Short Communication

Detection and identification of *Leishmania* isolates from patients with Cutaneous Leishmaniasis (CL) in Isfahan (central region of Iran) by PCR method

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ABSTRACT

Leishmaniasis is caused by parasitic protozoa of the genus *Leishmania*. Cutaneous leishmaniasis (CL) is a complex disease with wide spectrum of clinical manifestations. In order to identify leishmania species causing CL in Isfahan by a definite molecular technique (PCR method), this study was undertaken over 2010- 2011. 124 Patients with suspicious lesion of Leishmaniasis and positive direct smear from lesion were selected. Samples were cultured in NNN and RPMI 1640 media Negative and positive control and clinical samples was applied for PCR in the same condition. In the next step, standard PCR was carried out using classic protocol. From 124 patients, 111 (89.51%) cases were infected as *L. major* and 12 (9.67%) cases were infected by *L. tropica*, However only in one patient simultaneous infectious with both *L. major* and *L. tropica* was identified by PCR techniques which could not be possible in microscopy. *L.major* was the most prevalent species in the studied patients (p-value<0.001).

Keywords: Characterization, Cutaneous Leishmania, L. major, L. tropica, PCR

INTRODUCTION

Leishmaniasis is caused by parasitic protozoa of the genus *Leishmania*. Humans are infected via the bite of phlebotomine sandflies, which breed in forest areas, caves, or the burrows of small rodents. Cutaneous leishmaniasis (CL) is a complex disease with wide spectrum of clinical manifestations. More than 90 percent of CL cases live in the following countries: Afghanistan, Saudi-Arabia, Aljazeera, Brazil, Iran, Iraq and Syria. Meanwhile 350 million people are exposed

a number of limitations. Microscopic examination of

to the parasite. The number of new cases of CL has

reached to 1.5 million people in the word (World

Health Organization Control of leishmaniases 2010).

In Iran, CL distributes in some geographical locations

such as north-east (Hajjaran *et al* 2004), center (Sharifi *et al* 2011), west, east and south (Farahmand *et al* 2011, Fazaeli *et al* 2009). Two species of *Leishmania* are involved in CL infections in Iran. *L. major* is causative agent of zoonotic cutaneous leishmaniasis (ZCL) and *L. tropica* causes anthroponotic CL (ACL). The classic diagnostic techniques for cutaneous leishmaniasis have

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skin scrapings has limited sensitivity, particularly in chronic lesions. In vitro culture techniques are susceptible to microbiologic contamination and since certain strains grow better than others in vitro, the dominant strains can be inadvertently selected when culturing mixed infections (Navin et al 1990). The Montenegro skin test detects specific cutaneous delayed-type hypersensitivity but cannot distinguish between current and past infection. Serologic diagnostic techniques present drawbacks that include the cross-reactivity of leishmanial antigens with antibodies induced by other kinetoplastids (Armijos et al 1990) as well as poor sensitivity due to the low antibody titer characteristic of cutaneous leishmaniasis. Species identification has been conventionally achieved using isoenzyme electrophoresis (zymodeme analysis) monoclonal antibodies (serodeme (Kreutzer et al 1980). Zymodeme analysis is a lengthy and expensive process that requires large-scale cultivation of parasites. Monoclonal antibodies are useful for identification of species in cultured strains but are not as amenable to direct analysis of clinical specimens. Recently, conventional parasitological and serological techniques have been integrated by the more sensitive and specific polymerase chain reaction (PCR) assay (Grimaldi et al 1987, Leontides et al 2002, Manna et al 2008, Strauss-Ayali et al 2004, Schonian et al 2003). PCR has been shown to overcome problems such as the low sensitivity found with microscopic examination of tissue smears, and the limited predictive value of serology where the results may be affected by persistent antibodies immunosuppression (Leontides et al 2002). During the past two decades epidemiological aspects of cutaneous leishmaniasis have been increasingly changed in Iran. These changes have been motivated through the population movement between the urban and rural areas, the migration of the neighboring countries including Afghanistan and Iraq. The rural cutaneous leishmaniasis is one of the most important health challenges in Iran as in most of the rural regions; the disease is prevalent in 17 provinces of 30 provinces in

Iran. Isfahan and TurkmanSahra were the most important centers of cutaneous leishmaniasis of the villagers in Iran (Rassi *et al* 2008, Yaghoobi-Ershadi 1997).

