Essential oil Composition and Antioxidant Activity of *Levisticum* officinale Koch. at Various Phenological Stages

Mohamadi N (Ph.D. Student)¹, Rajaei P (Ph.D.)²*, Moradalizadeh M (Ph.D.)³, Amiri MS (Ph.D.)⁴

1- Pharmaceutics Research Center, Institute of Neuropharmacology, Kerman University of Medical Sciences, Kerman, Iran

2- Departments of Biology, Kerman Branch, Islamic Azad University, Kerman, Iran

3- Departments of Chemistry, Kerman Branch, Islamic Azad University, Kerman, Iran

4- Department of Biology, Payame Noor University, Tehran, Iran

* Corresponding author: Departments of Biology, Kerman Branch, Islamic Azad University, Kerman, Iran

Tel: +98-034-31325245, Fax: +98-034-31325215

Email: rajaeipeyman@gmail.com

Received: 12 June 2016

Accepted: 25 Dec. 2016

Abstract

Background: The leaves, roots and seeds of Lovage (*Levisticum officinale* Koch) being used as an herb, vegetable and spice.

Objective: The aim of this study is evaluation of the changes of phytochemical compound in aerial parts of the Lovage plants at different developmental stages as well as antioxidant activity.

Methods: The plants were harvested from Hezar Mountain. After drying, essential oils were extracted by steam distillation. To identify the essential components GC and GC-MS was used. Antioxidant activity of samples was examined by diphenylpicrylhydrazyl (DPPH) assay.

Results: The average yield of essential oil was respectively, 2.3% and 3.1% and 1.5% respectively, in vegetative, Flowering and seed stage. The oil analysis results showed that 21 compounds were identified in the vegetative stage. β -phellandrene (10.7%) and α -Terpinyl acetate (% 38.9) and Curzerene (% 10.6) were the major compound. in the reproductive stage, 22 compounds were identified including β -phellandrene (20.3%) and α -Terpinyl acetate (% 20.4) and γ -Cadinene (12.1%). in the seed stage β -phellandrene (21.1%) and α -Terpinyl acetate (% 25.3) and Sabinene (10.2%) were the highest. Analysis of variance showed that the effect of harvesting time has significant effect on plant height, stem branch number and yield of oil. The Higher antioxidant power was observed respectively in the flowering stage (83%), vegetative stage (68%) and seed stage (60%).

Conclusion: The phytochemical and antioxidant compounds in *Levisticum officinale* depend on the phenological stage.

Keywords: Lovage (*Levisticum officinale* Koch.), Essential oils, Antioxidant activity, vegetative stage, flowering stage, seed stage



Essential oil ...

Introduction

Lovage (Levisticum officinale Koch.) is known as "Karafse kuhi" in Persian. It is a perennial plant belongs to the family Apiaceae. The traditional usage of Levisticum officinale in several diseases has been documented in previous resources and in folk tradition. The lovage root is utilized as a diuretic drug [1]. A lot of phytochemical studies have been done to determine the chemical composition of the essential oil of lovage. Terpenes and phthalides, βphellandrene, α -terpinyl acetate, and Zligustilide are the main components of lovage oil that present in different value in the various plant organs. The Essential oil compounds of the hairy roots of lovage were falcarinol, (Z)ligustilide, (Z)-3-butylidenephthalide, trans-bfarnesene, b-phellandrene. n-octanal. gelemene and n-heptanal [2]. In other reports, ligustilide, a natural phthalide, is the most abundant bioactive component in essential oils of L. officinale roots [3-6]. L. officinale exhibits diverse pharmacological activities, containing estrogenic [7], apoptotic [8], and antimycobacterial activities [9].

The chemical composition of different extracts of Levisticum officinale revealed more than 190 volatile compounds, frequently monoterpenes and phtalide[10] .Eskin and Tamir found that n-butylidene-4,5dihydrophtalide is One of the major components at 67% concentration range [11]. Cichy et al. (1984) and Bradley (2006) found two phtalide dimers, levistolide A and levistolide B in lovage roots [12]. Up to 70% of the oil is composed of alkylphtalides which are responsible for the characteristic odour [13].



