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Research Article

Salmonellosis and Related Risk Factors in Broiler Flocks in Mazandaran Province, Northern Iran

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Abstract

Background: Prevention of foodborne pathogens is essential to control infectious diseases; *Salmonella* spp. is referred to as the most common causative agent of foodborne illnesses.

Objectives: The current study aimed to determine the prevalence of *Salmonella enterica* subsp. *enterica* in broiler flocks in Mazandaran province, north of Iran and find the potential risk factors including: age, size of flock, strain, season, vaccination program and use of antibiotics.

Materials and Methods: From March 2012 to December 2013, a total of 50 flocks were selected in slaughterhouse and 20 cloacal samples were collected from each flock. Every five samples were pooled and investigated for *Salmonella* spp. using polymerase chain reaction (PCR).

Results: Thirteen flocks out of 50 (26%) were positive for *Salmonella* species. Chances of *Salmonella* spp. detection was higher in flocks with lower age (P = 0.41). Increasing flock population was associated with increased chance of *Salmonella* spp. isolation (P = 0.21). The risk of salmonellosis in broiler flocks was increased when no antibiotics were given to day-old chicks. There was no significant difference (P = 0.30) in the prevalence of salmonellosis among different broiler strains.

Conclusions: In the current study, six risk factors were assessed for *Salmonella* spp. contamination in broiler flocks. Some of these factors contributed to the risk of salmonellosis in broiler flocks.

Keywords: Broiler Flocks, Risk Factor, Iran, Salmonella spp

1. Background

Prevention of food hazards in the first part of the food chain is essential to prevent illness of consumers. Salmonella spp. is cited as the most common causative agent of foodborne diseases (1). The genus Salmonella contains two species: S. enterica and S. bongori. Six subspecies are differentiated within S. enterica based on their biochemical and genomic characteristics, that one of them is S. enterica subsp. enterica. With regard to food safety, S. enterica subsp. enterica serogroups should be considered more than others, since they are known to cause 99% of *Salmonella* spp. infections in humans (2,3). The usual route of infection in chickens is the oral uptake of Salmonella spp. from the environment. Contaminated food and water, for instance, are important sources of salmonellosis in chickens (4, 5). The vertical transmission of these bacteria can also be an important issue in poultry (6, 7). Different prevalence rate of these bacteria are reported between countries, nearly 0 in Sweden, 68.2% in Hungary (8), 76.9% in Canada (9); 69.8% in France (10); 41.3% in Turkey

(11) and 25% in Denmark (12). Some management and environmental risk factors are associated with the incidence of salmonellosis in the flocks. The environmental persistence of *Salmonellae* spp. is a significant factor in the epidemiology of these bacteria in poultry by creating opportunities for horizontal transmission of infection within and between flocks (13). The role of rodents, flies, beetles and wild birds as vectors in *Salmonella* spp. transfer is extensively discussed (14-17). Stress has an immunosuppressive effect in egg laying hens, which can have negative consequences with respect to salmonellosis and shedding (18, 19). There are few studies on the role of vaccination programs and using antibiotics in rearing the period with salmonellosis.

2. Objectives

Mazandaran province in Northern Iran is one of the major poultry production areas. No study is conducted to determine the prevalence of salmonellosis in poultry flocks in this region. Therefore, the present study aimed to determine the prevalence of *Salmonella* spp. in broiler

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flocks in Mazandaran province and its possible associations with some potential risk factors including age, size of flock, strain, season, vaccination programs, and use of antibiotics during the production period.

3. Materials and Methods

A cross-sectional study was conducted to determine the prevalence of salmonellosis and related risk factors in broiler flocks.

3.1. Sample Collection

A total of 1000 cloacal swab samples were randomly collected from 50 broiler flocks (20 swabs per each flock) on the slaughter line of four abattoirs of Mazandaran province from March 2012 to December 2013. Sampling was randomly done in a systematic manner, which means each cloacal swab was taken from one of each five birds in the slaughter line.

