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Silicone Polymers in Controlled Drug Delivery Systems: A Review

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ABSTRACT

n this paper some of the latest studies and research works conducted on silicone-based drug delivery systems (DDS) are reviewed and some of more specific and important novel drug delivery devices are discussed in detail. An overview on rapidly growing developments on silicone-based drug delivery systems is provided by presenting the necessary fundamental knowledge on silicone polymers and a literature survey including an introductory account on some of the drugs that are diffused through silicone polymers. The results based on vast investigations over a period of a decade indicate that intravaginal and transdermal routes of administration of the drugs using silicone-based DDS are more developed. It is also found that silicone polymers are suitable candidates for the release of hormonal steroids for controlling the estrous cycle. Finally, some commercially available silicone-based DDS are described.

CONTENTS

Key Words:

silicone; veterinary devices; drug delivery systems; intravaginal devices; transdermal devices.

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INTRODUCTION

Controlled drug delivery field is one of the most attractive and challenging areas in medical sciences, chemistry, materials science, engineering, pharmaceutics, and other related biological sciences. Its application also covers diverse fields including medicine, agriculture, and biotechnology [1,2]. Recent growing interest and efforts of scientists in this area are due to DDS vitality in achievement of a better quality of life and health care for human beings. In addition to research on new drugs, a significant amount of pharmaceutical research works has been focused on designing novel methods of making drug dosages more effective. Many definitions of controlled drug delivery system are found in the literature, but all refer to a system consisting of two essential components of a drug and a drug carrier with the following purposes: the maintenance of drug levels within a desired therapeutic range, delivery of the drug locally to specific sites of a particular body compartment, to avoid possible toxicity, to minimize side effects, increasing patient compliance and comfort by decreasing the dosing frequency, and reducing manufacturing costs [3-5].

A variety of biomaterials such as polymeric materials have been designed and developed that can be used as a drug carrier. To use these materials in controlled drug delivery devices, they should meet the following conditions:

- Chemical inertness,
- Biocompatibility and minimum undesirable degradation byproducts,
 - Non-leachability,
 - Ease of fabrication and sterilization,
- To possess suitable mechanical properties [5,6]. Some of the more important polymers which have been recently developed and used for controlled drug delivery systems are as follows:

Poly(urethanes), poly(vinyl alcohol), poly(acrylic acid), polyamide, poly(methacrylic acid), poly(n-vinyl pyrrolidone), polylactides, poly(lactide-co-glycolides), polyanhydrides, polyorthoesters, and silioxanes [6].

For a drug to be successfully used in a controlled drug delivery system, the administration route plays a vital role. The choice of a delivery route depends on some factors including:

- Patient acceptability,
- Properties of the drug (such as its solubility),
- Access ability to the treatment site,
- Effectiveness in dealing with a specific disease [7]. At present, the most accepted delivery routes as alternative to the traditional intravenous route are the nasal, transdermal, pulmonary buccal, ocular, vaginal, rectal, and oral. These administrations have been the focus of much academic and industrial research works [8].

The release of the drug occurs from a delivery system on the basis of diffusion, degradation, or swelling followed by diffusion. In a given DDS, the release may occur through any or all of these mechanisms [2].

When a drug is released through a polymer, diffusion is usually observed. In degradation mechanism, release of the drug within the body occurs through biological processes which eliminate or remove the carrier system. In swelling control mechanism, water penetration into polymer causes swelling which controls drug release [1,9]. Usually, the release of a drug through silicone rubber as a carrier system occurs based on diffusion mechanism [5].

SILICONE RUBBER POLYMERS

Polysiloxanes or silicones are important class of organo-silicon synthetic materials based on molecular chains of alternate silicon and oxygen atoms. Silicones as high technology materials have been used in many novel applications due to their unique combination of high temperature stability and low temperature elastomeric properties [10-13]. These compounds are produced in a wide range of properties all the way from water thin substances through heavy oil-like fluids to greases, rubbers, and solid resins by changing polymer chain length and the organic groups attached to silicon atoms. Silicone rubber is usually produced by hydrolysis of appropriate silanes and cross-linking the components produced via one of the methods described below [11]:

Polydimethyl Siloxane Cross-linking Methods

Polydimethyl siloxane (PDMS) is a commercially

ROOR
$$\longrightarrow$$
 2RO.

$$\begin{array}{c}
CH_3 \\
-Si - O \\
CH_3
\end{array}
+ RO.

+ ROH$$

$$\begin{array}{c}
CH_2 \\
-CH_3
\end{array}
+ ROH$$

$$\begin{array}{c}
CH_3 \\
-Si - O \\
-CH_2
\end{array}
+ ROH$$

$$\begin{array}{c}
CH_3 \\
-Si - O \\
-CH_2
\end{array}
+ CH_2$$

$$\begin{array}{c}
CH_2 \\
-CH_2
\end{array}
+ CH_2$$

$$\begin{array}{c}
CH_2 \\
-CH_2
\end{array}
+ CH_3$$

$$\begin{array}{c}
CH_2 \\
-CH_2
\end{array}
+ CH_3$$

Scheme I. Peroxide curing mechanism for PDMS [17].

available polymer with rapidly increasing applications in DDS which has two methyl groups attached to each silicon atom. Cross-linking of PDMS is enhanced in polysiloxanes structure by a wide range of methods, including: (i) use of tri- or tetra-functional siloxane comonomers during polymerization, (ii) incorporation of a thermal initiator which would abstract hydrogen atoms from PDMS and result in cross-linking or, (iii) the exposure of PDMS to high energy irradiation [14].

As some of the drugs are sensitive to heat and moisture, they may impose limitations in DDS applications on the choice of curing methods for cross-linking process of silicone rubber. Recently, a curing method based on the use of platinum as catalyst has attracted considerable attention, due to its cost-effectiveness and safety [15]. This method is FDA approved, because of the absence of organic compounds or volatile by-products in the device [16].

Peroxide

Peroxide curing is used to produce high temperature vulcanized (HTV) silicone rubber. This process is

carried out on the basis of free radical mechanism, and a given amount of heat is necessary to generate organic peroxides free radicals for initiating the curing reaction (Scheme I) [17].

Addition

In room temperature vulcanization (RTV) method, cross-linked silicone rubber is produced via Si-H group attachment to double bonds. Platinum, palladium, or rhodium salts or their complexes can be used as the catalyst. By using platinum-olefin complexes as the catalyst, curing process takes place at room temperature (Scheme II) [17].

Condensation

There are two systems for condensation curing: oneand two-component systems. For one-component system the catalyst and cross-linking agent are incorporated in the base compound at the time of manufacturing; whereas in two-component system the cross-linker and catalyst must be added to the base compound consecutively just before use. The catalysts used for the above reactions are of organo-tin comounds.

Scheme II. Addition curing mechanism of silicone rubber [17].

Scheme III. Condensation curing mechanism of silicone rubber [17].

Condensation curing takes place at room temperature based on the mechanisms depicted in Scheme III. The silicone rubbers produced by this method are suitable for sealing, bonding, and coating applications due to their unique property [17].

