



Research Article



Study the Efficacy of Antimicrobial Activities of Eight Clinically Applied Disinfectants against Clinical Isolated of Enterococci and *Pseudomonas aeruginosa*

Morteza Zareniya¹, Somayeh Hallaj-Nezhadi², Farideh Dinmohamadi¹, Fakhri Haghi³, Maryam Hassan^{1*}

¹Zanjan Pharmaceutical Biotechnology Research Center, Zanjan University of Medical Sciences, Zanjan, Iran.

²Drug Applied Research Center and Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran.

³Department of Microbiology, Zanjan University of Medical Sciences, Zanjan, Iran.

Article Info

Article History:

Received: 22 July 2016

Accepted: 14 December 2016

ePublished: 30 June 2017

Keywords:

-Hospital disinfectants -
 Enterococci -*Pseudomonas*
aeruginosa -Clinical
 isolates -Antibiogram

ABSTRACT

Background: Hospital-acquired infections are among the most significant reasons of human mortality world-wide which can be controlled by efficient application of suitable disinfectant for hospital setting. The main goal of the present study was to determine the efficacy of eight routinely used hospital disinfectants against clinical isolates.

Methods: In our descriptive study, in the first step the antibiogram assay of 99 clinical isolates enterococci and *Pseudomonas aeruginosa* were determined. Then, minimum inhibitory concentration and minimum bactericidal concentration of isolates against Povidone Iodine 10%, Ethanol 70%, Savlon 3.2%, Deconex51Gastro, Procept Floor, Septo med, Surfanious and Gigasept AF were evaluated. Furthermore, the efficacy of disinfectants was reevaluated in presence of 5% (w/v) Bovine Serum Albumin.

Results: The results showed that Septo med and Surfanious are the most and less potent disinfectants against clinical isolates, respectively. It is also resulted that Povidone Iodine is the most effective choice among the conventional disinfectants in this study. Clearly, addition of 5% organic substances reduced the efficacy of selected disinfectants significantly.

Conclusion: Novel quaternary ammonium compounds are the most applicable choice for disinfection of hospital surfaces and instruments in this study.

Introduction

Resistance to antimicrobial agents among bacteria is one the major world-wide challenge since the discovery of penicillin. Biocides are from antimicrobial agents which are divided to antiseptics, disinfectants and preservatives. The resistance level of various groups of bacteria to biocides were studied and characterized as well. Bacterial spores, mycobacteria and Gram-negative bacteria are insusceptible to biocides due to the presence of impermeable cell layers. Gram-positive bacteria, especially cocci are the most vulnerable types of microbes to biocides.^{1,2}

Enterococcus spp. which are resistant to antimicrobial agents play an important role as nosocomial pathogens in hospital outbreaks.³ As US nosocomial infections claimed, enterococcal

disease ranked in among 3 or 4 most prevalent hospital-acquired disease. It is clear that enterococci are notable among hospitalized patients due to their pathogenicity by carrying virulence factors. Enterococcal urinary tract infections (UTIs), bacteremia and uncomplicated wound infections are the most common and meningitis as well as endocarditis is less frequent infections caused by these microorganisms.^{4,5}

P. aeruginosa has emerged as an important cause of nosocomial infections among hospitalized patients, including pneumonia (hospital-acquired, healthcare-associated and ventilator-associated), burn infections, UTIs, meningitis and bacteremia. Unfortunately, developing antimicrobial resistance in *P. aeruginosa* due to acquisition of resistance genes on plasmids as well as intrinsic resistance

*Corresponding Author: Maryam Hassan, E-mail: mhassan@zums.ac.ir

©2017 The Authors. This is an open access article and applies the Creative Commons Attribution (CC BY), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited. No permission is required from the authors or the publishers.

make treatment of severe infections of *P. aeruginosa* quite problematic. Hence, selection of suitable antimicrobial agents is essential in health-care centers.^{4,6,7}

In hospitals and health care facilities, biocides are widely used to prevent and control hospital-acquired infections. Disinfectants and antiseptics are applied to strongly decontaminate the microbial count of surfaces (solid surfaces and skin). However, there is a growing concern regard to maintaining the sanitary condition in hospitals. Because of the emergence of resistant bacteria to different antibiotics and silver compounds as well as widespread usage of disinfectants in hospitals, more researches are necessary to evaluate the efficacy of active biocidal substances not only on standard bacteria but also on clinical isolates.^{4,8}

