

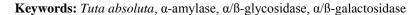
Research Article

Biochemical characterization of digestive carbohydrases of the tomato leaf miner, *Tuta absoluta* (Lepidoptera: Gelechiidae) larvae in response to feeding on six tomato cultivars

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Abstract: The tomato leaf miner, Tuta absoluta (Meyrick) is an imported pest and serious threat to tomato production in farms and greenhouses of Iran. Use of genetically engineered plants expressing carbohydrase inhibitors is one of the nonchemical methods for controlling insect pests, and knowledge about enzymatic properties of carbohydrases will help us to achieve this goal. Therefore, in present study we characterized biochemical properties of digestive carbohydrases in the midgut of last larval instar of T. absoluta fed on different tomato cultivars (Kingston, Riogrande, Super Luna, Super Chief, Super strain B and Calj). While the highest amylolytic activity was on Super strain B, the lowest was on Super Chief. The optimal pH and temperature for α-amylase were found to be at pH 9.0 and 45 °C, respectively. As calculated from Lineweaver-Burk plots, the highest K_m and V_{max} values for α -amylase obtained in Super Chief and Super Luna cultivars were 0.565 ± 0.11 mM and 2.287 ± 0.4 mM/min, respectively. The effects of different compounds on amylolytic activity indicated that CaCl₂, MgCl₂, NaCl and KCl increased amylase activity, whereas EDTA, ZnCl2 and BaCl2 decreased the enzyme activity in Super Luna cultivar. The highest activity of α -/ β -glucosidases was observed at pH 6.0 and 7.0, respectively, whereas the optimal pH for α/βgalactosidases was at 5.0. The highest specific activity of α -/ β -glucosidases was determined in Riogrande-fed larvae, whereas the highest α/β-galactosidases activity was in the larvae fed on Riogrande and Calj cultivars, respectively. By the native- PAGE, two bands were clearly detected for α-amylase. Since the larvae reared on Kingston showed lowest carbohydrase activities, this cultivar could possibly be suggested as the least suitable host for feeding of *T. absoluta*.



Introduction

Insects and plants are considered just as a part of the complex interactions in the

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*Corresponding author, e-mail: fs_nasab@yahoo.com Received: 29 January 2017, Accepted: 25 August 2017 Published online: 16 September 2017 natural ecosystem (Mello and Silva-Filho, 2002). In these complex interactions, various defense mechanisms are developed by plants to protect them against insect attacks. These defensive mechanisms include defensive protein production (Birkett *et al.*, 2000), secondary metabolites (Baldwin, 2001) and trichome density (Fordyce and Agrawal, 2001). On the other hand, various

mechanisms have also been developed by insects in encountering the deleterious effects of the proteinase inhibitors existing plants. their host These counter mechanisms include enhancement of their digestive enzyme activities or production of less sensitive enzymes to inhibitors by the insects (Paulillo et al., 2000); hydrolyzing inhibitors by different types of digestive proteases (Girard et al., 1998); and overproduction of proteinases insensitive to inhibitors using up regulation to compensate for the inhibited ones (Mello and Silva-Filho, 2002).

The tomato leaf miner, Tuta absoluta (Meyrick), is one of the most destructive pests of tomato in different countries including Iran that mainly attacks solanaceous crops. This pest has also been considered as one of the important insect pests of tomato crops since 2010 at different regions of Iran (Baniameri and Cheraghian, 2012). The chemical control as the primary method was used for restricting and control of this pest, unfortunately some populations of T. absoluta have developed resistance against the applied insecticides (Roditakis et al., 2013; Silva et al., 2011).

The larvae of many lepidopteran species have become capable of feeding on various host plants to get the essential nutrients for their optimal development and growth. The food sources quality and the secondary metabolites or enzymes inhibitors present in host plants, are the two determinant factors that influence the digestive enzymes of the insects. Poor utilization of nutrients and developmental retardation can occur as the consequence of any interference in digestive enzymes' activity by the enzyme-inhibitors of the host plant (Jongsma and Bolter, 1997).

