## Comparison of Induced Gene Response to Stressful Treatment in Spinacia oleracea and Brassica napus

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## **Abstract**

To study of some photosynthetic and defense genes expression in Spinacia Oleracea (cv. Vienna) and Brassica napus (cv. Falcon) seeds were grown at growth room. In a series of primary experiment a varied range of stress treatment with different concentration were compared. Finally, three treatments included methyl viologen, silver nitrate and 3-amin-trizoil have been selected for further experiment. All treatments were sprayed on the leaves at the maximum of vegetative growth stage. A CRD (completely Randomized Design) order has been used with four replicates. In order to assess cellular oxidative levels the TBARM assay has been used. Sampling has been done at intervals 6, 12, 24, 48 and 72 hours from the leaf treated. To analyzed gene expression and northern hybridization, sampling has been done only 48 hours after the leaves had sprayed. Since of so many results, here we have reported all results included percentage of cell death, TBARM amount and gene expression, for all the treatments only 48h after treatment sprayed. The results showed all treatments have altered the amount of cell death, TBARM and gene expression via increase of reactive oxygen species (ROS). The expression of photosynthetic gene RBCS declined by increasing of stressful treatment concentration. Sprayed pre-treatment ascorbic acid caused relative gene activity increase up to 50%. Also, this improved the physical damage and decline of TBARM by quenching of ROS. For the rest of genes although there were some differences among gene expression in depend on which plant or treatment has been checked out, but in general all genes showed a positive response to stressful treatments. For these genes with no exception using pretreatment of ascorbic acid caused some altered toward control expression.

Keywords: Gene expression, Spinach, Brassica napus, Ascorbic acid, Antioxidant

## References

Amory, A. M., Ford, L., Pammenter, N. W. and Cresswell, C. F. 1992. The use of 3-amino-1,2,4-triazole to investigate the short-term effects of oxygen toxicity on carbon assimilation by Pisum sativum seedlings. Plant Cell and Environment 15: 655–663

Bowler, C., Van Montagu, M. and Inze, D. 1992. Superoxide dismutase and stress tolerance. Annual Review of Plant Physiology and Plant Molecular Biology 43: 83-116

Butt, A., Mousley, K., Morris, K., Beynon, J., Can, C., Holub, E., Greenberg, J. T. and Buchanan-Wollaston, V. 1998. Differential expression of a senescence-enhanced metallothionein in *Arabidopsis* in response to isolates of *Peronospora parasitica* and *Psendomonas syringae*. Plant Journal 16 (2): 209-221

Choi, H. W., Kim, Y. J., Lee, S. C., Hong, J. K. and Hwang, B. K. 2007. Hydrogen peroxide generation by the pepper extracellular peroxidase CaPO2 activates local and systemic cell death and defense response to bacterial pathogens. Plant Physiology 145: 890–904

Christopher, M., Salah, E., Kathryn, A., Alexander, M., Jared, R., Jeffrey, A., Krishna, K. and Marinus, P. 2009. Copper Delivery by the Copper Chaperone for Chloroplast and Cytosolic Copper/Zinc-Superoxide Dismutases: Regulation and Unexpected Phenotypes in an *Arabidopsis* Mutant. Molecular Plant 2 (6): 1336-50

Clijsters, H., Cuypers, A. and Vangronsveld, J. 1999. Physiological responses to heavy metals in higher plants; defence against oxidative stress. Zew Naturforsch 54: 730-734

Doke, N. and Miura, Y. 1995. *In vitro* activation of NADPH-dependent O<sub>2</sub><sup>-</sup> generation system in a plasma membrane-rich fraction of potato tuber tissues by treatment with an elicitor from *Phytophthora infestans* or with digitonin. Physiology and Molecular Plant Pathology 46: 17-28

Epple, P., Apel, K. and Bohlmann, H. 1995. An *Arabidopsis thaliana* thionin gene is inducible via a signal transformation pathway different from that for pathogenesis-related proteins. Plant Physiology 109: 813-820

Foley, R. C., Liang, Z. M. and Singh, K. B. 1997. Analysis of type 1 metallothionein cDNA in *Vicia faba*. Plant Molecular Biology 33: 583-591

Golden, T. A., Hinerfeld, D. and Melov, S. 2002. Oxidative stress and aging: beyond correlation. Aging Cell 117–123