This study has been conducted with the purpose of identifying dominant *Leishmania* parasite species in Isfahan using the PCR method because any effective control programmes should be based on the accurate baseline information about pathogen species.

MATERIALS AND METHODS

Sample collection. This study was conducted from September 2010 to September 2011. All procedures were performed under consent of patients. Sampling was accomplished by scraping doubtful cutaneous lesion of 124 patients. The research was done on the population referring to the Sidiqay Taheray center for skin disease and Leishmaniasis and population in this area with suspicious lesion of Leishmaniasis with positive direct smear from lesion was found. Patients with suspicious lesion of Leishmaniasis and positive direct smear from lesion were selected.

DNA extraction and PCR. Samples were cultured in NNN and RPMI 1640 media (John et al 2006). Total DNA was extracted from RPMI 1640 cultured samples and promastigote of Leishmania. To extract DNA, Phenol-chloroform method was applied. 500µl lysis buffer was added to parasite sediment (Surcrose 0.32 M, SDS 1%, Tris-Hcl 10mM, Mgcl2 5mM) and it was placed in water bath at 37°C. Then, an equal volume of the solution, 500µl phenol was added and it was centrifuged for 5min at 8000rpm. After adding 500µl chloroform to the supernatant and centrifuging at 8000 rpm, the supernatant was removed 0.1 of the liquid, 3 M sodium and two volumes of 100% ethanol were added and it was kept in freezer -20 °C. Then, it was centrifuged for 10 min at 12000 rpm and 100µl alcohol 80% was added and it was centrifuged for 2 min at 12000 rpm. After removing the supernatant, the sample was added for 5-10 min in incubator and then 30µl sterile deionized distilled water was added. Negative and positive control and clinical samples was applied

for PCR in the same condition. Positive control was prepared from Isfahan skin diseases and leishmaniasis research center. In the next step, standard PCR was carried out using classic protocol. Primer pair (Primer1: 5' TCGCAGAACGCCCCTACC 3' and Prime2: 5' AGGGGTTGGTGTAAAATAGGC 3') was used that generate products with size of 620 bp for L.Major and 830bp for L. tropica and was specific for conserved sequences of kDNA of Leishmania (Mahboudi et al 2001). For amplification, 2.5 µl of DNA sample was added to 22.5 µl of reaction mixture containing KCl 50 mM, Tris buffer 10 mM (pH 8.3), 0.2 mM each deoxynucleotide triphosphate, MgCl2 1.5 mM, DMSO 10.5%, tetramethyl ammonium chloride 50 mM, betaine 0.6 M, dithiothreitol 1mM, LU-5A probe 0.4 μM, 0.2 μM each 3' primer (LM-3A, LB-3C and LC-3L) and, Tag DNA polymerase 0.04 U/µl (Sigma). Amplification was incubated in a Thermolyne-Amplitron II thermal cycler, using an initial denaturation step of 95 °C x 5 min, followed by 35 cycles of 95°C x 30 s, 54 °C x 45 s, and 72°C x 30 s with a final extension at 72 °C x 5 min. Finally, 8 µl of amplification products were analyzed by electrophoresis on 1.5% agarose gel in TBE buffer (89 mM Tris borate, 2 mM EDTA, pH 8.3) containing ethidium bromide 0.5 µg/ml. The amplification products were visualized under UV light and the gels documented by EPS-600Z cameras.

RESULTS AND DISCUSSION

From 124 patients, 111 (89.51%) cases were infected as *L. major* and 12 (9.67%) cases were infected by *L. tropica*, However only in one patient simultaneous infectious with both *L. major* and *L. tropica* was identified. *L. major* was the most prevalent species in the studied patients (p-value<0.001). Epidemiological results related to the information obtain from the questionnaire and determined species showed that: from 124 patient from whom the samples were taken, 10 of them were less than 10 years old. 48 patients were in the age group of 10-19. 32 patients were between 20-29

years old.13 of them was between 30-39 and 7 patient was 40-49 years old and 14 patient was >50 years old among these ages. One month old was the least and maximum age was 65 and the mean age was 29 and the maximum pottion was in 10-19 years old group (Table 1).

