DESERT

DESERT Online at http://jdesert.ut.ac.ir

DESERT 15 (2010) 119-125

Effects of NaCl and Na₂SO₄ on germination and initial growth phase of *Halostachys caspica*

M.H. Assareh^a, B. Rasouli^{b*}, B. Amiri^c

^a Professor, Institute of Forests & Rangelands, Iran ^bAssistant Professor, Islamic Azad University, Rasht Branch, Iran ^c Assistant Professor,, Islamic Azad University, Firouz Abad Branch, Iran

Received: 2 October 2009; Received in revised form: 19 February 2010; Accepted: 15 November 2010

Abstract

Current research on effect of increasing concentrations (0 (control), 100, 200, 300, 400 and 500 mM) of different salts including NaCl and Na₂SO₄ on germination and initial growth phase of *Halostachys caspica* were studied. The experimental design was completely randomized design with three replications. Characters of percentage, speed and index of germination, seed healthy index, radicle, plumule and plant length were measured. For analyzing of results were used of ANOVA, Duncan test and parried T- test sample in SPSS software. It was compared with germination under control condition, the most of characters of NaCl salt were not affected by 100 mM NaCl but were affected significantly different by 100 mM Na₂SO₄ and index of germination and seed healthy index the different aren't significantly. The results showed that germination percentage were severely inhibited by 500 mM NaCl but no affected severely by 500 mM Na₂SO₄. However, the results showed that *Halostachys caspica* in growth characters such as percentage, peed and index of germination is more sensitive to Na₂SO₄ salt. At least we can put *Halostachys caspica* chloridephyte and sulfatephyte group of halophytes.

Keywords: Halostachys caspica; NaCl; Na₂SO₄; Germination; Chloridephyte; Sulfatephyte

1. Introduction

World population is increased continuously and over 800 million hectares of land throughout the world are salt effected (FAO, 2005) then the halophytes plants are more worthy than before for us. Optimum and sustainable utilization of halophytes would play an important role to cope with the reduction of yield due to salinity by providing different raw materials for food, chemical industry and medicinal purposes in addition to elimination of soil and wind erosion (Flowers, 1999 & Larcher,

1995). Salt stress is probably the first

environmental factors that organisms face to it in evolution stages. Halophyte plants are founded of salty and arid regions of world. Biosalinity researches lead our studies to resistant plants with salinity and are impacted to use of salt waters such as sea, salinity lakes and drainage waters. Seed of halophytes under natural conditions are usually subjected to salt stress dominated by NaCl, however, other chloride, sulfate and carbonate salts, singly as well as can also affect seed germination and other factors significantly (Duan, 2004), (Khan, 2002) and (Beweley, 1994). Saline soils of Iran are formed from the accumulation of various chloride and sulfate salt dominated by NaCl and Na₂SO₄ salts (Jafari, 1993 & Szabolic, 1992). Important difficulties of salinity for plants are due to increasing of NaCl (Glenn, 1997) and Na₂SO₄ (Martin, 1993) salts. Halostachys caspica is halophyte (Chenopodiaceae) that in

^{*} Corresponding author. Tel.: +98 21 44196575, Fax: +98 21 44196575.