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- Gupta, A. S., Webb, R., Holaday, A. S. and Allen, R. 1993. Overexpression of Superoxide Dismutase Protects Plants from Oxidative Stress. Plant Physiology 103: -1073
- Hagege, D., Nouvelot, A., Boucard, J. and Gaspar, T. 1990. Malondialdehyde titration with thiobarbiturate in plant extracts: avoidance of pigment interference. Phytochemistry Anall 1: 86-89
- Kolupaev, Y. U., Karpets, Y. V., and Kosakivska, I. V. 2008. The importance of reactive oxygen species in the induction of plant resistance to heat stress. Plant Phisology 34: 251-266
- Larkindale, J. D., Hall, J. R., Knight, M. and Vierling, E. 2005. Heat stress phenotypes of *Arabidopsis* mutants implicate multiple signaling pathways in the acquisition of thermotolerance. Plant Physiology 138: 882–897
- Lewis, N. G. and Yamamoto, E. 1990. Lignin-Occurrence biogenesis and biodegradation. Annual Review Plant Physiology and Plant Molecular Biology 41: 455-496
- Mackerness, S. A. H., Jordan, B. R. and Thomas, B. 1999. Reactive oxygen species in the regulation of photosynthetic genes by ultraviolet-B radiation (UV-B: 280-320 nm) in green and etiolated buds of pea (*Pisum sativum L.*). Journal Photochemistry and Photobiolgy 48: 180-188
- Mackerness, S. A. H. 2000. Plant responses to ultraviolet-B (280-320 nm) stress. What are the key regulators? Plant Growth Regulation 32: 27-39
- Mackerness, S. A. H., John, C. F. Jordan, B. and Thomas, B. 2001. Early signaling components in ultraviolet-B responses: distinct roles for different reactive oxygen species and nitric oxide. FEBS Letter 489: 237-242
- Manning, V., Chu, A., Steeves, J., Wolpert, T. and Ciuffetti, L. 2009. A host-selective toxin of *Pyrenophora tritici-repentis*, ptr toxA, induces photosystem changes and reactive oxygen species accumulation in sensitive wheat. American Phytopathological Society 22 (6): 665–676
- Navabpour, S., Morris, K., Allen, R., Harrison, E., Mackerness, S. and Buchanan- Wollaston, V. 2003. Expression of senescence–enhanced genes in response to oxidative stress. Experimental Botany 54 (391): 2285 -2292
- Navabpour, S., Bagherieh –Najjar, M. B. and Soltanloo, H. 2007. Identification of novel genesexpressed in *Brassica* napus during leaf senescence and in response to oxidative stress. Plant production 1(1): 35 -44
- Neill, S. J., Desikan, R., Clarke, A., Hurst, R. and Hancock, J. T. 2001. Hydrogen peroxide and nitric oxide as signalling molecules in plants. Journal Experimetal Botany 52:9-17
- Orozco-Cardenas, M. L., Narvaez-Vasquez, J. and Ryan, C. A. 2001. Hydrogen peroxide acts as a second messenger for the induction of defense genes in tomato plants in response to wounding, systemin, and methyl jasmonate. Plant Cell 13: 179-191
- Peyvast O. A. 2002. Olericulture. Agricultural Sciences Press. 2<sup>nd</sup> ed. 248P
- Pitzschke, A., Forrzani, C. and Hirt, H. 2006. Reactive oxygen species signaling in plants. Antioxid Redox Signal 8: 1757–1764
- Qi- lin, D., Chen, C. H., Bin, F., Ting-Ting, L., Yuan, G., Ying-Kun, S., Jin, W. and Shi-Zhang, D. 2009. Effects of NaCL treatment on the antioxidant enzymes of oilseed rape (*Brassica napus*) seedlings. African Journal of Biotechnology 8 (20): 5400 -5405
- Robertson McClung, C. 1997. Regulation of catalases in *Arabidopsis*. Free Radical Biology and Medicine 23 (3): 489-496
- Robinson, N. j., Tommey, A. M., Kuske, C. and Jackson, P. 1993. Plant metallothioneins. Biochemistry Journal 295: 1-10
- Sanchez-Casas, P. and Klessig, D. F. 1994. A salicylic acid-binding activity and a salicylic acid-inhibitable catalase activity are present in a variety of plant species. Plant Physiology 106: 1675-1679
- Shariati S. H. and Shahnizadeh Q. 2000. Kolza. Statistic and Information Institute (Agricultural Affeirs). 115P
- Sturgeon, B. E., Sipe, H. J., Barr, D. P., Corbett, J. T., Martinez, J. G. and Mason, R. P. 1998. The fate of the oxidizing tyrosyl radical in the present of glutathione and ascorbate. Journal Biology Chemistry 273: 30116-30121

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