Archive of SLD PYRROLIZIDINE ALKALOIDS FROM HELIOTROPIUM DISSITIFLORUM BOISS.

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ABSTRACT

Heliotropium/dissitiflorum Boiss. collected from Rud-e-Shur Tehran Province, Iran was investigated for Pyrrolizidine alkaloids (PAs). Four alkaloids namely: heliotrine I, heliotrine N-oxide II, europine III and 5'-deoxylasiocarpine IV were identified. The structures of all compounds were confirmed by spectroscopic methods.

Key Words: Heliotropium dissitiflorum, Heliotrine, Heliotrine N-oxide, Europine, 5'-Deoxylasiocarpine

INTRODUCTION

Plant containing pyrrolizidine alkaloids (PAs) are widely distributed in the world (1). These alkaloids have been isolated from several plant families such as Boraginaceae including Heliotropium genus (2). The genus Heliotropium is among the genera of plants which are known to be rich in PAs (2,3). Numerous intoxications in animals and human which are caused by the consumption of certain plants containing PAs has promoted extensive studies on these alkaloids (1, 4). The source of human poisoning can be the contaminated food stuffs or traditional herbal medicine (4). The chemistry of these alkaloids in relation to their toxic and therapeutic effects has been the subject of several investigations (2,3).In continuation of our ongoing research on heliotropium species of Iran (5-7), the isolation and structural elucidation of PAs from H. dissitiflorum Boiss. is reported. Literature review showed that except a brief report by authors no detailed phytochemical studies on these species has been reported and this is the first report on the isolation of four PAs from this species (8). Of the four alkaloids, isolation of the alkaloid IV from this species is reported for the first time .

MATERIALS AND METHODS

Plant material

Plant material was collected from Rud-e-Shur 60 Km of Southwest of Tehran in Jun 1998. The plant was identified by the Herbarium department of the Faculty of Pharmacy, Tehran University of Medical Sciences, where a voucher specimen is deposited under number 6525 TEH. The whole plant was air dried in the shade, powdered and kept in tightened light protected containers.

Instrumentation: FT-IR spectra were obtained on a Nicolet 550 spectrometer. EIMS spectra were measured by a Finnigan TSQ at 70 eV. ¹H NMR (400 MHz) and ¹³C NMR (100 MHz) Varian were used to record spectra. The optical rotation was measured on a Perkin-Elmer 241 polarimeter. Solvents and silica gel were purchased from Merck. *Extraction and Isolation Procedures*

Powdered plant material (3600 g) was percolated with MeOH at room temperature and the extract was evaporated under reduced pressure below 50°C. The residue was partitioned between ethyl acetate and 2N HCl in water. The pH of the aqeous phase was adjusted to 2 by addition of 2N HCl and to it was added 0.5 g NaCl for every 15 ml of the solution. After 24 hr, the gummy nonalkaloid residue was filtered and the pH of the filtrate was adjusted to 10 by addition of 25% NH₄OH and extracted with CHCl₃ (7×300 ml). The chloroform extract was evaporated to give 18 g crude alkaloid (part A).

To investigate the presence of the N-oxide of PA, the pH of the aqueous layer was adjusted to 2 with H_2SO_4 , and after addition of Zn dust (10 g) to the solution , the mixture was stirred for 48 h. The mixture was filtered, the solution was made alkaline and treated as above (part B). The crude extract of part A was applied to column chromatography over silica gel (Merck), eluted conescutively with following solvents in the order of increase in polarity:petroleum-ether,

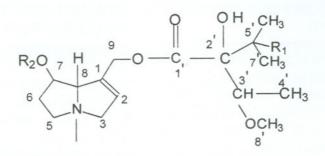
CHCl₃ and MeOH (1%). The obtained fractions were then applied to thick layer chromatography for further purification, using CHCl₃, MeOH; NH₄OH (17; 3.8; 0.25) as eluent. Part B was purified similarly.

RESULTS AND DISCUSSION

The total alkaloid content of the dried plant was 0.5%. The whole plant of *H. dissitiflorum* furnihed four PAs, namely heliotrine I, heliotrine N Noxide II, europine III, 5'-deoxylasiocarpine IV.