E-mail address: beh_rasooli@yahoo.com

order to resistant with high salt measure is absorbed and storied a lot of water and has succulent stems. Large canopy cover is effected to conservation and fixed of soil and salt special adage of lakes and wet land. The plant could be for improvement and sustainable used development in salty regions due to excellent germination and easily generation with correct management (Moghimi, 2004). Halostachys caspica is not used in initial growth and green stage as forage but could be used as forage for livestock after seeding special in autumn and winter season with preference value respectfully camel, goat and sheep (Zhao, 2001). Researchers reported that Halostachys caspica is one of the dominant species of salty community and region such as Urmia and Qom lakes. (Asri, 1997) and (Akhani & Ghorbanli 1993). (Strogonov, 1964) divided halophyte plants in two groups as chloridephyte and sulfatephyte and put Atriplex verruciferum, Atriplex canescens in chloridephyte group and Suaeda glaea, Haloxylon sp in sulfatephyte group. Of course, seed germination and seedling emergence are critical to the survival of plants and salt- affected area (Khan, 2002). Then Current research was studied on the resistant of salts stress (NaCl and Na₂SO₄) on germination of Halostachys caspica. (De-yu, 2007) showed that germination of Suada salsa inhibition was in the following order $Na_2SO_4 > NaCl$ but (Dashtkeyan, 2000) on Rubia tinctorum, showed that germination was in the following order NaCl>Na₂SO₄+NaCl>Na₂SO₄. (Shalka, 2006) on Urochondra setulosa and Indulkar on Sorghom showed that germination was in the following order NaCl>Na₂SO₄. Seed of halophytes respond to salinity stress the initial germination process is delayed under salt stress (Ungar & Keiffer, 1997). Reddy and Vora (1983) showed that seed germination of Bara delayed and radicle and plumule and decreased significantly with increased salinity of Na₂SO₄, NaCl and KCl. Jie (2005) reported that germination percentage of Halostachys caspica was not affected by 100 mM NaCl, while severely inhibited by 500 mM NaCl. (Assareh, 2003) on three species *Eculyptus camaldulensis*, *Eculvptus salubris* and *Eculvptus tetragona* and (Tajbakhsh, 2001) on Hordeum showed that percentage, speed, index of germination, seed healthy index, radicle, plumule and plant length decreased significantly with increased salinity of NaCl. Results of such as same researches could be used for improvement and sustainable development in salty regions in addition to elimination of soil and wind erosion and forage for livestock in arid and saline regions.

2. Materials and Methods

Seeds of Halostachys caspica were collected from Qom Lake. The seeds were surface sterilized with %70 alcohol for 15 second and washed with distilled water three times with germination experiment being started immediately. Then the seeds were surface sterilized with 1000 ppm for 15 minutes and washed with distilled water three times. Germination were carried out in Petri dishes (9 cm in diameter) on two layers of filter paper moistened with 5 ml distilled water (0 (control), 100, 200, 300, 400 and 500 mM NaCl and Na₂SO₄) solution and covered Petri dishes with Paraffin. Three replicates of 30 seeds were used for each treatment. Germination experiment was carried out 20°C temperature with an 8-h dark, 16-h light photoperiod. Germination was recorded every three day along 30 days. Characters of percentage speed and index of germination, seed healthy index, radicle, plumule and plant length were measured. Before statistical analysis in order to ensure homogeneity of variance was used Kolmogrovsmirnov test. Statistical analysis was carried out using SPSS 10. One way ANOVA was carried out to determine differences among treatment groups of characters. Treatment means were compared by Duncan test to determine whether differences among means were significant between treatments within each salinity concentration of NaCl and Na2SO4. Parried T-Test sample was carried out to determine differences among same concentration treatment of NaCl and Na₂SO₄. Characters were calculated with following formulas

Germination index (GI= $(\sum T_i N_i)/S$ where T_i is I days after started experiment, N_i is number of seed germinated along I day, S is total of seeds (30).

Germination speed (GS= $\sum n_i/D_i$ where n_i is the number of germination in special day and D_i is number days after started experiment).

Seed healthy index (SHI= (plant length (mM) *germination percentage) /100

3. Results

Kolmogrov-Smirnov test showed that all groups variance are homogene. Results of one way ANOVA and Duncan'test of NaCl salt are shown in Tables 1 and 2 and Na_2SO_4 salt in Tables 3 and 4. Parried T-test sample of same concentration treatment of NaCl and Na_2SO_4 carried out in Table 5.

Characters	Source of variables	Sum of squares	df	F
	Between groups	126.81	5	10.63**
Radicle length	Within groups	28.63	12	
	Total		17	
	Between groups	35.82	5	12.1**
Plumule length	Within groups	7.1	12	
	Total		17	
_	Between groups	223.32	5	10.37**
Plant length	Within groups	51.71	12	
	Total		17	
	Between groups	21344.41	5	24.53**
Germination percentage	Within groups	2088.8	12	
	Total		17	
	Between groups	349.89	5	20.76**
Seed healthy index	Within groups	40.45	12	
	Total		17	
	Between groups	213.51	5	24.54**
Germination speed	Within groups	20.88	12	
	Total		17	
	Between groups	19.21	5	24.52**
Germination index	Within groups	1.88	12	
_	Total		17	