Irradiation

This curing method is suitable for radiation-induced cross-linking of elastomers. There are numerous published works describing the material properties of the irradiated silicone rubber. Nearly 50 years ago, silicone rubber was cross-linked by using high energy electron beam or gamma rays. The mechanical properties of irradiated silicone rubber have been investigated by Bopp et al. [18], Warrick [19], Przybyla [20], Vokal [21], Basfer [22] and more recently by Frounchi [23]. In addition, many scientists have studied the radiation chemistry of silicone rubber [14,

24-26], and a mechanism has been proposed for cross-linking reaction which is based on the scission of C-H and C-Si bonds in the side chains (Scheme IV) [14]. Somehow, the curing methods by irradiation are not as popular as other methods due to lack of knowledge on radiation/drug interactions.

SILICONE-BASED DRUG DELIVERY SYSTEMS

Silicones or in more scientific term "siloxanes", have been used in a wide range of biomedical applications during the past four decades, due to their physiological inertness, high blood compatibility, low toxicity, good thermal and oxidative stability, and low modulus [27,28]. In addition to drug delivery devices, medical applications based on silioxanes are as

Scheme IV. Irradiation curing mechanism of silicone rubber [14].

follows: blood pumps, cardiac pacemaker leads, mammary prostheses, drainage implants in glaucoma, artificial skin, maxillofacial reconstruction, replacement oesophagus, contact lenses, oxygenators, medical adhesives, finger joints, coating for cochlear implants, and catheters [29].

Due to unique structural features of silicone, this polymer plays an important role as matrix, membrane, and coating for DDS. Using siloxanes in controlled release delivery systems goes back to 1960s. The first work on the permeability of silicone to ordinary gases was published in 1957 by Kammermeyer [30].

In addition, it is found that oil soluble dye in powder form can diffuse through the silicone rubber tubing walls over a period of a few months. Further works in this field have led to the general conclusions that low molecular weight and lipophilic drugs easily diffuse through silicone rubber [31-35], while substantial efforts have been devoted to the release of high molecular weight and polar species via diffusion through silicone rubber [36,37].

The main structural feature of silicone system is the large atomic volume of the silicon atom itself as well as the size and position of the constituent groups which lead to complete freedom of rotation around the Si-O-Si bonds. Silicone polymers form helices, and the silicon-oxygen bond angles create large amounts of free volume in silicone elastomers. This free volume and the high compressibility of silicone compounds are responsible for their permeability to certain gases and liquids. The gas permeability of silicone rubber is around 100 times greater than that of the natural or butyl rubbers [38]. Also, silicone is cheap and easy to fabricate into many shapes by moulding and also it is curable without excessive heating. It is also chemically inert and non-toxic. This polymer as a drug carrier matrix does not degrade the drug due to its chemical inertness. The application of silicone based controlled release device also improves the patient's comfort for its simple administration and better compliance. In addition, pharmaceutical formulator parametric functions are based on silicone type and device size according to anatomical insertion inside the body to make optimum use of the polymer [39].

Silicone rubber performs suitably in releasing steroid molecules due to their relatively high solubility in the hydrophobic silicone and their relatively low molecular weight/volume which allow a relatively rapid molecular diffusion. In particular, it is found that drug release depends primarily on the silicone elastomer diffusivity and solubility of the drug molecule, as it is shown by cumulative releases (eqns (1) and (2)) for matrix and reservoir-type devices [5,40]:

$$Q = \sqrt{D_p C_p (2A - C_p)t} \tag{1}$$

$$Q = \frac{D_p C_p}{h} t \tag{2}$$

where,

Q: cumulative amount of drug released per unit area (mg/cm²),

t: time (days),

A: initial amount of drug loading per unit volume in a matrix system (mg/cm³),

D_p: diffusion coefficient of the drug in the polymer (cm²/day),

C_p: solubility of the drug in the polymer (mg/cm³), and

h: thickness of the sheet layer in the reservoir-type system (cm).

Extensive work has been conducted on the measurements of D_p and C_p , and many researchers have studied the drug release behaviour using silicone matrices which lead to different products (Table 1).

Modification of Silicone Rubber in Drug Delivery Systems

Though physico-chemical properties of silicone rubber makes it a good candidate for medical applications, but due to its hydrophobic properties, it is not suitable for some medical applications, so it should be modified to acquire some hydrophilic characteristics. PDMS rubber is used as a soft tissue substitute, because some problems occur when the silicone devices are kept implanted for a long time. For example, percutaneous devices are not immuned towards infection because there is a dead-space between the tissue and the implanted device. Such dead-space would not exist if the material surface could bond to the skin tissue [6,41,42].

For some biomedical applications surface and bulk modifications are necessary via suitable method to

Table 1. Silicone as drug carrier in DDS.

Drug	Drug function	System type	Ref. No.
Progesterone	Control of the estrous	Matrix-type	[76-80]
-	cycle(Antidepressants/Anxiolytics)		
Norgestomet	Control of the estrous cycle	Matrix-type	[81]
Desoxycorticosterone acetate (DCA)	Mineralocorticoid Hormone	Matrix-type	[82]
Indomethacin	Non-steroidal anti-inflammatory	Matrix-type	[83]
Brilliant blue	Model drug	Composite	[84]
Bovine serum albumin (BSA), Antipyrine	Model drugs	Matrix-type	[85]
(ANP), Indometacin (IDM), and			
Ketoprofen (KP)			
Hydrocortisone, Dapsone, and Estradiol	Anti-inflammatory, acne treatment, hormone	Tuneable silicones	[86]
Timolol maleate	Anti high blood pressure	Reservoir device	[87]
Acular	Nonsteroidal anti-inflammatory	Contact lens	[88]
Ketotifen	Antihistamine	Pressure sensitive adhesive	[89]
		(PSA) matrices	
Nonoxynol-9-α-cyclodextrin	Spermicidal	Matrix-type	[90]
Genencor enzymes	Model drugs	Pressure sensitive adhesive	[91]
Vancomycin	Antibiotic	Amino-polysiloxane matrixes	[92]
Indomethacin and Dexamethasone	Non-steroidal and steroidal	Rods made of silicone	[93]
	anti-inflammatory	rubber-polylactide	
Ibuprofen (IBU), Methyl paraben (MP)	Anti-inflammatory and antifungal	Colloidal microgels(butyl	[94]
and Propyl paraben (PP),		acrylate (10%) co-polyNIPAM	
		(90%))	
Ethynodiol diacetate	Hormonal contraceptives	Matrix-type	[95]
1,3-Bis(2-chloroethyl)-1-nitrosourea	Anticancer	Membrane	[96]
(BCNU)			
Gentamicin	Aminoglycoside antibiotic	Rod implant	[97,98]
Phenol	Model drug	Interpenetrating polymer network (IPN)	[99]
Testosterone	Hormone	Membranes	[100]
Nitroglycerine and scopolamine	Prevent angina and motion sickness	Transdermal	[101]
Levonorgestrel	Hormone	Matrix-type	[102]
Rifampicin	Antibiotic	Matrix-type	[103]
Salicylic acid and propranolol	Model drugs	Matrix-type	[104]
hydrochloride			
2-Pyridine aldoxime chloride (PAM-CI)	Antidote	Matrix-type	[105-106
Clindamycin, 17β -estradiol, 17β -estradiol-	Model drugs	Intravaginal ring devices of	[107-108
3-acetate, 17β-estradiol diacetate,		matrix-type & reservoir	
metronidazole, norethisterone, norethis-			
terone acetate, and oxybutynin			
Oxybutynin	Anticholinergics	Intravaginal ring (IVR) of	[109]
		reservoir design	