Furthermore, presence of organic materials such as mucous membranes and wounds is inevitable in health-care settings and as it is evidenced, the effectiveness of most of biocidal agents would reduce in contact to organic substances. Hence, the influence of such substances should be assessed in practical use of biocides.^{9,10} There are many surveys on antibiotic resistance of bacteria while less research was focused on resistance to biocides. Therefore, the main goal of the present study was to determine and evaluate susceptibility of isolated enterococci and *P. aeruginosa* from hospitals to eight widely used disinfectants and biocides, which routinely used for disinfection of skins, surfaces, floors and facilities in the same medical centers both in presence and without organic substances.

Materials and Methods

Bacterial isolates

In our descriptive study, fifty isolates of enterococci and forty-nine isolates of *P. aeruginosa* were obtained from patients who were hospitalized at two state hospitals of Zanjan city of Iran from August 2012 to May 2013. Enterococci as well as *P. aeruginosa* were identified according to standard bacteriologic methods. All of enterococcal samples were recultured at our laboratory by incubated on bile esculin sodium azide agar for 18 h at 37 °C.¹¹ Samples of *P. aeruginosa* were reidentified by cultured on cefrimide agar and P-agar.^{12,13} In the next step, the single colonies saved in 30% glycerol at -80 °C as stocks.

Antimicrobial susceptibility pattern of clinically isolated enterococci and *P. aeruginosa*

The susceptibility pattern of isolated enterococci and *P. aeruginosa* were evaluated by paper disc method.^{14,15} Ten µl of (0.5 McFarland) enterococcal suspension was spread on surface of Mueller-Hinton agar plates. Then, the following discs of antibiotic purchased from Himedia, India were applied for enterococci: vancomycin (30 µg),

ampicillin (10 µg), ciprofloxacin (5 µg), gentamicin (10 µg), chloramphenicol (30 µg), erythromycin (15 µg), and streptomycin (10 µg). The antibiotics used for determination of susceptibility pattern of *P. aeruginosa* were tobramycin (10 µg), gentamicin (10 µg), ceftazidime (30 µg), imipenem (10 µg), ticarcillin (75 µg), levofloxacin (5 µg), kanamycin (30µg), piperacillin (100 µg) and aztreonam (30 µg). Subsequently, inhibition zones around disks were measured after incubation for 18 -24 hours. Quality control of tests was done by standard strains of *P. aeruginosa* PTCC 1310 and *E. faecalis* PTCC 1237.

Disinfectants and susceptibility testing

Eight different disinfectants which were used broadly in Zanjan state hospitals were obtained: Povidone Iodine 10% (PI), Ethanol 70% (Et), Savlon 3.2% (Sa), Deconex51Gastro (DG), Procept Floor (PF), Septo med (Sm), Surfanious(Sf) and Gigasept AF (Gi). The MICs of the above mentioned disinfectants were evaluated by broth micro-dilution method (microtiter assay) with reference to the protocol of the CLSI.¹⁵

Determination of bactericidal activity: MBCs

The MBCs were measured by plating and incubating 10 µl of four final clear diluted wells of each disinfectant at 37° C for 48 hours. MBC was defined as minimum concentration of disinfectants, which killed desired microorganisms.¹⁰

Measurement of bactericidal activities in the presence of organic material

The MBC test was repeated as described above in presence of 5% (w/v) Bovine Serum Albumin. It was applied to simulate presence of organic materials and evaluate bactericidal activity. The experiments were repeated three times on different days.¹⁰

Results

The antibiotic resistance pattern of 50 enterococcal strains isolated from patients was tested 82% of isolates were resistant to up to seven applied antibiotics. Regarding resistance pattern of antibiotics, seven different resistance patterns were shown in this survey. As shown in graph 1, 41 (82%) of enterococcal isolates were multi-drug resistant. Interestingly, the lowest prevalence of multi-drug resistance was 10% to three antibiotics. Resistance to five antibiotics (30%) was widely distributed among isolates, followed by 20% to four and 16% to two. As it is obvious from Figure 1 80% of *P. aeruginosa* were multi-drug resistant. Interestingly, 30% of isolates were resistant to 5 out of 9 different antibiotics. The most important result to get from the data is that 10% of isolates were resistant to almost all of the tested antibiotics.