**. Indicated significant difference at p=0.01

Table 2. Results of Duncan test of characteristic of *Halostachys caspica* due to NaCl salt concentrations (Different letters in the same column indicate significant difference at p=0.01)

(Difference at p=0.01)							
	Radicle	Plumule	Plant	Germination	Seed	Germination	Germination
	length	length	length	percentage	healthy	speed	index
0(control)	7.22a	4.66a	11.89a	95.56a	11.47a	9.56a	2.87a
100 mM	7.44a	4.89a	12.33a	100a	10.34a	10a	3a
200 mM	8.44a	3.43bc	11.89a	83.33ab	9.74a	8.33ab	2.5ab
300 mM	5.11bc	2.77c	7.89bc	65.56b	5.13b	6.56b	1.98b
400 mM	2.44cd	2.66c	5.11cd	23.33c	1.18c	2.33c	0.71c
500 mM	0.67d	0.67d	2.44d	11.11c	0.44c	1.11c	0.23c

One way ANOVA indicated significant (p=0.01) effect of various concentration (0,100, 200, 300, 400 and 500 mM NaCl) on all characters measured of Halostachys caspica (Table 1). Duncan test indicated except of plumule, compared with the control, all characters were not affected by 200 mM NaCl and the plant tolerate easily salt stress by 200 mM NaCl (Table 2). While Duncan test indicated except of plumule, compared with the control, all characters were severely inhibited by 400 and 500 mM NaCl (Table 2). Maximum seed and speed germination was obtained in 100 mM NaCl (Table 2). Rate of radicle, plumule and plant length were not decreased with increase of concentration in 300 by 400 mM NaCl but percentage, speed and index of germination, seed healthy index were decreased with increase of concentration in 300 by 400 mM NaCl (Table 2). Rate of percentage, speed and index of germination and plumule were not decreased with increase of concentration in 200 by 300 mM NaCl but, radicle, plant length and seed healthy index were decreased with increase of concentration in 200 by 300 mM NaCl (Table2). However results showed that Halostachys caspica in characters of percentage, speed and index of germination,

seed healthy index and plant length tolerate easily salt stress by 200 mM NaCl and increased rate of their by 100 mM NaCl (Table 2). Results indicated that radicle, plumule and plant length (growth characters) are more sensitive than germination percentage, speed and index of germination and seed healthy index (generation characters) to increase of NaCl salt.

One way ANOVA indicated significant (p=0.01) effect of various concentration (0,100, 200, 300, 400 and 500 mM Na₂SO₄) on all characters measured of Halostachys caspica (Table 3). Duncan test indicated compared with the control, characters of radicle, plumule, plant length and seed healthy index were severely inhibited by 100 mM Na₂SO₄ salt and showed that the plant is very sensitive to Na₂SO₄ salt but germination index and speed were not decreased with increased by 200 mM Na₂SO₄ salt and showed the plant rather tolerate by 200 mM Na_2SO_4 salt (Table 4). Results indicated that radicle, plumule and plant length (growth characters) and seed healthy index are more sensitive than germination percentage, speed and index (generation characters) with increased of Na2SO4 salt (Table 4).

Results showed that effect of each same concentration of NaCl and Na₂SO₄ salt (100,

200, 300 and 400 mM) on percentage, speed and index of germination and seed healthy index characters have similar responses and were not significantly different but these characters in concentration 500 mM of NaCl was significantly lower in comparison with 500 mM of Na₂SO₄. Rate of radicle and plant length in all each same concentrations of NaCl and Na_2SO_4 salt (100, 200, 300, 500 mM) except 400 mM and Plumule in all each same concentrations (100, 200, 300 mM) except 400 and 500 mM in Na_2SO_4 salt was significantly lower in comparison NaCl salt. However the plant in Plumule, radicle and plant length is more sensitive to Na_2SO_4 than NaCl salt.

