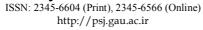


# Poultry Science Journal





# Effect of Zinc Sulfate and Organic Chromium Supplementation on the Performance, Meat Quality and Immune Response of Japanese Quails under Heat Stress Conditions

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#### Abstract

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#### Keywords:

Zinc Chromium Japanese quail Meat quality Immune response This study was conducted to investigate the effects of different levels of zinc sulfate and chromium picolinate on the performance, immune response and meat quality of Japanese quails under heat stress condition. The birds (n = 540; 7-d-old) were randomly assigned to 9 treatment groups consisting of 3 replicates of 20 birds each in a 3 × 3 factorial arrangement of treatments [zinc (0, 40, 80 mg/Kg); chromium (0, 500, 1000  $\mu$ g/Kg)]. Birds were kept in floor cages in a temperature controlled room and subjected to heat stress for 8 h/d (9.00 to 17.00) from 7 d to the end of the study. Zn was supplemented to the basal diet as ZnSO4 and Cr as chromium picolinate. A linear decrease in feed intake and an improvement in feed conversion ratio were found by supplementing Zn and Cr to the quails diets. Chromium supplementation increased daily weight gain linearly. By increasing the level of dietary Zn and Cr in the diet, heterophil to lymphocyte ratio and meat quality parameters were improved linearly. The results of this study revealed positive effects of Zn and Cr supplementation on the performance, immune responses and meat quality of quails under heat stress condition. It seems that supplementation of the quail's diets with 80 mg/Kg Zn and 1000 μg/Kg Cr can be greatly helpful for improving the adverse effects of heat stress.

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#### Introduction

When birds are exposed to a high ambient temperature, a series of consecutive events appears in their body that finally damages their products (Niu et al., 2009). Ward and Peterson (1973) perceived that plasma level of zinc was lower in broilers exposed to high ambient temperatures (33-35°C) rather than those reared under the thermoneutral condition (18-22°C). Since the production of free radicals escalates under heat stress conditions (Mujahid et al., 2005), it seems to be beneficial to use elements in the poultry diets that someway take part in clearance of the oxidant agents. One of the major functions of zinc is participation in the antioxidant defense system of the body (Prasad and Kucuk, 2002). The mechanism whereby zinc plays the role is not well recognized yet, but it is thought that Zn increases synthesis of metallothionein protein as the scavenger of free radicals (Oteiza et al., 1996). In pancreas, Zn plays a protective role against oxidative damages and aids this organ to secrete digestive enzymes, which leads to the increased nutrients digestibility (Sahin et al., 2009). Under the heat stress conditions, excretion of zinc from the body is intense (Belay and Teeter, 1996). Zinc influences utilization of feed via participating in the metabolism of carbohydrates, fats and proteins advantageously (McDonald, 2011). Zinc is an important element in all aspects of immunity (Chandra and Dayton, 1982; Sherman, 1992) and is essential for integrated cellular immunity response (Dardenne et al., 1985). Zinc deficiency decreases the cellular immunity (Prasad and Kucuk, 2002) and adversely affects functions of the thymus (Fraker et al., 1977) and spleen (Luecke et al., 1978) and interleukin-2 production (Dowd et al., 1986).

Conflicting reports have been published regarding the effect of zinc on growth performance of poultry under the heat stress conditions. Bartlett and Smith (2003) report that different dietary levels of zinc have no effects on the performance and plasma concentration of zinc in broilers under the heat stress conditions while Sahin *et al.* (2005) stated that the performance and carcass quality of quails fed a zinc supplemented diet were improved under the heat stress condition.

Insulin, a hormone that plays an important role in metabolism of nutrients requires chromium (Cr) to do its function properly. Chromium participates in the structure of an organometallic molecule called the glucose tolerance factor (GTF) that stimulates insulin receptors to pair with this hormone (Mertz, 1993). Many other functions have been reported for chromium such as participation in the metabolism of lipids, proteins and nucleic acids. It has been established that chromium shows anti-oxidative properties from itself in the body (Tezuka *et al.*, 1991). Furthermore, chromium improves health and subsequent performances via aiding the immune system (Mallard and Borgs, 1997). During the occurrence of environmental stresses, elevation of plasma level of corticosterone intensifies free radicals production that leads to suppression of the immune system (McIntosh and Sapolsky. 1996). It has been shown that supplementation of laying hens diet with Cr decreases plasma level of corticosterone under the cold stress conditions (Sahin

et al., 2001; Hanafy, 2011). Chromium is not a particularly toxic element in its trivalent form, and a wide margin of safety exists between the normal amounts ingested and those likely to produce deleterious effects (McDonald et al., 2011). Stress depletes the body stores of chromium and increases the need for this element (Hayirli, 2005). Heterophile to lymphocyte ratio as a sensitive and important index for a variety of stresses (Gross and Sigel, 1983; Mashaly et al., 2004) increases during heat stress (McFarlane and Curtis, 1989; Mashaly et al., 2004). Adding chromium as a supplement to the diet of broilers that were reared under the heat stress condition decreased this ratio (Ebrahimzadeh et al., 2012). Finally, some reports exist that confirm the positive effects of chromium on digestibility of nutrients in animals (Kim et al., 1997; Sahin et al., 2001; Onderci et al., 2003; Emami et al., 2012).

This experiment was conducted to investigate the effects of different levels of zinc sulfate and chromium picolinate on performance, immune response and meat quality of Japanese quails under the heat stress conditions.

### Materials and Methods

#### Birds, treatments and management

Five hundred of forty 7-day-old Japanese quails (mixed sex) were housed in floor cages in a temperature controlled room and subjected to heat stress (to reach panting) for 8 h/d (0900 to 1700 h) from 7 d to the end of the study (35 d). Birds fed a corn-soybean meal basal diet (control) or the basal diet supplemented with either Zn as ZnSO4 or Cr as chromium picolinate (CrPic).