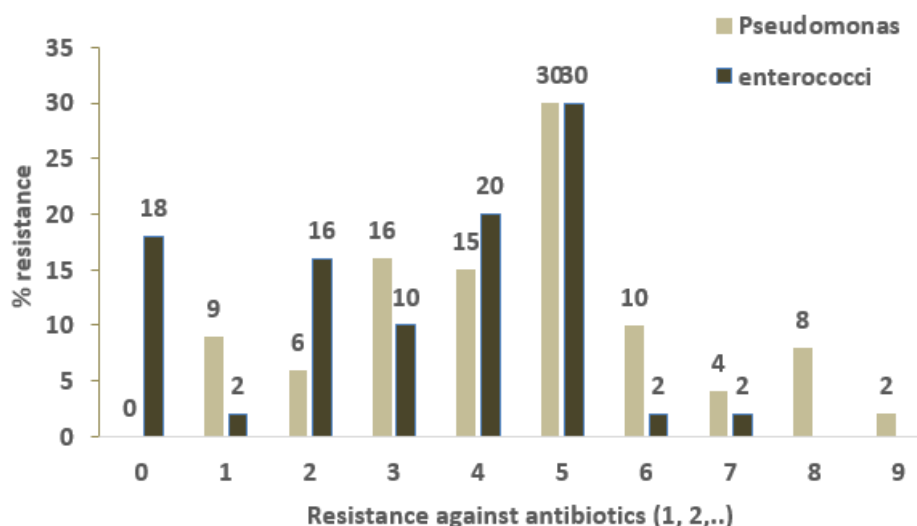


Figure 1. Antimicrobial resistance pattern of *Enterococcus* spp. and *Pseudomonas aeruginosa* isolated from clinical samples.

Table 1. Distribution of MIC (%), MBC (%) and MBCal (%) of various disinfectants by microtiter method.

Disinfectant	Number of strains at each MIC (%) of disinfectant											
	3.1×10^{-2}	1.5×10^{-2}	7.8×10^{-3}	3.9×10^{-3}	1.9×10^{-3}	9.7×10^{-4}	4.8×10^{-4}	2.4×10^{-4}	1.2×10^{-4}	6.1×10^{-5}	3×10^{-6}	1.5×10^{-5}
Sf -P	0	2	3	3	11	19	6	2	0	0	0	0
Sf -E	0	2	1	5	14	15	9	3	2	0	0	0
PF-P	1	0	1	1	0	10	16	14	0	0	0	0
Pf-E	0	0	0	0	2	8	21	11	6	2	0	0
Gi-P	0	1	1	3	6	18	13	4	0	0	0	0
Gi-E	0	0	0	0	8	13	17	9	2	1	0	0
DG-P	0	1	2	2	6	12	16	5	0	0	0	0
DG-E	0	0	0	2	7	9	17	9	4	2	0	0
Sm-P	0	0	0	1	2	1	2	3	14	14	6	1
Sm-E	0	0	0	0	0	1	3	4	20	13	6	3

Disinfectant	Number of strains at each MBC (%) of disinfectant											
	6.2×10^{-2}	3.1×10^{-2}	1.5×10^{-2}	7.8×10^{-3}	3.9×10^{-3}	1.9×10^{-3}	9.7×10^{-4}	4.8×10^{-4}	2.4×10^{-4}	1.2×10^{-4}	6.1×10^{-5}	3×10^{-6}
Sf -P	0	4	9	17	14	3	0	0	0	0	0	0
Sf -E	3	4	14	16	10	2	1	0	0	0	0	0
PF-P	0	2	2	10	17	11	4	1	0	0	0	0
Pf-E	0	0	2	8	17	13	7	3	0	0	0	0
Gi-P	0	2	5	16	17	6	1	0	0	0	0	0
Gi-E	0	0	5	12	18	12	2	0	0	0	0	0
DG-P	0	3	6	14	14	8	2	0	0	0	0	0
DG-E	0	2	7	9	16	9	6	1	0	0	0	0
Sm-P	0	0	0	2	3	6	10	21	5	0	0	0
Sm-E	0	0	0	1	2	4	17	17	5	4	0	0