Continued

Drug	Drug function	System type	Ref. No.
Polyethylene glycols and Norethisterone	Different drug	Matrix (ring intravaginal	[110]
acetate, Estradiol, Triclosan, Oleyl alcohol,	_	drug delivery device)	
and Oxybutynin			
Progestins	Hormone	Modelling	[111]
Timolol, Dexamethasone, and	Model drugs	Composite	[112]
Dexamethasone 21-acetate			
Nonoxynol-9 (N-9)	Spermicide	Matrix-type	[113]
Metronidazole, Acyclovir, Clotrimazole,	Antimicrobial agent	Matrix (ring intravaginal	[114]
Fluconazole, Terconazole, Azithromycin,		drug delivery device)	
Erythromycin, Doxycycline, Tetracycline,			
Minocycline, Clindamycin, Famcyclovir,			
Valacyclovir, and Clarithromycin			
Ethynyl estradiol, Estradiol	Hormone	Matrix-type	[115]
Estradiol, Progesteron, Vitamin B6,	Model drugs	Silicone ring matrix	[116]
Vitamin D, and Vitamin E			
Fentanyl	Opioid analgesics	Reservoir- type	[117]
Metronidazole, Miconazole, Ciprofloxacin,	Microbicidal intravaginal	Silicone ring matrix	[118]
and others			

make silicone rubber hydrophilic in character. In controlled drug delivery systems bulk modification is preferred for drug transport throughout the polymer. Blending, copolymerization, formation of interpenetrating polymer network (IPN), and functionalization process are employed in bulk modification methods [43].

In IPN technique two interpenetrating three-dimensional networks are formed which are not covalently bonded [43]. An interpenetrating polymer network (IPN) is defined as an intimate combination of two or more polymers, both in network form; at least one of which is synthesized and/or cross-linked in the immediate presence of the other. There is a very promising technology based on hydrogel/PDMS IPN for transdermal drug delivery devices [44,45].

Another technique to make silicone rubber hydrophilic is to disperse hydrophilic polymer-hydrogel powder in the silicone rubber matrix. Many reports have been published that deal with the preparation and characterization of composites of PDMS having fine particles of hydrogels as dispersed phase. Silicone rubber and hydrogels based composite materials are two-phase polymeric systems. The composites show good mechanical properties of its compart-

ments: ease of fabrication of silicone rubber and hydrophilicity and water-permeable of hydrogels [46-54].

Alternative Silicone Applications

PDMS Coatings

PDMS coatings are alternative silicone polymer applications in controlled drug release. One of the most important goals of pharmaceutical coatings is to control drug release by the use of polymeric materials. Surface coating provides an effective way to control the drug release. A drug-containing core or tablet is surrounded by a coated film, and the release rate of the drug is controlled by its diffusion through the film [55-58].

The use of PDMS as pharmaceutical tablet coating has shown potential for possible zero-order release (i.e., the highly desirable delivery of a constant drug dosage per unit of time). Drug release in these formulations is mainly dependent on the dispersion of domains of water-soluble substances such as low molecular weight polyethylene glycols within the PDMS films. Their dissolution provides channels for the release of drug molecules, thus a relatively large amount of water-soluble substances is necessary to

assure connectivity of the generated paths [59-68].

PDMS in Drug-eluting Stents

Silicone rubber is also used as drug bearing coating in drug eluting stents [69]. Usually several components are combined to make up the drug-coated stent platform. In drug-coated metallic stent with a polymer, the polymer functions as a drug carrier and a mixture of drug and polymer is applied as a coating on the abluminal surface of the metal stent. In drug-eluting stents, it is necessary to place a top-coat over the base coat. This top-coat is made up of another layer of polymer and acts as a diffusion barrier. Using this barrier a concentration gradient is created from stent to artery for controlled drug delivery [70]. It should be mentioned that the first organic material used for stenting was silicone rubber. However, silicone has low tensile and coil strengths and small inner to outer diameter ratio [71]. There are some reports on silicone or its copolymers with polyurethane drugeluting stent for drug release (e.g., dexamethasone) [72-75].

DEVELOPMENTS OF SILICONE-BASED DRUG DELIVERY SYSTEMS

Silicone-based drug delivery devices offer a new area with broad opportunities for scientists who are looking for novel methods of administrating known drugs. In spite of substantial works and efforts that have been devoted to this rapidly growing field of research, most of these systems still remain as a laboratory curiosity. Some examples of studied silicone-based DDS are presented in Table 1.

COMMERTIALLY AVAILABLE SILICONE-BASED DRUG DELIVERY DEVICES

A number of drug delivery devices have been developed for administration of drugs for human and livestock in different ways. It is interesting to note that most of the therapeutic systems have been derived from fundamental studies in laboratory. Some routes of the drug administrations are presented in the following sections.

Intravaginal Drug Delivery Devices

In designing intravaginal drug delivery systems, anatomical and physiological considerations should be taken into account. The human trials have been focused primarily on the systemic delivery of contraceptives. Currently, the intravaginal delivery of therapeutic peptides and proteins has gained considerable attention. The main advantage of intravaginal drug delivery is that the hepatic first-pass metabolism is avoided by this route. General intravaginally platforms include creams, foams, pessaries, gels, tablets, and particulate systems. Typical examples of specifically designed intravaginal delivery systems are those employing solid polymeric systems, such as elastomers and hydrogels [8].

Vaginal rings, also known as intravaginal rings, or V-rings, are polymeric drug delivery devices designed in the shape of a doughnut to provide controlled release of drugs to the vagina over long periods of time. The main advantages of the vaginal rings are cost-effectiveness, patient compliance, and therapeutic efficacy due to the ability and versatility of these systems in providing long-term and continuous release of drug at constant predetermined rates. In spite of subdermally implantable systems which need surgical operation for placement, these rings can be placed in the vagina by woman herself [8,119]. Several types of human vaginal ring products are currently available: Estring and Femring [120-122].

Estradiol vaginal ring such as Estring is a slightly opaque ring with a whitish core containing a drug reservoir with 2 mg estradiol. A combination of estradiol, silicone, and barium sulphate forms the ring. When Estring is placed in the vagina, estradiol is released approximately 7.5 mcg per 24 h in a consistent stable manner over 90 days. The dimensions of Estring are: outer diameter 55 mm, cross-sectional diameter 9 mm, and core diameter 2 mm [121,123].

Estradiol acetate vaginal ring, Femring, is an off-white, soft, flexible ring with a central core containing estradiol acetate. Femring is made of cured silicone elastomer containing dimethyl polysiloxane, silanol, silica (diatomaceous earth), normal propyl orthosilicate, stannous octoate, barium sulphate, and estradiol acetate. The ring dimensions are: outer diameter 56 mm, cross-sectional diameter 7.6 mm, and core diameter 2 mm [122,124].

