Original Article

Effects of dietary vitamin C supplementation on some oxidative status biomarkers in erythrocytes of common carp (*Cyprinus carpio*)

Hamideh Ghodrati Azadi*¹, Davar shahsavani², Hasan Baghshani¹

¹Department of basic Sciences, School of Veterinary Medicine, Ferdowsi University of Mashhad, Mashhad, Iran. ²Department of Food Hygiene and Aquaculture, School of Veterinary Medicine, Ferdowsi University of Mashhad, Mashhad, Iran.

Abstract: Regarding to the high content of polyunsaturated fatty acids in fish tissues, improving the fish antioxidant status seems to be necessary and may be associated with beneficial effects on fish health. The present study aimed to investigate the effects of dietary vitamin C supplementation (20 mg/kg body weight, 4 weeks) on some oxidative status biomarkers in RBC of common carp (*Cyprinus carpio*). The results showed that the activities of antioxidant enzymes including catalase (CAT), glutathione peroxidase (GPx) and superoxide dismutase (SOD) were not changed significantly following dietary vitamin C supplementation in comparison to the control group. Moreover, dietary vitamin C supplementation for 28 days significantly lowered malondialdehyde (MDA) concentration in erythrocyte haemolysate by approximately 26% compared to that of the control group. In conclusion, dietary vitamin C supplementation appears to be able to protect carp erythrocytes against oxidative stress by decreasing lipid peroxidation.

Article history: Received 17 Aug 2017 Accepted 20 October 2017 Available online 25 October 2017

Keywords: Vitamin C Antioxidant enzymes Lipid peroxidation Cyprinus carpio

Introduction

A variety of reactive oxygen species (ROS), derived from reduction of oxygen, ionizing radiations, reactive metals. and other environmental initiators, permanently threatens living organisms (Birben et al., 2012). Oxidative stress is a consequence of an increased generation of free radicals and/or reduced physiological activity of antioxidant defenses against them. Oxidative stress impairs DNA, causes enzymes and membrane disorders, changes the activity of the immune system, and alters the structure of basic biopolymers, which, cause various disorders (Abd Ellah, 2010). In both mammals and fish, insufficient dietary antioxidants have been followed by a decrease in antioxidative defense and increased susceptibility to oxidative stress (Sies et al., 2005; Welker and Congleton, 2009). The antioxidant capacity of fish may be insufficient in captivity (Mohebbi et al., 2011). Regarding to increased environmental pollutants and thus increased risk of oxidative stress, administration of dietary antioxidants would be beneficial for cultured fish. Moreover, in view of the high content of

polyunsaturated fatty acids in aquaculture feeds and fish tissues, improving antioxidant capacity of fish seems to be necessary and may be associated with many beneficial effects on fish health (Mohebbi et al., 2011).

Fish tissues are characterized by high concentrations of polyunsaturated fatty acids and may therefore be particularly susceptible to lipid peroxidation. Both oxidative responses and antioxidant potential of fish differ according to species habitat and feeding behavior (Yonar and Sakin, 2011). Dietary antioxidant vitamins may play an important role in protecting fish against oxidant damages. Vitamin C is important for many enzymatic reactions and also acts as a freeradical scavenger. Vitamin C deficiency has been shown to retard growth and impair wound healing in O. mykiss and Oncorhynchus kisutch (Zhou et al., 2003). Vitamin C supplementation in diet promoted growth in sea bass (Lates calcarifer) and common carp (Gouillou-Coustans et al., 1998). On the other hand, it has been documented that vitamin C affects metabolism of lipids and carnitine, which is essential

^{*}Corresponding author: Hamideh Ghodrati Azadi E-mail address: ghodrati@ferdowsi.um.ac.ir

for the β -oxidation of long-chain fatty acids (Feller and Rudman, 1988).

Potential functions of vitamin C in RBC include maintenance of plasma ascorbate concentrations by ascorbate or dehydroascorbic acid efflux from red blood cells, transmembrane electron transfer from erythrocyte ascorbate, and antioxidant functions to protect erythrocytes from oxidative damage or to recycle membrane tocopherol (Li et al., 2012). It has been reported that dietary vitamin C supplementation could significantly increase growth performance and antioxidant status in some aquatic animals (Asaikkutti et al., 2016; Gouillou-Coustans et al., 1998).