Carbohydrases have major role in digestive physiology and study on their inhibitors has become an important issue. Great potential has been shown by the plant α -amylase inhibitors as a control tool for making plants resistant to pests. These inhibitors are proteins that can be found in

diverse types of plants. Moreover, these proteins are considered as vital elements in natural defenses, particularly against pests feeding on starchy food. Insect α-amylase inhibitors are an effective device for controlling of insect pests (Shade et al., 1994). There is a little information about carbohydrases in the insects that feed on various host plants. The digestive enzymes activities were investigated in Helicoverpa armigera (Hübner) (Lep.: Noctuidae) that fed on six tomato cultivars (Nemati Kalkhoran et al. 2013). The larvae reared on the leaves of Cal JN3 and Korral cultivars showed lowest amylolytic activity and the highest amylolytic activity was determined in the larvae fed on the leaves of SUN 6108 f1.

To achieve a better perception and knowledge about the digestive physiology of tomato leaf minor, the present research has attempted to identify the biochemical properties of the carbohydrate hydrolyzing enzymes in the *T. absoluta* midgut after feeding on six tomato cultivars.

Materials and Methods

Chemicals

Starch, acrylamide, bisacrylamide, ammonium per sulphate p-Nitrophenol and bovine serum albumin were purchased from Merck (Darmstadt, Germany). p-Nitrophenyl- α -D-glucopyranoside (pN α G), p-Nitrophenyl- β -D-glucopyranoside (pN α G), p-Nitrophenyl- α -D-galactopyranoside (pN α Ga), and p-Nitrophenyl- β -D-galactopyranoside (pN α Ga) were obtained from sigma (St. Louis, USA).

Host plant and insect rearing

Six cultivars of tomato including Kingston, Riogrande, Super Luna, Super Chief, Super strain B and Calj, were used in this study.

The plants were separately planted in plastic pots (19 cm diameter, 14 cm depth) and maintained in insecticide free conditions in a greenhouse at Urmia Agricultural Research Institute, West Azerbaijan, Iran. *T. absoluta* larvae were originally collected

from tomato fields in west Azerbaijan province, Urmia, Isakan (N 37°32'15.1" E 45°14'38.5"). Newly emerged adults were transferred to rearing cages ($50 \times 50 \times 80$ cm) containing 2-3 insect free potted tomato plants at a sex ratio of 2 females:3 males and fed with 10% sugar solution. The tomato plants infested with T. absoluta eggs were placed in an insect-proof rearing cage to allow larval development. The insects reared separately on each cultivar for three generations and synchronized 4th-instar larvae were used for experiments. All experiments were conducted under laboratory conditions at $26 \pm 1^{\circ}$ C, 60-70% RH and 16:8 h light: dark photoperiod.

Sample preparation and enzyme assays

Fourth-instar larvae reared on different tomato cultivars were immobilized on ice and dissected under a stereomicroscope. Midgut was collected and transferred to a freezer (-80 °C). For measuring the enzyme activity, the samples were homogenized with a hand-held glass grinder on ice, and the homogenates were centrifuged at $12000 \times g$ for $10 \, \text{min}$ at $4 \, ^{\circ}\text{C}$.

α-amylase activity and its kinetic parameters

The dinitrosalicylic acid (DNS) procedure was used to determine the activity of α -amylase (Bernfeld, 1955). The supernatant (10µl) was added to a tube containing 40 µl of the buffer (50 mM; phosphate, boric-citric acid and glycine) and 50µl of 1% (w/v) starch and incubated for 30 min. The reaction was stopped by the addition of 100 µl DNS. The mixture was heated in boiling water for 10 min. Finally, the absorbance was recorded at 540 nm with a microplate reader model Stat Fax®3200 (Awareness Technology Inc.). Appropriate blanks were included in the experiments as well as. All assays were performed in triplicate.

Catalytic activities (kinetic parameters) of the enzymes were investigated at different concentrations of starch in a range of 0.03-1% (w/v) in 40μ l universal buffer, pH 9.0. The

Michaelis-Menten constant (K_m) and maximal velocity (V_{max}) were estimated from the Lineweaver- Burk plots. The kinetic values are the averages of three experiments.

