Original Article

Acute toxicity of nonylphenol ethoxylate-6 to whiteleg shrimp, *Penaeus vannamei* (Boone, 1931) (Decapoda, Penaeidae)

Azin Ahmadi¹, Ahmad Noori^{1*}, Mahdi Banaee²

¹Department of Fisheries Science, Faculty of Marine Science and Technology, University of Hormozgan, Bandar Abbas, Iran. ²Department of Aquaculture, Faculty of Natural Resources and Environment, Behbahan Khatam Al-Anbia University of Technology, Iran.

Abstract: Nonylphenol ethoxylate-6 (NP6EO) is widely used in industrial and domestic products and easily detected in the environment. The toxicity and estrogenic potency of alkylphenols have been investigated in several studies. However, to the best of our knowledge available, acute toxicity data about the effects of NP6EO on decapod and aquatic ecosystem in literature is yet scarce. Therefore, in this study the adult male and female whiteleg shrimp, *Penaeus vannamei*, were exposed to various concentrations of NP6EO (0.04, 1, 5, 25, 125, 625 μ L L⁻¹) for four days. Acute toxicity potential of NP6EO on adult *P. vannamei* was assessed by calculating LC₅₀ for different times. Median lethal concentration (LC₅₀) of NP6EO at 96 hours was 7.017 μ L L⁻¹. The LC₅₀ of this compound revealed a positive correlation between shrimp mortality and exposure periods. The data exhibited that NP6EO was considered as "toxic" to *P. vannamei* and further toxicity assessment to other species is strongly recommended.

Article history: Received 24 April 2016 Accepted 15 December 2016 Available online 25 February 2017

Keywords: Nonylphenol ethoxylate-6 Shrimp *Penaeus vannamei* Acute toxicity

Introduction

Man-made chemicals are an important part of the modern life. Human beings as well as wildlife populations cannot avoid coming into contact with many of chemicals employed in variety of industries like food production (plants and meat), pathogen control (insecticides), production of modern materials (plastics), or in the built environment (insulations and fire retardants) (Bergman et al., 2012). Considering the importance of these compounds and their widespread presence in the environment, it is important that comprehensive strategies are developed to preclude widespread contamination environmental with endocrine disruptors (EDs) and protect environment (David et al., 2009; Bergman et al., 2012).

Nonylphenol ethoxylate-6 (NP6EO) is used in countless number of applications and because of its extensive use, discharged to the sewer system and make their way into wastewater and aquatic systems (Ying et al., 2002). NP6EO is a nonionic surfactant that is used in a wide range of industrial applications and consumer products, such as laundry detergents, dust-control agents and deicers, industrial liquid soaps and cleaners, cosmetics, paints, and as the dispersing agents in pesticides and herbicides (Jobling and Sumpter, 1993). Concern has recently increased about the use of alkylphenol ethoxylates (APEs) because of the relative stability of their metabolites such as nonylphenol (NP), octylphenol (OP) and nonylphenol ethoxylate-1-3 (NP1-3EO) in the environment (Giger et al., 1984) and their estrogenic effects on organisms (Ying et al., 2002) which is considered as EDs.

Testing strategies employed acute toxicity studies to evaluate and measure the effect(s) of one or more pollutants on one or more species. This implies that tests at high doses will inform us about low-dose exposures (Reish and Oshida, 1986; Bergman et al., 2012). The lethality of the EDs was used as the endpoint in an aquatic acute toxicity testing system (Faheem and Lone, 2013). In general, determination of lethal concentrations, such as the median lethal concentration (LC₅₀), is recognized as the first step for risk assessment of synthetic and natural chemicals (Johnson and Finley, 1980; Ura et al., 2002). These data assist in the development and application of water quality criteria for the protection of the aquatic environment.

In spite of evidence that proved the toxicity of NPnEO on many aquatic animals (Dorn et al., 1993; Lussier et al., 2000; Hirano et al., 2004; Oliveira-Filho et al., 2005; Ricciardi et al., 2008; Liu et al., 2011), there is no attempts have been made to determine the impacts of NP6EO on *Penaeus vannamei*. Hence, this study aims to determine and compare the acute toxicity of NP6EO to *P. vannamei* upon modification of the exposure conditions.

Materials and Methods

Chemicals: NP6EO (CAS No.9016-45-9) was obtained from Kimiagaran Emroz Company (Tehran, Iran). Stock solution of the NP6EO was prepared by dissolving appropriate concentration in 96% ethanol as solvent. Required concentrations were obtained by serial dilution and stored in dark at 4° C until usage. Solvent concentration was kept at 0.01% (v/v) for all treatments.