Disinfectant	Number of strains at each MBC al (%) of disinfectant											
	1.2×10^{-1}	6.2×10^{-2}	3.1×10^{-2}	1.5×10^{-2}	7.8×10^{-3}	3.9×10^{-3}	1.9×10^{-3}	9.7×10^{-4}	4.8×10^{-4}	2.4×10^{-4}	1.2×10^{-4}	
Sf -P	1	4	10	16	14	2	0	0	0	0	0	0
Sf -E	1	4	4	19	18	2	2	0	0	0	0	0
PF-P	0	2	4	10	15	12	4	0	0	0	0	0
Pf-E	0	0	3	7	13	18	7	2	0	0	0	0
Gi-P	0	2	6	21	13	5	0	0	0	0	0	0
Gi-E	0	1	5	8	20	13	3	0	0	0	0	0
DG-P	0	3	3	14	17	7	3	0	0	0	0	0
DG-E	0	2	6	8	15	12	6	1	0	0	0	0
Sm-P	0	0	2	5	7	11	16	5	1	0	0	0
Sm-E	0	0	1	2	4	15	19	6	4	0	0	0

Gastro (DG), Procept Floor (PF), Septo med (Sm), Surfanious (Sf) and Gigasept AF (SG), (P) *Pseudomonas*, (E) *enterococci*.

Table 2. Distribution of MBCs of various disinfectants by Agar-plate method.

Disinfectant	Number of strains at each MIC (%) of disinfectant						
	35	17.5	8.75	4.3			
Et-P	2	12	29	3			
Et-E	3	18	25	4			
	4×10^{-1}	2×10^{-1}	1×10^{-1}	5×10^{-2}	2.5×10^{-2}	1.2×10^{-2}	6.2×10^{-3}
Sa-P	2	5	11	21	6	2	0
Sa-E	0	2	3	16	16	10	3
	1.25	6.2×10^{-1}	3.1×10^{-1}	1.5×10^{-1}	7.8×10^{-2}	3.9×10^{-2}	
PI-P	2	6	22	15	2	0	
PI-E	0	4	17	19	8	2	

Disinfectant	Number of strains at each MBC (%) of disinfectant						
	70	35	17.5	8.75			
Et-p	15	23	8	1			
Et-E	12	32	5	1			
	8×10^{-1}	4×10^{-1}	2×10^{-1}	1×10^{-1}	5×10^{-2}	2.5×10^{-2}	1.2×10^{-2}
Sa-p	1	3	14	20	7	2	0
Sa-E	0	1	3	16	22	6	2
	2.5	1.25	6.2×10^{-1}	3.1×10^{-1}	1.5×10^{-1}	7.8×10^{-2}	
PI -p	2	5	28	10	2	0	
PI-E	0	2	18	21	7	2	

Disinfectant	Number of strains at each MBC al (%) of disinfectant						
	70	35	17.5	8.75			
Et-P	43	4	0	0			
Et-E	44	6	0	0			
	8×10^{-1}	4×10^{-1}	2×10^{-1}	1×10^{-1}	5×10^{-2}	2.5×10^{-2}	1.2×10^{-2}
Sa-p	2	12	25	7	1	0	0
Sa-E	1	2	16	24	6	1	0
	5	2.5	1.25	6.2×10^{-1}	3.1×10^{-1}	1.5×10^{-1}	7.8×10^{-2}
PI -p	1	7	22	14	3	0	0
PI-E	0	1	17	21	9	2	0

Povidone Iodine 10% (PI), Ethanol 70% (Et), Savlon 3.2% (Sa), (P) Pseudomonas, (E) enterococci.

It was understood from Table 1 and 2 that the distribution of MICs of DG, PF, Sm, Sf, SG, PI, Et and Sa for 100 clinical isolates of enterococci and *P. aeruginosa*. From the data in Table 1, it is apparent that the hospital disinfectants are significantly more killing on isolated enterococci than *P. aeruginosa*. As Table 1 shows, Sm is the most potent disinfectant which strongly inhibits the growth of clinical isolates. Furthermore, the growth of approximately 78% of isolates was inhibited at 9.7×10^{-4} % by the rest of disinfectants except Sm. However, Sf is the less potent ones among hospital disinfectant. It can be understood from Table 2 that PI is the most effective choice among the conventional disinfectants in this study.

The bactericidal activity of eight selected disinfectants was evaluated by agar plate assay after addition of neutralizer to the microtiter plate. The data of MBCs from Table 1 and 2 can be compared with data of MICs in Table 1 and 2, which shows that bactericidal concentration is approximately eight times less than inhibitory concentrations. Interestingly, Sm revealed high potency for bactericidal activity against clinical isolates.