The essential oil content in lovage roots is determined by the harvesting time. Harvesting the leaves during the vegetation season has a negative effects on the gathering of essential oil in the roots [14]. Keeping shoot of Levisticum until autumn significantly increased the amount of essential oil from 0.52 to 0.85%. literature reviews present following composition of lovage root oil as % of oil yield - α -terpinyl acetate: 0.1-0.2, βphellandrene: 1.7-15.5, α -phellandrene: 0.2-0.5, myrcene: 0.3-0.9, (Z)-ligustilide: 37.0-67.5 and pentylcyclohexadiene: 7.4-29.3 [15]. Acording to Samiee et al research, the main constituents in the oil of Levisticum officinale, were α -terpinyl acetate (40.5%) and βphellandrene (16.7%) were the maior components, but in the extract, β -phellandrene (23.0%), naphtalene (20.6%) and γ -terpinene (12.1%) [16]. The main components of the oil in the other report were β -phellandrene (42.5%), cc-terpineol (27.9%), cis-ocimene (7.5%) [17].

It seemed, different developmental stages of *L. officinale* would have different oil compositions therefore, in this study we report the variation of the essential oil composition from the aerial parts of the plant harvested at different developmental stages as well as antioxidant activity. These results can be used to investigate the optimal harvesting time of this plant for relevant industries.

Materials and Methods Plant material and oil isolation

The aerial parts of *Levisticum officinale* were collected at different developmental stages (vegetative, full flowering and seed stage) from Hezar mt. (29°30/ N, 57°20/ E), Kerman province at an altitude of 3440 m. Voucher specimen was deposited at the Herbarium of Islamic Azad University, Kerman branch. The essential oil of all airdried samples was isolated by hydrodistillation for 3 h, using a Clevenger-type apparatus. The distillated oils were dried over anhydrous sodium sulfate and stored at 4 °C until analysis.

GC and GC/MS analysis

GC-MS analysis was performed using a Hewlett-Packard 5973 mass spectrometer coupled to a Hewlett-Packard 6890 gas chromatograph equipped with a HP-5MS capillary column (5 % phenylmethylpolysiloxane, 30 m × 0.25 mm, film thickness 0.25 μ m). The carrier gas was helium and the chromatographic conditions were as above. All mass spectra were acquired in electron-impact (EI) mode with an ionization voltage of 70 eV.

Identification of compounds

N-alkanes (C6–C24) were applied as reference points in the computation of relative retention indexes. The essential oils components were identified by comparing of the retention times of their mass spectra with the mass spectra library data search [18]. To determine the value of relative area percentages obtained by FID were utilized without the usage of correction factors.

Antioxidant activity

Antioxidant activity was calculated by a spectrophotometric method based on the

reduction of a methanol solution of 1,1diphenyl-2-picrylhydrazyl (DPPH) [19]. Samples were replicated 3 times. 50 μ l of sample was added to 5 ml of methanolic solution of DPPH (0.02 mM). The absorbance was measured after 30 min incubation period at room temperature at 517 nm. Methanol was used as blank. Butylated hydroxytoluene (BHT) and the solvent were used as positive and negative controls respectively. The percentage of inhibition was calculated according to formula: 1%=(A_b-A_s/A_b) ×100.

Where A_b is absorbance of the control mixture (containing all reagents except the test Sample), and A_s is the absorbance of the test Sample.

Results

Soil characteristics of the study area

Experimental results show that soil samples of the study area was sandy loam that is not suitable for agriculture. Soil phosphorus was 6.9 mg per kg while the optimal value is between 7-10 mg per kg (Table 1).

Chemical composition of the essential oil

The essential oil yields (w/w %) based on the dry weight of the plant, in different harvesting times of developmental growth stages were in the order of: flowering (3.1%)> and vegetative (2.3%) > seed stage (1.9%). In total, 21, 22 and 19 constituents were identified and quantified in the subsequent stages, respectively (Table 2). At vegetative stage the highest amount of compound belonged to α -Terpinyl acetate (38.9), β phellandrene (10.7) and Curzerene (10.6) while α-Terpinyl acetate (20.4),β-



www.SID.ir

Essential oil ...

phellandrene (20.3) and γ -Cadinene (12.1) were highest amount in full flowering stage. At seed stage, α -Terpinyl acetate (21.1), β - phellandrene (25.3) and Sabinene (10.2) were highest value.

Table 1- Soil characteristics of the study area							
Clay%	Silt%	Sand%	P (mg/kg)	K (mg/kg)	OC%	РН	EC (ds/m)
10	17	73	460	48	1.86	6.9	1.8

No	Compound	R.I	Vegetative stage	Flowering stage	Seed stage
1	α-Thujene	930	-	0.2	0.2
2	α-Pinene	939	1.2	0.6	0.7
3	Camphene	954	0.2	0.4	-
4	Sabinene	975	2.2	4.3	10.2
5	β-Pinene	979	1.3	1.1	1.1
6	Myrcene	991	0.7	1.3	0.9
7	α -Phellandrene	1003	0.5	0.6	1.2
8	α-Terpinene	1017	0.2	0.4	0.3
9	<i>p</i> -Cymene	1025	1.3	2.4	1.5
10	β-phellandrene	1030	10.7	20.3	21.1
11	z-β-Ocimene	1037	7.0	6.4	-