The birds were randomly assigned to 9 treatment groups consisting of 3 replicates of 20 birds each in a 3 × 3 factorial arrangement of treatments [zinc (0, 40, 80 mg/Kg); chromium (0, 500, 1000  $\mu$ g/Kg)]. The basal diet was formulated to meet the NRC requirements (NRC, 1994) of meat Japanese quails as shown in Table 1. Feed (in mash form) and fresh water were offered *ad libitum* throughout the experiment.

#### Sample and data collection

Growth performances (feed intake, body weight gain and feed conversion ratio) were recorded weekly. On days 35 of the experimental period, blood samples (via cervical cutting) of one male bird per cage (three birds per treatment) was collected and transferred to tubes containing EDTA as an anticoagulant. Blood films were made, air dried and then stained with Giemsa - Wright's stain. Differential white blood cell (WBC) counts were performed by using standard avian guidelines of Ritchie *et al.* (1994).

After slaughtering (on 35 d), breast and thigh muscles were obtained and kept frozen at -21°C for 40 days. Breast muscle lipid peroxidation was assessed as thiobarbituric acid-reactive substance concentrations in samples by the method of Tarladgis *et al.* (1960). Values were reported as the concentration of malondialdehyde

(MDA). Meat pH was measured by the method of Parizadian *et al.* (2011). The samples of breast muscles were used for MDA and pH analyses. Water holding capacity (WHC), driploss (DL) and cooking loss (CL) tests were done for thigh samples, using Grau and Hamm (1953) procedure.

Table 1. Ingredients and composition of the basal diet

Ingredients	Amount (g/Kg)
Corn	457.0
Soyabean meal (44% CP)	472.6
Soybean oil	40.0
Oyster shell	13.0
Dicalcium phosphate	7.5
Salt	3.6
DL-Methionine	1.3
Vitamin premix <sup>1</sup>	2.5
Mineral premix <sup>2</sup>	2.5
Chausical commonition	X
Chemical composition	2027
ME (Kcal/Kg)	2937
Crude protein (g/Kg)	242.8
Lysine (g/Kg)	13.9
Methionine (g/Kg)	5.03
Methionine + Cyctine (g/Kg)	8.97
Calcium (g/Kg)	8.07
Available Phosphorus (g/Kg)	3.06
Sodium (g/Kg)	1.54

 $^1$ provided the following per Kg of diet: retinol acetate 3.1 mg, thiamine 1.8 mg, riboflavin 6.6 mg, niacin 30 mg, pantothenic acid 10 mg, pyridoxine 3 mg, folic acid 1 mg, cyanocobalamine 15 μg, biotin 0.1 mg, cholecalciferol 0.05 mg, alpha-tocopherol acetate 18 mg, menadion 2 mg, choline chloride 0.4 g.  $^2$ provided the following per Kg of diet; Fe 50 mg, Mn 100 mg, , Zn 85mg, Cu 10 mg, Se 0.2 mg and I 1 mg.

#### Statistical analyses

The data were analyzed using the GLM procedure of SAS software (SAS Institute, 2008). Duncan multiple range test was used to detect (P<0.05) differences among treatment means. Linear and quadratic polynomial contrasts were used to evaluate the effect of different levels of zinc and chromium.

#### Results

#### Performance parameters

The effects of supplemental Zn and Cr on growth performance of Japanese quails are shown in Table 2. Our results showed that Zn supplementation decreased feed intake (FI) quadratically and improved feed conversion ratio (FCR)

linearly but had no significant effect on body weight gain (BWG). Cr supplementation decreased FI and FCR while increased BWG linearly.

A significant interaction between Zn and Cr was observed for FI and FCR. Chicks fed diet contained 1000  $\mu$ g Cr and 40 mg Zn per Kg, showed the lowest FI and FCR. In general, dietary combined supplementation of Zn and Cr decreased FI and FCR to a greater degree compared to supplementation with Zn and Cr separately.

## White blood cells and immunity organs

The effect of Zn supplementation on heterophile, lymphocyte and heterophile/lymphocyte ratio was significant (*P*<0.01, Table 3). As the dietary level of Zn increased, the percentages of heterophile and heterophile/lymphocyte ratio were decreased and the percentage of lymphocyte increased, linearly. The percentages of monocyte, eosinophil and basophil were not affected by different levels of Zn. The relative weight of bursa, spleen and thymus did not show any significant responses to Zn supplementation.

In the present study, the percentages of heterophile, lymphocyte and heterophile/ lymphocyte ratio were significantly affected by Cr (P < 0.01). The relative weight of the thymus was also positively affected by Cr supplementation (P < 0.05). The percentage of heterophile and heterophile/lymphocyte ratio decreased linearly but the percentage of lymphocyte increased linearly as the dietary level of Cr increased. The percentages of monocyte, eosinophil and basophil and relative weight of the spleen were not affected by Cr. Supplementation.

The interaction between Zn and Cr was significant for heterophile, eosinophil and basophil (P<0.01). The lowest percentage of heterophile was pertained to the birds that were supplemented with a combination of 80 mg/Kg Zn and 1000  $\mu$ g/Kg Cr, whereas the highest rate was related to the control treatment. The highest percentage of eosinophil was observed in the birds that were supplemented with a combination of 40 mg/Kg Zn and 500  $\mu$ g/Kg Cr, whereas the birds that were supplemented only by 500  $\mu$ g/Kg Cr had the lowest percentage of eosinophil. Supplementation of quails with a combination of 80 mg/Kg Zn and 500  $\mu$ g/Kg Cr or a combination of 40 mg/Kg Zn and 1000  $\mu$ g/Kg Cr caused the highest percentage of basophil.

#### Meat quality

Some meat related quality parameters are shown in Table 4. The effect of Zn supplementation on drip loss (DL), cooking loss (CL), water holding capacity (WHC), pH and malondialdehyde value (MDAV) of meat were significant (*P*<0.01). As dietary levels of Zn increased, DL, CL and MDAV levels of the meat decreased and WHC and pH increased linearly.

The effect of Cr was significant on all of the meat quality parameters (DL, CL, WHC, pH and MDAV) (P<0.01). As the dietary level of Cr increased, DL, CL and MDAV level of the meat decreased and WHC and pH increased linearly. A significant interaction was observed for MDAV (P<0.01). The highest level of MDAV was related to the birds fed with control diet and the lowest level was related to birds that were supplemented with a combination of 80 mg/Kg Zn and 1000  $\mu$ g/Kg Cr.

