



Adipose Tissue Derived- Stem Cells: Applications and Benefits in Tissue Regeneration

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Received 2017 May 04; Accepted 2017 June 12.

Keywords: Adipose Derived Stem Cell, Proliferation, Growth Factor

Mesenchymal stem cells (MSCs) are the proper hotspot for the regeneration of tissues due to of self-renew and multi-potent differentiation properties. Furthermore, adipose derived stem cells (ADSCs) are multi-potent stem cells assigned to the generic nomenclature (1) and can easily be isolated from subcutaneous liposuction. These are the foremost advantages of ADSCs compared to other stem cells sources. In addition, these cells have no ethical and political issues compared to embryonic stem cells. They make ADSCs become more acceptable for tissue and organ transplantation in regenerative medicine and clinical studies (2).

There are some cell surface markers used to characterize ADSCs, using flow cytometry, however, a unique single marker has yet to be identified. It is positive at the first passage of culture expression of ADSCs for CD34, however, the expression for hematopoietic antigens such as CD14, CD31, CD45 and CD144 is negative (3).

For in vitro purpose, it takes 2 to 5 days for cultured ADSC to double. Under specific culturing conditions, ADSCs differentiated into multiple cells such as chondrocytes, myocytes, adipocytes, osteoblast, and neural cells; therefore, it makes these cells proper for multiple clinical applications (4).

Nowadays transplantation of ADSCs is used in researches that focus on therapy of a variety of diseases including diabetes, trauma (5, 6). Additionally, differentiation into neural cells made these cells appropriate for treatment of neurodegenerative diseases (7). For example, Parkinson disease (PD) is the second most common progressive neurodegenerative disorder in the world and transplantation of dopaminergic neurons open the windows to the recreation of the nigrostriatal pathway and treatment of PD; therefore, the first step in therapy is a production of functional dopaminergic neurons (8). In ischemic tissues, mature adipocytes die easily, however, AD-

SCs survive in this condition and can secrete angiogenic factors (9, 10).

On the other hand, ADSCs implantation also supports cell attachment, proliferation, and differentiation in the specific clinical application and can be used in wound healing applications. Some researches have been shown that grafted an acellular dermal matrix construct seeded with human ADSCs into a murine injury model can enhance the wound healing at day 7 (11, 12).

Another potential of ADSCs is the osteogenic application, for example, bone tissue regeneration after injection or congenital malformations. For purpose of bone growth, many in vivo studies have combined ADSCs with biodegradable scaffold materials (13). In examining ADSCs for the cartilage tissue, the engineering is required to remedy osteoarthritis (OA) (14, 15).

Finally, multiple growth factors are secreted by ADSCs, for example, basic fibroblast growth factor (bFGF), vascular endothelial growth factor (VEGF), insulin-like growth factor 1 (IGF), hepatocyte growth factors (HGF), and transforming growth factor (TGF)- β 1. Some evidence has been shown the protective roles of these factors are secreted by ADSCs on epithelial cells and prevent cell death during oxidative injury (16).

At the result, these features of ADSCs made them suitable for research on treating various diseases and ultimately, these efforts can lead to widespread clinical applications of these cells.

Acknowledgments

The author of this article appreciates the department of histology of Zahedan University of Medical Sciences (ZA-UMS), Zahedan, Iran, for the assistance in preparing this article.