Animal maintenance and exposure to NP6EO: Adult P. vannamei (both sexes; body weight: 25.89±0.79 g, total length: 14.26±0.16 cm) were obtained from shrimp farms located at southern coast lines of Iran and were transferred to Kolahi Aquatic Restock Center (KARC). Adult shrimps were acclimated in 300L fiberglass tanks containing ultraviolet-filtered recirculating water (pH=7.77±0.017) for 2 weeks prior to the experiment. Feeding was done on daily basis in four times at the rate of 2.5% of the body weight by commercial feed until 24 hrs prior to the initiation of the test. After the acclimation period, 240 adult shrimp were randomly distributed among 24 tanks, comprising control, vehicle control (ethanol with a final concentration of 1:1,000 v/v water), 0.04, 1, 5, 25, 125, 625 µL L⁻¹ of NP6EO.

The experiment was run in triplicate, without feeding and water exchange during the experiment. The mortality of shrimps in each treatments were counted and recorded over the exposure period at 24, 48, 72 and 96 hrs. Dead shrimps were removed from treatments immediately. The study was approved by the Iranian Society for the Animal Welfare.

Data analysis and statistics: Data are expressed as mean with the corresponding standard error (SE). LC values and 95% confidence intervals (95% CI) were calculated using probit analysis. Shapiro-Wilk and Levene's tests were used to check the normality of data distribution and the homogeneity of variances, respectively (Zar, 2010). If data support the prerequisites for parametric analysis, one-way analysis of variance (ANOVA) followed by Tukey's multiple range test was applied. Otherwise, Kruskal-Wallis and Mann-Whitney U test were applied to determine the statistical significance (Zar, 2010). All the analysis was performed by SPSS 16.0. The significant level in all analysis was set at $P \le 0.05$.

Results

The percent mortality of *P. vannamei* after exposure to various concentrations of NP6EO for 24, 48, 72 and 96 hrs has been depicted in Figure 1. Mortality increased with increasing concentrations and exposure time (Fig. 1). The LC₅₀ values were $437.052 \pm 326.250 \mu$ L L⁻¹ for 24 hrs, 33.627 ± 15.443 μ L L⁻¹ for 48 hrs, $10.816 \pm 3.936 \mu$ L L⁻¹ for 72 hrs and $7.017 \pm 2.391 \mu$ L L⁻¹ for 96 hrs.

The LC values, their upper and lower confidence limits and slope functions for NP6EO have been given in Table 1.

Discussion

Acute toxicity tests provide a measure of the toxicity of the given compounds to experimental species under specific environmental conditions (Reish and Oshida, 1986). They also reflect the severe and rapid damage caused by sudden exposure to lethal concentrations of contaminants (Alam and Maughan, 1993).

In the present study, calculated LC₅₀-96 hrs value

Table 1. Effective dose, confidence limits, and slope function for nonylphenol ethoxylate-6 (NP6EO) at different intervals for the whiteleg shrimp, *Penaeus vannamei.*

Exposure periods	Effective dose	SE	limits		Slope	't'	Hotomasonsit
	(µL/L)		LCL	UCL	function	ratio	Heterogeneity
	LC1=0.006	0.009	0.000	0.052			
	LC ₅ =0.161	0.150	0.012	0.683			
24 hrs	LC10=0.922	0.647	0.144	2.895	0.479±0.081	5.920	1.135
	LC ₂₀ =7.642	3.976	2.346	20.537			
	LC ₅₀ =437.052	326.250	131.656	3241.25			
	LC ₈₀ =*	*	3340.90	*			
	LC ₉₀ =*	*	*	*			
	LC ₉₅ =*	*	*	*			
	LC99=*	*	*	*			
48 hrs 72 hrs	LC1=0.001	0.002	0.000	0.010	0.530±0.070 7		1.540
	LC ₅ =0.026	0.023	0.003	0.107		7.548	
	$LC_{10}=0.128$	0.089	0.022	0.400			
	$LC_{20}=0.866$	0.447	0.255	2.124			
	LC ₅₀ =33.627	15.443	14.568	95.011			
	$LC_{80} = 1305.031$	1032.11	362.622	9758.932			
	LC ₉₀ =8834.585	8962.67	1755.602	*			
	LC ₉₅ =*	*	6344.154	*			
	LC ₉₉ =*	*	*	*			
	LC ₁ =0.002	0.002	0.000	0.042			
	$LC_5 = 0.028$	0.020	0.000	0.247	0.635±0.075	8.436	2.903
	$LC_{10}=0.104$	0.063	0.001	0.664			
	$LC_{20}=0.512$	0.237	0.022	2.438			
	LC ₅₀ =10.816	3.936	2.243	74.971			
	$LC_{80}=228.489$	127.492	39.623	*			
	$LC_{90}=1125.541$	802.962	133.766	*			
	$LC_{95}=4199.877$	3578.53	348.322	*			
	LC ₉₉ =*	*	1984.653	*			
96 hrs	LC ₁ =0.003	0.002	0.000	0.035	0.680±0.078 8.7		
	$LC_5=0.027$	0.018	0.000	0.193		8.755	2.538
	$LC_{10}=0.091$	0.052	0.002	0.495			
	$LC_{20} = 0.406$	0.182	0.028	1.673			
	LC ₅₀ =7.017	2.391	1.708	33.301			
	$LC_{80}=121.319$	59.905	26.836	2593.424			
	$LC_{90}=538.201$	338.229	87.163	*			
	$LC_{95}=1841.874$	1382.53	219.975	*			
	$LC_{99}=*$	*	1182.462	*			

