

The Genetic Heterogeneity of Common Variable Immunodeficiency (CVID)

Vassilios Lougaris¹, Alessandro Plebani^{1*}

¹Pediatrics Clinic and Institute for Molecular Medicine A. Nocivelli, Department of Clinical and Experimental Sciences, University of Brescia, ASST Spedali Civili of Brescia, Italy

Abstract

CVID represents the most frequent symptomatic primary humoral immunodeficiency. Clinical presentation includes hypogammaglobulinemia, recurrent infections, autoimmune phenomena and increased lymphoma and cancer risk. While the first cases were reported in the early 50's, the first genetic cause of CVID was described after 5 decades.

After the first description, and also thanks to the advances in the field of biomedical research, several additional genetic causes of CVID have been described. The current genetic landscape of CVID includes numerous genetic alterations that may cause or contribute to the development of CVID, underscoring the complexity and heterogeneity of this disorder.

Keywords: common variable Immunodeficiency, lymphoma, hypogammaglobulinemia, autoimmunity

* Corresponding author: Alessandro Plebani

Pediatrics Clinic and Institute for Molecular Medicine A. Nocivelli, Department of Clinical and Experimental Sciences, University of Brescia, ASST Spedali Civili of Brescia, Italy

E-mail: alessandro.plebani@unibs.it

Introduction

CVID is classically characterized by low immunoglobulin serum levels of at least two isotypes (IgG and/or IgA and/or IgM) with defective antibody response to recall immunizations, in the presence of normal peripheral B cell numbers (in the majority of cases). The clinical manifestations of CVID are variable and may include recurrent infections, mainly of the respiratory and gastrointestinal tract, autoimmune manifes-

tations, splenomegaly and lymphadenopathies, granulomata and increased susceptibility to cancer and lymphomas (1). The age of onset is variable, with a higher prevalence during the second and third decade of life; both sexes are involved in an equal manner. The complex and heterogeneous clinical and immunological phenotype of CVID, has long suggested that the pathogenesis of this disorder may be related to immunological alterations related to different genetic defects. In

Archive of SID

the last two decades, the genetic studies in patients with CVID have confirmed this hypothesis, underscoring a significant molecular and immunological complexity in CVID.

ICOS deficiency

While the first case of CVID was described in the early 50's (2), it was only after almost 5 decades that the first genetic cause of CVID was identified. In 2003, biallelic null mutations in ICOS were described in four adult-onset CVID patients resulting in lack of expression of ICOS (Inducible T-cell Costimulator), a T cell receptor that binds ICOS-ligand (ICOSL) expressed on B cells (3). Lymph node analysis from an ICOS deficient patient revealed important alterations in the formation and architecture of the germinal centers (4).

TACI deficiency

Soon after the description of human ICOS deficiency in CVID, mutations in TNFRSF13B encoding for TACI, expressed on B cells, were identified in patients affected with CVID (5-6). In a cohort of 162 unrelated CVID patients, Salzer et al identified the homozygous p.C104R, p.S144X mutations in TACI that resulted in lack of TACI expression on B cells, with defective binding to APRIL but not to BAFF, since the expression of the BAFF receptor (BAFF-R) was unaltered. Mutated patients' B cells failed to class

switch upon in vitro stimulation with APRIL and BAFF. Heterozygous variants (p.C104R, p.A181E, p.S194X and p.R202H) were also identified in CVID patients and were reported to be associated with defective antibody production (6). In parallel, a second group identified TNFRSF13B mutations in a small number of patients affected with CVID and SIgAD supporting the hypothesis that CVID and SIgAD may share similar pathogenetic mechanisms. Of note, in both studies, the features of patients' B cells did not reproduce the phenotype of the mouse model, confirming the existence of important differences between the animal and the hu-

man immune systems (5, 7).

Screening of larger cohorts of patients for TACI mutations revealed additional genetic variations (5, 6, 8-11), frequently in the heterozygous state, rendering difficult their interpretation in terms of pathogenicity. In a cohort of 564 CVID patients, the p.C104R monoallelic TACI mutation was present in 4.6% of affected patients and in 0.9% of 675 healthy controls, and resulted associated with an elevated risk for the development of hypogammaglobulinemia, lymphoproliferation and autoimmunity (11). Available data suggest that while biallelic TNFRSF13B variants that abrogate TACI on B cells are responsible for CVID, the role of heterozygous variants is still in debate.