7.4

0.4

0.1

0.6

1.7

0.2

38.9

0.2

4.3

10.6

4.3

94

8.9

1.4

0.5

1.6

2.2

0.5

20.4

0.4

0.8

8.2

12.1

95

8.0

0.4

-

0.5

0.5

0.7

25.3

0.4

-

9.9

5.8

88.7

1060

1089

1132

1177

1189

1237

1349

1359

1400

1499

1514

Table 2- Composition of essential oil of Levisticum officinale Koch at different developmental stages



γ-Terpinene

α-Terpinolene

allo-Ocimene

Terpinene-4-ol

α-Terpineol

α-Terpinyl acetate

N-Buthyl Phthalide

Methyl Eugenol

Pulegone

Curzerene

γ-Cadinene

Total

12

13

14

15

16

17

18

19

20

21

22



Fig. 1- GC-MS Chromatogram of Levisticum officinale Koch essential oil in the vegetative stage





Fig. 2- GC-MS Chromatogram of Levisticum officinale Koch essential oil in the flowering stage





Fig. 3- GC-MS Chromatogram of Levisticum officinale Koch essential oil in the seed stage

Antioxidant activity

The results of DPPH inhibition assay of different phenological stages samples of *L. officinale* have shown in Table 3. The

Higher antioxidant power was observed respectively in the flowering stage (83%), vegetative stage (68%) and seed stage (60%).

Table 3- Inhibition percentage of Levisticum officinale Koch esse	ential oil at different
developmental stages	

Phenological stages	Inhibition persentage (%) (Mean±SE)
vegetative stage	68 ± 3.2^{b}
flowering stage	$83{\pm}5.1^{a}$
seed stage	$60\pm2.8^{\circ}$



have different composition depending on the different factors [37].

Morphological study

The highest stem height (103 cm) was observed in the seed stage. The highest number of branches and the highest essential oil was obtained from the flowering stage. The number of branches from 12 at the vegetative stage reached to 18 at the flowering stage and at the seed stage remains the same number.

Discussion

From an agricultural perspective, carbon and organic matter is very important that the analyzed samples in this area have been largely poor. Further, the supporting results were also observed earlier in other members of the family Apiaceae. The essential oil content of Foeniculum vulgare (fruit) was evaluated at four different growth stages (immature, premature mature and full mature). Results showed that essential oil content declined with fruit maturity [25]. Similarly, in Coriandrum sativum L., the green fruit stage produced more essential oil than the brown fruit stage [26]. The results of Karimi et al study showed that the different phonological stages did not influence on chemical composition but has effects on the essential oil content [27]. Results showed that monoterpenes are the major portion of all samples. In Bylaite et al report, β -phellandrene was the most abundant component in all parts of lovage except the roots [11]. They showed seeds and flowers possessed the highest yield of oil. α -Terpinyl acetate was the highest compound in leaves

and stems, β -phellandrene in seeds and flowers, Z-ligustilide was found as a major lovage phthalide in the stem's essential oils depending on the harvesting time [28]. The results of Nejad Ebrahimi et al are consistent with the present research. They reported quality and quantity of essential oil components were different in various developmental stages including vegetative, floral budding, full flowering and seed stage. Flowering and floral budding stages have the highest percentage of essential oil and vegetative stage has the lowest one [29]. Marotti et al found different ontogenetic stages (early, late waxy and ripe seed) influence on the chemical composition and biological activity of Foeniculum vulgare [30]. The variation in the quantity and quality of Origanum majorana L. essential oil in different phonological stages has been demonstrated by Sellami et al. They found bioactive compounds were in maximum value in later vegetative stage [31]. Effect of developmental stage on yield and components of essential oil has been confirmed in other plants such as Salvia officinalis [32], Ribes nigrum [33], Mentha piperita [34] and Coriandrum sativum [35]. Plant metabolism is different in various stage of growth; therefore the production of bioactive components of essential oils could be affected [36]. The antioxidant activity depends on different factors. Then the chemical composition of the essential oil potentially influence on the biological activities of it. Shafaghat indicated from different that extracts parts of L. persicum possess considerable antioxidant activity. The highest radical scavenging



Journal of Medicinal Plants, Volume 16, No. 61, Winter 2017 activity was detected in seed [37]. The changes in chemical composition of plant essential oil depends on different factors such as genotype, Climate, plant organs, geographical locations, season of sampling, developmental stages. Thus, the essential oils of the same species probably have different composition depending on the different factors [38].