Table 3. One way ANOVA of characteristic of Halostachys caspica due to Na2SO4 salt concentrations and their interaction

Characters	Source of variable	Sum of squares	df	F
	Between groups	59.04	5	12.83**
Radicle length	Within groups	11.05	12	
	Total		17	
	Between groups	20.44	5	12.51**
Plumule length	Within groups	3.92	12	
	Total		17	
	Between groups	137.31	5	12.24**
Plant length	Within groups	26.93	12	
	Total		17	
	Between groups	5316.87	5	7.58**
Germination percentage	Within groups	1688.53	12	
	Total		17	
	Between groups	164.56	5	11.22**
Seed healthy index	Within groups	35.02	12	
	Total		17	
	Between groups	53.24	5	7.58**
Germination speed	Within groups	16.85	12	
	Total		17	
	Between groups	4.78	5	7.55**
Germination index	Within groups	1.52	12	
	Total		17	

**. Indicated significant difference at p=0.01

Table 4. Results of Duncan test of characteristic of *Halostachys caspica* due to Na_2SO_4 salt concentrations (Different letters in the same column indicate significant difference at p=0.01)

in the sume et	Julii inalea	te significant e	interence at j	5 0.01)			
	Radicle	Plumule	Plant	Germination	Seed healthy	Germination	Germination
	length	length	length	percentage	index	speed	index
0(control)	7.22a	4.66a	11.89a	95.57a	11.47a	9.56a	2.87ab
100 mM	2.55b	1.67b	4.22b	98.89a	4.18b	9.89a	2.97a
200 mM	2.55b	1.89b	4.44b	98.89a	4.36b	9.89a	2.97a
300 mM	2.66b	1.89b	4.55b	75.55bc	3.47b	7.55bc	2.27bc
400 mM	2.11b	1.9b	4.77b	62.22c	2.99b	6.22c	1.87c
500 mM	2.11b	1.7b	4.44b	57.22c	2.59b	5.78c	1.73c

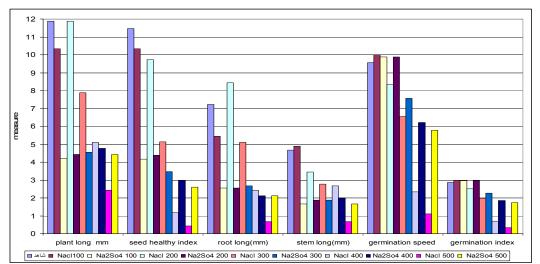


Fig.1. Mean final all characters measured (except germination percentage) of *Halostachys caspica* in different concentrations of NaCl and Na₂SO₄ salts

Sources	Compared of treatments	Different of variance treatment	t	df	significant
Radicle length	NaCl100- Na2SO4100	2.72	3.194	8	0.013*
	NaCl200- Na2SO4200	2.57	6.871	8	0.001**
	NaCl300- Na ₂ SO ₄ 300	1.94	3.773	8	0.005**
	NaCl400- Na ₂ SO ₄ 400	0.71	1.414	8	0.195ns
	NaC1500- Na2SO4500	1.13	3.833	8	0.005**
	NaCl100- Na2SO4100	1.48	6.526	8	0.001**
	NaCl200- Na2SO4200	1.67	2.8	8	0.023*
Plumule length	NaCl300- Na2SO4300	0.61	4.438	8	0.002**
	NaCl400- Na2SO4400	1	2	8	0.081ns
	NaC1500- Na2SO4500	1.32	2.268	8	0.053ns
	NaC1100- Na2SO4100	3.55	5.163	8	0.001**
	NaCl200- Na2SO4200	2.56	8.741	8	0.001**
Plant length	NaCl300- Na2SO4300	2.45	4.082	8	0.004**
•	NaCl400- Na2SO4400	2.78	.359	8	0.729ns
	NaC1500- Na2SO4500	2.61	2.309	8	0.05*
	NaCl100- Na ₂ SO ₄ 100	1.92	1	2	0.423ns
Germination	NaCl200- Na2SO4200	11.71	2.3	2	0.148ns
	NaCl300- Na2SO4300	23.34	0.742	2	0.535ns
percentage	NaCl400- Na2SO4400	21.43	3.143	2	.088ns
	NaC1500- Na2SO4500	8.81	9.168	2	0.012*
	NaC1100- Na2SO4100	2.48	4.292	2	0.05*
Seed healthy	NaCl200- Na2SO4200	1.01	9.123	2	0.012*
5	NaCl300- Na2SO4300	1.98	1.451	2	.0284ns
index	NaCl400- Na ₂ SO ₄ 400	0.96	3.271	2	0.082ns
	NaC1500- Na2SO4500	0.50	7.406	2	0.018*
	NaC1100- Na2SO4100	0.19	1	2	0.423ns
Comination	NaCl200- Na ₂ SO ₄ 200	1.17	2.296	2	0.149ns
Germination speed	NaCl300- Na ₂ SO ₄ 300	2.34	0.739	2	0.537ns
	NaCl400- Na ₂ SO ₄ 400	2.14	3.142	2	0.088ns
	NaC1500- Na ₂ SO ₄ 500	0.88	9.198	2	0.012*
	NaCl100- Na ₂ SO ₄ 100	5.77	1	2	0.423ns
Cominatio	NaCl200- Na ₂ SO ₄ 200	0.35	2.302	2	0.148ns
Germination	NaCl300- Na ₂ SO ₄ 300	0.7	0.742	2	0.525ns
index	NaCl400- Na ₂ SO ₄ 400	0.65	3.143	2	0.088ns
-	NaCl500- Na ₂ SO ₄ 500	0.26	9.165	2	0.012*