In the veterinary field, the major market for the application of drug delivery devices includes the fertility control via delivery of hormones. There are some older families of silicone based veterinary DDS. Progesterone releasing intravaginal device (PRID) is the first progesterone-releasing device of this kind for use in dairy cows. The PRID comprises a strip of stainless steel covered on both sides with a matrix of silicone and progesterone (1.55 g) in the form of a coil with a diameter of 4 cm and length of 12 cm (Figure 1a). It is manufactured by injection moulding a progesterone/silicone mixture around a stainless steel strip set in a dye such that both sides of the strips are coated by a matrix with defined and uniform thickness [125,126].

Controlled internal drug release (CIDR) is another device for release of progesterone from silicone matrix. CIDR devices are developed in different sizes; the so called CIDR-B and -G, for cattle and goats, respectively. CIDR-B is a T-shaped device with a wing tip-to-tip distance of 15 cm and body length of 13 cm comprising a silicone/progesterone (1.9 g progesterone) matrix that is injection moulded over a nylon spine (Figure 1b). Using this device, progesterone is released into the cow's bloodstream, and the ability to synchronize anestrous cows is substantially increased. The CIDR-G contains 0.33 g progesterone in silicone matrix. The T-shaped device is similar to CIDR-B with a 5.7 cm wing tip-to-tip width (Figure 1c) [127,128].

CueMate is (Figure 1d) a veterinary intravaginal

drug delivery system. In this device progesterone (1.56 g) is homogenously distributed throughout silicone that is manufactured by injection moulding and then attached onto a wishbone shaped spine to facilitate drug administration, retention in the vaginal and removal after implantation [129].

Subdermal Drug Delivery Devices

A drug delivery method involves implanting drug delivery system beneath the skin, subdermal or subcutaneous, and targeting specific tissues. In case of subdermal implants, flexible capsules are placed under the skin, and various hormones are administered in this way [130].

Norplant is a human contraceptive device that consists of six capsules of crystalline levonorgestrel encapsulated in silicone rubber (Figure 2). The rod-shaped devices are implanted in a fan-shaped array just below the dermis, usually in the upper arm over the triceps or biceps. Norplant is approved for use for 5 years. Levonorgestrel levels reach contraceptive strength within 24 h of insertion and decline slowly thereafter [131].

Compudose and Crestar are two commercially available animal ear implants. Compudose is implanted in the ear using the appropriate applicator. Compudose is a controlled release device made by coating non-medicated silicone rubber core with a thin layer of silicone rubber that contains oestradiol-17 β . It has a surface area of 4.84 cm² and has been shown to release drug by a square-root-of-time

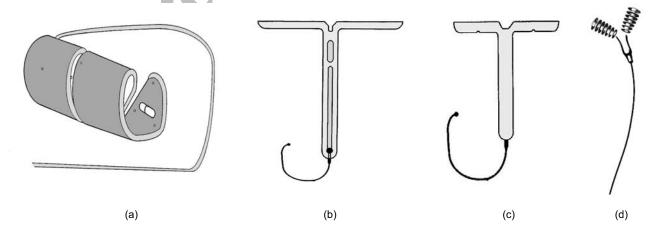


Figure 1. Commercially available controlled release intravaginal drug delivery systems in veterinary field: (a) PRID device, (b) CIDR -B, (c) CIDR-G, and (d) CueMate device.



Figure 2. Norplant.

mechanism. This is designed in four products range: Compudose 400, 200, 100, and -G to provide controlled release of drug over a period of 400, 200, 100, and 90-100 days, respectively [132,133].

Crestar is an ear implant used for the control of estrous cycle in cattles. It is a small rod-shaped device containing 3 mg norgestomet dispersed throughout a silicone matrix. It is marketed with a 2 mL injection of 3 mg norgestomet and 5 mg estradiol valerate [134,135].

Transdermal Drug Delivery Devices

The human skin is a readily accessible surface for drug delivery. Transdermal drug delivery systems (TDDS) are able to systemically deliver medicinal drugs through the skin. Skin is a protective membrane for body and consists of several complex layers which stratum corneum is the most impermeable of these layers. Skin penetration is very difficult and highly dependent on the particularity of drugs. Therefore, a few numbers of the drugs can be effectively delivered transdermally. Several penetration enhancers, such as oleic acid, oleic acid esters, and poly(ethylene glycol) have been added to transdermal formulation as an aid to pass skin [125,136].

Generally, the drug is contained in a transdermal patch which is applied to the skin similarly to a band aid. A patch system consists of four layers including: backing layer, membrane, adhesive layer, and liner [137]. Two major types of transdermal drug delivery system products are:

- Reservoir: the active ingredient is in a solution or suspension which is held between the backing layer and a rate-controlling membrane (Figure 3a).

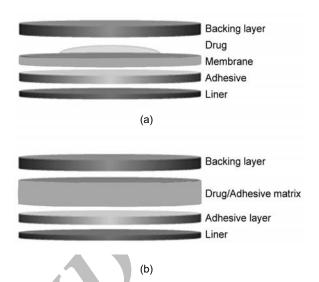


Figure 3. Schematic of thransdermal drug delivery systems: (a) reservoir and (b) matrix-type.

- Matrix: the drug as a suspension is dispersed in polymer and may be in contact with skin by using an adhesive applied to the perimeter of the system to be held to the skin (Figure 3b). Another refinement is drug-in-adhesive matrix in which the polymer is an adhesive containing the drug as dispersion [137].

For avialable drugs, transdermals may be considered as novel delivery systems. Only drugs which are given in relatively small daily doses can be administered through patches. Examples of such drugs include nitroglycerin (for chest pain), scopolamine (for motion sickness), nicotine (for smoking cessation), clonidine (for high blood pressure), fentanyl (for pain relief), and estradiol [139].

Pressure sensitive adhesives (PSAs) are important components of TDDS, because they ensure intimate contact between the drug releasing area of a TDDS and the skin surface which is critical for controlled-drug delivery [137]. Usually, the bond formed between PSA and skin can be broken with a measurable force which leaves the substrate relatively free from damage [142]. The major classes of medical PSAs are acrylics, silicones, and polyisobutylenes. Transdermal patches have entered the pharmaceutical market since the early 1980s. The first transdermal patch known as Transderm Scop, marketed by CIBA Co., releases, scopolamine, the anti-motion sickness drug [138,139].

BIO-PSA 355 silicone pressure-sensitive adhesive

is well-suited as a contact adhesive in reservoir-type delivery systems. Its properties are somewhat compromised, however, when co-formulated with aminofunctional agents. BIO-PSA Q7-2920 was developed to perform as amine resistance. PSA either functions as contact adhesive or may potentially act as drugloaded adhesive matrix is conceptually simple, but technologically complex drug delivery system [140,141]. There are several transdermal patches that silicone adhesive is incorporated in their formulation: Vivelle-Dot (for estradiol release) by Noven Pharmaceuticals/Novartis Co. [142], Matrifen® transdermal pain patch (for fentanyl release) by Nycomed Co. [143], Duragesic (for fentanyl release) by Alza/Janssen Pharmaceutica Co. [144].