* Values more than 10000 are not shown

of NP6EO for *P. vannamei* was assessed as 7.017 μ L L⁻¹, which was in agreement with the results reported for other species have been tested with NP and NPnEO (Dorn et al., 1993; Mann and Bidwell, 2000; Oliveira-Filho et al., 2005; Ricciardi et al., 2008). The acute and chronic toxicity of alkylphenol ethoxylates and their metabolites have been investigated for several freshwater and marine species (Servos, 1999; Staples et al., 2004). Previous studies showed that species sensitivity varies from 17 μ g L⁻¹ of para-nonylphenol (PNP) for winter

flounder (*Pleuronectes americanus*) (Lussier et al., 2000) to 9.2 mg L⁻¹ of NP8EO for *Litoria adelaidensis* (Mann and Bidwell, 2000). Median lethal concentrations of 4 to 6.6 mg L⁻¹ of NP9EO was reported for fathead minnow, *Pimephales promelas* (Dorn et al., 1993; Staples et al., 1998). Also in the other similar studies, LC₅₀-96 hrs for NP, NP1EO and NP2EO for fathead minnow were 136, 218, and 323 μ g L⁻¹, respectively (TenEyck and Markee, 2007). The varying data of available toxicity tests resulted as a function of ethoxy chain *WWW.SID.U*

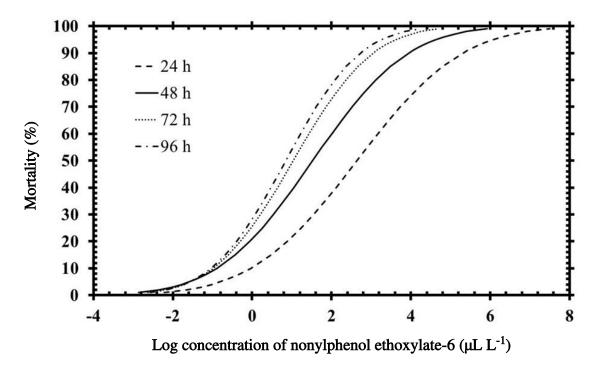


Figure 1. Percent mortality of *Penaeus vannamei* after 24, 48, 72 and 96 hrs exposure to different concentrations of nonylphenol ethoxylate-6 (NP6EO) (μ L L⁻¹).

length, the type of test used, and the species tested.

Considering the relative toxicity values, or NP toxic equivalency factors (TEFs) which calculated for nonylphenol compounds, toxic concentrations for different nonylphenolic compounds such as NPnEO with various EO chain length could be matched with similar endpoints for NP for the same species. If TEF for NP be considered as 1, for example, toxic equivalency factors for NPnEO (1 < n < 8) will be considered as 0.5 (Environment Canada, 2002). Based on the present results, it can be proposed that in *P. vannamei* sensitivity to NP toxicity is approximately the same as other studied species.

According to the USEPA toxicity categories (U.S. EPA., 2015), chemicals with LC_{50} values ranging from 1 to 10 mg L⁻¹ are considered as "toxic to aquatic species with long lasting effects". Therefore, our result strongly indicate that NP6EO is toxic for *P. vannamei*.