Table 1. The frequency of *L. tropica* and *L. major* based on different ages.

| Infection with Ltropica and Lmajor | Infection with L tropica | Infection with L | Number of patients | Age of patients |
|------------------------------------|--------------------------|--------------------|--------------------|-----------------|
| number | number | number | | |
| 0 | 0 | 10 | 10 | 10> |
| 0 | 2 | 46 | 48 | 10-19 |
| 1 | 6 | 25 | 32 | 20-29 |
| 0 | 1 | 12 | 13 | 30-39 |
| 0 | 1 | 6 | 7 | 40-49 |
| 0 | 2 | 12 | 14 | 50> |

From 124 patient, 75 were male and 49 female, from 75 male patient 67 were infect with L. major and 8 of them infected with L. tropica and from 49 female patient 45 had L. major and 4 had L. tropica (Table 2). Out of 124 patients, 66 had lesion on hand, 32 in the leg, 18 patients had lesion in the face and neck and 8 patients had lesion in trunk (Table 3). Leishmaniasis is caused by parasitic protozoa of the genus Leishmania and it is essential to identify the species of the parasite in epidemiological and clinical studies. All the species are similar physically (even by electron microscope) except a little difference in the size of the species. According to the considerable studies performed, it was defined that various species (despite the physical similarity), they are different in terms of molecular, biochemical and antigenic characteristics. Advance techniques as PCR are proposed to identify various species. PCR is more precise compared to the microscopic observation of blood extension and other methods in identification of *Leishmania* species.

Table 2. The frequency of *L. tropica* and *L. major* among males and females.

| Infection with L.tropica Infection with L. tropica and L.major | | Infection with <i>L.major</i> | Number of patients | sex |
|---|--------|----------------------------------|--------------------|------|
| number | number | number | • | |
| 0 | 8 | 67 | 75 | male |
| | | | | |

PCR method is precise in discriminating pathogenic and non-pathogenic species. Brustoloni et al. 2007 evaluated the sensitivity and specificity of PCR method compared to other experimental methods. PCR showed the highest sensitivity (92.3%) and had good specificity (97.5%) (Brustoloni *et al* 2007).

Table 3. The frequency of *L.tropica* and *L.major* based on different part of body.

| different j | part of body | ٧. | | | |
|--|--------------------------|-------------------------|--------------------|---------------|-----------------|
| Infection with <i>L. tropica</i> and <i>L. major</i> | Infection with L. ropica | Infection with L. major | Number of patients | | Lesion location |
| number | number | number | _ | | |
| 1 | 5 | 60 | 66 | hand | |
| 0 | 3 | 29 | 32 | leg | |
| 0 | 0 | 8 | 8 | trunk | |
| 0 | 4 | 14 | 18 | Face and neck | |
| | | | | | |



Figure 1. Polymerase chain reaction (PCR) profiles obtained from the *Leishmania* stocks: Lane1 negative control; lane 2 positive control for *L. tropica*; lane 3 positive control for *L.major*; Lane 4,5,6,10,11 are *L.major*; lane 8, 9 are *L.tropica*; lane7 is infection with both *L. tropica and L. major*.

As it was shown in frequency distribution of the patients in age group, 10 of the subjects were less than 10 years old. 48 patients were in the age group of 10-19. 32 patients were between 20-29 years old.13 of them was between 30-39 and 7 patients were 40-49 years old and 14 patients were>50 years old among these ages. From 124 patients, 75 were male and 49 female. Out of 124 patients, 66 people had lesion on hand, 32 in the leg. 18 patients had lesion in the face and neck and 8 patients had lesion in trunk. The major injury was observed in the hands and the result was consistent with the results of the study performed by Shiie et al. 2012. By the investigation of 137 infected subjects showed that there were 74 (54%) men and 63 (46%) women. The results of the study showed that the mean lesion was 2 and the infection duration was 97 days and 62.8 % of the cases were dedicated to lesion on hand (Shiee et al 2012). Azizi et al. 2012 by microscopic and Nested-PCR detected the smear species of cutaneous leishmaniasis. They showed that the only smear species in southern province of Iran was L.major (Azizi et al 2012). Based on the results of amastigot species by PCR, from 124 patients, 111 (89.51%) cases were infected as L.major and 12 (9.67%) cases were infected by L.tropica, However only one patient (0.8%) was infected by both L.major and L.tropica. The results of the present study showed that L.major was the most prevalent species and it showed that L.tropica is local in Isfahan region. It can be said that population changes, migration and climatic changes caused that clinical forms of the disease were different. Mesgarian et al. 2010, applied PCR method to detect Leishmania species