SUMMARY

Some of the latest research works and studies on silicone-based drug delivery systems and biomedical applications such as novel drug delivery devices are the main focus covered in this review. With this in mind, drug delivery systems administration routes: subdermal, transdermal, and vaginal and the mechanisms of drug release are introduced and their differences are highlighted. Silicone rubber as a suitable drug carrier candidate in DDS is introduced and critical curing parameters, such as heat and moisture which play important roles in designing and processing of drug delivery devices are studied. Modifications of silicone rubber in drug delivery systems to impart some physico-chemical properties to make it suitable for specific medical applications are discussed, as well. The most informative part of this paper includes a survey on the developments of silicone-based drug delivery systems in relation to drug categories, their functions, and types. The last section includes a brief introductory of commercially available silicone-based drug delivery systems consisting of intravaginal, subdermal, and transdermal drug delivery devices. Transdermal drug delivery devices including reservoir and matrix types are discussed in more detail. It can be concluded that silicone polymers as important drug carriers have made great impact on the advancement of drug delivery science and technology.

REFERENCES

- Lee PI, Good WR, Controlled-Release Technology Pharmaceutical Applications, ACS Washington DC, 1987.
- Fan LT, Singh SK, Controlled Release, A Quantitative Treatment, Springe Verlag, Berlin, 9-83, 1989.
- 3. Brannon-Peppas L, Polymers in Controlled Drug Delivery, *Med Plast Biomater*, **4**, 34-44, 1997.
- 4. Kost J, Controlled drug delivery systems. *In: Polymer Materials Encyclopedia*, Salamone JC (Ed), CRC, New York, 1509-1517, 1996.
- 5. Kim CJ, Controlled Release Dosage Form Design, Technomic, Lancaster, PA, 49-74, 2000.
- 6. Uchegbu IF, Schätzlein AG, *Polymers in Drug Delivery*, CRC, New York, 2006.
- 7. Kaparissides C, Alexandridou S, Kotti K, Chaitidou S, Recent advances in novel drug delivery systems, *J Nano Tech*, **2**, 1-11, 2006.
- 8. Rathbone MJ, Hadgraft J, Roberts MS, *Modified-Release Drug Delivery Technology*, Informa Health Care, New York, 2003.
- 9. Ganji F, Vasheghani-Farahani E, Hydrogels in controlled drug delivery systems, *Iran Polym J*, **18**, 63-88, 2009.
- 10. Rahimi A, Shokrolahi P, Rezaie M, Inorganic polymers: novel applications, *5th Iran Sem Polym Sci Technol (ISPST)*, Tehran, 12th-14th Sep, 2000.
- 11. Rahimi A, Shokrolahi P, Application of inorganic polymeric materials, I: polysiloxanes, *Int J Inorg Mat*, **3**, 843-847, 2001.
- 12. Rahimi A, Inorganic Polymers, *Proceedings of 6th Iran Sem Polym Sci Technol (ISPST 2003)*, Tehran, Iran, 12th-15th May, 2003.
- 13. Rahimi A, Inorganic and organometallic polymers: a review, *Iran Polym J*, **13**, 149-164, 2004.
- 14. Hill DJT, Preston CML, Whittaker AK, Hunt S, The radiation chemistry of poly(dimethylsiloxane), *Macro Mol Symp*, **156**, 95-102, Tokyo, Japan, 5th-9th Sep, 1999.
- 15. Heiner J, Stenberg B, Persson M, Crosslinking of siloxane elastomers, *Polym Test*, **22**, 253-257, 2003.
- 16. Warbrik JS, Boylan JC, *Encyclopedia of Pharmaceutical Technology*, Informa Health Care, New York, 266, 2004.

- 17. Morton M, *Rubber Technology*, Van Nostrand, New York, 375-409, 1987.
- 18. Bopp CD, Sisman O, Radiation stability of plastics and elastomers, *Nucleonics*, **13**, 28-33, 1955.
- 19. Warrick EL, Effects of radiation on organopolysiloxanes, *Ind Eng Chem*, **45**, 2388-2393, 1955.
- Przybyla RL, Preliminary investigation of the feasibility of radiation vulcanization of silicone rubber, *Rubber Chem Technol*, 47, 285-288, 1974.
- 21. Vokal A, Kourim P, Sussmilchova J, Heidingsfeldova M, Kopecky B, Comparison of thermal and radiation curing of silicone rubber, *Radiat Phys Chem*, **28**, 497-499, 1986.
- 22. Basfer AA, Hardness measurents of silicone rubber and polyurethane rubber cured by radiation, *Radiat Phys Chem*, **50**, 607-610, 1997.
- 23. Frounchi M, Dadbin S, Panahinia F, Comparison between electron beam and chemical crosslinking of silicone rubber, *Nucl Inst Meth B*, **243**, 354-358, 2006.
- 24. St Pierre LE, Dewhurst HA, Bueche AM, Swelling and elasticity of irradiated polydimethylsiloxanes, *J Polym Sci*, **36**, 105-111, 1959.
- 25. Delides CG, Shepherd IW, Dose effects in the crosslinking of irradiated polysiloxane, *Radiat Phys Chem*, **10**, 379-385, 1977.
- 26. Hill DJT, Preston CML, Salisbury DJ, Whittaker AK, Molecular weight changes and scission and crosslinking in poly(dimethyl siloxane) on gamma radiolysis, *Radiat Phys Chem*, **62**, 11-17, 2001.
- 27. Abbasi F, Mirzadeh H, Katbab AA, Modification of polysiloxane polymers for biomedical applications: a review, *Polym Int*, **50**, 1279-1287, 2001.
- 28. Nikolaev OO, Urhanov VB, Britov VP, Babaev AD, Bogdanov VV, Mirzadeh H, A thermoplastic rubber composition for medical purposes, *Iran Polym J*, **10**, 9-14, 2001.
- 29. McMillin CR, Elastomers for biomedical applications, *Rubber Chem Technol*, **67**, 417-446, 1994.
- 30. Kammermeyer K, Silicone rubber as a selective barrier: gas and vapor transfer, *Ind Eng Chem*, **49**, 1685, 1957.
- 31. Robb WL, Thin silicone membranes-their permeation properties and some applications, *Ann N Y*

- Acad Sci, 146, 119-137, 1968.
- 32. Folkman J, Long DM, Drug pacemakers in the treatment of heart block, *Ann N Y Acad Sci*, **111**, 857-868, 1964.
- 33. Bates BL, Ragheb AO, Stewart JM, Bourdeau WJ, Choules BD, Purdy JD, Fearnot NE, Coated implantable medical device, *US Patent 6,918,927*, (Jul 19, 2005).
- 34. Chen CH, Doan PD, Chitre Y, Helland JR, Method of manufacturing a drug-eluting endocardial lead utilizing, *US Patent 7,174,221*, (Feb 6, 2007).
- 35. Sano A, Maeda H, Kajihara M, Tani S, Sugie T, Long time drug-sustained release preparation, *US Patent 6*,756,048 (Jun 29, 2004).
- 36. Ragheb AO, Bates BL, Fearnot NE, Kozma TG, Voorhees WD, Gershlick AH, Coated implantable medical device, *US Patent* 7,410,665, (Aug 12, 2008).
- 37. Chen CH, Doan PD, Chitre Y, Helland JR, Method of molding silicone elastomer drug carrier in an endocardial, *US Patent 7,363,091*, (Apr 22, 2008).
- 38. Hof M, Bruner S, Freedman J, Comparing silicone pressure-sensitive adhesives to silicone gels for transdermal drug delivery, LLC *33rd Ann Meet Expo Control Rel Soc*, Vienna, Austria, 22th-26th July, 2006.
- 39. Ogle CR, Pickering KL, Rathbone MJ, Burggraaf S, Bunt C, Application of materials science to drug delivery, *The Inst Prof Eng New Zealand (IPENZ) Ann Conf*, **2**, Wellington, New Zealand, 7th-10th Feb, 1997.
- 40. Chien WY, Novel drug delivery systems. *In: Drugs and the Pharmaceutical Sciences*, Marcel & Dekker, New York, **50**, 43-137, 1992.
- 41. Keshvari H, Mirzadeh H, Mansoori P, Orang F, Khorasani MT, Collagen immobilization onto acrylic acid laser-grafted silicone for using as artificial skin: in vitro, *Iran Polym J*, **17**, 171-182, 2008.
- 42. Okada T, Ikada Y, Modification of silicone surface by graft polymerization of acrylamide with corona discharge, *Makromol Chem*, **192**, 1705-1713, 1991.
- 43. Abbasi F, Mirzadeh H, Katbab AA, Bulk and surface modification of silicone rubber for biomed-