References

1. Strem BM, Hicok KC, Zhu M, Wulur I, Alfonso Z, Schreiber RE, et al. Multipotential differentiation of adipose tissue-derived stem cells. *Keio J Med.* 2005;54(3):132–41. doi: [10.2302/kjm.54.132](https://doi.org/10.2302/kjm.54.132). [PubMed: 16237275].
2. Cawthorn WP, Scheller EL, MacDougald OA. Adipose tissue stem cells: the great WAT hope. *Trends Endocrinol Metab.* 2012;23(6):270–7. doi: [10.1016/j.tem.2012.01.003](https://doi.org/10.1016/j.tem.2012.01.003). [PubMed: 22417866].
3. Keung EZ, Nelson PJ, Conrad C. Concise review: genetically engineered stem cell therapy targeting angiogenesis and tumor stroma in gastrointestinal malignancy. *Stem Cells.* 2013;31(2):227–35. doi: [10.1002/stem.1269](https://doi.org/10.1002/stem.1269). [PubMed: 23132810].
4. Katz AJ, Tholpady A, Tholpady SS, Shang H, Ogle RC. Cell surface and transcriptional characterization of human adipose-derived adherent stromal (hADAS) cells. *Stem Cells.* 2005;23(3):412–23. doi: [10.1634/stemcells.2004-0021](https://doi.org/10.1634/stemcells.2004-0021). [PubMed: 15749936].
5. Radtke C, Schmitz B, Spies M, Kocsis JD, Vogt PM. Peripheral glial cell differentiation from neurospheres derived from adipose mesenchymal stem cells. *Int J Dev Neurosci.* 2009;27(8):817–23. doi: [10.1016/j.ijdevneu.2009.08.006](https://doi.org/10.1016/j.ijdevneu.2009.08.006). [PubMed: 19699793].
6. Safford KM, Hicok KC, Safford SD, Halvorsen YD, Wilkison WO, Gimble JM, et al. Neurogenic differentiation of murine and human adipose-derived stromal cells. *Biochem Biophys Res Commun.* 2002;294(2):371–9. doi: [10.1016/S0006-291X\(02\)00469-2](https://doi.org/10.1016/S0006-291X(02)00469-2). [PubMed: 12051722].
7. Asemi Rad A, Hasan Heidari M, Aliaghaei A, Eskandarian Broujeni M, Shojaei A, Abbaszadeh HA, et al. In vitro differentiation of adipose derived stem cells into functional dopaminergic neurons. *Biomed Pharmacol J.* 2017;10(2):595–605. doi: [10.13005/bpj/1146](https://doi.org/10.13005/bpj/1146).
8. Mariani E, Facchini A. Clinical applications and biosafety of human adult mesenchymal stem cells. *Curr Pharm Des.* 2012;18(13):1821–45. doi: [10.2174/138161212799859666](https://doi.org/10.2174/138161212799859666). [PubMed: 22352750].
9. Suga H, Eto H, Aoi N, Kato H, Araki J, Doi K, et al. Adipose tissue remodeling under ischemia: death of adipocytes and activation of stem/progenitor cells. *Plast Reconstr Surg.* 2010;126(6):1911–23. doi: [10.1097/PRS.0b013e3181f4468b](https://doi.org/10.1097/PRS.0b013e3181f4468b). [PubMed: 21124131].
10. Lee EY, Xia Y, Kim WS, Kim MH, Kim TH, Kim KJ, et al. Hypoxia-enhanced wound-healing function of adipose-derived stem cells: increase in stem cell proliferation and up-regulation of VEGF and bFGF. *Wound Repair Regen.* 2009;17(4):540–7. doi: [10.1111/j.1524-475X.2009.00499.x](https://doi.org/10.1111/j.1524-475X.2009.00499.x). [PubMed: 19614919].
11. Choi JH, Gimble JM, Lee K, Marra KG, Rubin JP, Yoo JJ, et al. Adipose tissue engineering for soft tissue regeneration. *Tissue Eng Part B Rev.* 2010;16(4):413–26. doi: [10.1089/ten.TEB.2009.0544](https://doi.org/10.1089/ten.TEB.2009.0544). [PubMed: 20166810].
12. Ito R, Morimoto N, Liem PH, Nakamura Y, Kawai K, Taira T, et al. Adipogenesis using human adipose tissue-derived stromal cells combined with a collagen/gelatin sponge sustaining release of basic fibroblast growth factor. *J Tissue Eng Regen Med.* 2014;8(12):1000–8. doi: [10.1002/term.1611](https://doi.org/10.1002/term.1611). [PubMed: 22997068].
13. Zanetti AS, Sabliov C, Gimble JM, Hayes DJ. Human adipose-derived stem cells and three-dimensional scaffold constructs: a review of the biomaterials and models currently used for bone regeneration. *J Biomed Mater Res B Appl Biomater.* 2013;101(1):187–99. doi: [10.1002/jbm.b.32817](https://doi.org/10.1002/jbm.b.32817). [PubMed: 22997152].
14. Schmidt CE, Leach JB. Neural tissue engineering: strategies for repair and regeneration. *Annu Rev Biomed Eng.* 2003;5:293–347. doi: [10.1146/annurev.bioeng.5.011303.120731](https://doi.org/10.1146/annurev.bioeng.5.011303.120731). [PubMed: 14527315].
15. Salgado AJ, Reis RL, Sousa NJ, Gimble JM. Adipose tissue derived stem cells secretome: soluble factors and their roles in regenerative medicine. *Curr Stem Cell Res Ther.* 2010;5(2):103–10. doi: [10.2174/157488810791268564](https://doi.org/10.2174/157488810791268564). [PubMed: 19941460].
16. Ghorabi MT, Aliaghaei A, Sadeghi Y, Shaerzadeh F, Rad AA, Mohamadi R, et al. Evidence supporting neuroprotective effect of adipose derived stem cells on PC12 cells against oxidative stress induced by H₂O₂. *Cell Mol Biol (Noisy-le-grand).* 2017;63(3):1–6. doi: [10.14715/cmb/2017.63.3.1](https://doi.org/10.14715/cmb/2017.63.3.1). [PubMed: 28466808].