TACI, together with BAFFR and BCMA, are members of a TNF receptor superfamily, that bind to BAFF and APRIL (BAFF-R binds only BAFF). Based on available knockout animal data, and after the identification of TACI mutations in patients with CVID, the candidate gene approach led to further investigation of the above mentioned genes.

BAFFR deficiency

Analysis of the TNFRSF13C gene encoding for BAFF-R in 48 patients (12) revealed the presence of three novel variants, all at the heterozygous state: p.P21R, p.G64V and p.H159Y. Of interest, the p.P21R variant was recently shown to alter the polymerization of BAFF-R on the surface of B cells, contributing therefore to the pathogenesis of CVID (13). Recently, biallelic mutations in TNFRSF13C, the gene encoding for BAFF-R, were identified in two siblings with adult-onset CVID and low peripheral B cell counts (14). They both carried a homozygous 24bp in-frame deletion (del89-96) located in exon 2 of the TNFRSF13C gene leading to the lack of BAFFR on B cell surface. Both patients presented hypogammaglobulinemia (low IgG and IgM, normal IgA serum levels). They also did not mount a T-independent immune response against pneumococcal cell wall polysaccharides.

The role of BAFF, BCMA and APRIL

Analysis of the genes encoding for the other members of this TNF receptor superfamily revealed a single novel synonymous variant, p.V63V, in a single patient in the heterozygous state in the TNFSF13B gene encoding for BAFF (15). Analysis of the genes encoding for BCMA and APRIL in CVID patients revealed the presence of the p.S81N, p.T159T, p.T175T, p.K179Q (BCMA) and p.G67R, p.N96S (APRIL) variants, at the same frequency as observed in healthy individuals, thus likely not involved in the pathogenesis of CVID (16).

CD19 deficiency

Following the description of mutations in TACI and BAFFR associated with the pathogenesis of CVID, CD19 deficiency was identified in four patients. Three patients harboured the homozygous deletion 1384del(ga), while one harboured the homozygous insertion 972ins(a). Both mutations led to a premature stop codon and thus lack of CD19 expression on the B cell surface. CD27⁺ memory B cells were decreased in affected patients, as typically observed in CVID (17). A fifth patient with CD19 deficiency was identified after the initial description, carrying a compound heterozygous mutation in the gene encoding for CD19 (18). More in detail, mutation analysis of CD19 revealed a mutation in the splice acceptor site of intron 5 (IVS5- 1G>T) of the maternal allele, resulting in skipping of exon 6, and a truncated protein product. The paternal allele was disrupted by a gross deletion encompassing at least the ATP2A1, CD19 and NFATC2IP genes (18).

CD20 deficiency

Upon CD19 deficiency, CD20 deficiency was identified in a single patient with hypogammaglobulinemia due to a compound mutation of the noncanonical splice donor sequence of exon 5 of the CD20 gene. The patient showed defective antibody formation upon T-independent antigen stimulation, similarly to what observed in the CD20 knockout mice (19).

CD81 deficiency

CD81 deficiency was subsequently identified in a patient with hypogammaglob, severe nephropathy and lack of CD19 expression on B cells was described. The lack of CD19 expression was due to CD81 deficiency, a member of the tetraspanin family that interacts with CD19 and CD21 on the B cell surface, due to a homozygous c.561+1G>A mutation in the CD81 gene resulting in a complete lack of CD81 expression. Memory B cells were reduced. As observed in CD19 deficiency, BCR cross-linking failed to activate properly B cells; on the contrary, T cell defects were not observed (20).

CD21 deficiency

An adult patient with recurrent infections and hypogammaglobulinemia presented with lack of CD21 expression on B cells during immunological work-up. Sequence analysis revealed a biallelic deleterious mutation in the CR2 gene (encoding CD21) (c.1225+1G>C; p.W766X). B cell maturation evaluation showed reduced class-switched memory B cells. In vitro experiments revealed absent co-stimulatory activity of C3d in enhancing suboptimal B-cell receptor stimulation. While vaccination responses to protein antigens were normal, the response to pneumococcal polysaccharide vaccination was moderately impaired (21).