- ical applications, *Polym Int*, **51**, 882-888, 2002.
- 44. Kim SJ, Shin MS, Synthesis and characteristics of interpenetrating polymer network hydrogels based on silicone and poly(vinyl alcohol), *Funct Biomat Key Engineering Materials*, **198**, 301-307, 2001.
- 45. Dillon ME, Microporous polytetrafluoroethylene and polydimethylsiloxane IPN membrane, *Polym Mat Sci Eng Proc ACS Div Polym Mat Sci Eng*, **62**, 814-818, Boston, USA, 1st April, 1990.
- 46. Lopour P, Vondracek P, Janatova V, Sulc J, Vacik J, Silicone rubber-hydrogel composites as polymeric biomaterials. II: hydrophilicity and permeability to water-soluble low-molecular-weight compounds, *Biomaterials*, 11, 397-402, 1990.
- 47. Lednicky F, Janatova V, Lopour P, Vondracek P, Silicone rubber-hydrogel composites as polymeric biomaterials. III: an investigation of phase distribution by scanning electron microscopy, *Biomaterials*, **12**, 848-852, 1991.
- 48. Lopour P, Plichta Z, Volfova Z, Hron P, Vondracek P, Silicone rubber-hydrogel composites as polymeric biomaterials. IV: silicone matrix-hydrogel filler interaction and mechanical properties, *Biomaterials*, **14**, 1051-1055, 1993.
- 49. Lopour P, Janatova V, Silicone rubber-hydrogel composites as polymeric biomaterials. VI: transport properties in the water-swollen state, *Biomaterials*, **16**, 633-640, 1995.
- Hron P, Slechtova J, Smetana K, Dvorankova B, Lopour P, Silicone rubber-hydrogel composites as polymeric biomaterials. IX: composites containing powdery polyacrylamide hydrogel, *Biomaterials*, 18, 1069-1073, 1997.
- 51. Cifkova P, Lopour P, Vondracek P, Jelinek F, Silicone rubber-hydrogel composites as polymeric biomaterials. I: biological properties of the silicone rubber-poly(HEMA) composite, *Biomaterials*, **11**, 393-396, 1990.
- 52. Hron P, Hydrophilisation of silicone rubber for medical applications, *Polym Int*, **52**, 1531-1539, 2003.
- 53. Fallahi D, Mirzadeh H, Khorasani MT, Physical, mechanical, and biocompatibility evaluation of three different types of silicone rubber, *J Appl Polym Sci*, **8**, 2522-2529, 2003.
- 54. Mashak A, In vitro drug release from silicone

- rubber-polyacrylamide composite, *Silicon Chem*, **3**, 295-301, 2008.
- 55. Wu Z, Jiang Y, Kim T, Lee K, Effects of surface coating on the controlled release of vitamin B1 from mesoporous silica tablets, *J Control Rel*, **119**, 215-221, 2007.
- 56. Altinkaya SA, Yenal H, In vitro drug release rates from asymmetric membrane tablet coatings: prediction of phase-inversion dynamics, *Biochem Eng J*, **28**, 131-139, 2006.
- 57. Nahrup JS, Gao ZM, Mark JE, Sakr A, Poly(dimethylsiloxane) coatings for controlled drug release-polymer modifications, *Int J Pharm*, **270**, 199-208, 2004.
- 58. Gao Z, Nahrup JS, Mark JE, Sakr A, Poly(dimethylsiloxane) coatings for controlled drug release. I: preparation and characterization of pharmaceutically acceptable materials, *J Appl Polym Sci*, **90**, 658-666, 2003.
- 59. Gao Z, Nahrup JS, Mark JE, Sakr A, Poly(dimethylsiloxane) coatings for controlled drug release. II: mechanism of the crosslinking reaction in emulsion, *J Appl Polym Sci*, **91**, 2186-2194, 2004.
- 60. Gao Z, Nahrup JS, Mark JE, Sakr A, Poly(dimethylsiloxane) coatings for controlled drug release. III: drug release profiles and swelling properties of the free-standing films, *J Appl Polym Sci*, **96**, 494-501, 2005.
- 61. Li LC, Peck GE, Water based silicone elastomer controlled release tablet film coating. I: free film evaluation, *Drug Dev Ind Pharm*, **15**, 65-95, 1989.
- 62. Li LC, Peck GE, Water based silicone elastomer controlled release tablet film coating. II: formulation considerations and coating evaluation, *Drug Dev Ind Pharm*, **15**, 499-531, 1989.
- 63. Li LC, Peck GE, Water based silicone elastomer controlled release tablet film coating. III: drug release mechanisms, *Drug Dev Ind Pharm*, **15**, 1943-1968, 1989.
- 64. Li LC, Peck GE, Water based silicone elastomer controlled release tablet film coating. IV: process evaluation, *Drug Dev Ind Pharm*, **16**, 415-435, 1990.
- 65. Li LC, Peck GE, Water based silicone elastomer controlled release tablet film coating. V: a statis-

- tical approach, *Drug Dev Ind Pharm*, **17**, 27-37, 1991.
- 66. Li LC, Peck GE, Water based silicone elastomer controlled release tablet film coating. VI: the effect of tablet shape, *Drug Dev Ind Pharm*, **18**, 333-343, 1992.
- 67. Tan EL, Liu JC, Chien, YW, Controlled drug release from silicone coated tablets: preliminary evaluation of coating techniques and characterization of membrane permeation kinetics, *Int J Pharm*, **42**, 161-169, 1988.
- 68. Dahl TC, Sue IT, Mechanisms to control drug release from pellets coated with a silicone elastomer aqueous dispersion, *Pharm Res*, **9**, 398-405, 1992.
- 69. Padsalgikar AD, Nguyen X, Griffiths IM, Evaluation of Biostable Polyurethane Silicone Copolymers (Elast-EonTMs) as Drug Eluting Polymers for Stent Coating, *7th Biomat Cong*, Sydney, Australia, 17th-21th May, 2004.
- 70. Shapiro M, Hanon S, Misra D, Drug-eluting stents, *Hosp Physician*, **40**, 11-20, 2004.
- 71. Lim L, Biocompatibility of stent materials, *MURJ*, **11**, 33-37, 2004.
- 72. Ding N, Method of applying drug-release coatings, *US Patent* 5,980,972 (Nov 9, 1999).
- 73. Tijsma EJ, Bachert C, Hissong JB, Watelet JB, Drug-releasing sinus stent, *US Patent* 2007/0014830 A1, (Jun 8, 2005).
- 74. Ding N, Helmus MN, Drug release stent coating process, *US Patent* 5,837,313, (Nov 17, 1998).
- 75. Lanzer P, *Mastering Endovascular Techniques:* A Guide to Excellence, Lippincott Williams & Wilkins, Philadelphia, 132-133, 2006.
- 76. Taghizadeh SM, Mashak A, Jamshidi A, Imani M, Study of additive effect on mechanical properties, drug release behaviour and mechanisms in a monolithic system, *Iran Polym J*, 12, 407-412, 2003.
- 77. Mashak A, Taghizadeh SM, In vitro progesterone release from γ-irradiated cross-linked polydimethylsiloxane, *Radiat Phys Chem*, **75**, 229-235, 2006.
- 78. Mashak A, Taghizadeh SM, The effect of radiation type and dose on drug release from RTV silicone elastomer, *7th Iran Sem Polym Sci Technol*, Tehran, Iran, 27th-29th Sep, 2005.