Common carp (*Cyprinus carpio*) has great commercial importance because it is widely consumed all over the world. Although cyprinids appear to be able to synthesize vitamin C at rates sufficient to meet its physiological needs, dietary supplementation with vitamin C might be beneficial when metabolic demand exceeds endogenous supply (Combs, 1998). Therefore, the present study was conducted to assess the effects of dietary vitamin C supplementation on some oxidative status biomarkers in RBC of common carp.

Materials and Methods

Vitamin C was purchased from Nature Made (California, USA). Commercial enzyme kits for superoxide dismutase (Ransod, RANDOX/SD-125) and glutathione peroxidase (Ransel, RANDOX/RS-505) were obtained from Randox Laboratories (Crumlin, UK). MS-222 (Ethyl 3-aminobenzoate methanesulfonate, Tricaine) and 2-thiobarbituric acid (TBA) was purchased from Sigma Chemical Co. (St. Louis, MO, USA). The rest of the utilized chemicals were of analytical grade and were supplied by Sigma (St Lewis, MO, USA) or Merck (Darmstadt, Germany).

Common carp (total n=60), weighing 60-80 g, were obtained from a local commercial farm. They were held in four glass aquaria, each containing 250 L freshwater. The fish were acclimatized for 7 days (Naeiji et al., 2013) before the commencement of the experiment and were daily fed with commercial fish feed at 3% of total body weight at a fixed time. The aquaria were aerated continuously. Physicochemical characteristics of the water during the experimental period were: dissolved oxygen, 5.5-6 ppm: temperature, 25±1°C; pH, 7±0.5; photoperiod, 12:12 light-dark. The aquaria water was renewed every 48 h (Naeiji et al., 2013). The fish were divided randomly into two groups of 30 each. Group 1 fish were held in two aquaria (each containing 15 fish) and fed with basal diet; served as control. Group 2 fish were held in two aquaria (each containing 15 fish) and fed the basal diet supplemented with vitamin C (20 mg/kg body weight, daily (NRC, 1993)) for 4 weeks. The vitaminsupplemented diet was prepared by spraying vitamin C solution onto the pellet (Treves-Brown, 2000).

At the end of the experimental period, twenty fish were sampled randomly from each aquarium and anesthetized in diluted MS-222. Blood samples were taken by cardiac puncture using heparinized syringes and tubes. After plasma separation by centrifugation at 750×g for 20 min (Nazifi et al., 2010), erythrocyte pellet was washed three times with normal saline solution. The washed centrifuged erythrocytes were hemolyzed by the addition of an equal volume of ice-cold redistilled water (Nazifi et al., 2010) and prepared hemolysate aliquots were stored at -70° C until analysis.

Glutathione peroxidase (GPx) activity was measured using RANDOX-Ransel enzyme kit. In this method, GPx catalyzes the oxidation of GSH by cumene hydroperoxide. In the presence of glutathione reductase (GR) and NADPH, the oxidized glutathione (GSSG) is immediately converted to the reduced form, with a concomitant oxidation of NADPH to NADP⁺. The decrease in absorbance at 340 nm was measured spectrophotometrically, and the results were expressed as units per gram hemoglobin. Hemoglobin (Hb) concentration was measured by cyanmethemoglobin method (Prakash and Banerji, 1972).

Superoxide dismutase (SOD) activity was determined by a modified method of iodophenyl nitrophenol phenyltetrazolium chloride using the RANDOX-Ransod enzyme kit. This method employs xanthine and xanthine oxidase to generate superoxide

Parameter	Control	Vitamin C
Catalase (U/g Hb)	273.41±30.95	310.25±24.74
Superoxide dismutase (U/g Hb)	1573.67±68.53	1557.90±53.67
Glutathione peroxidase (U/g Hb)	744.96±38.37	836.4±38.86
Malondialdehyde (nmol/g Hb)	129.29 ± 10.37 a	95.25±7.40 ^b

Table 1. Mean±SEM of measured oxidative status biomarkers in experimental groups (n= 20 in each group).