This study assessed the acute toxicity of NP6EO on *P. vannamei*, which is the first report of LC_{50} values of this chemical on this species. The work has revealed that NP6EO has a potentially harmful impact on *P. vannamei* and probably other aquatic

crustaceans and invertebrates. The response of various species to same chemicals are different at the same time of exposure. Moreover, the potential of NP6EO for endocrine disruption at even lower concentrations may cause further concerns for aquatic fauna. Additional long term studies are needed to study other impacts of NP6EO on aquatics. The findings of this research suggest that the policy makers must take necessary measures to prevent the crucial damage of this compound on aquatic organisms and human beings.

Acknowledgments

The authors wish to thank M. Darvishi for providing shrimp. Also the authors thank Mr. Sirpoor for providing the facilities and KARC staffs for their helps.

References

- Alam M.K., Maughan O.E. (1993). Acute toxicity of selected organophosphorus pesticides to *Cyprinus carpio* and *Barilius vagra*. Journal of Environmental Science and Health, Part B, 28: 81-89.
- Bergman Å., Heindel J., Jobling S., Kidd K., Zoeller R.T. (2012). State of the science of endocrine disrupting

chemicals. Toxicology Letters, 211: S3.

- David A., Fenet H., Gomez E. (2009). Alkylphenols in marine environments: distribution monitoring strategies and detection considerations. Marine Pollution Bulletin, 58: 953-960.
- Dorn P.B., Salanitro J.P., Evans S.H., Kravetz L. (1993). Assessing the aquatic hazard of some branched and linear nonionic surfactants by biodegradation and toxicity. Environmental Toxicology and Chemistry, 12: 1751-1762.
- Environment Canada (2002). Canadian environmental quality guidelines for nonylphenol and its ethoxylates. Scientific supporting document (water, sediment, and soil). Environment Canada, Environmental Quality Branch, National Guidelines and Standards Office, Ottawa.
- Faheem M., Lone K.P. (2013). Acute toxicity and Behavioural response of Cirrhinus mrigala fingerlings to Bisphenol-A. International Journal of Open Scientific Research, 1: 28-37.
- Giger W., Brunner P.H., Schaffner C. (1984). 4-Nonylphenol in sewage sludge: accumulation of toxic metabolites from nonionic surfactants. Science, 225: 623-625.
- Hirano M., Ishibashi H., Matsumura N., Nagao Y., Watanabe N., Watanabe A., Onikura N., Kishi K., Arizono K. (2004). Acute toxicity responses of two crustaceans, Americamysis bahia and Daphnia magna, to endocrine disrupters. Journal of Health Science, 50: 97-100.
- Jobling S., Sumpter J.P. (1993). Detergent components in sewage effluent are weakly oestrogenic to fish: An in vitro study using rainbow trout (Oncorhynchus mykiss) hepatocytes. Aquatic Toxicology, 27: 361-372.
- Johnson W.W., Finley M.T. (1980). Handbook of acute toxicity of chemicals to fish and aquatic invertebrates: Summaries of toxicity tests conducted at Columbia National Fisheries Research Laboratory, 1965-78US Fish and Wildlife Service. 98 p.
- Liu Y., Tam N.F.Y., Guan Y., Yasojima M., Zhou J., Gao B. (2011). Acute toxicity of nonylphenols and bisphenol A to the embryonic development of the abalone Haliotis diversicolor supertexta. Ecotoxicology, 20: 1233-1245.
- Lussier S.M., Champlin D., LiVolsi J., Poucher S., Pruell R.J. (2000). Acute toxicity of para-nonylphenol to

saltwater animals. Environmental Toxicology and Chemistry, 19: 617-621.