3.2. Data Collection

To evaluate potential risk factors, a questionnaire that structured the parameters such as: age, size of flock, strain, season, vaccination programs and antibiotics used in the rearing period was completed for each flock.

3.3. Enrichment of the Samples

Every five samples from each flock were pooled, and were pre-enriched in buffered peptone water in 1:10 sample/broth ratio at 37°C for 24 hours. One milliliter of this pre-enrichment broth was used to inoculate 10 mL of cysteine-selenite broth (Merck, Germany).

3.4. DNA Extraction

DNA extraction was performed through the protocol described by Khoshbakht et al. (20). One milliliter of each enriched fecal sample was transferred to a 1.5 mL microtube and centrifuged at 10,000 rpm for two minutes. Pellets were re-suspended in 570 μ L of Tris-EDTA (TE) buffer (10 mM Tris-HCl pH = 8;1 mM Na₂ ethylenediaminetetraacetic acid: EDTA), 30 μ L 10% sodium dodecyl sulphate and 4 μ L proteinase K (Fermentas, Germany) in a concentration of 20 mg/mL. Samples in micro-tubes were mixed vigorously before incubation at 56°C in a water bath for one hour. One hundred microliters of 5 M and One hundred microliters of CTAB/NaCl (cetyltrimethylammonium bromide: CTAB 10%, NaCl 0.6 M) were added and mixed. After incubation at 65°C in water bath for 12 minutes, 500 μ L of phenol/chloroform/isoamyl alcohol (25:24:1) was added and vortexed. Then samples were centrifuged at 13,000 rpm

for seven minutes. Five hundred-fifty microliters of supernatant was transferred to a fresh micro-tube and equal volume of chloroform/isoamyl alcohol (24:1) was added. The samples were mixed and centrifuged as above. The supernatant was transferred to a new micro-tube and 300 μ L isopropanol was added to each tube, then centrifuged at 10,000 rpm for three minutes and the pellet was washed in 70% ice-cold ethanol and centrifuged again at 10,000 rpm for three minutes. The final pellet was re-suspended in 50 μ L of TE buffer and stored at -20°C until polymerase chain reaction (PCR) was performed.

3.5. PCR Reaction

PCR amplifications were performed in a final volume of 25 μ L. The PCR reaction mixtures consisted of 2 μ L of the DNA template, 2.5 μ L 10X PCR buffer (75 mM Tris-HCl, pH 9.0, 2 mM MgCl₂, 50 mM KCl, 20 mM [NH4]₂SO₄), 1 μ L dNTPs (50 μ M), 1 μ L (1 U Ampli Taq DNA polymerase), 1 μ L (25 pmol) from each forward and reverse primers (Cinna-Gen, Iran) in the total volume of 25 μ L using distilled deionized water. The forward (S139) and reverse (S141) primer sequences were 5'GTGAAATTATCGCCGCCACGTTCGAA3' and 5'TCATCGCACCGTCAAAGGAACC 3' (21), respectively. The thermal cycler (MJ mini, BioRad, USA) was adjusted as follows: initial denaturation at 94°C for four minutes, followed by 33 cycles of denaturation at 94°C for one minute, annealing at 57°C for one minute and extension at 72°C for one minute. Final extension was carried out at 72°C for seven minutes. Previously identified Salmonella strains isolated from animals, in faculty of veterinary medicine of Amol, were used as positive control. Amplified products were separated by electrophoresis on 1.5% agarose gel stained with ethidium bromide. Visualizations were undertaken using a UV transilluminator (BTS-20, Japan), and the 100 bp DNA ladder was used as molecular size marker.

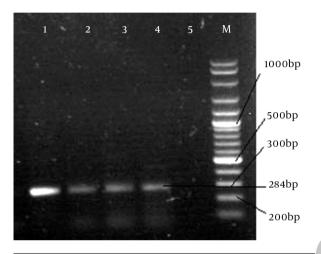
3.6. Statistical Analysis

Prevalence of infection and corresponding 95% confidence interval (CI) were estimated. Possible association between the contamination with *Salmonella* spp. in the flock and the risk factors was investigated using univariate and multivariate logistic regression analyses. Variables with P value equal or less than 0.30 were included in the multivariate logistic regression analysis. Final logistic model was fitted based on stepwise backward elimination procedure and significance of Wald statistics. A P value less than 0.05 was considered statistically significant in the final model.