The comparison of bactericidal activity of the eight disinfectants for enterococci and *P. aeruginosa*

under the so-called conditions (clean and dirty) showed that the MBC values were increased up to half of the in-use concentrations in most of the disinfectants in presence of 5% BSA (Table 1 and 2). However, there was no significant difference between MBC of DG, GI and PF in dirty and clean conditions for isolated enterococci.

Discussion

Nosocomial infections caused by enterococci and *P. aeruginosa* are the main problems in hospitals since 1980 and as a result of this issue medical expenses are a burden for health care systems.¹⁶ The studies show that efficacy of antibiotics and disinfectants are gradually reduced.^{17,18} Inappropriate usage, inaccurate concentration and lack of sufficient training for preparation and storage of hospital disinfectants are among the important reasons of prevalence of resistance of disinfectants.¹⁸⁻²⁰ In comparison to many antimicrobial resistance researches about antibiotics, there is not so many global researches regard to resistance to biocides. Because of the clinical importance of enterococci and *P. aeruginosa* the efficacy of eight hospital disinfectants were evaluated against clinical isolates of enterococci and *P. aeruginosa*.

The results showed that Gram-positive bacteria in this study (enterococci) were more susceptible to disinfectants in comparison to Gram-negative bacteria (*P. aeruginosa*). The outer membrane of Gram-negative bacteria inhibits or seriously reduces the penetration of molecules of disinfectants into cells.²¹ Interestingly, *pseudomonas* genus contains structural factors, enzymes and toxins which are the main reasons of resistance to antibiotics and disinfectants.²²

Septomed showed the strongest bactericidal activity against isolated enterococci and *P. aeruginosa* followed by DG, PF and Gi.

Overall, ethanol exhibited the highest MBC against isolates. High MBC in this study corroborates earlier findings by Mansouri (2006), Mitiku (2014), Alkolaibe (2015) which suggested ethanol as weakest disinfectant in comparison to cetrimide-C and Betadin.²³⁻²⁵

As it is obvious, the best activity was observed from new generation of disinfectants. This finding is in agreement with Amini's (2012), Saharkhizan's (2014) and Azma's (2015) findings, which found QACs such as Deconex, Descocid and Decocept as strongest antibacterial agents in clinic, therefore it is highly recommended that QACs should be applied for disinfection of instruments and critical surfaces in hospitals.²⁶⁻²⁸

These compounds were manufactured in 1990s and today they considered as compounds which are strongly effective against types of microorganisms. Interestingly, they have several unique characteristics such as broad-spectrum activity, without smell, colorless, heat resistant, with low toxicity and considered as very good detergents.

It is interesting to note that as previous studies showed the rate of resistance in Zanjan hospitals is notably low in comparison to hospitals of other cities of Iran such as Uromiyeh, Tabriz, Tehran or other countries such as Cuba and Pakistan.²⁹⁻³³ Our investigations hypothesized that there are some main reasons regard to this occurrence. Firstly, majority of disinfectants which are prepared and used in hospitals of Zanjan are strong and fast acting ones. Secondly, the staffs changed the type of disinfectants used for different purposes (facilities, instruments, floors, etc.) every 6-9 months. Thirdly, maintaining of appropriate and sufficient education for precise preparation and practical application of disinfectants to the staffs.

Conclusion

In conclusion, the results of MIC and MBC of applied disinfectants in two state hospitals of Zanjan against clinical isolates of enterococci and *P. aeruginosa* showed that Septomed (Sm) is the best antimicrobial agents applied in two state hospitals of Zanjan and conventional antimicrobial agents such as Povidone Iodine 10%, Ethanol 70%

and Savlon 3.2% are amongst the less effective ones.

It is recommended that further research be undertaken to evaluate the applicability of these disinfectants against other important clinical microorganisms such as *Escherichia coli* and *Staphylococcus aureus*. Furthermore, it would be interesting to assess required contact time for each disinfectant.

Acknowledgments

This work was supported fully by Pharmaceutical Biotechnology Research Center, Zanjan University of Medical Sciences, Zanjan, Iran. This is a report of a database from thesis entitled "Inhibition evaluation of commercial disinfectants against enterococci and pseudomonas isolated from clinical samples" registered in Zanjan University of Medical Sciences.

Conflict of interests

The authors claim that there is no conflict of interest.