- Mann R.M., Bidwell J.R. (2000). Application of the FETAX protocol to assess the developmental toxicity of nonylphenol ethoxylate to Xenopus laevis and two Australian frogs. Aquatic Toxicology, 51: 19-29.
- Oliveira-Filho E., Geraldino B., Grisolia C., Paumgartten F. (2005). Acute toxicity of endosulfan, nonylphenol ethoxylate, and ethanol to different life stages of the freshwater snail Biomphalaria tenagophila (Orbigny, 1835). Bulletin of environmental contamination and toxicology, 75: 1185-1190.
- Reish D.L., Oshida P.S. (1986). Manual of methods in aquatic environment research, Part 10: short-term static bioassays. FAO. 62 p.
- Ricciardi F., Matozzo V., Marin M.G. (2008). Effects of 4-nonylphenol exposure in mussels (Mytilus galloprovincialis) and crabs (Carcinus aestuarii) with particular emphasis on vitellogenin induction. Marine Pollution Bulletin, 57: 365-372.
- Servos M.R. (1999). Review of the aquatic toxicity, estrogenic responses and bioaccumulation of alkylphenols and alkylphenol polyethoxylates. Water Quality Research Journal of Canada, 34: 123-177.
- Staples C., Mihaich E., Carbone J., Woodburn K., Klecka G. (2004). A weight of evidence analysis of the chronic ecotoxicity of nonylphenol ethoxylates, nonylphenol ether carboxylates, and nonylphenol. Human and Ecological Risk Assessment, 10: 999-1017.
- Staples C.A., Weeks J., Hall J.F., Naylor C.G. (1998). Evaluation of aquatic toxicity and bioaccumulation of C8-and C9-alkylphenol ethoxylates. Environmental Toxicology and Chemistry, 17: 2470-2480.
- TenEyck M.C., Markee T.P. (2007). Toxicity of Nonylphenol, Nonylphenol Monoethoxylate, and Nonylphenol Diethoxylate and Mixtures of these Compounds to Pimephales promelas (Fathead Minnow) and Ceriodaphnia dubia. Archives of Environmental Contamination and Toxicology, 53: 599-606.
- U.S. EPA. (2015). Chemical hazard classification and labeling: comparison of opp requirements and the GHS. US Environmental Protection Agency. 22 p.
- Ura K., Kai T., Sakata S., Iguchi T., Arizono K. (2002). Aquatic acute toxicity testing using the nematode Caenorhabditis elegans. Journal of health science, 48: 583-586.

- Ying G.-G., Williams B., Kookana R. (2002). Environmental fate of alkylphenols and alkylphenol ethoxylates—a review. Environment International, 28: 215-226.
- Zar J.H. (2010). Biostatistical analysis. Fifth edition. Pearson Prentice Hall. Upper Saddle River, New Jersey, 947 p.

چکیدہ فارسی

تاثیر تعیین سمیت حاد نونیل فنل ۶–اتوکسیلات بر روی میگوی پاسفید غربی Penaeus vannamei (Boone, 1931) (Decapoda, Penaeidae)

آذين احمدي^۱، احمد نوري^۱*، مهدي بنايي^۲

^آگروه شیلات، دانشکده علوم و فنون دریایی، دانشگاه هرمزگان، بندرعباس، ایران. ^۲گروه شیلات، دانشکده منابع طبیعی و محیط زیست، دانشگاه صنعتی خاتم الانبیاء (ص) بهبهان، بهبهان، ایران، کدپستی: ۴۷۱۸۹-۶۳۶۱۶

چکیدہ:

نونیل فنل ۶-اتوکسیلات کاربرد وسیعی در تولید محصولات خانگی و صنعتی داشته و به همین دلیل به راحتی در محیط زیست قابل سنجش می باشد. سمیت و خاصیت شبه استروژنی ترکیبات آلکیل فنل در بسیاری از مطالعات مورد ارزیابی قرار گرفته است. با این وجود کمبود اطلاعات در خصوص تاثیرات سمیت حاد نونیل فنل ۶-اتوکسیلات بر روی دهپایان و همچنین اکوسیستمهای آبی به خوبی احساس میگردد. به همین منظور، در مطالعه حاضر هر دو جنس نر و ماده بالغ میگوی پاسفید غربی (Penaeus vannamei) در یک دوره چهار روزه در معرض غلظتهای مختلف نونیل فنل ۶-اتوکسیلات (۲۰۱۰، ۱، ۵، ۲۵، ۲۵ و ۲۵ میکرولیتر بر لیتر) قرار گرفت. سمیت حاد نونیل فنل ۶-اتوکسیلات با توجه به مقدار غلظت در زمانهای مختلف ارزیابی شد. با توجه به نتایج حاصل، مقدار غلظت میال در این ۶-اتوکسیلات برای ۹۶ ساعت برابر با ۲۰۱۷ میکرولیتر بر لیتر محاسبه شد. غلظت ۵. LCم میکرولیتر بر لیتر) قرار گرفت. سمیت حاد نونیل فنل ۶-اتوکسیلات با توجه به مقدار غلظت میکرولیتر بر لیتر محاسبه شد. غلظت ۵. LCم محاسبه شده در این تحقیق بیانگر رابطه مستقیم بین بروز مرگ و میر در میگوها و مدت زمان تاثیر گذاری بود. نتایج نشان داد که نونیل فنل ۶-اتوکسیلات برای میگوی پاسفید غربی ترکیبی سمی به شمار میرود و به همین دلیل ارزیابی سمیت

کلمات کلیدی: نونیل فنل ۶-اتوکسیلات، میگو، پاسفید غربی، سمیت حاد.