4. Results

Of the 1000 cloacal samples analyzed by PCR, the prevalence of *Salmonella* spp. contamination in the broiler was 26%. Thirteen out of 50 flocks were positive for *Salmonella* species (Figure 1). Summary of statistics for broiler flocks variables according to the rate of contamination with *Salmonella* spp. are shown in Table 1.

Figure 1. PCR Final Product for Differential Detection of Salmonella Species



Lane 1, positive control; Lanes 2 - 4, positive samples; Lane 5, negative control; Lane M. DNA size marker

 $\textbf{Table 1.} \ \ \textbf{Summary of Statistics for Broiler Flocks Variables According to the Rate of Contamination With Salmonella spp. \ ^a$

Variable	Positive Flocks (N = 13)	Negative Flocks (N = 37)	P Value
Age at slaughter, d	52.6 ± 6.1	58.1 ± 3.6	0.41
Flock size (bird)	15245 ± 7840	10231 ± 4551	0.21

 $^{^{\}mathrm{a}}$ Values are expressed as mean \pm SD.

Age of flocks at slaughtering time was lower in positive flocks, but not significant (P = 0.41). The mean of bird population in negative flocks was slightly lower than that of the positive ones, but the difference was not statistically significant (P = 0.21). One or more kinds of antibiotics were used in nearly 90% of the flocks during the rearing period, and enrofloxacin was used most widely. According to the statistical analysis, there was no relationship between the use of enrofloxacin and isolation of *Salmonella* spp. (P = 0.17); but, the risk of salmonellosis in broiler flock was increased when no antibiotics were given to day-old chicks (P = 0.04). All of the studied broiler flocks had been vac-

cinated against avian influenza virus (AIV), Newcastle disease (ND), and infectious bursal disease (IBD) according to the regional vaccination program; however, only 45% received infectious bronchitis virus (IBV) vaccine. Surprisingly, the isolation of Salmonella spp. was insignificantly higher in flocks which received IBV vaccines (P = 0.23). There was no significant difference (P = 0.30) among different strains (Ross, Cobb, and Arbor acres) regarding the contamination with Salmonella spp. According to the season, the frequency of Salmonella spp. isolation were higher and lower in flocks sampled in winter (17.8%) and in spring (9.2%) compared to those of autumn and summer (7.1% and 6.9%, respectively); but differences were not statistically significant (P = 0.09). Four variables including seasons of sampling (P = 0.09), broiler strains (P = 0.30), using antibiotic on day one (P = 0.04) and vaccination against IBV (P = 0.04)= 0.23) were included in the logistic model (Table 2). Results showed that odds of infection increased with using antibiotic on day 1 (P = 0.04) and vaccination against IBV (P = 0.23).

 $\label{lem:continuous} \textbf{Table 2.} \ \ Results \ of the Final Logistic Regression \ Model for \ Contamination \ With \ \textit{Salmonella} \ \text{spp. in Broiler Flocks in Iran}$

Variable	P Value	Standard Error	Odds Ratio
Season	0.09	1.41	9.1
Strain	0.30	1.01	15.3
Use of antibiotics on day one			
Yes	NA	NA	1
No	0.04	0.89	7.1
Vaccination against IBV			
Yes	0.23	1.77	7.23
No	NA	NA	1

Abbreviations: IBV, infectious bronchitis virus; NA, not available.