References

- McDonnell G, Russell AD. Antiseptics and disinfectants: Activity, action, and resistance. *Clin Microbiol Rev.* 1999;12(1):147-79.
- Hassan M, Kjos M, Nes IF, Diep DB, Lotfipour F. Natural antimicrobial peptides from bacteria: Characteristics and potential applications to fight against antibiotic resistance. *J Appl Microbiol.* 2012;113(4):723-36. doi:10.1111/j.1365-2672.2012.05338.x
- Hassan M, Javadzadeh Y, Lotfipour F, Badomchi R. Determination of comparative minimum inhibitory concentration (mic) of bacteriocins produced by enterococci for selected isolates of multi-antibiotic resistant enterococcus spp. *Adv Pharm Bull.* 2011;1(2):75-9. doi:10.5681/apb.2011.011
- Abreu AC, Tavares RR, Borges A, Mergulhao F, Simoes M. Current and emergent strategies for disinfection of hospital environments. *J Antimicrob Chemother.* 2013;68(12):2718-32. doi:10.1093/jac/dkt281
- Kampf G, Hofer M, Wendt C. Efficacy of hand disinfectants against vancomycin-resistant enterococci in vitro. *J Hosp Infect.* 1999;42(2):143-50. doi:10.1053/jhin.1998.0559
- Russell AD. Bacterial resistance to disinfectants: Present knowledge and future problems. *J Hosp Infect.* 1999;43:S57-68. doi:10.1016/s0195-6701(99)90066-x
- Fonseca EL, Vieira VV, Cipriano R, Vicente AC. Class 1 integrons in pseudomonas aeruginosa isolates from clinical settings in amazon region, brazil. *FEMS Immunol Med*