Table 5. Parried T-test sample compared of same concentration treatment of NaCl and Na₂SO₄

**. Indicated significant difference at p=0.01 *. Indicated significant difference at p=0.05

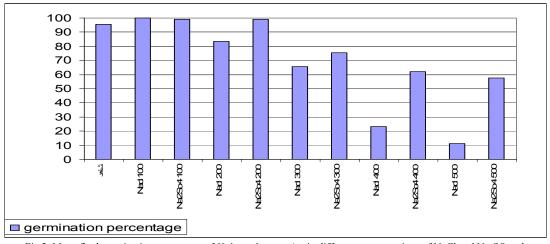


Fig.2. Mean final germination percentage of Halostachys caspica in different concentrations of NaCl and Na₂SO₄ salt

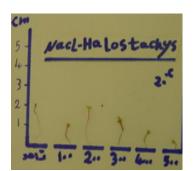
Rate of germination percentage, index and speed in all same of concentrations Na₂SO₄ salt was more than with comparison NaCl salt and radicle, plant length and seed healthy index in all same of concentrations (except 500 mM) Na₂SO₄ salt was lower with comparison NaCl salt (Fig1,2). Rate of germination percentage in concentrations 100 and 200 mM of NaCl and

Na₂SO₄ salts was more than with comparison controlled (Fig. 2). However rate of germination percentage in concentrations 400 and 500 mM of NaCl was severely inhibited but in concentrations 400 and 500 mM Na₂SO₄ salt was not severely affected and is almost 60% (Fig. 2).

4. Discussion and Conclusion

Results showed that Compared with control condition percentage, speed and index of germination were increased by 200 mM NaCl and Na₂SO₄ salts. Jie. Song (2005) on Halostachys caspica and Shariat (2000) on *Poterium sanguisorba* showed that were increased germination percentage, speed and index by 100 to 150 mM of NaCl salt too. But Assareh (2003) on three species Eculyptus camaldulensis, Eculyptus salubris and Eculyptus tetragona and Tajbakhsh (2001) on Hordeum, Poresmail(2000) on Nitraria schoberi, Suaeda fruticosa, Bagheri (1998) on Kochia prostrata, Eurotia ceratoides, Elymus junceus, Afzali (2000) on Melilotus officinalis, Trifolium fragiferum, Farkhah(2000) on Salsola dendroides pall, Alhaji persarum, Aleluropus lagopoides and Assadian (1987) on Medicago showed that compared with control condition germination percentage, speed, and index, seed healthy index, radicle, plumule and plant length decreased significantly with increased salinity of NaCl salt. The results showed that germination percentage and speed were severely inhibited by above 400 mM NaCl such as Jie. Song. (2005) on Halostachys caspica. E.B.kurkova, (2002) on Seidlitzia rosmarinus, Shalka, (2006) on Urochondra setulosa and Poresmail (2000) on Nitraria schoberi. Suaeda fruticosa showed that germination percentage and speed were severely inhibited by above 400 mM NaCl salt. The results showed that germination percentage and speed were not severely inhibited by above 400 mM Na₂SO₄ salt and is almost 60 % but Shalka, (2006) on Urochondra setulosa showed that germination percentage and speed were severely inhibited by above 400 mM Na2SO4 salt. However, the results showed that Halostachys caspica in germination characters such as germination percentage, speed and index is more sensitive to NaCl than Na₂SO₄ salt such as Dashtkeyan (2000) on Rubia tinctorum, Shalka, (2006) on Urochondra setulosa, Strogonov (1964) on