Conclusion

Present study clearly demonstrated that the

References

1. Yarnell E. Botanical medicines for the urinary tract. *World Journal of Urology* 2002; 20: 285-293.

2. Santos PA, Figueiredo AC, Oliveira MM, Barroso JG, Pedro LG, Deans SG, *et al.* Growth and essential oil composition of hairy root cultures of Levisticum officinale WDJ Koch (lovage). *Plant Science* 2005; 168: 1089-1096.

3. Kemzūraitė PR, Venskutonis A, Baranauskienė R and Navikienė D Optimization of supercritical CO 2 extraction of different anatomical parts of lovage (Levisticum officinale Koch.) using response methodology and evaluation surface of composition. The Journal of extracts Supercritical Fluids 2014; 87: 93-103.

4. Gijbels M, Scheffer J and Baerheim SA. Phthalides in the essential oil from roots of Levisticum officinale. *Planta Medica* 1982; 44: 207 - 211.

5. Penka M and Kocabova J. [Contribution to the study on variations in the essential oil content in the plant levisticum officinale

optimal time for harvesting of *Levisticum* is at flowering stage, because of the highest content of bioactive compounds in this period. The essential oil of lavage show strong antioxidant activity special in flowering stage, so it can be used as a natural preservative in food industries. GC-MS analysis displayed that the oil of lovage is rich in case of bioactive compounds such as β phellandrene and α -Terpinyl acetate and Curzerene, γ -Cadinene, Sabinene.

Koch.]. Ceskoslovenska Farmacie 1962; 11: 229 - 233.

6. Segebrecht S and Schilcher H. Ligustilide: guiding component for preparations of Levisticum officinale roots. *Planta Medica* 1989; 55: 572 - 573.

7. San Martin R. The estrogenic activity of various plant species; the activity of Levisticum officinale. *Farmacognosia; anales del Instituto José Celestino Mutis.* 1958; 18: 179.

8. Bogucka-Kocka A, Smolarz H and Kocki J. Apoptotic activities of ethanol extracts from some Apiaceae on human leukaemia cell lines. *Fitoterapia* 2008; 79: 487 - 97.

9. Schinkovitz A, Stavri M, Gibbons S, Bucar F. Antimycobacterial polyacetylenes from Levisticum officinale. *Phytotherapy Research* 2008; 22: 681 - 684.

10. Musitelli S. A Brief Historical Survey of Anaesthesia from Homer (9th-8th Century BC) to the 19th Century. Research 2014.

11. Eskin M and Tamir S. Dictionary of nutraceuticals and functional foods: CRC



Essential oil ...

Press 2005, P. 520.

12. Cichy M, Wray V and Hofle G. NEW CONSTITUENTS OF LEVISTICUM-OFFICINALE KOCH. *Liebigs Annalen Der Chemie* 1984; 397-400.

13. Bisset N and Wichtl M. Herbal Drugs and Phytopharmaceuticals Medpharm GmbH Scientific Publishers. In: Stuttgart, CRC Press, Boca Raton; 1994, P. 708.

14. Andruszczak S. Wplyw sposobu zalozenia plantacji i terminu zbioru lisci na plonowanie lubczyku ogrodowego [Levisticum officinale Koch.]. *Annales Universitatis Mariae Curie-Skłodowska. Sectio E, Agricultura.* 2004; 3: 1049 - 1056.

15. Hogg C, Svoboda K, Hampson J and Brocklehurst S. Investigation into the composition and bioactivity of essential oil from lovage (Levisticum officinale WDJ Koch). *International Journal of Aromatherapy* 2001; 11: 144 - 151.

16. Samiee K, Akhgar MR, Rustaiyan A and Masoudi S. Composition of the volatiles of Ferulago carduchorum Boiss. et Hausskn. and Levisticum officinale Koch. obtained by hydrodistillation and extraction. *Journal of Essential Oil Research* 2006; 18: 19 - 22.

17. Reza VRM and Abbas H. The essential oil composition of Levisticum officinalis from Iran. *Asian J. Biochem.* 2007; 2: 161 - 163.

18. Adam R. Identification of essential oil components by gas chromatography/quadrupole mass spectroscopy. Publisher, *Allured: Carol Stream, IL.* 2001.

19. Cuendet M, Hostettmann K, Potterat O and Dyatmiko W. Iridoid glucosides with free radical scavenging properties from Fagraea

blumei. *Helvetica Chimica Acta* 1997; 80: 1144 - 1152.

20. Telci I, Demirtas I and Sahin A. Variation in plant properties and essential oil composition of sweet fennel (*Foeniculum vulgare* Mill.) fruits during stages of maturity. *Industrial Crops and Products* 2009; 30: 126-130.