Effect of temperature on α-amylase activity

The activity of α -amylase was determined by incubating the reaction mixture at 15, 25, 35, 45, 55 and 65 °C for 30 min. After the incubation time, remain activity was measured as mentioned above.

$\alpha\text{-}$ and $\beta\text{-}glucosidase$ and $\alpha\text{-}$ and $\beta\text{-}galactosidase$ activities

The activities of α - and β -glucosidases and α and β-galactosidases were measured with p-Nitrophenyl-α-D-glucopyranoside (pNαG), pnitrophenyl-β-D-glucopyranoside (pNβG), pnitrophenyl- α -D-galactopyranoside (pN α Ga) p-nitrophenyl-β-D-galactopyranoside (pNβGa) as substrates, respectively, based on Low et al. (1986). Homogenates were incubated for 30 min at 37 °C with 45 µl of substrate (25mM) and 115µl of 50mM phosphate, boric-citric acid and glycine mixed buffer. The reaction was stopped by addition of 600µl of NaOH (0.25M). Optical density was measured at 405nm using microplate reader Stat Fax®3200 (Awareness Technology Inc.). Blanks without enzymes were included.

Effect of pH on enzyme activity

The effect of different pH on the activities of α -amylase, α - and β -glucosidases and α -and β -galactosidases were determined at room temperature in a mixed buffer containing phosphate, boric-citric acid and glycine (50mM of each) adjusted to various pH values (pH 3.0 to 12.0) by adding HCl or NaOH for acidic and basic pH values, respectively.

Protein concentration

Protein concentrations were estimated as originally described by Bradford (1976), using bovine serum albumin as standard.

Effect of activators and inhibitors on α -amylase activity

The effects of various metal ions including Na^+ , K^+ , Mg^{2+} , Ba^{2+} , Zn^{2+} , Ca^{2+} and ethylene diamine tetraacetic acid (EDTA) (20 mM) on α -amylase activity were determined. After 30 min of incubation of ions and EDTA with enzyme at room temperature, remain activity was measured as mentioned above.

Polyacrylamide gel electrophoresis and zymogram analysis

The visualization of α -amylase activity present in homogenates of larval midgut fed on different tomato cultivars was carried out using non-denaturing polyacrylamide gel electrophoresis (PAGE) based on the procedure of Davis (1960). Electrophoresis was conducted in an 8% (w/v) separating gel and a 5% stacking gel with 100 V at 4 °C. After running, the gel was incubated in 2.5% (v/v) Triton X-100 for 30 min at room temperature with mild agitation. After incubation, the gel was rinsed with deionized water to remove the triton x-100. Gel was then incubated in 1% starch for 120 min, and gel strips were stained with lugol's solution to detect α -amylase activity as white bands in a dark background.

Statistical analysis

The data were compared by one-way analysis of variance (ANOVA) followed by Tukey's test when significant differences were found at $P \le 0.05$ using SAS software (SAS Institute 1997).

Results

$\alpha\text{-amylase}$ activity and effect of pH and temperature on its activity

Amylolytic activity in midgut extracts from larvae fed on the different tomato cultivars are presented in Table 1. Among the tomato cultivars, larvae reared on Super strain B had the highest and those on Super chief showed the lowest levels of amylolytic activity. Results showed that the optimal pH for α -amylase in the digestive system was 9.0 (Fig. 1). The *T. absoluta* α -amylase has an optimum activity at 45 °C (Fig. 2).

Kinetic parameters of α-amylase

Alpha- amylases revealed a Michaelis-Menten type kinetics when hydrolyzing starch at their optimum pH. The K_m and V_{max} values for α -amylase hydrolyzing starch as substrate are presented in Table 2.