- 79. Taghizadeh SM, Mashak A, Jamshidi A, Imani M, Preparation of polymeric matrix containing progesterone and release study by a novel analytical method, *Proce 30th Ann Meet Expo Control Rel Soc*, Glasgow, UK, 19th-23th July, 2003.
- 80. Taghizadeh SM, Mashak A, Jamshidi A, Release study of progesterone from silicone by a novel analytical method, *6th Iran Sem Polym Sci Technol*, Tehran, Iran, 12th-15th May, 2003.
- 81. Favero RJ, Henderson EA, Kesler DJ, Ability of norgestomet impregnated polydimethylsiloxane implants to maintain pregnancy subsequent to ovariectomy, *Polym Mat Sci Eng Proc ACS Div Polym Mat Sci Eng*, **59**, 807, 1988.
- 82. Chien YW, Rozek LF, Lamberg HJ, In vitro- in vivo correlation on controlled release of desoxy-corticosterone acetate from matrix-type silicone devices, *ACS Div Org Coat Plast Chem Prepri*, **36**, 329-330, 1976.
- 83. Maeda H, Sano A, Kawasaki H, Kurosaki Y, Study on accelerated evaluation system for release profiles of covered-rod type silicone formulation using indomethacin as a model drug, *J Control Rel*, **94**, 337-349, 2004.
- 84. Hu Z, Wang C, Nelson KD, Eberhart RC, Controlled release from a composite silicone/hydrogel membrane, *ASAIO J*, **46**, 431-434, 2000.
- 85. Maeda H, Ohashi E, Sano A, Kawasaki H, Kurosaki Y, Investigation of the release behavior of a covered-rod-type formulation using silicone, *J Control Rel*, **90**, 59-70, 2003.
- 86. Gantner DC, Klykken PC, Raul VA, Schalau GK, Thomas X, Tuneable silicone matrices for sustained release of actives, *Trans-7th World Biomat Cong*, Sydney, Australia, 856, 17th-21th May, 2004.
- 87. Sutinen R, Paronen P, Saano V, Urtti A, Wateractivated, pH-controlled patch in transdermal administration of timolol. II: drug absorption and skin irritation, *Eur J Pharm Sci*, **11**, 25-31, 2000.
- 88. Karlgard C, Moresoli C, Uptake and release of acular from silicone-hydrogel and conventional hydrogel contact lens materials, *Optometry Vision Sci*, 77, 179, 2000.

- 89. Inoue K, Ogawa K, Okada J, Sugibayashi K, Enhancement of skin permeation of ketotifen by supersaturation generated by amorphous form of the drug, *J Control Rel*, **108**, 306-318, 2005.
- 90. Whang H, Hunt MA, Wrench N, Hockney JE, Farin CE, Tonelli AE, Nonoxynol-9-α-cyclodextrin inclusion compound and its application for the controlled release of nonoxynol-9 spermicide, *J Appl Polym Sci*, **106**, 4104-4109, 2007.
- 91. Klykken PC, Gantner DC, Thomas X, Gebert MS, Mazeaud I, Saldajeno M, Bott R, Sustained release of active enzymes from silicone matrices, *Trans-7th World Biomat Cong*, Sydney, Australia, 17th-21th May, 2004.
- 92. Gonzalez B, Colilla M, Vallet-Regi M, Time-delayed release of bioencapsulates: a novel controlled delivery concept for bone implant technologies, *Chem Mater*, **20**, 4826-4834, 2008.
- 93. Uurto I, Isotalo T, Mikkonen J, Martikainen PM, Kellomaki M, Tormala P, Tammela TLJ, Talja M, Salenius JP, Tissue biocompatibility of new biodegradable drug-eluting stent materials, *J Mater Sci Mater Med*, **18**, 1543-1547, 2007.
- 94. Lopez VC, Raghavan SL, Snowden MJ, Colloidal microgels as transdermal delivery systems, *React Funct Polym*, **58**, 175-185, 2004.
- 95. Chien TW, Mares SE, Berg J, Huber S, Lambert HJ, King KF, Controlled drug release from polymeric delivery devices. III: in vitro- in vivo correlation for intravaginal release of ethynodiol diacetate from silicone devices in rabbits, *J Pharm Sci*, **64**, 1776-1781, 1975.
- 96. Ueno N, Refojo MF, Liu LHS, Controlled release rate of a lipophilic drug (BCNU) from a refillable silicone rubber device, *J Biomed Mater Res*, **16**, 669, 1982.
- 97. Tiffany J, Drug diffusion-through silicone membranes, *Transac Ann Meet of the Soc Biomater Conjunc with the Intern*, **8**, San Diego, CA, USA, 24th-28th April, 1985.
- 98. Anderson JM, Niven H, Pelagalli J, Olanoff LS, Jones RD, Role of the fibrous capsule in the function of implanted drug-polymer sustained release systems, *J Biomed Mater Res*, **15**, 889-902, 1981.
- 99. Boileau S, Bouteiller L, Khalifa RB, Liang Y, Teyssie D, Polycarbonate-polysiloxane based