^{a, b} Mean±SEM in each row with no common superscript differ significantly (P<0.05).

radicals which react with 2-(4-iodophenyl)-3-(4nitrophenol)-5-phenyltetrazolium chloride (INT) to form a red formazan dye. The SOD activity was then measured by the degree of inhibition of this reaction. One unit of SOD was considered a 50% inhibition of reduction of INT under the condition of the assay. The results were expressed as units per gram hemoglobin.

Catalase (CAT) activity was measured in the RBC hemolysate by the method described by Claiborne (1986) and expressed as units per gram hemoglobin. The decomposition of H₂O₂ can be directly followed by the decrease in absorbance at 240 nm. The difference in absorbance at 240 nm per time unit allows determining the CAT activity. 50 µl of the diluted hemolysate was mixed with 1.9 ml of the phosphate buffer (0.05 M, pH 7) and 1 ml of 30 mM H₂O₂. The decrease in absorption was measured at 240 nm for 1 min. Activity of catalase was calculated using the extinction coefficient of 43.6 M⁻¹cm⁻¹. One unit of catalase activity is equal to the amount of enzyme that will decompose 1 µmol H₂O₂ per minute.

Lipid peroxidation was assayed by measurement of malondialdehyde (MDA). Determination of MDA concentration was based on spectrophotometry of the pink-colored product of thiobarbituric acid reactive substances, as described by Latha and Pari (2003). The concentration of MDA was calculated using a molar extinction coefficient value of 156,000 M^{-1} cm⁻¹. The results were expressed as nanomoles of MDA per gram hemoglobin.

Statistical analysis: All experimental values have been represented as mean \pm standard error of mean (SEM). The obtained data were analyzed using Student's t-test. The level of significance was set at *P*<0.05. All calculations were performed using SPSS/PC software, version 18.

Results

The values (mean±SEM) of the measured erythrocyte oxidative status biomarkers in experimental groups are presented in Table 1. The activities of antioxidant enzymes including CAT, GPx and SOD were not changed significantly following dietary vitamin C supplementation in comparison to the control group. As shown in Table 1, dietary vitamin C supplementation for 28 days significantly lowered the MDA concentration in erythrocyte hemolysate by approximately 26% compared to concentration in the control group.

Discussion

Measurement of circulatory biomarkers of oxidative stress has emerged as a reliable method for screening putative antioxidative agents (Balasenthil et al., 2000). The extent of lipid peroxidation is most frequently measured by estimating MDA levels (Latha and Pari, 2003). Fish erythrocytes have been proposed as a useful model to investigate oxidative stress, since their membranes are rich in long chain n-3 polyunsaturated fatty acids, which are oxidized under oxidative stress conditions (Roche and Boge, 1993; Gabryelak et al., 2000; Nagasaka et al., 2004). Moreover, repeatedly exposure to high concentration of oxygen or presence of iron renders erythrocytes highly susceptible to peroxidative damage (Clemens and Waller, 1987). The effect of vitamin C in decreasing MDA levels in the present study suggest that this vitamin may provide an effective protection against lipid peroxidation. Similar to this finding, it has been reported that the level of thiobarbituric acid reactivesubstances (TBARS) in the hepatopancreas and muscle of common carp was decreased following vitamin C supplementation (Hwang and Lin, 2002). Moreover, Chien and Hwang (2001) indicated that vitamin C can prevent the increase in liver lipid peroxidation due to high water temperature in thorn fish *Terapon jarbua*. Similarly, it has been reported that dietary vitamin C supplementation caused a significant decrease in the levels of tissue lipid peroxides in chicken erythrocytes (Aydemir et al., 2000).

Vitamin C as a strong reducing reagent has a direct reactive oxygen scavenger action. Additionally, depletion of tissue GSH is one of the primary factors that permit lipid peroxidation and vitamin C has been increase intracellular shown to glutathione concentrations (Hwang and Lin, 2002; Johnston, 1993). Another criterion, which might contribute to the lowering effect of vitamin C on peroxidation of hydrophobic regions of the cells may be its ability to reduce the semi-stable β -tocopheroxyl radical (vitamin E radical form after performing the antioxidant role), thus regenerating the metabolically active form of the lipid antioxidant vitamin E (Kontush et al., 1996).