5. Discussion

Overall, thirteen out of 50 flocks (26%) were positive for *Salmonella* species. This finding was in agreement with the results of Chadfield et al. (12), and Gutierrez et al. (22) who reported 25% in Denmark and 27% in Ireland, respectively. Ansari-Lari et al. (23) reported that 22.5% of the broiler flocks of Shiraz were positive for *Salmonella* spp. The difference observed between the results of the current study and those of the latter study may be due to different detecting procedures; they used culture methods. Excretion of *S. enterica* decreases with age (24). In the current study,

the positive flocks had lower age, but the age of the chickens was not a significant variable in the present investigation. Skov et al. (25) showed that the risk of salmonellosis increased when the flock size was larger, which is in line with results of the current study. In the current study, the prevalence of Salmonella spp. was the highest (17.8%) in winter and the lowest (6.9%) in summer. These results contradict with those of the studies by Bouwknegt et al. (26), Mollenhorst et al. (27), Namata et al. (28) and Huneau-Salaun et al. (29); but in accordance with those of Wales et al. (30) and Upadhyaya et al. (31) that reported a seasonable effect in their studies. The chance of Salmonella isolation in flocks sampled in winter was significantly higher than that of the flocks sampled in the other seasons. A high density of animals is a well-known risk factor for contamination with Salmonella spp. (8, 29). Furthermore, the air quality in flocks seems to be lower in winter (32, 33). This can cause stress in broilers, leading them from a Salmonellacarrying state to a Salmonella-shedding state. The results showed no relationship (P = 0.30) between broiler strain and salmonellosis, which were in agreement with the results obtained by Skov et al. (25) and Huneau-Salaun et al. (29). According to the results, using antibiotics in day-old chicks reduces the chance of Salmonella spp. isolation. This finding was supported by the results of another study (11). Prophylactic usage of antibiotics against mortality during the first day of life can reduce the number of colonized and shed bacteria (34-36). In Iran, there are different opinions among farmers about the use of IBV vaccine. Although IBV vaccination is used in a nationwide program in Iran, a significant proportion of farm owners do not use IBV vaccination in their flocks due to their undesirable personal experience with this vaccine in the field (35). Results of this study showed an association between IBV vaccination and salmonellosis. Volkova et al. (36) showed that increased dosage of IBV vaccine delivered via spray to the one-day-old birds was linked to a higher probability of Salmonella spp. isolation from the flock, which is in line with the current study findings.

In the current study, six risk factors were assessed for contamination with *Salmonella* species in broiler flocks. This is the first time that such results are obtained in Iran. Further studies on *Salmonella* serotypes during rearing period are necessary.

Footnotes

Authors' Contribution: All authors contributed equally in this article, especially in design, laboratory analysis, statistical analysis and manuscript writing.