21. Ramezani S, Rasouli F and Solaimani B. Changes in essential oil content of Coriander (*Coriandrum sativum* L.) aerial parts during four phonological stages in Iran. *Journal of Essential Oil Bearing Plants* 2009; 12: 683-689.

22. Karimi N, Yari M and Ghasmpour HR. Identification and comparison of essential oil composition and mineral changes in different phenological stages of *Satureja hortensis* L. *Iranian J Plant Physiol.* 2012; 3: 577 - 582.

23. Bylaite E, Roozen JP, Legger A, Venskutonis RP and Posthumus MA. Dynamic headspace-gas chromatography-olfactometry analysis of different anatomical parts of lovage (*Levisticum officinale* Koch.) at eight growing stages. *Journal of Agricultural and Food Chemistry* 2000; 48: 6183 - 6190.

24. Bylaite E, Venskutonis RP and Roozen JP. Influence of harvesting time on the composition of volatile components in different anatomical parts of lovage (Levisticum officinale Koch.). Journal of Agricultural and Food Chemistry 1998; 46: 3735-3740.

25. Ebrahimi SN, Hadian J, Mirjalili M, Sonboli A and Yousefzadi M. Essential oil composition and antibacterial activity of Thymus caramanicus at different phenological stages. *Food Chem.* 2008; 110: 927 - 931.



26. Marotti M, Piccaglia R, Giovanelli E, Deans SG and Eaglesham E. Effects of variety and ontogenic stage on the essential oil composition and biological activity of fennel (*Foeniculum vulgare* Mill.). *Journal of Essential Oil Res.* 1994; 6: 57-62.

27. Sellami IH, Maamouri E, Chahed T, Wannes WA, Kchouk ME and Marzouk B. Effect of growth stage on the content and composition of the essential oil and phenolic fraction of sweet marjoram (*Origanum majorana* L.). *Industrial Crops and Products* 2009; 30: 395 - 402.

28. Heiran ghamari SM, Ghasem nejad Gh and Ghanbari AR. Evaluation of Phytochemical Composition of Sahandian Savory (*Satureja sahendica* Bornm.) Essential Oils at Different Phenological Stages (In Persian). *Journal of Agroecolog.* 2016; 8: 1 - 16.

29. Mirjalili MH, Salehi P, Sonboli A and Vala MM. Essential oil variation of *Salvia officinalis* aerial parts during its phenological cycle. *Chemistry of Natural Compounds* 2006; 42: 19 - 23.

30. Ahmadi L MM. Evaluation of different stages of growth, the chemical composition of *Salvia officinalis* (In persian). *Journal of Soil and Water Sciences* 1999; 2: 93-100.

31. Dvaranauskaitė A, Venskutonis P, Raynaud C, Talou T, Viškelis P and Sasnauskas A. Variations in the essential oil composition in buds of six blackcurrant (*Ribes nigrum* L.) cultivars at various development phases. *Food Chem.* 2009; 114: 671 – 679. **32.** Rohloff J, Dragland S, Mordal R and Iversen T-H. Effect of harvest time and drying method on biomass production, essential oil yield, and quality of peppermint (*Mentha piperita* L.). *Journal of Agricultural and Food Chem.* 2005; 53: 4143 - 4148.

33. Telci I and Hişil Y. Biomass yield and herb essential oil characters at different harvest stages of spring and autumn sown Coriandrum sativum. *European Journal of Horticultural Science* 2008; 73(6): 267-272.

34. Alirezaei N AM, Nemati SS, Rezvani MP and Rezazadeh S. Variation of some Phytochemical Compound in Shoot and Root of Rumex turcomanicus Czerep. at Different Phenological Stages. *Journal of Medicinal Plants* 2016; 2: 25 - 36.

35. Hussain AI, Anwar F, Sherazi STH and Przybylski R. Chemical composition, antioxidant and antimicrobial activities of basil (Ocimum basilicum) essential oils depends on seasonal variations. *Food Chemistry* 2008; 108: 986 - 995.

36. Shafaghat A. Chemical constituents, antimicrobial and antioxidant activity of the hexane extract from root and seed of Levisticum persicum Freyn and Bornm. *Journal of Medicinal Plants Res.* 2011; 5: 5127 - 5131.

37. Verma RS, Padalia RC and Chauhan A. Chemical composition variability of essential oil during ontogenesis of *Daucus carota* L. subsp. sativus (Hoffm.) Arcang. *Industrial Crops and Products* 2014; 52: 809 - 814.



www.SID.ir