Haloxylon sp and Indulkar on Sorghom showed that germination of was in the following order NaCl>Na₂SO₄+NaCl> Na₂SO₄ but De-vu. (2007) showed that germination of Suada salsa inhibition was in the following order Na₂SO₄>NaCl. Halostachys caspica were not decreased significantly with increased salinity of NaCl in growth characters such as radicle. plumule and plant length by 300 mM but more concentration of 300 mM NaCl occurred a shock for plant and compared with control condition were decreased significantly by 500 mM while in the growth characters salt the shock occurred by and more concentration of mM 100 of Na₂SO₄ compared with control condition were decreased significantly. Rate of radicle was affected lower than plumule in each of NaCl and Na₂SO₄ salts but Jafari (1993) showed that plumule was affected lower than radicle. Ungar & Keiffer, (1997) showed that Seed of halophytes in the initial germination process is delayed and decreased under salt stress but this research showed that Halostachys caspica germination percentage and speed increased by 200 mM of NaCl and Na₂SO₄ salts. Halostachys caspica with comparison control was increased the growth characters in NaCl salt and increased the germination characters in Na₂SO₄ Then, we can put this plant in obligate halophyte groups such as Moghim (2004), Meiri (1984) and Zhao, (2005) reported that Halostachys caspica is obligate halophyte and has been adapted to saline and wet land regions. Results showed that Halostachys caspica has different interaction to NaCl and Na₂SO₄ salts; however Halostachys caspica in the germination characters has been adapted to Na₂SO₄ and in the growth characters adapted to NaCl salt. Then we can put *Halostachys caspica* chloridephyte and sulfatephyte group of halophytes. Halostachys caspica plant has excellent power germination about 100% (Moghimi, 2004) and could be used for improvement and sustainable development, elimination of soil and wind erosion and forage for livestock in arid and saline regions.



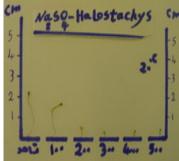


Fig.3. Picture of plant length in different concentrations of NaCl and Na₂SO₄

References

- Akhani, H. and Ghorbanli, M. (1993). In: A contribution to the halophytic vegetation and flora of Iran. Lieth, H. Al Massoom, A. (eds). Towards the rational use of high salinity tolerant plants. Vol. 1: 35-44. Kluwer Academic publishers, Netherlands.
- Asare. M. H., Shariat. A. 2003. Resistant of Eculyptus camaldulensis, Eculyptus *salubris* and *Eculyptus tetragona* on salt stress in Germination and initial Growth phase. , institute of forest & range researches of Iran publication house, vol 13, 4: 385-399
- Asri. Y. 1997.Plant cover of saline region of Uromia lake, institute of forest & range researches of Iran publication house. 191: 385-399
- Assadian, N. W. and Miyamoto, S. (1987). Salt effects on alfalfa seedling emergence. *Agron. J.* 76:710-714
- Bagheri. A. 1998. Study of Resistant Khochia prostrate, Eurotia ceratoidesT 'Elymus junceus to salt and drought.
- Bewley, J.D. Black, M. 1994: seeds: physiology of development germination - Plenum prees. London.
- Dashtkeyan. K. 2000. The effect of physical and chemical properties of soil in *Rubia tinctorum L* composition.
- FAO. 2005. Global Network on Integrated Soil Management for Sustainable Use of Salt-affected Solis. Rome, Italy: FAO Land Plant Nutrition Management Service.
- Farkhah. A. 2001. Study of Resistant physiological properties in *Alhaji persarum, Aleuropus lagopoieles, Salsola dendroides.*
- Flowers, T. J., Flowers, S. A. And Greenway, H. (1985). Effects of sodium chloride on tobacco plant. *Plant Soil Environment*.9:645-651.
- Flowers, T. J. (1999). Salinisation and horticultural production. Scient. Hort. 77:1-4.
- Jafari. M. 1993.Investigation tolerate of some of Iran range land grasses plant to salt stress, institute of forest & range researches of Iran publication house. Num: 67
- Glenn, E. P. Brown, J. and Jamal-Khan, M. (1997). Mechanisms of salt tolerance in higer plants. The university of Arizona, PP:83-110
- Indulker, B. S. and More, S. D. (1984). Response if sorghum to phosphorus application in presence of chloride and sulphate salinity. *Current. Agric.* 8(1-2):81-85
- Ismail, S. Malcom, C. V. and Ahmad, R. (1990). A bibliography of forage halophytes and trees for saltaffected land: Their uses, culture and physiology. Department of Botany, University of Karachi, Pakistan.
- Jie. Song, Gu. Feng, Fusuo Zhang, (2006). Salinity and temperature effects on germination for three saltresistant euhalophytes, *Halostachys caspica*, *Kalidium foliatum* and *Halocnemum strobilaceum*. Plant and Soil, 279:201-207

