ANTI-INFLAMMATORY AND ANALGESIC ACTIVITY OF METHANOLIC EXTRACTS OF AERIAL PARTS OF STACHYS SCHTSCHEGLEEVII SOSN. AND STACHYS BALANSAE BOISS. AND KOTSCHY EX BOISS IN RATS.

¹SHAMSALI REZAZADEH, ²ABBAS KEBRYAEEZADEH, ¹MORTEZA PIRALI-HAMEDANI, ¹ABBAS SHAFIEE, ²SAEED GHARUNI ISFAHANI

¹Department of Medicinal Chemistry, ²Department of Pharmacology and Toxicology, Faculty of Pharmacy and pharmaceutical Sciences Research Centre, Tehran University of Medical Sciences, Tehran, Iran

ABSTRACT

Extracts of the flowering aerial parts of *Stachys schtschegleevii* Sosn. and *S. balansae* Boiss. and Kotschy ex Boiss have been used in Iranian folk medicine as remedy for rheumatic and other inflammatory disorders and anti-inflammatory and analgesic effects of some species of *Stachys* e.g. *Stachys inflata* have been reported. In this study, the anti-inflammatory and antinociceptive properties of total methanolic extracts of the flowering aerial parts of two *Stachys* species in rat were investigated by carrageenan-induced paw edema and formalin test. Intraperitoneal injection of the extracts, 60 min before induction of inflammation, resulted in inhibition of carrageenan-induced rat paw edema in dose dependant manner (doses 50, 100 and 200 mg/kg). In the formalin test, the extract (50, 100 and 200 mg/kg) had low effect in the first phase (0–5 min) of the formalin-induced pain, but all three doses showed analgesic and anti nociception effects significantly. In conclusion the methanolic extracts of *Stachys schtschegleevii* and *Stachys balansae* have analgesic and anti-inflammatory effects in formalin test and carrageenan-induced paw edema.

Keywords: Anti-inflammatory; Analgesic; Stachys schtschegleevii; Stachys balansae

INTRODUCTION

The use of natural products is growing in the world especially in developing countries such as China, India and Arabic countries. For several centuries some Stachys species have been used traditionally for their health benefits (1). About three hundred Stachys species have been reported (2); 34 of them are found in Iran: of which 13 are endemic (3, 4). The plants of Stachys are widely distributed in tropical and subtropical countries. Stachys schtschegleevii and Stachys balansae are native plants of Iran (4). In Iranian traditional medicine the extracts of the aerial parts of Stachys schtschegleevii (traditionally named Poulk) have been used in infectious, rheumatic and respiratory inflamamatory diseases. Phytochemical investigations on Stachys species have shown the presence of phenylethanoid glycosides (5, 6), terpenoids and steroids (7, 8) diterpenes (9) and flavonoids (10, 11). Also the compositions of the essential oils of some species have been reported (12, 13). Pharmacological studies have shown that extracts of some Stachys species have antiinflammatory, anti-toxic (14, 15), anti-nephritic (16, 17), antihepatitis (18) and anti-anoxia (19)

properties. Since there are no reports on the pharmacology and phytochemistry of *Stachys schtschegleevii* and *Stachys balansae* Boiss, in this study, anti-inflammatory and analgesic properties of methanolic extracts of these species were evaluated by carrageenan-induced paw edema and formalin tests in rat (20, 21).

MATERIALS AND METHODS

Plant material

Aerial flowering parts of these two *Stachys* species were collected during flowering period (June and July 2001), identified in Central Herbarium of Iran (Research Institute of Forest and Rangelands, Tehran) and a voucher specimen of both species (*S. balansae* and *S. schtschegleevii*) (83579 TARI and 83580 TARI respectively) were deposited in herbarium. Plants were dried in shadow before extraction.

Chemicals

Carrageenan was prepared from Sigma- Aldrich Company (Germany); indometacin was obtained from Hakim Pharmaceutical Company (Iran); all other solvents were prepared from Panreac Company (Spain).

Correspondence: Abbas Kebryaeezadeh, Department of Pharmacology and Toxicology Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran. Email: kebriaee@sina.tums.ac.ir

Extract preparation

200 g of crushed aerial parts of plant was subjected to Soxhlet extraction with methanol (99.9%) for 3 hours. Methanolic extracts were rotary evaporated to volume of 200 ml at 50°C, and after addition of 50 ml of water, the solution was extracted with 3 x 100 ml ether (40-60°C) (3x100ml). The extracts were then filtered and filtrates were evaporated at maximum 50 °C to an amorphous gum under reduced pressure.