Effect of pHs on α - and β -glucosidase and α and β -galactosidase activities

The effect of pHs on the hydrolytic activity of α - and β -glucosidase and α -and β -galactosidase towards pN α G, pN β G, pN α Ga and pN β Ga was tested using a mixed buffer containing phosphate, boric-cirtic acid and glycine (50 mM of each) (pH 3.0-12.0). Maximum activity in the midgut was observed at pH 6.0 and 7.0 for α - and β -glucosidase, respectively, whereas, the optimal pH for both α -and β -galactosidase activity was 5.0 (Fig. 1).

Table 1 The effects of tomato cultivars on specific activity (mM/min/mg protein) of digestive carbohydrases in *Tuta absoluta* larvae.

Enzymes	Specific activity (mM/min/mg protein) (Mean \pm SE)					
	Calj	Riogrande	Kingston	Super Luna	Super strain B	Super Chief
α-amylase	$4.14 \pm 0.38ab$	$5.05 \pm 0.17a$	5.11 ± 0.29a	$4.18 \pm 0.47ab$	$5.29 \pm 0.07a$	$3.19 \pm 0.16b$
α -glucosidases	$14.24\pm1.9ab$	$19.42\pm0.62a$	$7.16 \pm 0.71c$	10.58 ± 0.83 bc	$9.60 \pm 0.38c$	$7.41 \pm 0.50c$
β-glucosidases	$3.77\pm0.33b$	$12.21 \pm 0.34a$	$3.09\pm0.09b$	$3.15 \pm 0.14b$	$3.84 \pm 0.26b$	$3.58\pm0.05b$
α -galactosidase	$0.99 \pm 0.04b$	$1.82 \pm 0.17a$	0.75 ± 0.02 bc	0.44 ± 0.01 cd	0.67 ± 0.03 bc	$0.27\pm0.04d$
β-galactosidase	$1.65\pm0.11a$	$1.54\pm0.26a$	$0.85 \pm 0.08b$	$1.62 \pm 0.05a$	$1.01\pm0.07ab$	$1.11 \pm 0.14ab$

Different letters show significant differences among values using Tukey's test at $p \le 0.05$.

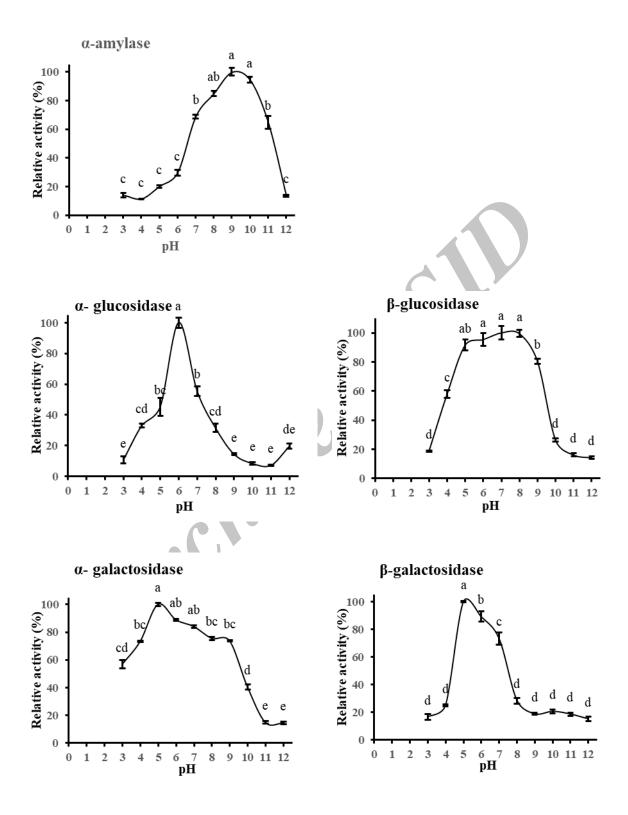


Figure 1 The effect of pHs on the activities of digestive carbohydrases extracted from the midgut of *Tuta absoluta* larvae.

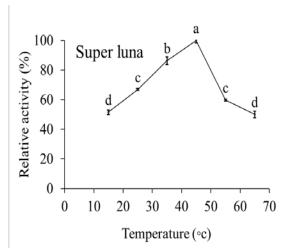


Figure 2 The effect of temperature on the activity of α -amylase extracted from the midgut of *Tuta absoluta* larvae.