- interpenetrating networks, ACS Polym Prepri, Div Polym Chem, **39**, 457, 1998.
- 100. Tojo K, Sun Y, Ghannam MM, Chien YW, Characterization of a membrane permeation system for controlled drug delivery studies, *AIChE J*, **31**, 741-746, 1985.
- 101. Walters PA, Medical-grade silicone in new controlled release medication, *Elastomerics*, 117, 26, 1985.
- 102.Ma JT, Wu L, Qi GG, Lui JH, Tang PL, Zhang YL, Tong TZ, Yao KD, Radiation crosslinked poly(vinylmethylsiloxane) for levonorgestrel delivery system, *J Polym Sci C Polym Lett*, **26**, 195, 1988.
- 103. Schierholz J, Jaenicke L, Pulverer G, In-vitro efficacy of an antibiotic releasing silicone ventricle catheter to prevent shunt infection, *Biomaterials*, **15**, 996-1000, 1994.
- 104. Andreopoulos AG, Plytaria M, Biomedical silicone elastomers as carriers for controlled release, *J Biomater Appl*, **12**, 258-271, 1998.
- 105.Gupta DC, Sumana G, Jain R, Pant BP, Jaiswal DK, Effect of silica and poly(ethylene glycol) on the in vitro release rate of 2-pyridine aldoxime chloride from polysiloxane matrices, *Polym Int*, **49**, 329-333, 2000.
- 106. Gupta DC, Sumana G, Agarwal S, Polysiloxanes as matrix materials for slow release of 2-pyridine aldoxime chloride, *Polym Int*, **45**, 211-216, 1998.
- 107.Malcolm K, Woolfson D, Russell J, Tallon P, McAuley L, Craig D, Influence of silicone elastomer solubility and diffusivity on the in vitro release of drugs from intravaginal rings, *J Control Rel*, **90**, 217-225, 2003.
- 108. Woolfson AD, Elliott GRE, Gilligan CA, Passmore CM, Design of an intravaginal ring for the controlled delivery of 17β-estradiol as its 3-acetate ester, *J Control Rel*, **61**, 319-328, 1999.
- 109. Woolfson AD, Malcolm RK, Gallagher RJ, Design of a silicone reservoir intravaginal ring for the delivery of oxybutynin, *J Control Rel*, **91**, 465-476, 2003.
- 110. Malcolm RK, McCullagh S, Woolfson AD, Catney M, Tallon P, A dynamic mechanical method for determining the silicone elastomer

- solubility of drugs and pharmaceutical excipients in silicone intravaginal drug delivery rings, *Biomaterials*, **23**, 3589-3594, 2002.
- 111. Hukka TI, Pakkanen TT, Modeling the interaction between two- and four-ring progestin models and a silicone-based polymer model: a density functional theory study, *J Chem Inf Model*, **47**, 535-546, 2007.
- 112. Kim J, Conway A, Chauhan A, Extended delivery of ophthalmic drugs by silicone hydrogel contact lenses, *Biomaterials*, **29**, 2259-2269, 2008.
- 113. Malcolm K, Woolfson D, Russell J, Andrews C, In vitro release of nonoxynol-9 from silicone matrix intravaginal rings, *J Control Rel*, 91, 355-364, 2003.
- 114. Malcolm K, Woolfson D, Elliott G, Shephard M, Intravaginal drug delivery devices for the administration of an antimicrobial agent, *US Patent 6,951,654* (Oct 4, 2005).
- 115. Nabahi S, Intravaginal drug delivery device, *US Patent 6,103,256* (Aug 15, 2000).
- 116. Nabahi S, Intravaginal drug delivery device, *US Patent 5,788,980* (Aug 4, 1998).
- 117. Brown TA, Osborne J, Rudella M, Hunt B, Malik V, Compositions and medical device for transdermal delivery of a drug, *US Patent* 7,247,315 (Jul 24,2007).
- 118. Shalaby SW, Hilas GT, Multicomponent bioactive intravaginal ring, *US Patent* 11,974,140 (Oct 11, 2007).
- 119. Malcolm RK, Woolfson AD, Toner CF, Morrow RJ, McCullagh SD, Long-term, controlled release of the HIV microbicide TMC120 from silicone elastomer vaginal rings, *J Antimicrob Chemother*, **56**, 954-956, 2005.
- 120. Foye WO, Lemke TL, Williams DA, *Foye's Principles of Medicinal Chemistry*, Lippincott Williams & Wilkins, Philadelphia, 1329, 2007.
- 121.ESTRING, http://www.pfizer.com/files/prod-ucts/uspi_estring.pdf, Aug 2008.
- 122. Touitou E, Bary BW, Enhancement in Drug Delivery, CRC, 449-450, 2006.
- 123. Ansel HC, Allen LV, Popovich NG, *Pharmaceutical Dosage Forms and Drug Delivery systems*, 7th ed, Lippincott Williams & Wilkins, Philadelphia, 661, 1999.

- 124.Femring, http://www.fda.gov/cder/foi/label/2003/21367 femring lbl.pdf.
- 125. Mathiowitz E, *Encyclopedia of Controlled Drug Delivery*, Wiley, New York, 1006-1037, 1999.
- 126.Rothen-Weinhold A, Dahn M, Gurny R, Formulation and technology aspects of controlled drug delivery in animals, *PSTT*, **3**, 222-231, 2000.
- 127.Rathbone MJ, Gurny R, Controlled Release Veterinary Drug Delivery: Biological and Pharmaceutical Consideration, Elsevier Science, New York, 180-183, 2000.
- 128.Rathbone MJ, Macmillan KL, Inskeep K, Burggraaf S, Bunt CR, Fertility regulation in cattle, *J Control Rel*, **54**, 117-148, 1998.
- 129. Rathbone MJ, Macmillan KL, Applications of controlled release science and technology: progesterone, *CRS*, *Newsletter*, **21**, 8, 2004.
- 130. Wilkins WL, *Lippincott's Nursing Procedures*, Lippincott Williams & Wilkins, Philadelphia, 337, 2008.
- 131. Wells BG, Dipiro JT, Schwinghammer TL, Hamilton CW, *Pharmacotherapy Handbook*, McGraw-Hill, New York, 302, 2005.
- 132. Swarbrick J, Boylan JC, *Encyclopedia of Pharmaceutical Technology*, 2nd ed, Informa Health Care, New York, 816, 2002.
- 133.Hardee GE, Baggot JD, Development and Formulation of Veterinary Dosage Forms, Marcel & Dekker, New York, 220-221, 1998.
- 134. Tregaskes LD, Broadbent PJ, Dolman DF, Grimmer SP, Franklin MF, Evaluation of Crestar, a synthetic progestogen regime, for synchronising oestrus in maiden heifers used as recipients of embryo transfers, *Vet Rec*, **134**, 92-94, 1994.
- 135.Crestar, http://www.intervet.com.au/binaries /82 103700.pdf.
- 136.Guy RH, Hadgraft J, *Transdermal Drug Delivery*, Informa Health Care, New York, 5-7, 2002.
- 137.Shah SH, Shah D, Transdermal drug delivery technology revisited: recent advances, *Pharmaceut Rev*, **6**, 2008.
- 138. Taghizadeh SM, Lahootifard F, Transdermal excipients effect on adhesion strength of a pressure sensitive adhesive, *Iran Polym J*, **12**, 243-

248, 2003.

- 139. Dumitriu S, *Polymeric Biomaterials*, CRC, New York, 703-710, 2001.
- 140.Ranade VV, Hollinger MA, *Drug Delivery Systems*, CRC, New York, 230-231, 2003.
- 141.www.dowcorning.com, Silicone adhesives: for transdermal and topical drug delivery, medical devices and wound care, DowCorning Corporation, 2006.
- 142.http://www.pharma.us.novartis.com/product/pi/pdf/Vivelle DOT.pdf, August 2004.
- 143. Hair PI, Keating GM, McKeage K, Transdermal matrix fentanyl membrane patch (Matrifen®): in severe cancer-related chronic pain, *Drugs*, **68**, 2001-2009, 2008.
- 144. Venkatraman S, Gale R, Skin adhesives and skin adhesion. I: transdermal drug delivery systems, *Biomaterials*, **19**, 1119-1136, 1998.