- Microbiol. 2005;44(3):303-9. doi:10.1016/j.femsim.2005.01.004
8. Namba Y, Suzuki A, Takeshima N, Kato N. Comparative study of bactericidal activities of six different disinfectants. Nagoya J Med Sci. 1985;47(3-4):101-12.
 9. Khedmat S, Aligholi M, Sadeghi S. Influence of bovine serum albumin on the antibacterial activity of endodontic irrigants against enterococcus faecalis. Iran Endod J. 2009;4(4):139-43.
 10. Kawamura-Sato K, Wachino J, Kondo T, Ito H, Arakawa Y. Reduction of disinfectant bactericidal activities in clinically isolated acinetobacter species in the presence of organic material. J Antimicrob Chemother. 2008;61(3):568-76. doi:10.1093/jac/dkm498
 11. Hassan M, Diep DB, Javadzadeh Y, Dastmalchi S, Nes IF, Sharifi Y, et al. Prevalence of bacteriocin activities and bacteriocin-encoding genes in enterococcal clinical isolates in iran. Can J Microbiol. 2012;58(4):359-68. doi:10.1139/w11-136
 12. Peymani A, Naserpour Farivar T, Mohammadi Ghanbarlou M, Najafipour R. Dissemination of pseudomonas aeruginosa producing bla imp-1 and bla vim-1 in qazvin and alborz educational hospitals, iran. Iran J Microbiol. 2015;7(6):302-9.
 13. Consuelo M, Lehman D, Manuselis G. Textbook of diagnostic microbiology. 5th ed. New York:Saunders; 2014.
 14. Abamecha A, Wondafrash B, Abdissa A. Antimicrobial resistance profile of enterococcus species isolated from intestinal tracts of hospitalized patients in jimma, ethiopia. BMC Res Notes. 2015;8(1):213. doi:10.1186/s13104-015-1200-2
 15. Clinical and laboratory standards institute. Performance standards for antimicrobial susceptibility testing: Seventeenth informational supplement m100-s23 USA: Wayne, PA; 2013.
 16. Tripathi A, Shukla SK, Singh A, Prasad KN. Prevalence, outcome and risk factor associated with vancomycin-resistant enterococcus faecalis and enterococcus faecium at a tertiary care hospital in northern india. Indian J Med Microbiol. 2016;34(1):38-45. doi:10.4103/0255-0857.174099
 17. Suleyman G, Zervos MJ. Safety and efficacy of commonly used antimicrobial agents in the treatment of enterococcal infections: A review. Expert Opin Drug Saf. 2016;15(2):153-67. doi:10.1517/14740338.2016.1127349
 18. Rayner D. MRSA: An infection control overview. Nurs Stand. 2003;17(45):47-53. doi:10.7748/ns2003.07.17.45.47.c3424
 19. Dettenkofer M, Wenzler S, Amthor S, Antes G, Motschall E, Daschner FD. Does disinfection of environmental surfaces influence nosocomial infection rates? A systematic review. Am J Infect Control. 2004;32(2):84-9. doi:10.1016/j.ajic.2003.07.006
 20. Di Muzio M, Cammilletti V, Petrelli E, Di Simone E. Hand hygiene in preventing nosocomial infections: A nursing research. Ann Ig. 2015;27(2):485-91. doi:10.7416/ai.2015.2035
 21. Sheldon AT. Antiseptic resistance: What do we know and what does it mean? Clin Lab Sci. 2005;18(3):181-7.
 22. Khan FZ, Khan A, Kazmi SU. Prevalence and susceptibility pattern of multi drug resistant clinical isolates of pseudomonas aeruginosa in karachi. Pak J Med Sci. 2014;30(5):951-4. doi:10.12669/pjms.305.5400
 23. Mansouri SH, Moshafi MH, Nojumi F. Inhibitory effects of povidone- iodine and cetrimide-c against antibacterial resistance isolates of escherichia coli and enterococci. J Kerman Uni Me Sci. 2006;13(3):152-8.
 24. Mitiku M, Ali S, Kibru G. Antimicrobial drug resistance and disinfectants susceptibility of pseudomonas aeruginosa isolates from clinical and environmental samples in jimma university specialized hospital, southwest ethiopia. Am J Biomed Life Sci. 2014;2(2):40-5. doi:10.11648/j.ajbls.20140202.12
 25. Alkolaibea AM, AL-Ameri GA, Alkadasi MN, Zaid AA. Study of the efficacy of disinfectant against bacterial contamination in burns unit – alqumhory and international yemen hospitals in taiz city. Int J Res Stud Biosci. 2015;3(3):26-33.
 26. Amini F, Yunesian M, Dehghani MH, Jazani NH, Nodehi RN, Arjomandi MM. Comparison of antiseptics' efficacy on pseudomonas aeruginosa, staphylococcus epidermidis and enterobacter aeruginosa in hospital of imam khomeini (urmia). Iran. J. Health & Environ. 2012;5(1):88-97.
 27. Sharkhizan M, Yousefi Mahouf R, Balalifard S, Esmaeili R. Evaluation of efficacy of new disinfectants of sanosil, alprocide, bibfort and javel-dose compared with micro 10 and deconex on isolated organisms from dentistry units. Pajouhan Sci J. 2014;12(4):43-9.
 28. Azma E, Sadeghi Khanjani M, Kazemnejad Leili E, Baghernia M. Comparison of the antimicrobial effects of iranian disinfectant dissept with disinfectants helvemed forte and micro10 enzyme. J Mash Dent Sch. 2015;39(1):35-42.
 29. Sharifi Y, Hasani A, Ghotaslou R, Naghili B, Aghazadeh M, Milani M, et al. Virulence and antimicrobial resistance in enterococci isolated from urinary tract infections. Adv Pharm Bull. 2013;3(1):197-201. doi:10.5681/apb.2013.032

30. Medell M, Hart M, Batista ML. [in vitro antimicrobial susceptibility in enterococcus faecalis and enterococcus faecium isolated from hospitalized patients]. Biomedica. 2014;34(Suppl 1):50-7. doi:10.1590/s0120-41572014000500007
31. Balaei Gajan E, Shirmohammadi A, Aghazadeh M, Alizadeh M, Sighari Deljavan A, Ahmadpour F. Antibiotic resistance in enterococcus faecalis isolated from hospitalized patients. J Dent Res Dent Clin Dent Prospect. 2013;7(2):102-4. doi:10.5681/joddd.2013.018
32. Pourakbari B, Aghdam MK, Mahmoudi S, Ashtiani MT, Sabouni F, Movahedi Z, et al. High frequency of vancomycin-resistant enterococcus faecalis in an iranian referral children medical hospital. Maedica (Buchar). 2012;7(3):201-4.
33. Bao L, Peng R, Ren X, Ma R, Li J, Wang Y. Analysis of some common pathogens and their drug resistance to antibiotics. Pak J Med Sci. 2013;29(1):135-9. doi:10.12669/pjms.291.2744