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References

- Cardinale E, Gross-Claude J, Rivoal K, et al. . Epidemiologycal analysis
 of Salmonella enterica spp. serovar Hadar, Brancater and Enteriditis
 in humans and broilers chicken in Senegal using Pulsed Field Gel Electrophoresis and antibiotic susceptibility. J Food Protec. 2005;60:1312-7.
- Brenner FW, Villar RG, Angulo FJ, Tauxe R, Swaminathan B. Salmonella nomenclature. J Clin Microbiol. 2000;38(7):2465-7. [PubMed: 10878026].
- 3. Bell C, Kyriakides A. Salmonella: a practical approach to the organism and its control in foods. New Jersey: John Wiley & Sons; 2008.
- Cox NA, Bailey JS, Mauldin JM, Blankenship LC, Wilson JL. Extent of salmonellae contamination in breeder hatcheries. *Poult Sci.* 1991;70(2):416-8. [PubMed; 2027849].
- Heyndrickx M, Vandekerchove D, Herman L, Rollier I, Grijspeerdt K, De Zutter L. Routes for salmonella contamination of poultry meat: epidemiological study from hatchery to slaughterhouse. *Epidemiol Infect.* 2002;129(2):253-65. [PubMed: 12403101].
- Poppe C. Salmonella infections in the domestic fowl. In: Wray C, Wray A, editors. Salmonella in Domestic animals. New York, USA: CAB International; 2000. pp. 107–32.
- Liljebjelke KA, Hofacre CL, Liu T, White DG, Ayers S, Young S, et al. Vertical and horizontal transmission of salmonella within integrated broiler production system. *Foodborne Pathog Dis.* 2005;2(1):90–102. doi:10.1089/fpd.2005.2.90. [PubMed: 15992303].
- Le Bouquin S, Allain V, Rouxel S, Petetin I, Picherot M, Michel V, et al. Prevalence and risk factors for Salmonella spp. contamination in French broiler-chicken flocks at the end of the rearing period. *Prev* Vet Med. 2010;97(3-4):245–51. doi: 10.1016/j.prevetmed.2010.09.014. [PubMed: 20970209].
- Chambers JR, Bisaillon JR, Labbe Y, Poppe C, Langford CF. Salmonella prevalence in crops of Ontario and Quebec broiler chickens at slaughter. Poult Sci. 1998;77(10):1497-501. [PubMed: 9776057].
- Rose N, Beaudeau F, Drouin P, Toux JY, Rose V, Colin P. Risk factors for Salmonella enterica subsp. enterica contamination in French broiler-chicken flocks at the end of the rearing period. *Prev Vet Med*. 1999;39(4):265-77. [PubMed: 10327442].
- Carli KT, Eyigor A, Caner V. Prevalence of Salmonella serovars in chickens in Turkey. J Food Prot. 2001;64(11):1832-5. [PubMed: 11726169].
- Chadfield M, Skov M, Christensen J, Madsen M, Bisgaard M. An epidemiological study of Salmonella enterica serovar 4, 12:b:- in broiler chickens in Denmark. Vet Microbiol. 2001;82(3):233-47. [PubMed: 11470545].
- Smyser C, Adinarayanan N, Van Roekel H, et al. . Field and laboratory observations on Salmonella heidelberg infection in three chicken breeding flocks. Avian Dis. 1966;10(2):314–29.
- Guard-Petter J. The chicken, the egg and Salmonella enteritidis. Environ Microbiol. 2001;3(7):421-30. [PubMed: 11553232].
- Davies R, Breslin M. Environmental contamination and detection of Salmonella enterica serovar enteritidis in laying flocks. Vet Rec. 2001;149(23):699–704. [PubMed: 11787781].
- Kinde H, Castellan DM, Kerr D, Campbell J, Breitmeyer R, Ardans A. Longitudinal monitoring of two commercial layer flocks and their environments for Salmonella enterica serovar enteritidis and other Salmonellae. Avian Dis. 2005;49(2):189–94. doi:10.1637/7228-062704R. [PubMed: 16094821].
- Carrique-Mas JJ, Breslin M, Snow L, McLaren I, Sayers AR, Davies RH. Persistence and clearance of different Salmonella serovars in buildings housing laying hens. *Epidemiol Infect.* 2009;137(6):837-46. doi: 10.1017/S0950268808001568. [PubMed: 19017427].

- El-Lethey H, Huber-Eicher B, Jungi TW. Exploration of stress-induced immunosuppression in chickens reveals both stress-resistant and stress-susceptible antigen responses. *Vet Immunol Immunopathol*. 2003;95(3-4):91-101. [PubMed: 12963270].
- Humphrey T. Are happy chickens safer chickens? Poultry welfare and disease susceptibility. *Br Poult Sci.* 2006;47(4):379–91. doi: 10.1080/00071660600829084. [PubMed: 16905463].
- Khoshbakht R, Tabatabaei M, Hosseinzadeh S, Shekarforoush SS, Aski HS. Distribution of nine virulence-associated genes in Campylobacter jejuni and C. coli isolated from broiler feces in Shiraz, Southern Iran. Foodborne Pathog Dis. 2013;10(9):764-70. doi: 10.1089/fpd.2013.1489. [PubMed: 23789768].
- 21. Rahn K, De Grandis SA, Clarke RC, McEwen SA, Galan JE, Ginocchio C, et al. Amplification of an invA gene sequence of Salmonella typhimurium by polymerase chain reaction as a specific method of detection of Salmonella. *Mol Cell Probes.* 1992;**6**(4):271-9. [PubMed: 1528198].
- Gutierrez M, Fanning J, Murphy A, Murray G, Griffin M, Flack A, et al. Salmonella in broiler flocks in the republic of Ireland. *Foodborne Pathog Dis.* 2009;6(1):111–20. doi: 10.1089/fpd.2008.0163. [PubMed: 1906;369].
- Ansari-Lari M, Shekarforoush S, Mehrshad S, Safari H. Prevalence and risk factors for Salmonella spp. colonization in broiler flocks in Shiraz, southern Iran. Vet Res Forum. 2014;5(1):65-8. [PubMed: 25568695].
- 24. Bisgaard M, Haaning K, Velling G, editors. Poultry-borne salmonellosis: Prophylactic measures in the broiler production including bacteriological investigations before and after slaughter. 14 Nordic Veterinary Congress. 1982; Copenhagen (Denmark). Flagstad.
- Skov MN, Angen O, Chriel M, Olsen JE, Bisgaard M. Risk factors associated with Salmonella enterica serovar typhimurium infection in Danish broiler flocks. *Poult Sci.* 1999;78(6):848–54. [PubMed: 10438128].
- Bouwknegt M, Dam-Deisz WDC, Schouten JM, Wannet WJB, Van Pelt W, Visser G. Surveillance of zoonotic bacteria in farm animals in The Netherlands. Results from January 1998 until December 2000. 2003.
- 27. Mollenhorst H, van Woudenbergh CJ, Bokkers EG, de Boer IJ. Risk factors for Salmonella enteritidis infections in laying hens. *Poult Sci.*