in Gonbad Kavus. The results of the study showed that the tissues of the patients with cutaneous leishmaniasis were L.major (Mesgarian et al 2010). Rahbarian et al. 2009 in Gonbad-e Qabus, among 6990 inhabitants of 5 villages, 62.9% were identified as scars and 1.5% as active lesions. Individuals 11 to 20 years were the most highly infected age group. All the parasite isolates were Leishmania major. (Rahbarian et al 2009). In a study performed in Mashhad in 2010, of total 21 samples, 19 cases were L.tropica and 2 cases were L.major. The study showed that besides L.tropica species, L.major species is observed but L.tropica was prevalent (Mahmoodi et al 2010). The determination of the species of Leishmania in epidemiological studies to determine the main vector, definite source and human infection and their relationship with each other, the evaluation of different vaccines provided against leishmaniasis, the determination of treatment effect of various methods and the selection of a good strategy to control and prevent leishmaniasis are necessary.

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References

- Armijos, R.X., Chico, M.E., Cruz, M.E., Guderian, R.H., Kreutzer, R.D., Berman, J.D. et al. (1990). Human cutaneous leishmaniasis in Ecuador: identification of parasites by enzyme electrophoresis. *American Journal of Tropical Medicine and Hygiene* 42:424.
- Azizi, K., Soltani, A., Alipour, H. (2012). Molecular detection of Leishmania isolated from cutaneous leishmaniasis patients in Jask County, Hormozgan Province, Southern Iran. *Asian Pacific Journal of Tropical Medicine* 514-517.
- Brustoloni, Y.M., Lima, R.B., Cunha, R.V., Dorval, M.E., Oshiro, E.T., Oliveira, A.L., Pirmez, C. (2007). Sensitivity and specificity of PCR in Giemsa-stained slides for diagnosis of visceral leishmaniasis in children. *Memórias do Instituto Oswaldo Cruz* 102(4):497-500
- Farahmand, M., Nahrevanian, H., Atashi Shirazi, H., Naeimi, S., Farzanehnejad, Z. (2011). An overview of a diagnostic