Effect of tomato cultivars on activities of α and β -glucosidase and α - and β -galactosidase

Alpha and β -glucosidase activity of T. absoluta reared on six different tomato cultivars are shown in Table 1. The highest level of α -and β -glucosidase activity was present in Riogrande - fed larvae. The lowest α - and β -glucosidase activity was detected in midgut extracts larvae fed on Kingston (Table 1).

The specific activities of α -and β -galactosidase in midgut of fourth larval instar were determined. The obtained results showed that the highest α - and β - galactosidase activity was in Riogande and Calj -fed larvae, while, the lowest activity was in midgut extract from larvae fed with Super chief and Kingston, respectively (Table 1).

Effect of activators and inhibitors on α -amylase activity

CaCl₂, MgCl₂, NaCl and KCl enhanced α -amylase activity, whereas activity of α -amylase was inhibited by ethylene diamine tetra acetic acid (EDTA), ZnCl₂ and BaCl₂ (Table 3).

Zymogram analysis of α-amylase

The crude extracts of T. absoluta were analyzed by native PAGE. After α -amylase activity staining, two major isoforms of α -amylase were clearly detected (Fig. 3).

Table 3 shows effects of different compounds (20 mM) on digestive amylase activity in *T. absoluta* larvae fed on Super Luna cultivar.

Table 3 Effects of different compounds (20 mM) on digestive amylase activity in *Tuta absoluta* larvae fed on Super Luna cultivar.

Compounds	Relative activity (%)
Control (no addition)	100c
Na^+	$134.684 \pm 8.23b$
K^+	$127.342 \pm 0.81b$
${ m Mg^{2+}}$	$191.646 \pm 1.84a$
Ba^{2+}	$76.962 \pm 2.60d$
Zn^{2+}	$48.860 \pm 1.69e$
Ca^{2+}	$192.152 \pm 3.42a$
EDTA	65.569 ± 1.19 de

Different letters show significant differences among values using Tukey's test at $p \le 0.05$.

Table 2 Effect of tomato cultivars on kinetic parameters of α-amylases extracted from midgut of *Tuta absoluta* larvae.

K_m (mM)	V_{max} (mM/min)	K_m/V_{max}
0.282 ± 0.06 abc	1.786 ± 0.1 ab	0.158
$0.045 \pm 0.00c$	$0.971 \pm 0.01c$	0.046
$0.207 \pm 0.03bc$	$1.258 \pm 0.1bc$	0.165
$0.179 \pm 0.08bc$	$2.287 \pm 0.4a$	0.078
$0.385 \pm 0.06ab$	$1.240 \pm 0.1bc$	0.310
$0.565 \pm 0.11a$	$2.156 \pm 0.2ab$	0.262
	0.282 ± 0.06 abc 0.045 ± 0.00 c 0.207 ± 0.03 bc 0.179 ± 0.08 bc 0.385 ± 0.06 ab	$0.282 \pm 0.06abc$ $1.786 \pm 0.1ab$ $0.045 \pm 0.00c$ $0.971 \pm 0.01c$ $0.207 \pm 0.03bc$ $1.258 \pm 0.1bc$ $0.179 \pm 0.08bc$ $2.287 \pm 0.4a$ $0.385 \pm 0.06ab$ $1.240 \pm 0.1bc$

Different letters show significant differences among values using Tukey's test at $p \le 0.05$.

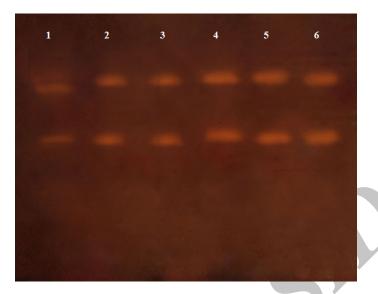


Figure 3 Zymogram of α -amylase extracted from the midgut of *Tuta absoluta* larvae reared on different cultivars (1 Calj, 2 Riogrande, 3 Kingston, 4 Super Luna, 5 Super Chief and 6 Super strain B.).