The genetic alterations described so far as associated or causative of CVID were related to receptors expressed on the lymphocyte (B or T) cell surface. Novel genetic defects affecting cytoplasmic proteins have been reported afterwards in patients with clinical and immunological phenotype compatible with CVID.

PRKCD deficiency

Salzer et al, (22) reported on a single patient born to consanguineous parents, that presented with hypogammaglobulinemia, progressive B cell lymphopenia and severe autoimmunity. A homozygous (c.1352+1G>A) mutation affecting

Archive of SID

a splice site of PRKCD led to absent expression of the encoded protein. Another case of PRKCD deficiency due to a p.R614W homozygous deleterious mutation was reported in a single patient with chronic, low-grade Epstein-Barr virus infection (23). The patient had chronic lymphadenopathy, splenomegaly, autoantibodies, elevated immunoglobulins and natural killer dysfunction. Interestingly, the homozygous p.G510S homozygous mutation in PRKCD was identified in three patients affected with systemic lupus erythematosus, with B cell defective apoptosis and hyperproliferation (24), suggesting that biallelic mutation in PRKCD may give rise to variable clinical and immunological phenotypes.

PIK3CD and PIK3R1 Deficiency

Dominant-activating germline mutations in the gene encoding the PI(3)K catalytic subunit p110 δ were recently identified in patients with CVID-like phenotype (APDS-1; Activated phosphoinositide 3-kinase d syndrome) (25, 26). Affected patients presented various features, including lymphopenia, hypogammaglobulinemia, variable B and T cell maturational defects and, of note, T cell senescence (25, 26). Affected patients present an increased prevalence of B-cell lymphomas (27).

In addition, and shortly after the identification of APDS-1, monoallelic activating mutations in PIK3R1, the gene encoding for the regulatory subunit p85a of PI3K, were identified in patients with a CVID-like phenotype (APDS-2). Affected patients presented recurrent respiratory infections, gut involvement, enlarged lymph nodes and tonsils, normal to elevated IgM with low IgG and IgA serum levels, and variable lymphopenia (28-30). Of note, evaluation of NK cells in both APDS-1 and -2 revealed important functional defects (31, 32), a rather uncommon feature for CVID. Since in both APDS-1 and -2 the underlying genetic defect leads to PI3K hyperactivation, targeted inhibition of this cascade, for example with rapamycin, an mTOR inhibitor, or specific

PI3K inhibitors, is under clinical consideration (25, 26, 28, 29).

TWEAK deficiency

Three family members (father, daughter and son) affected with classical CVID were found to carry a rare genetic variant (p.R145C) in the gene encoding for TWEAK leading to TWEAK deficiency. Functional in vitro experiments showed that the mutant protein caused inhibition of BAFF-dependent B-cell survival and proliferation, suggesting thus a pathogenetic role of this mutant for CVID in this family (33).

LRBA deficiency

LRBA (lipopolysaccharide responsive beige-like anchor protein) deficiency was recently identified in patients with CVID and/or autoimmune disorders adding further complexity to the pathogenesis of CVID. The first five CVID patients identified harboured homozygous mutations (p.I2657S, p.R1683X, p.E59X and homozygous deletion including exons 1 and 2) in the gene encoding for LRBA that led to loss of protein expression (34). All patients had early onset hypogammaglobulinemia and severe autoimmune manifestations. Immunological evaluation revealed disturbed B cell development, defective in vitro B cell activation, immunoglobulin secretion and proliferation, and defects in B cell autophagy (34). LRBA deficiency due to a homozygous deletion from exon 1 to exon 30 was recently reported in a single patient with autoimmunity but without hypogammaglobulinemia, underlying that LRBA defects may present with variable immunological phenotypes (35). Considering the complicated clinical course of LRBA deficiency, as underlined by long-term follow-up of relatively numerous affected patients (36), HSCT has been implemented in a small number of cases, with variable results (36, 37). Recent experimental data showed that LRBA deficiency leads to defective CTLA-4 expression on T regulatory cells (37, 38), explaining thus the increased prevalence of autoimmunity in this

Archive of SID

disorder. Based on these findings, treatment with abatacept, a fusion protein composed by the Fc of IgG1 and the extracellular domain of CTLA-4, has been shown to be a valid alternative to HSCT for a large part of LRBA deficient patients (39).