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(1) R1=R2=R3=H (Heliotrine)

(2)R1=R2=H, R3=O (Heliotrine N-oxide)

$(_3)R_2=R_3=H$, $R_1=OH$ (Europine)
(4)R ₁ =R ₃ =H, R ₂ = $\overset{H}{C}$ = $\overset{3''}{C}$ = $\overset{C}{C}$ $\overset{C}{C}$ (5' – deoxylasiocarpine)

Fig1.

Carbon	Ι	п	III	IV
2	5.65 (bs)	5.70 (bs)	5.78 (bs)	5.8 (bs)
3u	3.27(d)(J=16Hz)	4.37 (d)(J=16.4 Hz)	3.63 (d)	3.5 (d)
3d	3.8 (d)	4.88 (d)	4.2 (d)	4.18 (d)
5u	2.6(m)(J=12Hz)	3.6 (m)	2.81 (m)	3.00 (m)(J=6 Hz)
5d	3.3(m)	4.00 (m)	3.28 (m)	3.4 (m)
	1.87 (m)	2.079 (m)	1.92 (m)	1.8 (m)
6u	1.98 (m)	2.37 (m)	2.02 (m)	1.8 (m)
6d	4.09 (m)(J=6 Hz)	4.28 (m)	4.34 (m)	5.12 (m)
7	3.99 (bs)	4.73 (bs)	4.47 (s)	4.37 (bs)
8	4.63 (d)(J=13.2 Hz)	4.75 (d)(J=14.4)	4.75 (d) (J=14 Hz)	4.88 (d) (J=14.4 Hz)
9u	4.92 (d)	4.86 (d)	4.97 (d)	4.88 (d)
9d	3.57 (q)(J=6.4 Hz)	3.67 (q)(J=6Hz)	3.8 (q)(J=6 Hz)	3.7 (q) (J=6 Hz)
3'	1.08(d)	1.11 (d)	1.24 (d)	1.21 (d)
4'	0.87 (d) (J=6.8 Hz)	0.90 (d) (J=6.4 Hz)	1.26 (d)	0.95 (d)
	0.84 (d)	0.88 (d)	1.24 (d)	0.94 (d)
6'	3.27 (s)	3.24 (s)	3.33 (s)	3.22 (s)
7'	2.04 (hept)	1.92 (hept)	-	2.2 (hept)
8'				6.13(q)(J=7.2 Hz)
5'				1.95(d)(J=7.2 Hz)
3''				1.82(s)
-				1.02(0)
4''				
5''				

Table 1. ¹HNMR assignments of compound I-IV

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Heliotrine I: mp: 122-124°C; $[\alpha]_D = 63.8$ (CHCl₃); $R_f = 0.5$ (CHCl₃; MeOH; NH₄OH) (17; 3.8; 0.25); IR (CHCl₃): v cm⁻¹ 3377 (OH), 1736 (C=O), 1097 (C-O), 1280 (C-O)

EIMS m/z (% relative intensity) 314 [(MH⁺, 3)], 255 (1.4), 197 (4), 156 (12), 138 (100), 120 (7), 93 (100), 80 (25); ¹H NMR (see Table 1).

¹³CNMR (CDCl₃)(carbon's number) 136(1),
127.2 (2), 62 (3), 54 (5), 34 (6), 75.4 (7), 78.4 (8),
62.6 (9), 175 (1'), 82.4 (2'), 79.9 (3'), 12.4 (4'),
31.7 (5'), 17 (6'), 16.4 (7'), 57 (8').

<u>Heliotrine N-oxide II:</u> oil $[\alpha]_D$ =26.6 (EtOH); R_f = 0.26 (CHCl₃; MeOH; NH₄ OH)(17; 38,0.25); IR v cm⁻¹ (CHCl₃): 3300 (OH), 2389 (N-O), 1727 (C=O), 1229 (C-O)

EIMS m/z (% relative intensity): 314[(MH⁺-16)], 4.7), 240 (5), 138 (100), 120 (5), 93 (40), 83 (12), 59 (14); ¹H NMR (see Table 1). ¹³CNMR (CDCl₃)(carbon's number) 133.7 (1), 120.7 (2), 76.6 (3), 68 (5), 33 (6), 71.6 (7), 96.2 (8), 60.6 (9), 173.9 (1'), 83.1 (2'), 78.7 (3'), 11.4 (4'), 32.9 (5'), 16.9 (6'), 16.9 (7'), 56.5 (8').