Discussion

The activities of digestive enzymes are directly influenced by some important factors such as temperature, density of consumed diet, pH (Sivakumar et al., 2006) and secondary metabolites in the diet. These enzymes are necessary for providing nutrition and energy to the growing larva of insects. The optimal pH for a-amylase was obtained at alkaline pH in the present research. Regarding the lepidopteran insects, the alkaline pH range has been determined to be the suitable for the activity of α-amylase. In order to feed on tannin-rich plant materials, lepidopteran insects have adapted to alkaline pH in their midguts (Chapman 1998). In this condition, the digestion efficiency was increased as a result of less binding of tannin with proteins (Dow, 1986). The optimum pH was demonstrated in most of the studies, on lepidopteran α -amylase activity, to be at 8-12, reported e.g. pH 9.0 for α-amylase activity of Pieris brassicae (Linn), Chilo suppressalis (Walker), Tecia solanivora (Povolny) and H. armigera (Zibaee et al. 2008; Zibaee, 2012; Valencia Jime'nez et al. 2008; Ozgur et al. 2009). Esmaeily and Bandani (2015) reported the optimum pH of 8.0 for α-amylase activity of

T. absoluta. Therefore, the consumed chemical compounds and the kind of insect species are two main factors on the optimum pH for the α -amylase activity (Hori, 1971; Zeng and Cohen, 2000; Swart *et al.*, 2006).

While, the pH 6.0 and 5.0 were observed to be the optimum activity for α -glucosidase and α -galactosidase in midgut of T. absoluta, the β -glucosidase and β -galactosidase activity were shown to have the maximum activity at pH 7.0 and 5.0, respectively. According to Kaur et al. (2014), the phylogenetic relation or response of the insect to different diets may suggest the dissimilarities in the insect's maximum pH of α - and β -glucosidases and α - and β -galactosidases.

We also evaluated the α -amylase activity of T. absoluta fourth instar larvae fed on different cultivars. Our results showed that the food quality or the chemical and inhibitor compounds existing in food maybe essential factors in the activity of carbohydrases in midgut of insects as previously proved by Slansky (1982) and Mendiola-Olaya $et\ al.$ (2000). The results revealed that the highest and lowest amylolytic activity were detected in larvae reared on Super strain B and Super chief, respectively, compared to other cultivars. This

result suggests that, the responsible factor for any significant differences in the larvae's amylase activities can be due to dissimilarities in carbohydrate contents in different host plant cultivars. In a study, the influence of various tomato cultivars on the digestive enzymes activities of H. armigera larvae was investigated (Nemati Kalkhoran et al. 2013). The results of their study demonstrated that the highest amylolytic activity $(0.062 \pm 0.00004 \text{ mU mg}^{-1})$ was shown by the larvae reared on the leaves of SUN 6108 f1, while, the larvae fed on the leaves of Cal JN3 as well as Korral had the lowest activities of $0.027 \pm 0.00004 \text{ mU mg}^{-1}$ and $0.027 \pm$ 0.0001 mU mg⁻¹, respectively. Insect species and the physiological diversities in the tomato cultivars are possibly among the reasons for such disagreements.

Measuring activity of α -/ β -glucosidases and α -/ β -galactosidases in last larval instar of T. absoluta fed on various tomato cultivars showed significant differences in their specific activities. Availability of compounds containing β-glycosidic bonds in insect diet can influence the glucosidase and galactosidase activities (Gholamzadeh Chitgar et al. 2014). The previous studies on phytophagous lepidopterans showed the presence of α - and β -glucosidase in the salivary glands and in the midgut (Ghadamyari et al., 2010; Franzl et al., 1989; Marana et al., 2000). Comparing the tested carbohydrases, the highest activity was shown by α-glucosidase. The obtained findings agree with the results of Riseh et al. (2012). They reported that the activity of α -glucosidase was higher than that of other galactosidases and glucosidases in the Rhynchophorus ferrugineus Olivieri larvae's digestive system and female adult. Also, similar results were obtained for the Choreutis nemorana (Hübner) last larvae instar (Gholamzadeh Chitgar et al. 2014). Contrary to these findings, high activity of β-glucosidase was found by Ferreira et al (1998) in a number of insects belonging to diverse orders such as Tenebrio molitor (L.) (Col.: Tenebrionidae), Abracris flavolineata (De Geer) (Orth.: Acrididae) and Scaptotrigona bipunctata