NF- κ B1 and NF- κ B2 deficiency

Recently, monoallelic mutations in components of the NF- κ B pathway were identified in patients with CVID. Regarding the non-canonical NF- κ B pathway, monoallelic germline mutations in NFKB2 were described in a small number of patients affected with early onset hypogammaglobulinemia, recurrent infections, autoimmune features and adrenal insufficiency (40). The NFKB2 mutations identified lead to altered processing of p100, and therefore affect p52 nuclear translocation. Upon the initial description of NFKB2 deficiency, additional CVID patients with NFKB2 mutations have been described (41-43). Of interest, and similarly to what observed in APDS-1 and-2, NFKB2 mutated NK cells showed defective cytotoxic activity, a rather unusual feature for CVID (43).

Regarding the canonical pathway, monoallelic mutations in NFKB1 were initially identified in adult-onset CVID patients. Affected patients presented a classical CVID phenotype characterized by hypogammaglobulinemia, recurrent infections and variable autoimmune features. The identified mutations altered the normal processing of p105 and the nuclear localization of p50 (44). Evaluation of B cell maturation showed both early and late B cell developmental alterations in NFKB1 mutated patients (45). Following the identification of this genetic defect, additional cases have been described, broadening the clinical and immunological phenotype that now also includes EBV-driven lymphoproliferation and autoinflammatory manifestations (46, 47). Of interest, as observed in APDS-1, APDS-2 and NFKB2 deficiency, NFKB1 mutated NK cells revealed functional alterations as well as maturation perturbations (48).

CTLA-4 deficiency

A novel genetic form of CVID was recently

identified in monoallelic mutations in Cytotoxic T lymphocyte antigen 4 (CTLA-4). CTLA-4 is expressed in activated regulatory T cells and exhibits an inhibitory function on T cell biology. Affected patients present a complex syndrome of immune dysregulation characterized by variable features such hypogammaglobulinemia, lymphopenia, autoimmune cytopenias, lymphoproliferation and granulomas (49, 50), resembling LRBA deficiency (34, 36). They also seem to present an increased risk of gastric cancer, in accordance with recent experimental observations in animal models (51). Targeted treatments such as CTLA4-Ig, may become promising tools for patients affected with CTLA-4 or LRBA deficiency, since LRBA was recently shown to regulate CTLA-4 expression on T regulatory cells (37, 38).

ADA2 deficiency

ADA2 deficiency was originally described in patients with vasculitis and early-onset strokes (52). However, patients with a CVID-like phenotype were later identified with biallelic mutations in CECR1, causing ADA2 deficiency (53, 54), with important implications for prognosis and therapeutic strategies (55).

polymorphisms of MSH5

Besides the above mentioned genes, genes involved in the DNA repair process have also been implicated in the pathogenesis of CVID. This is the case of MSH5, a gene encoded in the central MHC class III region, which plays a critical role in regulating meiotic homologous recombination. Sekine et al. (56) presented evidence that the human MSH5 alleles containing two non-synonymous polymorphisms (p.L85F/p.P786S), may be involved in the pathogenesis of selective IgA deficiency and common variable immune deficiency (CVID).

RAG1/2 deficiency

Mutations in RAG1/2 are typically associated with severe combined immunodeficiency (57). However, hypomorphic mutations in these genes

Archive of SID

may be associated with a CVID-like phenotype in a limited number of patients (58- 60).

Conclusion

The scientific advances of the last two decades have allowed to shed light into the genetic alterations that contribute to the pathogenesis of CVID. These important findings underscore the fact that while patients affected with CVID may have a similar immunological phenotype, the underlying genetic causes may be diverse, with important implications in patients' clinical management, genetic counselling and prognosis.