Keiffer, C. H., and Ungar, I, A. (1997). The effect of

extended exposure to hyper saline condition on the germination of five inland halophyte species. American Journal of Botany. 84(1):104-111

- Khan, M.A., and Rizvi, Y. (1994). Effect of salinity, temperature and growth regulation in water early seedling growth of *Atriplex giriffithii*. Can. J. Bot. 72:475-479
- Khan, M. S. A., Hamid, A. and Karim, M. A. (1997). Effect on sodium chloride on germination and seeding characters of different types of rice (Oriza sativa L.). Crop S ci. 176:163-169
- Kurkova, E. B. LL. G. Kalinkina, O. K. Baburina, N. A. Myasoedov, and T. G. Naumova. (2002). Responses of *Seidlitzia rosmarinus* to Salt Stress, Biology Bulletin, vol. 29, No. 3, 221-228
- Larcher, W. (1995), Physiological plant ecology (3 rd). PP: 390. Springer Publishing.
- Makki, Y. M., Tahir, O. A. and Asif, M.I. (1987). Effect of drainage water on seed germination and early seeding growth of five group species. Biological wastes. 2: 133-137
- Martin, J. P., ElavumMoottil, O. C. and Moreno, M. L. (1993). Changes on protein expression associated with salinity tolerance in *Brassica* cell culture. Cell Biol. Intern. 17:839-845
- Meiri, A. (1984). Plant response to salinity: experimental methodology and application to fiels. In: salinity under Irrigation. I. Shainberg and J. Shalhavet (Ed) springer Verlag. New York. PP. 284-297
- Miller, T.R and Chapman, S.R, 1978. Germination response of three forage grasses to different concentrate ion of six salts. Journal of Range Management. 31(2):123-124
- Moghimi. J. Introduce of important rangeland plant of Iran. 2004
- Poresmail. M. 2001. Study of Resistant (Chenopodiaceae) *Suaeda fruticosa* (*L*.) Forssk) Zygophyllaceae) *Nitraria schoberi* L. on salt stress in Germination and initial Growth phase.
- Reddy, M. P. and Vora, A. B. (1983). Effect of salinity on germination and free proline content of Bara seedlinhgs. Proceedings of the Indian National Science Academy, B. 49(6): 702-705
- Strogonov, B. P. (1964). Physiological basis of salt tolerance of plants. Acad. Sci. USSR. Davey and Co. New York.
- Szabolic, I. (1992). Salinization of soil and water and its relation to desertification. Desertification Contr. Bull. 21:32-37
- Tajbakhsh. M. 2001. The effect of salt stress (NaCl) on *Hordeum sp.* 6th Conference of botany, Babolsar. Iran.
- Zhao, K, F. and Feng, L. 2001. Recourse of Halophytes in China. China Science Press, Beijing (in Chinese).
- Zhao, K, F. Fan, H, Song, J, Sun, M, X, Wang B, Z, Zhang, S, Q and Ungar, I, A. 2005, Two Na⁺ and Cl⁻ hyperaccumulators of the Chenopodiaceae. J.Integrative Plant Biol. 47, 311-318