(Lepeletier). (Hym.: Apidae). Additionally, according to Aghaali *et al.* (2012), comparing to other glucosidases and galactosidases, β -galactosidase activity was higher in the digestive system of *Osphranteria coerulescens* Redt. larvae than that of other carbohydrases.

The reaction rate of enzymatic-catalysis will increase with temperature due to the increase in kinetic energy as well as the interaction frequency of molecules. Kaur *et al.* (2014) found that α -amylase is active over a wide range of temperatures from 30-60 °C. Accordingly, the optimum temperature activity for α -amylase of *T. absoluta* was obtained at 45 °C.

The substrate's affinity of enzyme is determined by K_m . High amount of K_m value would results in less affinity between substrate and enzyme. In addition, as demonstrated by Kaur et al. (2014), when an enzyme is completely saturated with substrate, the number of converted substrate molecules into product by an enzyme in a unit time is revealed by V_{max} . In order to find the kinetic parameters of α amylase extracted from T. absoluta larvae in the presence of starch substrate, Linweaver-Burk analysis was applied. Regarding αamylase in Super Chief and Super Luna cultivars, the findings showed 0.565 ± 0.11 mM and 2.287 \pm 0.4 mM/min as the highest K_m and V_{max} values, respectively.

The K_m values for the α -amylase of *Bombyx mori* (L.), *P. brassicae* and *Mamestra brassicae* (L.), were obtained as 0.57, 1.37 and 0.33 mg/ml, respectively (Tanabe and Kusano, 1984; Sharifloo *et al.*, 2016; Matsumura, 1934). According to the report by Sharifi *et al.* (2011), using starch as substrate, the K_m and V_{max} values of 1.34 mg/ml and 1.52 μ mol/min/mg protein were obtained for α -amylase of *Xanthogaleruca luteola* (Müller), respectively. In addition, in another study by Ramzi and Hosseininaveh (2010), the K_m and V_{max} values of 0.77 mM and 0.064 μ mol min⁻¹were observed for the α -amylase in *Brachynema germari* Kolenati midgut, respectively.

Insect α -amylases are metallo-enzymes which need metal ions for maintaining integrity, structural stability and activity (Terra and

Ferreira, 2012; Kaur et al. 2014). Mg²⁺, Ba²⁺, Fe²⁺, Mo⁺, K⁺, Mg⁺, Cu⁺, Hg⁺, Zn²⁺, nitrite, urea, phytic acid, tannic acid, EDTA, SDS, tris, triton X-100, triethylene tetramine hexa acetic acid (TTHA), ethylene glycol tetra acetic acid (EGTA) inhibited α-amylase activity of various insects (Kaur et al. 2014). Two factors were shown to be the possible cause of the inhibition of amylase by some metal ions. First, this could be due to their binding to either catalytic residue. The other possibility is suggested to be the substitution of Ca²⁺ from the substrate binding site of α-amylase. In the present research ZnCl2, EDTA and BaCl2 decreased amylase activity in T. absoluta larvae significantly. The presence of CaCl₂, MgCl₂, NaCl and KCl were shown to increase the activities of the enzyme. These findings are in accordance with the results of previous studies (Zibaee et al., 2008; Asadi et al., 2010; Yezdani et al., 2010).

The Zymogram analysis showed two bands of α -amylase activity in midgut of T. absoluta. Considering other insects such as Spodoptera littoralis (Fabricius) and C nemorana, the presence of a mixture of diverse α -amylase isoenzymes has been stated (Bigham et al., 2013; Darvishzadeh et al., 2014). Thus, it can be argued that the presence of this diversity in α -amylase iso-enzymes could be associated with the significance of this enzyme in the food digestion of the insect.