The detoxification of ROS involves the cooperative action of the intracellular antioxidant enzymes SOD, CAT, and GPx. GPx contributes to the oxidative defense of animal tissues by catalyzing the reduction of hydrogen peroxide and lipid peroxides (Harvey, 1997). CAT has an equal importance to in the defense of human erythrocytes against H₂O₂ generating reactions (Harvey, 1997). Superoxide dismutase is also important in the antioxidant defense mechanism and protects against lipid peroxidation (Miller et al., 1993). Animal studies have shown that antioxidant enzymes' levels are variable depending on the availability of antioxidants in food (Meydani, 1993; Puangkaew et al., 2005). It has been reported that SOD and GPx activities increased but CAT activity decreased with an increase of plasma vitamin C levels (Monget et al., 1996). In chicken the activities of erythrocyte SOD and GPx increased following vitamin C supplementation although no significant variations were observed in the CAT activity (Aydemir et al., 2000). Similarly, the present results indicate no significant increment of the measured

antioxidant enzymes following vitamin C supplementation in carp erythrocytes.

The results of the present work suggest that dietary supplementation of vitamin C appears to be able to protect erythrocytes of carp against oxidative stress by decreasing lipid peroxidation. However, more investigations are required to elucidate the pharmacokinetic effects of this vitamin and also precise molecular basis of the beneficial effects of vitamin C in fish species.

Acknowledgments

This study was supported by grant from Ferdowsi University of Mashhad, Mashhad, Iran.

References

- Abd Ellah M.R. (2010). Involvement of free radicals in animal diseases. Comparative Clinical Pathology, 19: 615-619.
- Asaikkutti A., Bhavan P.S., Vimala K., Karthik M., Cheruparambath P. (2016). Effect of different levels of dietary vitamin C on growth performance, muscle composition, antioxidant and enzyme activity of *Macrobrachium rosenbergii*. Aquaculture Reports, 3: 229-236.
- Ashley L.M., Halver J.E., Smith R.R. (1975). The pathology of fishes. The University of Wisconsin Press, Wisconsin. pp: 769-786.
- Aydemir T., Oztürk R., Bozkaya L.A., Tarhan L. (2000). Effects of antioxidant vitamins A, C, E and trace elements Cu, Se on CuZn SOD, GSH-Px, CAT and LPO levels in chicken erythrocytes. Cell Biochemistry Function, 18(2): 109-115.
- Balasenthil S., Arivazhagan S., Nagini S. (2000). Garlic enhances circulatory antioxidants during 7, 12dimethylbenz[a]anthracene-induced hamster buccal pouch carcinogenesis. Journal of Ethnopharmacology, 72: 429-433.
- Birben E., Sahiner U.M., Sackesen C., Erzurum S., KalayciO. (2012). Oxidative stress and antioxidant defense.World Allergy Organization Journal, 5(1): 9-19.
- Chien L.T., Hwang D.F. (2001). Effects of thermal stress and vitamin C in lipid peroxidation and fatty acid composition in the liver of thornfish *Terapon jarbua*. Comparative Biochemistry Physiology, 128B: 91-97.
- Claiborne A. (1986). Catalase activity. In: R.A. Greenwald www.SID.ir

(Ed.). Handbook of methods for oxygen radical research, CRC Press, Boca Raton (FL). pp: 283–284