Conflict of interest

The authors declare no conflicts of interests in regard with this study.

Acknowledgment

This research received no grant from any financial organizations or funding agency in the public, commercial or not-for-profit sectors. Also, there is no Financial & competing interest disclosure.

References

1. Cunningham-Rundles C. The many faces of common variable immunodeficiency. *Hematology Am Soc Hematol Educ Program*. 2012;2012:301-5.
2. Bruton OC, Apt L, Gitlin D, Janeway CA. Absence of serum gamma globulins. *AMA Am J Dis Child*. 1952;84(5):632-6.
3. Grimbacher B, Hutloff A, Schlesier M, Glocker E, Warnatz K, Dräger R, et al. Homozygous loss of ICOS is associated with adult-onset common variable immunodeficiency. *Nature Immunology*. 2003;4(3):261-8.
4. Warnatz K, Bossaller L, Salzer U, Skrabl-Baumgartner A, Schwinger W, van der Burg M, et al. Human ICOS deficiency abrogates the germinal center reaction and provides a monogenic model for common variable immunodeficiency. *Blood*. 2006;107(8):3045-52.
5. Castigli E, Wilson SA, Garibyan L, Rachid R, Bonilla F, Schneider L, et al. TACI is mutant in common variable immunodeficiency and IgA deficiency. *Nat Genet*. 2005;37(8):829-34.
6. Salzer U, Chapel HM, Webster AD, Pan-Hammarstrom Q, Schmitt-Graeff A, Schlesier M, et al. Mutations in TNFRSF13B encoding TACI are associated with common variable immunodeficiency in humans. *Nat Genet*. 2005;37(8):820-8.
7. Yan M, Wang H, Chan B, Roose-Girma M, Erickson S, Baker T, et al. Activation and accumulation of B cells in TACI-deficient mice. *Nat Immunol*. 2001;2(7):638-43.
8. Pan-Hammarstrom Q, Salzer U, Du L, Bjorkander J, Cunningham-Rundles C, Nelson DL, et al. Reexamining the role of TACI coding variants in common variable immunodeficiency and selective IgA deficiency. *Nat Genet*. 2007;39(4):429-30.
9. Castigli E, Wilson S, Garibyan L, Rachid R, Bonilla F, Schneider L, et al. Reexamining the role of TACI coding variants in common variable immunodeficiency and selective IgA deficiency. *Nat Genet*. 2007;39(4):430-1.
10. Lougaris V, Gallizzi R, Vitali M, Baronio M, Salpietro A, Bergbreiter A, et al. A novel compound heterozygous TACI mutation in an autosomal recessive common variable immunodeficiency (CVID) family. *Hum Immunol*. 2012;73(8):836-9.
11. Salzer U, Bacchelli C, Buckridge S, Pan-Hammarstrom Q, Jennings S, Lougaris V, et al. Relevance of biallelic versus monoallelic TNFRSF13B mutations in distinguishing disease-causing from risk-increasing TNFRSF13B variants in antibody deficiency syndromes. *Blood*. 2009;113(9):1967-76.
12. Losi CG, Silini A, Fiorini C, Soresina A, Meini A, Ferrari S, et al. Mutational Analysis of Human BAFF Receptor TNFRSF13C (BAFF-R) in Patients with Common Variable Immunodeficiency. *Journal of Clinical Immunology*. 2005;25(5):496-502.
13. Pieper K, Rizzi M, Speletas M, Smulski CR,