Table 2. Effects of zinc sulfate and chromium picolinate on the performance of Japanese quails reared under the heat stress condition

Treatment	S					
Chromium Zinc		$FI^{1}\left( g/b/d\right)$	$BWG^2(g/b/d)$	FCR <sup>3</sup>		
(μg/Kg)	(mg/Kg)					
0	0	21.83a	6.80	3.21a		
	40	18.75c	6.62	2.84b		
	80	19.67 <sup>b</sup>	6.93	$2.84^{b}$		
500	0	19.35 <sup>b</sup>	6.89	2.81bc		
	40	18.39 <sup>cd</sup>	6.68	2.75bcd		
	80	18.32 <sup>d</sup>	6.78	2.70 <sup>cde</sup>		
1000	0	18.30d	6.94	2.64de		
	40	18.14 <sup>d</sup>	6.94	$2.62^{\rm e}$		
	80	18.54 <sup>cd</sup>	7.04	2.63de		
SEM		0.219	0.0363	0.0355		
Main effect means						
Zinc:						
0		19.83a	6.88	$2.88^{a}$		
40		18.43c	6.75	$2.74^{b}$		
80	_ ^	18.84b	6.92	2.72 <sup>b</sup>		
Chromium:	401					
0		20.08a	6.79 <sup>b</sup>	2.96a		
500		18.69b	$6.78^{b}$	2.75 <sup>b</sup>		
1000		18.33c	$6.98^{a}$	2.63c		
API	Probabilities					
Source of variation						
Zinc effect		< 0.01	0.0963	< 0.01		
Chromium effect		< 0.01	0.0338	< 0.01		
Zinc × Chromium effo	ect	< 0.01	0.5213	< 0.01		
Polynomial contrast						
Linear zinc		< 0.01	0.5986	< 0.01		
Quadratic zinc		< 0.01	0.0372	0.0216		
Linear chromium		< 0.01	0.0234	< 0.01		
Quadratic chromium		< 0.01	0.1652	0.1558		

<sup>&</sup>lt;sup>1</sup>Feed intake, <sup>2</sup>Body weight gain, <sup>3</sup>Feed conversion ratio.

a-c Means with different superscripts in the same column differ significantly (*P*<0.05).

Table 3. Immune parameters of Japanese quails supplemented with different levels of zinc sulfate and chromium picolinate reared under the heat stress condition

Treatme	ents	Hetero	I romanla		Mono	Eosino	Basop			
Chromium	Zinc	phile			cyte	phil	hil	Bursa <sup>2</sup>	Spleen <sup>2</sup>	Thymus <sup>2</sup>
(µg/Kg)	(mg/Kg)	(%)	yte (%)		(%)	(%)	(%)			-
0	0	43.00a	48.00	0.900	3.67	3.67ab	1.67ab	0.0733	0.0563	0.112
40 80	40	42.67a	50.67	0.843	3.33	2.33bc	1.00abc	0.0830	0.1063	0.155
	80	$42.00^{a}$	51.33	0.817	4.00	2.33bc	$0.33^{c}$	0.0887	0.0667	0.178
500	0	$41.67^{a}$	52.67	0.790	2.33	$2.00^{\circ}$	1.33abc	0.0807	0.0663	0.154
	40	39.67b	51.00	0.777	3.67	5.00a	0.67bc	0.1097	0.0877	0.190
	80	38.67bc	54.00	0.720	3.00	2.33bc	2.00a	0.0957	0.0873	0.208
1000	0	37.67c	55.67	0.677	3.67	2.33bc	0.67bc	0.0823	0.0883	0.202
	40	$34.67^{d}$	57.33	0.607	3.67	2.33bc	2.00a	0.1117	0.0933	0.235
	80	$32.00^{e}$	59.67	0.537	3.67	3.00 bc	1.67ab	0.1313	0.0943	0.244
SEM		0.719	0.706	0.0220	0.163	0.220	0.147	0.00528	0.00576	0.0110
Main effect me	eans									
Zinc:										
0		$40.78^{a}$	52.11b	$0.789^{a}$	3.22	2.67	1.22	0.0788	0.0703	0.156
40		39.00b	53.00b	$0.742^{b}$	3.56	3.22	1.22	0.1014	0.0958	0.193
80		37.56c	55.00a	0.691c	3.56	2.56	1.33	0.1052	0.0828	0.210
Chromium:										
0		42.56a	50.00c	0.853a	3.67	2.78	1.00	0.0817	0.0764	0.148b
500		40.00b	52.56b	$0.762^{b}$	3.00	3.11	1.33	0.0953	0.0804	0.184ab
1000		$34.78^{c}$	57.56a	$0.607^{c}$	3.67	2.56	1.44	0.1084	0.0920	$0.227^{a}$
				I	robabiliti	es				
Source of varia	ation:			_ \	1					
Zinc effect		< 0.01	0.0004	< 0.01	0.6302	0.2068	0.8954	0.0759	0.2401	0.0848
Chromium effo	ect	< 0.01	< 0.01	< 0.01	0.1792	0.3686	0.2619	0.1005	0.5474	0.0109
Zinc × Chromi	um effect	0.0086	0.0611	0.0945	0.4596	0.0015	0.0027	0.6380	0.6741	0.9956
Polynomial co										
Linear zinc		< 0.01	0.0001	< 0.01	0.4103	0.7761	0.6879	0.0365	0.4011	0.0320
Quadratic zinc		0.6701	0.2896	0.8250	0.6324	0.0833	0.8163	0.3638	0.1425	0.6085
Linear chromi		< 0.01	< 0.01	< 0.01	1.0000	0.5709	0.1198	0.0345	0.2966	0.0031
Quadratic chro	omium	0.0028	0.0274	0.0044	0.0674	0.1991	0.6430	0.9784	0.7665	0.8611

a-c Means with different superscripts in the same column differ significantly (*P*<0.05).

<sup>&</sup>lt;sup>1</sup> Heterophile/Lymphocyte ration; <sup>2</sup>Values are expressed as a percentage of live body weight.