<u>Europine III:</u> oil; $[\alpha]_D = +19.9^{\circ}$ (C=2.5 mg/cc in EtOH); $R_f = 0.31$ (CHCl₃; MeOH; NH₄OH)(17; 3.8; 0.25); IR v cm⁻¹ (CHCl₃): 3427 (OH), 1729 (C=O), 1991 (C-O), 1280 (C-O); EIMS m/z (% relative intensity): 329 (M⁺, 2.5), 240 (2), 138 (100), 120 (29), 93 (75), 80 (25); ¹H NMR (see Table 1).

¹³C NMR (CDCl₃) (carbon's number) 135 (1),
125 (2), 61.1 (3), 54.5 (5), 33.1 (6), 73.7 (7), 79.6
(8), 61.3 (9), 174 (1'), 82.4 (2'), 79.1 (3'), 13 (4'),
72.6 (5'), 26.1 (6'), 24.8 (7')

<u>5'-Deoxylasiocarpine IV</u>: yellow oil; picrate mp: 190-198; $[\alpha]_D = -13.7$ (EtOH); R_f = 0.8 (CHCl₃; MeOH; NH₄OH)(17; 3.8; 0.25); IR v cm⁻¹ (CHCl₃): 3414 (OH), 1722 (C=O), 1991 (C-O) EIMS m/z (% relative intensity) 396 (MH⁺, 2),

295 (5.5), 263 (1.1), 220 (80), 120 (83), 93 (68), 83 (32); ¹H NMR (see Table 1).

The structures of compound I-III were identified by comparison of their spectral data with those reported in the literatures (9,10). Compound IV showed a carboxyl absorption at 1722. The EIMS showed a molecular ion at $m/z 396 (M^++1)$ which corresponds to an empirical formula of C₂₁H₃₃NO₆. The molecular formula was 16 mass unit lower than lasiocarpine. Furthermore, the ¹HNMR (table1) was very similar to the spectra of lasiocarpine except that H-5' appeared as heptate at $\delta = 2.2$ which is in support of 5'deoxylasiocarpine. The fragmentation pattern in the mass spectrum of compound IV likewise support this conclusion. Compound IV has been isolated from Heliotropiume genus (11).

REFERENCES

- Swith, L.W., Culvenor, C.C.J. (1981) Plant sources of hepatotoxic pyrrolizidine alkaloids. J. Nat. Prod. 44: 129-152
- 2. Mattocks, A.R. (1986) Chemistry and Toxicology of Pyrrolizidine Alkaloids, Academic Press, Orlando, Florida, pp: 158-315
- 3. Anon, (1988) Pyrrolizidine Alkaloids, Environmental Health Criteria (80), 1st eds, World Health Organization, Geneva, pp: 38-54
- 4. Roeder, E. (1995) Medicinal plants in Europe containing pyrrolizidine alkaloids, Pharmazie 50: 83-96
- Yassa, N., Farsam, H., Shafiee, A., Rustaiyan, A. (1996) Pyrrolizidine alkaloids from *H. esfandiari*. Planta Med. 62: 583-584
- Yassa, N., Farsam, H., Rustaiyan, A., Shafiee, A. (1999) Alkaloids of Boraginaceae II [1], Pyrrolizidine alkaloids of *Heliotropium europaeum* L. Population Garmsar. J. Sci. I.R. Iran 10: 39-42
- Farsam, H., Yassa, N., Sarkhail, P., Shafiee, A. (2000), New Pyrrolizidine Alkaloids from *Heliotrop*ium crassifolium, Planta Med. 66: 389-391.
- Hammouda, F. M., Rizk, A. M., Ismail, S. I., Atteya, S. Z. et al. (1984) Poisonous plants contaminating edible ones and toxic substances in plant foods. Pharmazie 39: 703-5
- 9. Alan, J.; Culvenor, C.C.J., Smith, L.W. (1982) Pyrrolizidine alkaloids a carbon-13 N.M.R study. Aust. J. Chem. 35: 173-174
- Reina, M., Mericli, R., Gonzales, A. (1995) Pyrrolizidine alkaloids from H. Bovei. Phytochem. 38: 355-358
- 11. Farsam, H.; Yassa, N.; Shafiee, A.; Amanlou, M.; (1998) Pyrrolizidine alkaloids from *H. disciforme*. Pharm. Pharmacol. Lett (8) 4: 79-80