Conclusion

According to the results of the present research, the larvae fed on Kingston cultivar showed lowest carbohydrase activity. This could be because of changes in secondary chemicals or digestive system inhibitors existing in this cultivar, which rendered it as unsuitable host for *T.absoluta* larvae. It could beassumed that Kingston cultivar contains a range of inhibitors that will make it a good choice to develop from it *T. absoluta*-resistant transgenic plants. Of course, other biological data such as growth, development, survival and fecundity data are necessary to confirm resistance of this cultivar.

Due to the diversity in the activity of carbohydrate hydrolyzing enzymes in T. absoluta larvae, depending on their diet, and complex interactions between plant and insect digestive system, the inhibition of the insect digestive system should be considered in designing strategies for sustainable crop protection.

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تعیین خصوصیات بیوشیمیایی کربوهیدرازهای گوارشی در لارو شبپره مینوز گوجهفرنگی Tuta absoluta (Lep.: Gelechidae)

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چكيده: شبپره مينوز گوجهفرنگى، Tuta absoluta (Meyrick) تهديدى جدى براى توليد محصول گوجهفرنگی در مزارع و گلخانههای ایران محسوب میشود. یکی از روشهای غیرشیمیایی کنترل آفات استفاده از گیاهان تراریخته حاوی مهارکنندههای کربوهیدرازها میباشد، بنابراین مطالعه خصوصیات آنزیمی کربوهیدرازها برای دستیابی به این هدف به ما کمک خواهد کرد. در پژوهش حاضر ویژگیهای بیوشیمیایی کربوهیدرازهای گوارشی استخراج شده از روده میانی لارو سن آخر شبپره مینوز پرورش يافته روى ارقام مختلف گوجهفرنگي Kingston, Riogrande, Super Luna, Super Chief, Super strain (B, Calj تعیین شد. بیش ترین فعالیت ویژه آلفا-آمیلاز در لاروهای تغذیه کرده از رقم Super strain B و کمترین آن در رقم Super Chief مشاهده شد. اسیدیته و دمای بهینه برای فعالیت آلفا-آمیلاز بهتر تیب و ۴۵ درجه سلسیوس تعیین گردید. بیش ترین مقادیر K_m و K_m در لاروهای تغذیه کرده از ارقام ۹ کیلے مولار بر دقیقه) به دست ۲/۲۸۷ ± ۰/۴) Super Luna میلی مولار بر دقیقه) به دست آمد. اثر تركيبات مختلف روى فعاليت آميلوليتيك نشان داد كه تركيبات CaCl2, MgCl2, NaCl و CaCl به طور معنى دارى فعاليت آلفا-آميلاز را افزايش دادند، در حالي كه ZnCl2 ،EDTA وBaCl2 موجب كاهش فعالیت این آنزیم در رقم سوپر لونا شدند. بیش ترین فعالیت آلفا و بتا-گلوکوزیداز به تر تیب در اسیدیته ۶ و ۷ بهدست آمد، در حالی که مقدار اسیدیته بهینه برای فعالیت آلفا و بتا-گالاکتوزیداز برابر ۵ بود. بیش ترین فعالیت ویژه آلفا و بتا-گلوکوزیداز در لاروهای تغذیه کرده از رقم Riogrande مشاهده شد، درحالی که بیش ترین فعالیت ویژه آلفا و بتا-گالاکتوزیداز بهترتیب در لاروهای تغذیه کرده از ارقام Riogrande و Calj به دست آمد. تعداد باندهای مشاهده شده روی ژل برای آلفا-آمیلاز برابر ۲ بود. از آنجایی که لاروهای تغذیه کرده از رقم Kingston کمترین فعالیت کربوهیدرازی را نشان دادند احتمالاً این رقم میزبان نامناسبی برای شبیره مینوز گوجهفرنگی می باشد.

واژگان كليدى: Tuta absoluta ، آلفا-آميلاز، آلفا و بتا-گلوكوزيداز، آلفا و بتا-گالاكتوزيداز