Table 4. Meat quality parameters of Japanese quails supplemented with different levels of zinc sulfate and chromium picolinate reared under the heat stress condition

Treatments		$DL^1$	CL <sup>2</sup>	WHC <sup>3</sup>		MDAV4	
Chromium	Zinc	(%)		(%)	pН		
_(μg/Kg)	(mg/Kg)	( /0)	(%)	(70)	_	(mg/Kg)	
0	0	5.07	25.85	60.03	4.94	0.3447a	
	40	3.87	24.56	62.62	5.04	$0.2880^{b}$	
	80	3.19	23.58	62.58	5.29	$0.2007^{c}$	
500	0	3.22	23.63	62.60	5.29	0.2027c	
	40	2.98	22.91	64.29	5.35	$0.1460^{d}$	
	80	2.68	22.32	65.40	5.41	$0.1177^{de}$	
1000	0	2.68	22.40	66.66	5.54	0.1190 <sup>de</sup>	
	40	2.22	21.88	67.28	5.70	0.1110 <sup>de</sup>	
	80	1.88	21.45	68.83	5.80	$0.0907^{e}$	
SEM		0.187	0.280	0.534	0.0543	0.0165	
Main effect means							
Zinc:							
0		$3.66^{a}$	23.96a	63.10 <sup>c</sup>	5.25c	0.2221a	
40		$3.02^{b}$	23.11 <sup>b</sup>	64.73b	5.36b	$0.1817^{b}$	
80		$2.58^{c}$	22.45b	65.60a	5.50a	$0.1363^{\circ}$	
Chromium:							
0		$4.04^{a}$	24.66a	61.74c	$5.09^{c}$	$0.2778^{a}$	
500		$2.96^{b}$	22.95 <sup>b</sup>	64.10b	5.35 <sup>b</sup>	$0.1554^{b}$	
1000		$2.26^{c}$	21.91c	67.59a	5.68a	$0.1069^{c}$	
		Probabilities					
Source of variation							
Zinc effect		0.0002	0.0008	< 0.01	0.0002	< 0.01	
Chromium effect		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	
Zinc × Chromium effect		0.1056	0.5766	0.3004	0.2873	0.0019	
Polynomial contrast	44						
Linear zinc		< 0.01	0.0002	< 0.01	< 0.01	< 0.01	
Quadratic zinc		0.5789	0.7550	0.2678	0.7340	0.7724	
Linear chromium		< 0.01	< 0.01	< 0.01	< 0.01	< 0.01	
Quadratic chromiun	n	0.2838	0.2494	0.1037	0.3886	0.0003	

<sup>&</sup>lt;sup>1</sup>Drip loss; <sup>2</sup>Cooking loss; <sup>3</sup>Water holding capacity; <sup>4</sup>Malondialdehyde.

# Discussion

### Performance parameters

Dietary Zn and Cr supplementation under the heat stress condition improved growth performance of Japanese quails. The proven role of Zn and Cr in metabolism of carbohydrates, lipids, proteins and nucleic acids is in accrediting of this argument. Although the exact biochemical mechanism of how high dietary Zn decreases feed intake and subsequent growth is not clear yet, however the concentration of the element in brain does not appear to be a factor (Sandoval *et al.*, 1998). Reduced feed intake could be associated with losing of appetite (Brink *et al.*, 1950) or reduced palatability of the diets containing high levels of Zn (Fox, 1989) as

a-c Means with different superscripts in the same column differ significantly (P < 0.05).

well. Although there are no reports about the negative effect of high levels of chromium picolinate on the palatability of diet, but it can be considered in the investigation of the reasons for reduced FI. Zn plays a key role in cell division and proliferation (Rubin, 1972; Rubin and Koide, 1973) and activity of more than 300 enzymes in the body (Prasad and Kucuk, 2002). In the heat stress condition, the need for Zn intensifies, so, feed efficiency and growth performance will decline if this requirement is not to be supplied (Ensminger *et al.*, 1989). Furthermore, the proven role of Zn in increasing nutrients digestibility under the heat and cold stress (Onderci *et al.*, 2003; Sahin and Kucuk, 2003b) can play an important role in utilization of the consumed feed.

Improved performance parameters such as FCR in Cr supplemented quails can be related to Cr anti-stress properties and its booster effects on insulin, leading to more efficient utilization of nutrients and improved FCR. In addition, Cr increased nutrients digestibility by playing an antioxidant role in pancreas that results in decreased FCR. Cr also has an internal interplay with DNA templates which leads to stimulation of the RNA synthesis (Okada *et al.*, 1983).

The weight gain of the birds is mainly related to muscle tissue and most of this tissue is protein. Due to the proven role of RNAs in protein synthesis, the role of Cr in nucleic acids metabolism also can be another reason for the improved feed conversion ratio in Cr supplemented birds (Okada et al., 1983). According to our findings, it seems that Cr has been more effective than Zn in improving FCR. Bartlett and Smith (2003) investigated the effects of diets with different levels (34, 68 and 181 mg/Kg) of Zn on the performance of broilers either in a thermoneutral or heat stress condition and observed no improvement in this parameters. Sahin et al. (2005) used different levels (30 and 60 mg/Kg) of zinc sulfate and zinc picolinate in the diet of quails reared under the heat stress condition. They found an improvement in FI, BWG and FCR in Zn supplemented quails. Uyanik et al. (2005) reported that supplementation of the diet with 20, 40, 80 or 100 mg/Kg CrCl<sub>3</sub> had no effect on performance of Japanese quails but in a newer study that was conducted by Sahin et al. (2010), supplementation of diet with 400 and 800 µg/Kg CrCl<sub>3</sub> and CrPic improved the performance of Japanese quails under the heat stress condition.