- 2005;84(8):1308-13. [PubMed: 16156216].
- 28. Namata H, Meroc E, Aerts M, Faes C, Abrahantes JC, Imberechts H, et al. Salmonella in Belgian laying hens: an identification of risk factors. *Prev Vet Med.* 2008;83(3-4):323-36. doi: 10.1016/j.prevetmed.2007.09.002. [PubMed: 17961763].
- Huneau-Salaun A, Marianne C, Sophie le B, Francoise L, Isabelle P, Sandra R, et al. Risk factors for Salmonella enterica subsp. enterica contamination in 519 French laying hen flocks at the end of the laying period. *Prev Vet Med.* 2009;89(1-2):51-8. doi: 10.1016/j.prevetmed.2009.01.006. [PubMed: 19237216].
- Wales A, Breslin M, Davies R. Assessment of cleaning and disinfection in Salmonella-contaminated poultry layer houses using qualitative and semi-quantitative culture techniques. *Vet Microbiol.* 2006;116(4):283–93. doi:10.1016/j.vetmic.2006.04.026. [PubMed: 16730925].
- 31. Upadhyaya M, Poosaran N, Fries R. Prevalence and predictors of Salmonella spp. in retail meat shops in Kathmandu. *J Agric Sci Technol A*. 2012;2(9A):1094–106.
- Nimmermark S, Lund V, Gustafsson G, Eduard W. Ammonia, dust and bacteria in welfare-oriented systems for laying hens. *Ann Agric Environ* Med. 2009;16(1):103-13. [PubMed: 19630203].
- 33. Ellen HH, Bottcher RW, von Wachenfelt E, Takai H. Dust levels and control methods in poultry houses. *J Agric Saf Health*. 2000;**6**(4):275–82. [PubMed: 11217692].
- Chriel M, Stryhn H, Dauphin G. Generalised linear mixed models analysis of risk factors for contamination of Danish broiler flocks with Salmonella typhimurium. *PrevVet Med.* 1999;40(1):1–17. [PubMed: 10343330].
- Karimi-Madab M, Ansari-Lari M, Asasi K, Nili H. Risk factors for detection of bronchial casts, most frequently seen in endemic H9N2 avian influenza infection, in poultry flocks in Iran. Prev Vet Med. 2010;95(3-4):275-80. doi: 10.1016/j.prevetmed.2010.03.010. [PubMed: 20394998].
- Volkova VV, Wills RW, Hubbard SA, Magee D, Byrd JA, Bailey RH. Associations between vaccinations against protozoal and viral infections and Salmonella in broiler flocks. *Epidemiol Infect.* 2011;139(2):206-15. doi: 10.1017/S0950268810000804. [PubMed: 20426884].