Archive of SID

- Sic H, Kraus H, et al. A common single nucleotide polymorphism impairs B-cell activating factor receptor's multimerization, contributing to common variable immunodeficiency. *J Allergy Clin Immunol*. 2014;133(4):1222-5.
14. Warnatz K, Salzer U, Rizzi M, Fischer B, Gutenberger S, Böhm J, et al. B-cell activating factor receptor deficiency is associated with an adult-onset antibody deficiency syndrome in humans. *Proc Natl Acad Sci U S A*. 2009;106(33):13945-50.
15. Losi CG, Salzer U, Gatta R, Lougaris V, Cattaneo G, Meini A, et al. Mutational analysis of human BLyS in patients with common variable immunodeficiency. *J Clin Immunol*. 2006;26(4):396-9.
16. Salzer U, Neumann C, Thiel J, Woellner C, Pan-Hammarström Q, Lougaris V, et al. Screening of functional and positional candidate genes in families with common variable immunodeficiency. *BMC Immunol*. 2008;9:3.
17. Zelm MC, Reisli I, van der Burg M, Castaño D, van Noesel CJM, van Tol MJD, et al. An Antibody-Deficiency Syndrome Due to Mutations in the CD19 Gene. 2006;354(18):1901-12.
18. Kanegane H, Agematsu K, Futatani T, Sira MM, Suga K, Sekiguchi T, et al. Novel mutations in a Japanese patient with CD19 deficiency. *Genes Immun*. 2007;8(8):663-70.
19. Kuijpers TW, Bende RJ, Baars PA, Grumels A, Derks IA, Dolman KM, et al. CD20 deficiency in humans results in impaired T cell-independent antibody responses. *J Clin Invest*. 2010;120(1):214-22.
20. Zelm MC, Smet J, Adams B, Mascart F, Schandene L, Janssen F, et al. CD81 gene defect in humans disrupts CD19 complex formation and leads to antibody deficiency. *J Clin Invest*. 2010;120(4):1265-74.
21. Thiel J, Kimmig L, Salzer U, Grudzien M, Lebrecht D, Hagena T, et al. Genetic CD21 deficiency is associated with hypogammaglobulinemia. *J Allergy Clin Immunol*. 2012;129(3):801-10.e6.
22. Salzer E, Santos-Valente E, Klaver S, Ban SA, Emminger W, Prengemann NK, et al. B-cell deficiency and severe autoimmunity caused by deficiency of protein kinase C δ . *Blood*. 2013;121(16):3112-6.
23. Kuehn HS, Niemela JE, Rangel-Santos A, Zhang M, Pittaluga S, Stoddard JL, et al. Loss-of-function of the protein kinase C δ (PKC δ) causes a B-cell lymphoproliferative syndrome in humans. *Blood*. 2013;121(16):3117-25.
24. Belot A, Kasher PR, Trotter EW, Foray AP, Debaud AL, Rice GI, et al. Protein kinase c δ deficiency causes mendelian systemic lupus erythematosus with B cell-defective apoptosis and hyperproliferation. *Arthritis Rheum*. 2013;65(8):2161-71.
25. Angulo I, Vadas O, Garçon F, Banham-Hall E, Plagnol V, Leahy TR, et al. Phosphoinositide 3-kinase delta gene mutation predisposes to respiratory infection and airway damage. *Science*. 2013;342(6160):866-71.
26. Lucas CL, Kuehn HS, Zhao F, Niemela JE, Deenick EK, Palendira U, et al. Dominant-activating germline mutations in the gene encoding the PI(3)K catalytic subunit p110 δ result in T cell senescence and human immunodeficiency. *Nat Immunol*. 2014;15(1):88-97.
27. Kracker S, Curtis J, Ibrahim MAA, Sediva A, Salisbury J, Camp V, et al. Occurrence of B-cell lymphomas in patients with activated phosphoinositide 3-kinase δ syndrome. *The Journal of allergy and clinical immunology*. 2014;134(1):233-6.
28. Deau MC, Heurtier L, Frange P, Suarez F, Bole-Feysot C, Nitschke P, et al. A human immunodeficiency caused by mutations in the PIK3R1 gene. *J Clin Invest*. 2014;124(9):3923-8.
29. Lucas CL, Zhang Y, Venida A, Wang Y, Hughes J, McElwee J, et al. Heterozygous splice mutation in PIK3R1 causes human immunodeficiency with lymphoproliferation due to dominant activation of PI3K. *J Exp Med*. 2014;211(13):2537-47.