## White blood cells and immunity organs

Namra *et al.* (2008) reported that dietary supplementation of quails with 50 mg/Kg Zn, in organic and inorganic forms, had no effect on the relative weights of spleen and bursa. Bartlett and Smith (2003) reported that supplementation of broiler chicks with Zn did not affect the relative weights of bursa, spleen and thymus but improved the primary and secondary antibody titers. Furthermore, the percentage of macrophages was increased by Zn supplementation. Another study that was conducted by Bun *et al.* (2011) showed that dietary supplementation of Zn to broilers infected with *Eimeria tenella* led to an improved immune response. Zn

supplementation of broilers increased lymphocyte and proliferation in visceral blood (Yang et al., 2011).

Heteroplile/lymphocyte ratio is considered as a sensitive and important index for many of the stresses such as heat stress in chicks (Gross and Sigel., 1983; Mashaly et al., 2004). Under the heat stress condition H/L ratio increases that demonstrates the relationship between heat stress and non-specific immune cells (McFarlane and Curtis, 1989; Mashaly et al., 2004). Zn contributes to non-covalent reactions of cytoplasmic components by tyrosine kinase that is an essential protein in early stages of lymphocytes activity (Walsh et al., 1994). Moreover, Zn is an essential co-factor for thymulin that is a thymic hormone binds to surface receptors of lymphocyte T and leads to maturation and activation of these cells. Zn binds to thymulin by asparagine side chains and hydroxyl groups. It results in a structural change in thymulin and activates it (Dardenne et al., 1982). Therefore, supplementation of the diet with Zn may increase thymuline activity that leads to a more appropriate maturation and activity of T lymphocytes. In humans and animals, in both laboratory and field conditions, thymuline activity depends on plasma level of Zn (Dardenne et al., 1982; Prasad et al., 1988). Thymuline is present in the serum of zinc-deficient animals but is not active (Prasad et al., 1988). Thus, due to the increased T and B lymphocytes, the immune response of bird improves (Hudson et al., 2005). Zn also participates in lymphocyte cycle. It has been shown that Zn is required for G<sub>1</sub> phase in thymidine kinase expression and for transitional stage into S-phase (Chesters et al., 1993).

Ebrahimzadeh *et al.* (2012) used 200, 400 and 800  $\mu$ g/Kg Cr in the diet of broiler chicks that were reared under the heat stress conditions (33±2°C) and studied the immune response of these birds. The antibody titers against Newcastle disease and infectious bronchitis viruses in Cr supplemented birds on days 21 and 42 were higher than those fed control diet with no added Cr. Cr supplementation also decreased plasma level of cortisol. The chicks supplemented with Cr at level of 800  $\mu$ g/Kg, had a higher percentage of lymphocyte and a lower H/L ratio than the control group. Also, Bahrami *et al.* (2012) showed that Cr supplementation improved the immune response of broilers under the heat stress condition. Cr supplemented chicks had a higher antibody titer against Newcastle disease virus and a lower H/L ratio than those fed the control diet on days 18 and 30. No report is available about the effect of Cr supplementation on the immune response of quails.

The increase in the number of lymphocytes T and B in Cr supplemented birds can be attributed to Cr participation in RNA synthesis (Okada *et al.*, 1983), nuclear protein synthesis (Weser and Koolman, 1970) and DNA stability (Okada *et al.*, 1981). Another reason can be the increased glucose availability for lymphocytes as an energy source that is attributed to the improved insulin function (van Heugten and Spears, 1997). Gross and Sigel (1983) observed a greater number of

heterophiles in the blood of quails that were fed with corticosterone than those fed control diet. The increased percentage of lymphocyte and decreased H/L ratio in the heat stressed quails may be attributed to the decreased glucocorticoid secretion. The exact mechanism whereby Cr will strengthen the immune system is not yet known. Cortisol is the most important glucocorticoid that its role in suppression of immune system and prevention of white blood cells production and activity is proven (Roth and Kaeberle, 1982; Munck *et al.*, 1984).

#### Meat quality

Liu *et al.* (2011) reported a significant improvement in meat quality (redness value of breast muscle and pH of thigh muscle) in broiler chicks supplemented with different levels of Zn. Shear force of thigh muscle and drip loss of breast and thigh muscles were also decreased. The results of the study conducted by Yang *et al.* (2011) also show that supplementation of broiler chicks with Zn increases WHC of breast muscle.

Vakili and Rashidi (2011) used a combination of 50 mg Zn and 100 mg vitamin E per Kg of broiler diet under the heat stress condition and reported a decrease in thigh muscle level of MDAV. Sahin et al. (2005) used Zn supplementation in both inorganic (sulfate) and organic (picolinate) form at the levels of 30 and 60 mg/Kg in the diet of Japanese quails under the thermoneutral and heat stress conditions and reported that both forms of Zn supplementation decreased serum and liver levels of MDAV linearly, although ZnSO<sub>4</sub> did not have any effects on serum and liver MDAV under the thermoneutral condition. Zn positively affects meat quality by playing an antioxidant role. Heat stress causes free radicals production (Halliwell and Gutteridge, 1989) and decreases serum density of antioxidant vitamins (A, C and E) and minerals (Zn, Cr) (Feenster, 1985; Sahin and Kucuk, 2003a). Zn deficiency increases lipids peroxidation but this process can be inhibited by dietary Zn supplementation (Shaheen and Abd El-Fattah, 1995). In general, the antioxidant activity mechanism of Zn is divided to acute and chronic effects. The acute effects included long-term access of animal to Zn that induces synthesis of end antioxidants such as metallothionein. The interaction of Zn and vitamin E is one of the proposed mechanisms, so that in Zn-deficient animals, deficiency of vitamin E is also observed (Kim et al., 1998; Salgueiro et al., 2000). Zn deficiency causes oxidative damages through impressing the free radicals activity (Garfinkel, 1986; Powell et al., 1994; Salgueiro et al., 2000). Kim et al. (1998) reported that malabsorption of fat-soluble vitamins such as A and E, is probably due to the faulty construction of chylomicrons in the intestinal cells during the Zn deficiency period. Therfore, some of the oxidative damages in Zn-deficient animals may be related to the deficiency of vitamin E. In many cases, adding vitamin E has overcome some of these shortcomings. In the present study, Zn supplementation may be stimulating metallothionein production that leads to scavenging of free radicals. Improved WHC may be associated with the improved antioxidants

activity such as superoxide dismutase enzyme that benefits of Zn in its structure (Liu *et al.*, 2011).

