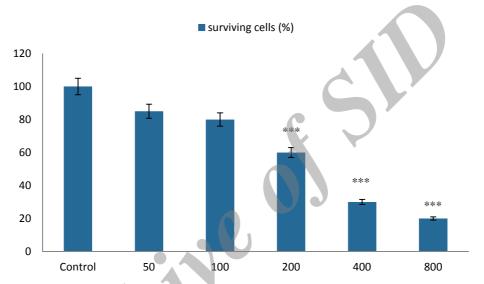


Figure 3. Effect of Artemisia lanata hydroalcoholic extract on proliferation of fibroblast L929 cell lines

The cells were cultured for 24 h in the medium containing vehicle (1% dimethyl sulfoxide) or 50-800 μ g/ml of A. lanata extract; Values are mean \pm standard error of mean (n = 8)

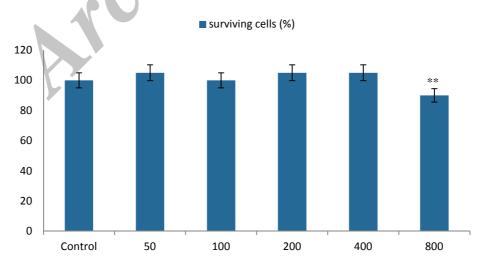


Figure 4. Effect of Artemisia lanata hydroalcoholic extract on proliferation of Chinese hamster ovary cell lines

The cells were cultured for 24 h in the medium containing vehicle (1% dimethyl sulfoxide) or 50-800 µg/ml of A. lanata extract; Values are mean ± standard error of mean (n = 8)

In conclusion, our results showed that continuous consumption of A. lanata in pregnancy may increase the risk of abortion and also may have toxic effect on some cells of body. Therefore, its continuous use is not recommended in pregnancy.

Ethical issues

The local ethics committee approved the study.

Conflict of interests

We declare that we have no conflict of interests.

Acknowledgments

We would like to thank Women's Reproductive Health Research Center that helped us in this study.

References

- 1. De Smet PA. Health risks of herbal remedies: an update. Clin Pharmacol Ther 2004; 76: 1-17.
- 2. Jordan SA, Cunningham DG, Marles RJ. Assessment of herbal medicinal products: challenges, and opportunities to increase the knowledge base for safety assessment. Toxicol Appl Pharmacol 2010; 243: 198-216.
- 3. Ramezani M, Behravan J, Yazdinezhad A. Composition and antimicrobial activity of the volatile oil of Artemisia kopetdaghensis Krasch., M.Pop. & Linecz ex Poljak from Iran. Flavour and Fragrance Journal 2006; 21: 869-71.
- 4. Mirdeilami SZ, Barani H, Mazandarani M, Heshmati GhA. Ethnopharmacological survey of medicinal pants in Maraveh Tappeh region, north of Iran. Iranian Journal of Plant Physiology 2011; 2: 325-36.
- 5. Ebrahimi M, Ramezani M, Tehrani SO, Malekshah OM, Behravan J. Cytotoxic effects of methanolic extract and essential oil of Artemisia Kopetdaghensis. Journal of Essential Oil-Bearing Plants 2010; 13: 732-7.
- 6. Ghorbani A, Hadjzadeh M, Rajaei Z, Zendehbad SB. Effects of Fenugreek Seeds on Adipogenesis and Lipolysis in Normal and Diabetic Rats. Pakistan Journal of Biological Sciences 2014; 17: 523-8.
- 7. Shafiee-Nick R, Ghorbani A, Vafaee BF, Rakhshandeh H. Chronic administration of a combination of six herbs inhibits the progression of hyperglycemia and decreases

- serum lipids and aspartate amino transferase activity in diabetic rats. Adv Pharmacol Sci 2012; 2012: 789796.
- 8. Akhila JS, Shyamjith S, Deepa D, Alwar MC. Acute toxicity studies and determination of median lethal dose. Current Science 2007; 93: 917-20.
- Forouzanfar F, Afkhami GA, Asadpour E, Ghorbani A, Sadeghnia HR. Protective Effect of Punica granatum L. against Serum/Glucose Deprivation-Induced PC12 Cells Injury. Evid Based Complement Alternat Med 2013; 2013: 716730.
- 10. Mortazavian SM, Ghorbani A. Antiproliferative effect of Viola tricolor on neuroblastoma cells in vitro. Australian Journal of Herbal Medicine 2012; 24: 93-6.
- 11. Mortazavian SM, Ghorbani A, Ghorbani Hesari T. Effect of hydro-alcoholic extract of Viola tricolor and its fractions on proliferation of uterine cervix carcinoma cells. Iranian Journal of Obstetrics, Gynecology and Infertility 2012; 15: 9-16.
- 12. Nordeng H, Havnen GC. Use of herbal drugs in pregnancy: a survey among 400 Norwegian women. Pharmacoepidemiol Drug Saf 2004; 13: 371-80.
- 13. Priya G, Saravanan K, Renuka C. Medicinal plants with potential antifertility activity- A review of sixteen years of herbal medicine research (1994-2010). International Journal of PharmTech Research 2012; 4: 481-94.
- 14. Montanari T, de Carvalho JE, Dolder H. Antispermatogenic effect of Achillea millefolium L. in mice. Contraception 1998; 58: 309-13.
- 15. Oyesola TO, Oyesola OA, Okoye CS. Effects of aqueous extract of Aspilia africana on reproductive functions of female Wistar rats. Pak J Biol Sci 2010; 13: 126-31.
- 16. Costa R, De Fina MR, Valentino MR, Rustaiyan A, Dugo P, Dugo G, et al. An investigation on the volatile composition of some Artemisia species from Iran. Flavour and Fragrance Journal 2009; 24: 75-82.
- 17. Rabl W, Katzgraber F, Steinlechner M. Camphor ingestion for abortion (case report). Forensic Sci Int 1997; 89: 137-40.
- 18. Linjawi SA. Effect of camphor on uterus histology of pregnant rats. Journal of King Abdulaziz University 2009; 16: 77-90.

Citation: Ainehchi N, Zahedi A. **Effects of Artemisia lanata Extract on Reproductive Parameters of Female Rats**. Crescent J Med & Biol Sci 2014; 1(2): 49-53.