30. Lougaris V, Faletra F, Lanzi G, Vozzi D, Marcuzzi A, Valencic E, et al. Altered germinal center reaction and abnormal B cell peripheral maturation in PI3KR1-mutated patients presenting with HIGM-like phenotype. *Clin Immunol*. 2015;159(1):33-6.
31. Lougaris V, Patrizi O, Baronio M, Tabellini G, Tampella G, Lanzi G, et al. p85 α is an intrinsic regulator of human natural killer cell effector functions. 2016;138(2):605-8. e3.
32. Ruiz-García R, Vargas-Hernández A, Chinn IK, Angelo LS, Cao TN, Coban-Akdemir Z, et al. Mutations in PI3K110 δ cause impaired natural killer cell function partially rescued by rapamycin treatment. *J Allergy Clin Immunol*. 2018;142(2):605-17.e7.
33. Wang HY, Ma CA, Zhao Y, Fan X, Zhou Q, Edmonds P, et al. Antibody deficiency associated with an inherited autosomal dominant mutation in TWEAK. *Proc Natl Acad Sci U S A*. 2013;110(13):5127-32.
34. Lopez-Herrera G, Tampella G, Pan-Hammarström Q, Herholz P, Trujillo-Vargas CM, Phadwal K, et al. Deleterious mutations in LRBA are associated with a syndrome of immune deficiency and autoimmunity. *Am J Hum Genet*. 2012;90(6):986-1001.
35. Burns SO, Zenner HL, Plagnol V, Curtis J, Mok K, Eisenhut M, et al. LRBA gene deletion in a patient presenting with autoimmunity without hypogammaglobulinemia. 2012;130(6):1428-32.
36. Gamez-Diaz L, August D, Stepensky P, Revel-Vilk S, Seidel MG, Noriko M, et al. The extended phenotype of LPS-responsive beige-like anchor protein (LRBA) deficiency. *J Allergy Clin Immunol*. 2016;137(1):223-30.
37. Bakhtiar S, Gamez-Diaz L, Jarisch A, Soerensen J, Grimbacher B, Belohradsky B, et al. Treatment of Infantile Inflammatory Bowel Disease and Autoimmunity by Allogeneic Stem Cell Transplantation in LPS-Responsive Beige-Like Anchor Deficiency. *Front Immunol*. 2017;8:52.
38. Alroqi FJ, Charbonnier LM, Baris S, Kiykim A, Chou J, Platt CD, et al. Exaggerated follicular helper T-cell responses in patients with LRBA deficiency caused by failure of CTLA4-mediated regulation. *J Allergy Clin Immunol*. 2018;141(3):1050-9 e10.
39. Lo B, Zhang K, Lu W, Zheng L, Zhang Q, Kanellopoulou C, et al. AUTOIMMUNE DISEASE. Patients with LRBA deficiency show CTLA4 loss and immune dysregulation responsive to abatacept therapy. *Science*. 2015;349(6246):436-40.
40. Kiykim A, Ogulur I, Dursun E, Charbonnier LM, Nain E, Cekic S, et al. Abatacept as a Long-Term Targeted Therapy for LRBA Deficiency. *J Allergy Clin Immunol Pract*. 2019;7(8):2790-800.e15.
41. Lindsley AW, Qian Y, Valencia CA, Shah K, Zhang K, Assa'ad A. Combined immune deficiency in a patient with a novel NFKB2 mutation. *J Clin Immunol*. 2014;34(8):910-5.
42. Lee CE, Fulcher DA, Whittle B, Chand R, Fewings N, Field M, et al. Autosomal-dominant B-cell deficiency with alopecia due to a mutation in NFKB2 that results in nonprocessable p100. *Blood*. 2014;124(19):2964-72.
43. Lougaris V, Tabellini G, Vitali M, Baronio M, Patrizi O, Tampella G, et al. Defective natural killer-cell cytotoxic activity in NFKB2-mutated CVID-like disease. *J Allergy Clin Immunol*. 2015;135(6):1641-3.
44. Fliegauf M, Bryant VL, Frede N, Slade C, Woon ST, Lehnert K, et al. Haploinsufficiency of the NF- κ B1 Subunit p50 in Common Variable Immunodeficiency. *Am J Hum Genet*. 2015;97(3):389-403.
45. Lougaris V, Moratto D, Baronio M, Tampella G, van der Meer JWM, Badolato R, et al. Early and late B-cell developmental impairment in nuclear factor kappa B, subunit 1-mutated common variable immunodeficiency disease. *J Allergy Clin Immunol*. 2017;139(1):349-52.e1.
46. Schipp C, Nabhani S, Bienemann K, Si-