Sahin *et al.* (2010) used Cr at the levels of 400 and 800  $\mu$ g/Kg in the diet of Japanese quails under the heat stress condition and reported a decrease in MDA level of serum and liver of Cr supplemented birds. Hanafy (2011) reported that Cr supplementation of cocks at the level of 1500  $\mu$ g/Kg decreased MDA level of semen in the winter season. It is proven that Cr reinforces the antioxidant defense system of the body (Preuss *et al.*, 1997). Decreased MDA level is probably due to the prevention of epinephrine production that occurs as the result of Cr effect on insulin and prevention of lipolysis process (Linder, 1991). Improvement of the other meat quality parameters is also probably due to the antioxidant effects of Cr, although researches conducted in this regard are extremely rare.

The results of this study revealed positive effects of Zn and Cr supplementation on the performance, immune responses and meat quality of quails under the heat stress condition. It seems that supplementation of the quails diets with 80 mg/Kg Zn and 1000  $\mu$ g/Kg Cr can be greatly helpful for improving the adverse effects of heat stress.

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#### References

- Bahrami A, Moeini MM, Ghazi SH & Targhibi MR. 2012. The effect of different levels of organic and inorganic chromium supplementation on immune function of broiler chicken under heat-stress conditions. Journal of Applied Poultry Research, 21: 209-215.
- Bartlett JR & Smith MO. 2003. Effects of different levels of zinc on the performance and immunocompetence of broilers under heat stress. Poultry Science, 82: 1580-1588.
- Belay T & Teeter RG. 1996. Effects of environmental temperature on broiler mineral balance partitioned into urinary and fecal loss. British Poultry Science, 37:423-433.
- Brink MF, Becker DE, Terrill SW & Jensen AH. 1950. Zinc toxicity in the weanling pig. Journal of Animal Science, 18: 836-842.
- Bun SD, Guo YM, Guo FC, Ji FJ & Cao H. 2011. Influence of organic zinc supplementation on the antioxidant status and immune responses of broilers challenged with *Eimeria tenella*. Poultry Science, 90: 1220-1226.
- Chandra RK & Dayton DH. 1982. Trace element regulation of immunity and infection. Nutrition Research, 2: 721-733.

- Chesters JK, Petrie L & Lipson KE. 1993. Two zinc-dependent steps during G1 to S phase transition. Journal of Cellular Physiology, 155: 445-451.
- Dardenne M, Pleau JM & Nabarra B. 1982. Contribution of zinc and other metals to the biological activity of the serum thymic factor. Proceeding of the National Academy of Sciences, 79: 5370-5373.
- Dardenne M, Savino W, Berrih S & Bach JF. 1985. A zinc-dependent epitope of the molecule of thymulin, a thymic hormone. Proceeding of the National Academy of Sciences, 82: 7035-7038.
- Dowd PS, Kelleher J & Guillou PJ. 1986. T-lymphocyte subsets and interleukin-2 production in zinc-deficient rats. British Journal of Nutrition, 55: 59-69.
- Ebrahimzadeh SK, Farhoomand P & Noori K. 2012. Immune response of broiler chickens fed diets supplemented with different level of chromium methionine under heat stress conditions. Asian-Australasian Journal of Animal Sciences, 25: 256-260.
- Emami A, Zali A, Ganjkhanlou M & Akbari Afjani A. 2012. Nutrient digestibility, carcass characteristics and plasma metabolites in kids fed diets supplemented with chromium methionine. Online Journal of Animal and Feed Research, 2: 127-132.
- Ensminger ME, Oldfield JE & Heinemann WW. 1989. Feeds and Nutrition. 2<sup>nd</sup> Ed. Prentice Hall Copyright. 1552 Pages.
- Feenster R. 1985. High temperatures decrease vitamin utilization. Misset World Poultry, 38: 38-41.
- Fox MRS. 1989. Zinc Excess. In: Mills CF. (Ed). Zinc in Human Biology. Springerverlag Berlin Heide1berg. New York. Pages, 365-370.
- Fraker PJ, Haas SM & Luecke RW. 1977. Effect of zinc deficiency on the immune response of the young adult A/J mouse. Journal of Nutrition, 107: 1889-1895.
- Garfinkel D. 1986. Is aging inevitable? The intracellular zinc deficiency hypothesis of aging. Medical Hypotheses, 19: 117-137.
- Grau R & Hamm R. 1953. Eine einfache methode zur bestimmung der wasserbindung im muskel. Naturwissenschaften, 40: 29-30.
- Gross WB & Siegel HS. 1983. Evaluation of the heterophil/lymphocyte ratio as a measure of stress in chickens. Avian Disease, 27: 972-979.
- Halliwell B & Gutteridge JMC. 1989. Free Radicals in Biology and Medicine. 2<sup>nd</sup> Ed. Oxford University Press. New York. 543 Pages.
- Hanafy MM. 2011. Influence of adding organic chromium in diet on productive traits, serum constituents and immune status of Bandarah laying hens and semen physical properties for cocks in winter season. Egyptian Poultry Science, 31: 203-216.
- Hayirli A. 2005. Chromium nutrition of livestock species. Nutrition Abstracts and Reviews: Series B 75: 1N-14N.