- and epidemiologic reappraisal of cutaneous leishmaniasis in Iran. *Brazilian Journal Infectious Diseases* 15(1): 17-21.
- Fazaeli, A., Fouladi, B., Sharifi, I. (2009). Emergence of cutaneous leishmaniasis in a border area at south-east of Iran: an epidemiological survey. *Journal Vector Borne Disease* 46(1): 36-42.
- Grimaldi, Jr.G., David, J.R., McMahon-Pratt, D. (1987). Identification and distribution of New World Leishmania species characterized by serodeme analysis using monoclonal antibodies. *The American Journal of Tropical Medicine and Hygiene* 36:270.
- Hajjaran, H., Mohebali, M., Razavi, M.R., Rezaei, S., Kazemi, B., Edrissian, G.h.H., et al. (2004). Identification of Leishmania species isolated from human cutaneous leishmaniasis, using Random Amplified Polymorphic DNA (RAPD-PCR). *Iranian Journal of Public Health* 33:8-15.
- John DT, Petri WA, Petri Jr WA, Markell EK, Voge M. *Markell and Voge's medical parasitology*: Saunders; 2006.
- Kreutzer, R.D., Christensen, H.A. (1980). Characterization of Leishmania spp. by isozyme electrophoresis. *The American Journal of Tropical Medicine and Hygiene* 29:199.
- Leontides, L.S., Saridomichelakis, M.N., Billinis, C., Kontos, V., Koutinas, A.F., Galatos, A.D., et al.(2002). A cross-sectional study of Leishmania spp. infection in clinically healthy dogs with polymerase chain reaction and serology in Greece. *Veterinary Parasitology* 109:19-27.
- Mahboudi, F., Abolhassani, M., Yaran, M., et al. (2001). Identification and Differentiation of Iranian Leishmania Species by PCR Amplification of kDNA. *Scandinavian Journal of Infectious Diseases* 33: 596–598.
- Mahmoodi, M.R., Tavakoli Afshar, J., Mohajeri, M., et al. (2010). Molecular Identification of Leishmania Species Causing Cutaneous Leishmaniasis in Mashhad, Iran. *Scientific Journal of Ilam University of Medical Sciences* 18(2):17-23.
- Manna, L., Reale, S., Vitale, F., Picillo, E., Pavone, L.M., Gravino, A.E. (2008).Real-time PCR assay in Leishmania-infected dogs treated with meglumine antimoniate and allopurinol. *Veterinary Journal* 177:279-82.
- Mesgarian, F., Rahbarian, N., Mahmoudi Rad, M., Hajaran, H. et al. (2010). Identification of Leishmania species isolated from human cutaneous Leishmaniasis in Gonbade-Qabus city using a PCR method during 2006-2007. *Tehran University Medical Journal* 68(4):250-256.
- Navin, T.R., Arana, B.A., Arana, F.E., Merida, A.M., Castillo, A.L., Pozuelos, J.L. (1990) Placebo-controlled clinical trial of meglumine antimonate (glucantime) vs.

- localized controlled heat in the treatment of cutaneous leishmaniasis in Guatemala. *The American Journal of Tropical Medicine and Hygiene* 42:43.
- Rahbarian, N., Mesgarian, A., Mahmoudi Rad, M., Hajaran, H., Shahbazi, F., Mesgarian, Z. et al. (2009). Identification of Leishmania Species Isolated from Human Cutaneous Leishmaniasis Using PCR Method. *Journal of Research in Health Sciences* 9:48-51.
- Rassi, Y., Sofizadeh, A., Abai, M., Oshaghi, M., Rafizadeh, S., Mohebail, M., et al. (2008). Molecular Detection of Leishmania major in the Vectors and Reservoir Hosts of Cutaneous Leishmaniasis in Kalaleh District, Golestan Province, Iran. *Iranian Journal of Arthropod-Borne Diseases* 2(2):21-7.
- Schonian, G., Nasereddin, A., Dinse, N., Schweynoch, C., Schallig, H., Presber, W., et al. (2003). PCR diagnosis and characterization of Leishmania in local and imported clinical samples. *Diagnostic Microbiology & Infectious Disease* 47:349-58.
- Sharifi, I., Fekri, AR., Aflatoonian, MR., Khamesipour, A., Leishmania tropica in rural communities of Bam district

- after the earthquake, Iran. *Tropical Medicine & International Health* 16(4): 510-513.
- Mahboudi, F., Dowlati, Y., et al. (2011). Emergence of a new focus of anthroponotic cutaneous leishmaniasis due to Shiee, M.R., Hajjaran, H., Mohebali, M., Doroodgar, A., et al. (2012). A molecular and parasitological survey on cutaneous leishmaniasis patients from historical city of Kashan in Isfahan province, center of Iran. *Asian Pacific Journal of Tropical Disease* 421-425.
- Strauss-Ayali, D., Jaffe, C.L., Burshtain, O., Gonen, L., Baneth, G. (2004). Polymerase chain reaction using noninvasively obtained samples, for the detection of Leishmania infantum DNA in dogs. *Journal of Infectious Diseases* 189:1729-33.
- World Health Organization Control of leishmaniases. Technical report series 949 of WHO Expert Committee, Geneva, 2010.
- Yaghoobi-Ershadi, M., Javadian, E. (1997). Studies on sandflies in a hyperendemic area of zoonotic cutaneous leishmaniasis in Iran. *The Indian Journal of Medical Research* 105:61-6.