- Clemens M.R., Waller H.D. (1987). Lipid peroxidation in erythrocytes. Chemistry and Physics of Lipids, 45: 251-268.
- Feller A.G., Rudman D. (1988). Role of carnitine in human nutrition. Journal of Nutrition, 118: 541-547.
- Gabryelak T., Filipiak A., Brichon G. (2000). Effects of zinc on lipids of erythrocytes from carp (*Cyprinus carpio* L.) acclimated to different temperatures. Comparative Biochemistry Physiology, 127C: 335-343.
- Combs Jr. G.F. (1998). The Vitamins: Fundamental Aspects in Nutrition and Health. 2nd ed. Academic Press, London. pp: 246-275.
- Gouillou-Coustans M.F., Bergot P., Kaushik S.J. (1998). Dietary ascorbic acid needs of common carp (*Cyprinus carpio*) larvae. Aquaculture, 161: 453-461.
- Harvey J.W. (1997). The erythrocyte: physiology, metabolism and biochemical disorders. In: J.J. Kaneko, J.W. Harvey, M.L. Bruss (Eds.). Clinical biochemistry of domestic animals. 5th edition, Academic Press, London. pp: 182-184.
- Hwang D.F., Lin T.K. (2002). Effect of temperature on dietary vitamin C requirement and lipid in common carp. Comparative Biochemistry and Physiology, 131B: 1-7.
- Johnston C., Meyer C., Srilakshmi J. (1993). Vitamin C elevates red blood cell glutathione in healthy adults. American Journal of Clinical Nutrition, 58: 103-105.
- Kontush A., Finckh B., Karten B., Kohlschutter A., Beisiegel U. (1996). Antioxidant and prooxidant activity of a-tocopherol in human plasma and low density lipoprotein. Journal of Lipid Research, 37: 1436-1448.
- Latha M., Pari L. (2003). Preventive effects of *Cassia auriculata* L. flowers on brain lipid peroxidation in rats treated with streptozotocin. Molecular and Cellular Biochemistry, 243: 23-28.
- Li H., Tu H., Wang Y., Levine M. (2012). Vitamin C in mouse and human red blood cells: an HPLC assay. Analytical Biochemistry, 426(2): 109–117.
- Meydani S.N. (1993). Vitamin, mineral supplementation, the aging immune response, and risk of infection. Nutrition Review, 51: 106-115.
- Miller J.K., Brzezinska-Slebodzinska E., Madsen F.C. (1993). Oxidative stress, antioxidants, and animal function. Journal of Dairy Science, 76: 2812-2823.

Mohebbi A., Nematollahi A., Ebrahimi Dorcheh E.,

Goodarzian Asad F. (2012). Influence of dietary garlic (*Allium sativum*) on the antioxidative status of rainbow trout (*Oncorhynchus mykiss*). Aquaculture Research, 43 (8): 1184-93.

- Monget A.L., Richard M.J., Cournot M. (1996). Effect of 6 months supplementation with different combinations of an association of antioxidant nutrients on biochemical parameters and markers of the antioxidant defence system in the elderly. European Journal of Clinical Nutrition, 50: 443-449.
- Nagasaka R., Okamoto N., Ushio H. (2004). Partial oxidative-stress perturbs membrane permeability and fluidity of fish nucleated red blood cells. Comparative Biochemistry and Physiology, 139C: 259-266.
- Naeiji N., Shahsavani D., Baghshani H. (2013). Effect of dietary garlic supplementation on lipid peroxidation and protein oxidation biomarkers of tissues as well as some serum biochemical parameters in common carp *Cyprinus carpio*. Fisheries Science, 79: 699-705.
- Nazifi S., Ghafari N., Farshneshani F., Rahsepar M., Razavi S.M. (2010). Reference values of oxidative stress parameters in adult Iranian fat-tailed sheep. Pakistan Veterinary Journal, 30(1): 13-16
- NRC (1993). Nutrient Requirements of Fish. National Academy Press, Washington, DC. USA. 114 p.
- Prakash N., Banerji H.N. (1972). Evaluation of cyanmethaemoglobin method for haemoglobin estimation. Indian Journal of Chest Diseases, 14(2): 102-105.
- Puangkaew J., Kiron V., Satoh S., Watanabe T. (2005). Antioxidant defense of rainbow trout (*Oncorhynchus mykiss*) in relation to dietary n-3 highly unsaturated fatty acids and vitamin E contents. Comparative Biochemistry and Physiology, 140C: 187-196
- Roche H., Boge G. (1993). Effects of Cu, Zn and Cr salts on antioxidant enzyme activities in vitro of red blood cells of a marine fish *Dicentrarchus labrax*. Toxicology in Vitro, 7(5): 623-629.
- Sies H., Stahl W., Sevanian A. (2005). Nutritional, dietary and postprandial oxidative stress. Journal Nutrition, 135: 969-972.
- Treves-Brown K.M. (2000). Applied Fish Pharmacology. Kluwer Academic Publishers, Dordrecht, pp: 3-15.
- Welker T.L., Congleton J.L. (2009). Effect of dietary alpha-tocopherol + ascorbic acid, selenium, and iron on oxidative stress in sub-yearling Chinook salmon (*Oncorhynchus tshawytscha* Walbaum). Journal Animal Physiology Animal Nutrition, 93: 15:25