- Hudson BP, Dozier WA & Wilson JL. 2005. Broiler live performance response to dietary zinc source and the influence of zinc supplementation in broiler breeder diets. Animal feed Science and Technology, 118: 329-335.
- Kim ES, Noh SK & Koo SI. 1998. Marginal zinc deficiency lowers the lymphatic absorption of α-tocopherol in rats. Journal of Nutrition, 128: 265-270.
- Kim JD, Han IK, Chae BJ, Lee JH, Park JH & Yang CJ. 1997. Effects of dietary chromium picolinate on performance, egg quality, serum traits and mortality rate of brown layers. Asian-Australasian Journal of Animal Sciences, 10: 1-7.
- Linder MC. 1991. Nutritional Biochemistry and Metabolism with Clinical Applications. 2<sup>nd</sup> Ed. Prentice-Hall. New York. 603 Pages.
- Liu ZH, Lu L, Li SF, Zhang LY, Xi L, Zhang KY & Luo XG. 2011. Effects of supplemental zinc source and level on growth performance, carcass traits, and meat quality of broilers. Poultry Science, 90: 1782-1790.
- Luecke RW, Siminol CE & Fraker PJ. 1978. The effect of restricted dietary intake on the antibody mediated response of the zinc deficient A/J mouse. Journal of Nutrition, 108: 881-887.
- MacDonald RS. 2000. The role of zinc in growth and cell proliferation. Journal of Nutrition, 130: 1500S-1508S.
- Mallard BA & Borgs P. 1997. Effects of supplemental trivalent chromium on hormone and immune responses of cattle. 13th Alltech Annual Symposium on the Biotechnology in the Feed Industry. Loughborough, UK. Pages, 241-250.
- Mashaly MM, Hendricks GL, Kalama MA, Gehad AE, Abbas AO & Patterson PH. 2004. Effect of heat stress on production parameters and immune responses of commercial laying hens. Poultry Science, 83: 889-894.
- McDonald P, Edwards RA, Greenhalgh JFD, Morgan CA, Sinclair LA & Wilkinson RG. 2011. Animal Nutrition. 7th Ed. Prentice Hall. London. 692 Pages.
- McFarlane JM & Curtis SE. 1989. Multiple concurrent stressors in chicks.: 3. Effects on plasma corticosterone and the heterophil:lymphocyte ratio. Poultry Science, 68: 522-527.
- McIntosh, LJ & Sapolsky RM. 1996. Glucocorticoids increase the accumulation of reactive oxygen species and enhance adriamycin-induced toxicity in neuronal culture. Experimental Neurology, 141: 201-206.
- Mertz W. 1993. Chromium in human nutrition: A review. Journal of Nutrition, 123: 626-633.
- Mujahid A, Yoshiki Y, Akiba Y & Toyomizu M. 2005. Superoxide radical production in chicken skeletal muscle induced by acute heat stress. Poultry Science, 84: 307-314.
- Munck A, Guyre PM & Holbrook NJ. 1984. Physiological functions of glucocorticoids in stress and their relation to pharmacological actions. Endocrine Reviews, 5: 25-44.

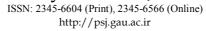
- Namra MMM, Fayek HM & Abdel Wahed HM. 2008. Devaluation of different sources of dietary zinc supplementation for Japanese quail: 1- growth performance. Egyptian Poultry Science, 28: 1023-1041.
- NRC (National Research Council). 1994. Nutrient Requirements of Poultry. 9<sup>th</sup> Rev. Ed., National Academy Press. Washington, DC. 176 Pages.
- Niu ZY, Liu FZ, Yan QL & Li WC. 2009. Effects of different levels of vitamin E on growth performance and immune responses of broilers under heat stress. Poultry Science, 88: 2101-2107.
- Okada S, Ohba H & Taniyama M. 1981. Alterations in ribonucleic acid synthesis by chromium (III). Journal of Inorganic Biochemistry, 15: 223-231.
- Okada S, Susuki M & Ohba H. 1983. Enhancement of ribonucleic acid synthesis by chromium (III) in mouse liver. Journal of Inorganic Biochemistry, 19: 95-103.
- Onderci M, Sahin N, Sahin K & Kilic N. 2003. The antioxidant properties of chromium and zinc: In vivo effects on digestibility, lipid peroxidation, antioxidant vitamins and some minerals under a low ambient temperature. Biological Trace Elements Research, 92: 139-150.
- Oteiza PL, Olin KL, Fraga CG & Keen CL. 1996. Oxidant defense systems in testes from zinc-deficient rats. Proceeding of the Society of Experimental Biology and Medicin, 213: 85-91.
- Parizadian B, Shams Shargh M & Zerehdaran S. 2011. Study the effect of different levels of energy and L- carnitine on meat quality and serum lipids of Japanese quail. Asian Journal of Animal and Veterinary Advances, 9: 944-952.
- Powell SR, Hall D, Aiuto L, Wapnir RA, Teichberg S & Tortolani AJ. 1994. Zinc improves postischemic recovery of isolated rat hearts through inhibition of oxidative stress. American Journal of Physiology, 266: H2497-H2507.
- Prasad AS & Kucuk O. 2002. Zinc in cancer prevention. Cancer Metastasis Reviews, 21: 291-295.
- Prasad AS, Meftah S & Abdallah J. 1988. Serum thymulin in human zinc deficiency. Journal of Clinical Investigations, 82: 1202-1210.
- Preuss HG, Grojec PL, Lieberman S & Anderson RA. 1997. Effects of different chromium compounds on blood pressure and lipid peroxidation in spontaneously hypertensive rats. Clinical Nephrology, 47: 325-330.
- Ritchie BW, Harrison JG & Harrison RL. 1994. Avian Medicine. Winger's Publishing Inc. Florida. USA. 1384 Pages.
- Roth JA & Kaeberle ML. 1982. The effect of glucocorticoids on the bovine immune system. Journal of the American Veterinary Medical Association, 180: 230-235.
- Rubin H. 1972. Inhibition of DNA synthesis in animal cells by ethylene diamine tetraacetate and its reversal by zinc. Proceedings of the National Academy of Sciences, 69: 712-716
- Rubin H & Koide T. 1973. Inhibition of DNA synthesis in chick embryo cultures by deprivation of either serum or zinc. Journal of Cell Biology, 56: 777-786.