- manovsky N, Kfir-Erenfeld S, Assayag-Ashe-rie N, et al. Specific antibody deficiency and autoinflammatory disease extend the clinical and immunological spectrum of heterozygous NFKB1 loss-of-function mutations in humans. *Haematologica*. 2016;101(10):e392-e6.
47. Boztug H, Hirschmugl T, Holter W, Lakatos K, Kager L, Trapin D, et al. NF- κ B1 Haploinsufficiency Causing Immunodeficiency and EBV-Driven Lymphoproliferation. *J Clin Immunol*. 2016;36(6):533-40.
48. Lougaris V, Patrizi O, Baronio M, Tabellini G, Tampella G, Damiani E, et al. NFKB1 regulates human NK cell maturation and effector functions. *Clin Immunol*. 2017;175:99-108.
49. Schubert D, Bode C, Kenefeck R, Hou TZ, Wing JB, Kennedy A, et al. Autosomal dominant immune dysregulation syndrome in humans with CTLA4 mutations. *Nat Med*. 2014;20(12):1410-6.
50. Kuehn HS, Ouyang W, Lo B, Deenick EK, Niemela JE, Avery DT, et al. Immune dysregulation in human subjects with heterozygous germline mutations in CTLA4. *Science*. 2014;345(6204):1623-7.
51. Miska J, Lui JB, Toomer KH, Devarajan P, Cai X, Houghton J, et al. Initiation of inflammatory tumorigenesis by CTLA4 insufficiency due to type 2 cytokines. *J Exp Med*. 2018;215(3):841-58.
52. Zhou Q, Yang D, Ombrello AK, Zavialov AV, Toro C, Zavialov AV, et al. Early-onset stroke and vasculopathy associated with mutations in ADA2. *N Engl J Med*. 2014;370(10):911-20.
53. Schepp J, Proietti M, Frede N, Buchta M, Hubscher K, Rojas Restrepo J, et al. Screening of 181 Patients With Antibody Deficiency for Deficiency of Adenosine Deaminase 2 Sheds New Light on the Disease in Adulthood. *Arthritis Rheumatol*. 2017;69(8):1689-700.
54. Schepp J, Bulashevskaya A, Mannhardt-Laakmann W, Cao H, Yang F, Seidl M, et al. Deficiency of Adenosine Deaminase 2 Causes Antibody Deficiency. *J Clin Immunol*. 2016;36(3):179-86.
55. Ombrello AK, Qin J, Hoffmann PM, Kumar P, Stone D, Jones A, et al. Treatment Strategies for Deficiency of Adenosine Deaminase 2. *N Engl J Med*. 2019;380(16):1582-4.
56. Sekine H, Ferreira RC, Pan-Hammarstrom Q, Graham RR, Ziembra B, de Vries SS, et al. Role for Msh5 in the regulation of Ig class switch recombination. *Proc Natl Acad Sci U S A*. 2007;104(17):7193-8.
57. Sobacchi C, Marrella V, Rucci F, Vezzoni P, Villa A. RAG-dependent primary immunodeficiencies. *Human mutation*. 2006;27(12):1174-84.
58. Abolhassani H, Wang N, Aghamohammadi A, Rezaei N, Lee YN, Frugoni F, et al. A hypomorphic recombination-activating gene 1 (RAG1) mutation resulting in a phenotype resembling common variable immunodeficiency. *J Allergy Clin Immunol*. 2014;134(6):1375-80.
59. Buchbinder D, Baker R, Lee YN, Ravell J, Zhang Y, McElwee J, et al. Identification of patients with RAG mutations previously diagnosed with common variable immunodeficiency disorders. *J Clin Immunol*. 2015;35(2):119-24.
60. Schröder C, Baerlecken NT, Pannicke U, Dörk T, Witte T, Jacobs R, et al. Evaluation of RAG1 mutations in an adult with combined immunodeficiency and progressive multifocal leukoencephalopathy. *Clin Immunol*. 2017;179:1-7.