- Winston G.W., Di Giulio R.T. (1991). Prooxidant and antioxidant mechanisms in aquatic organisms. Aquatic Toxicology, 19: 137-161.
- Yonar M.E., Sakin F. (2011). Ameliorative effect of lycopene on antioxidant status in *Cyprinus carpio* during pyrethroid deltamethrin exposure. Pesticide Biochemistry and Physiology, 99: 226-231.
- Zhou X., Xie M., Niu C., Sun R. (2003). The effects of dietary vitamin C on growth, liver vitamin C and serum cortisol in stressed and unstressed juvenile soft-shelled turtles (*Pelodiscus sinensis*). Comparative Biochemistry and Physiology, 135A(2): 263-270.

چکیدہ فارسی

تاثیر مکمل غذایی ویتامین C بر برخی بیومارکرهای وضعیت اکسیداتیو در گلبولهای قرمز کپور معمولی (Cyprinus carpio)

حميده قدرتي آزادي'* ، داور شاهسوني'، حسن باغشني'

^{اک}روه علوم پایه، دانشکده دامپزشکی، دانشگاه فردوسی مشهد، مشهد، ایران. ^۳گروه بهداشت مواد غذایی و آبزیان، دانشکده دامپزشکی دانشگاه فردوسی مشهد، مشهد، ایران.

چکیدہ:

با توجه به مقادیر بالای اسیدهای چرب غیر اشباع در بافتهای ماهی، بهبود وضعیت آنتی اکسیدانی در ماهی لازم به نظر میرسد که میتواند اثرات مفیدی در سلامت ماهی داشته باشد. تحقیق حاضر بهمنظور بررسی اثرات مکمل غذایی ویتامین C(۲۰ میلی گرم/کیلو گرم وزن بدن، ۴ هفته) بر برخی فراسنجههای وضعیت اکسیداتیو در خون ماهی کپور معمولی (*Cyprinus carpio*) انجام شد. نتایج نشان داد که فعالیت آنزیم های آنتی اکسیدان از جمله کاتالاز (CAT)، گلوتاتیون پراکسیداز (GPX) و سوپراکسید دیسموتاز (SOD) انجام شد. نتایج نشان داد که فعالیت آنزیم های آنتی شاهد تفاوت معنی در ماهی کپور معمولی (GPX) و سوپراکسید دیسموتاز (MDA) در حضور مکمل ویتامین C در مقایسه با گروه شاهد تفاوت معنیداری نداشته است. از سوی دیگر، مکمل ویتامین C باعث کاهش معنیداری در غلظت مالون دی آلدهید (MDA) گلبولهای قرمز تا حدود ۲۶ در مقایسه با گروه شاهد تفاوت معنیداری نداشته است. از سوی دیگر، مکمل ویتامین C باعث کاهش معنیداری در غلظت مالون دی آلدهید (MDA) گلبولهای قرمز تا حدود ۲۶ در مقایسه با گروه قرمز تا حدود ۲۶ در مقایسه با گروه شاهد تفاوت معنیداری نداشته است. از سوی دیگر، مکمل ویتامین C باعث کاهش معنیداری در غلظت مالون دی آلدهید (MDA) گلبولهای قرمز تا حدود ۲۶ درصد در مقایسه با گروه شاهد گردید. بر اساس نتایج این طرح بهنظر میرسد مکمل غذایی ویتامین C قادر به محافظت از گلبولهای قرمز ماهی کپور در برابر استرس اکسیدانیو با کاهش پراکسیداسیون چربی میباشد.

www.SID.ir