- Sahin K, Sahin N, Onderci M, Yaralioglu S & Kucuk O. 2001. Protective role of supplemental vitamin E on lipid peroxidation, vitamins E, A and some mineral concentrations of broilers reared under heat stress. Veterinary Medicine, 46: 140-144.
- Sahin K & Kucuk O. 2003a. Heat stress and dietary vitamin supplementation of poultry diets. Nutrition Abstract and Reviews, 73: 41R-50R.
- Sahin K & Kucuk O. 2003b. Zinc supplementation alleviates heat stress in laying Japanese quail. Journal of Nutrition, 133: 2808-2811.
- Sahin K, Smith MO, Onderci M, Sahin N, Gursu MF & Kucuk O. 2005. Supplementation of zinc from organic or inorganicsource improves performance and antioxidant status of heat-distressed quail. Poultry Science, 84: 882-887.
- Sahin K, Sahin N, Kucuk O, Hayirli A & Prasad AS. 2009. Role of dietary zinc in heat-stressed poultry: A review. Poultry Science, 88: 2176-2183.
- Sahin N, Akdemir F, Tuzcu M, Hayirli A, Smith MO & Sahin K. 2010. Effects of supplemental chromium sources and levels on performance, lipid peroxidation and proinflammatory markers in heat-stressed quails. Animal Feed Science and Technology, 159: 143-149.
- Salgueiro MJ, Zubillaga M, Lysionek A, Sarabia MI, Caro R, De Paoli T, Hager A, Weill R & Boccio J. 2000. Zinc as an essential micronutrient: A review. Nutrition Research, 20: 737-755.
- Sandoval M, Henry PR, Luo XG, Littell RC, Miles RD & Ammerman CB. 1998. Performance and tissue zinc and metallothionein accumulation in chicks fed a high dietary level of zinc. Poultry Science, 77: 1354-1363.
- Shaheen AA & El-Fattah AA. 1995. Effect of dietary zinc on lipid peroxidation, glutathione, protein thiols levels and superoxide dismutase activity in rat tissues. International Journal of Biochemistry and Cell Biology, 27: 89-95.
- Sherman AR. 1992. Zinc, copper and iron nutrition and immunity. Journal of Nutrition, 122: 604-609.
- SAS (Statistical Analysis System). 2008. SAS/STAT® 9.2. User's Guide. SAS Institute Inc. Cary, North Carolina.
- Tarladgis BG, Watts BM & Younathan MT & Dugan L. 1960. A distillation method for the quantitative determination of malonaldehyde in rancid foods. Journal of the American Oil Chemists Society, 37: 44-48.
- Tezuka M, Momiyama K, Edano T & Okada S. 1991. Protective effect of chromium (III) on acute lethal toxicity of carbon tetrachloride in rats and mice. Journal of Inorganic Biochemistry, 42: 1-8.
- Uyanik F, Eren M, Guclu BK & Sahin N. 2005. Effects of dietary chromium supplementation on performance, carcass traits, serum metabolites, and tissue chromium levels of Japanese quails. Biological Trace Elements Research, 103: 187-197.

- Vakili R & Rashidi AA. 2011. The effects of dietary fat, vitamin E and zinc supplementation on fatty acid composition and oxidative stability of muscle thigh in broilers under heat stress. African Journal of Agricultural Research, 6: 2800-2806.
- Van Heugten E & Spears JW. 1997. Immune response and growth of stressed weanling pigs fed diets supplemented with organic or inorganic forms of chromium. Journal of Animal Science, 75: 409-416.
- Walsh CT, Sandstead HH, Prasad AS, Newberne PM & Fraker PJ. 1994. Zinc: health effects and research priorities for the 1990S. Environmental Health Perspectives, 102: 5-46.
- Ward MA & Peterson RA. 1973. The effect of heat exposure on plasma uric acid, lactate dehydrogenase, chloride, total protein and zinc of the broiler. Poultry Science, 52: 1671-1673.
- Weser U & Koolman J. 1970. Reactivity of some transition metals on nuclear protein biosynthesis in rat liver. Cellular and Molecular Life Sciences, 26: 246-247.
- Yang XJ, Sun XX, Li CY, Wu XH & Yao JH. 2011. Effects of copper, iron, zinc, and manganese supplementation in a corn and soybean meal diet on the growth performance, meat quality, and immune responses of broiler chickens. Journal of Applied Poultry Research, 20: 263-271.



# Poultry Science Journal





# تاثیر استفاده از مکمل سولفات روی و کروم آلی بر عملکرد، کیفیت گوشت و پاسخ ایمنی بلدرچین ژاپنی در شرایط تنش گرمایی

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#### چکیده

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#### كلمات كليدى:

روی کروم بلدرچین ژاپنی کیفیت گوشت پاسخ ایمنی این آزمایش به منظور بررسی اثرات استفاده از سطوح مختلف سولفات روی و پیکولینات کروم بر عملکرد، پاسخ ایمنی و کیفیت گوشت بلدرچین ژاپنی در شرایط تنش گرمایی انجام گردید. پر عملکرد، پاسخ ایمنی و کیفیت گوشت بلدرچین ژاپنی در شرایط تنش گرمایی انجام گردید. برندگان بود، تقسیم شدند. آزمایش به صورت فاکتوریل ۳x۳ شامل سطوح مختلف مکمل روی (صفر، ۴۰ و ۸۰ میلی گرم/کیلوگرم) و کروم (صفر، ۵۰۰ و ۱۰۰۰ میکروگرم/کیلوگرم) انجام شد. پرندگان بر روی بستر پرورش داده شدند و از سن ۷ روزگی تا انتهای آزمایش روزانه به مدت ۸ ساعت (۹ تا ۱۷) در معرض تنش گرمایی قرار گرفتند، نوع مکمل روی و کروم مورد استفاده در آزمایش به ترتیب سولفات روی و پیکولینات کروم بود. با افزایش سطح مکمل روی و کروم در جیره، مصرف خوراک به طور خطی کاهش و ضریب تبدیل عذایی بهبود یافت. استفاده از مکمل کروم افزایش وزن روزانه را نیز به طور خطی افزایش داد. با افزایش سطح مکمل روی و کروم در جیره غذایی، نسبت هتروفیل به لنفوسیت و شخصهای کیفیت گوشت بهبود یافتند. بر اساس نتایج، به نظر میرسد استفاده از مکمل های روی و کروم در شرایط تنش گرمایی در بهبود عملکرد، پاسخ ایمنی و کیفیت گوشت بلدرچین موثر باشند. لذا مکمل کردن جیره بلدرچینها با ۸۰ میلی گرم/ کیلوگرم روی و و در جیره میتواند در کاهش اثرات زیانبار تنش گرمایی مفید باشد.

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