Carrageenan induced paw edema

The anti-inflammatory activities of the extracts were determined by the carrageenan-induced edema test in the hind paws of rats by the reported method (22). Male albino Wistar rats (150-200g) were fasted for 24 hours before the experiment with free access to water and then 150 µl of 1% suspension of carrageenan in saline, which was prepared 1 hour before each experiment, was injected into the plantar side of both hind paws of the rats. The extracts were dissolved in saline and passed through a weighed filter paper and the filtrate was used for intraperitoneal injection. Following filtration the filterate was dried and weighed again, in order to obtain the real concentration of the extract. Rats, were allocated randomly to groups of six; a) for controls; b) for extracts (50, 100 and 200 mg/kg) and c) for indometacin as the reference drug (5mg/kg). Indometacin solution was prepared in saline using tween 80 as dispersing agent. Saline and the extracts or indometacin in 0.5 ml of saline were injected intraperitoneally 60 minutes before induction of inflammation. The paw thickness was measured from the ventral to the dorsal surfaces using a dial caliper prior to carrageenan injection and then at 1 hour intervals for 5 hours. Data are expressed as a percentage increase in thickness compared with pre-injection values.

Formalin-test

Male Albino Wistar rats (n=6), weighing 180-220 g, were kept in Plexiglas cages with free access to food and water. Test was conducted in the middle of the light period of a 12-h light: 12h dark cycle. Each animal was tested once only. Plant extracts (50, 100 and 200 mg/kg) were dissolved in 0.9% saline and indometacin (5 mg/kg) was suspended in 0.9% (w/v) saline containing tween 80 and administered intraperitoneally in a volume of 1 ml. Control group received only vehicles (1 ml). The analgesic activities of the extracts were determined by the reported method for formalin test (23). One hour before testing, animals were placed in a standard cage (30×12×13 cm), that served as an observation chamber and then 60 µl of 5.0% formalin were injected to the dorsal

surface of the left hind paw. The rats were observed for 60 min after injection of formalin, and the amount of time that animals spent licking the injected hind paw was recorded. The first 5 min post formalin injection was assigned as the early phase and the period between 15 and 60 min as the late phase. Pain rate was calculated according to formula:

Pain rating =
$$\frac{1T_1 + 2T_2 + 3T_3}{Timebloke(second)}$$

 $T_{1,}$ T_{2} and T_{3} are every 15 seconds during 5 minutes, in which animals had 1, 2 or 3 type habit. The samples were administered 15 min before injection of formalin.

Statistical analysis

ANOVA followed Student-Newman-Keuls test was used to determine significant differences between groups and P < 0.05 were considered significant.

RESULTS

Anti-inflammatory effects of Stachys schtschegleevii

Induction of acute inflammation in control rats resulted in a prominent increase in paw thickness which began 1 hour after intraplantar injection of carrageenan and reached to a peak after 4 hours (fig. 1). Intraperitoneal injection of the extract of aerial flowering parts of Stachys schtschegleevii resulted in inhibition of carrageenan-induced paw edema dose dependently. All doses (50, 100 and 200 mg/kg) of extracts induced a significant (P<0.01) antiinflammatory effects 5 hours after carrageenan injection. The doses of 100 and 200 mg/kg showed more potent effects (P<0.001) at all time points. Indometacin (5 mg/kg, i.p) showed less activity than the extracts. The extracts (100 and 200 mg/kg) showed the maximum inhibitory effects at fourth hour.

Anti-inflammatory effects of Stachys balansae

Doses of the aerial flowering parts of Stachys balansae showed different anti-inflammatory in comparison with effects Stachys schtschegleevii. Dosage of 50mg/kg of extract showed no significant differences with controls during first 3 hours (fig. 2) but represented significant anti-inflammatory effect 4 hours after carrageenan injection (P<0.05). The dose of 100 mg/kg at first 2 hour showed inhibitory effect in carrageenan induced edema (P<0.01). The dose of 200 mg/kg showed significant antiinflammatory effect during all 5 hours (P<0.01) and reached to a maximum 4 hours after injection.

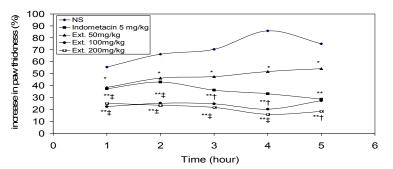


Figure 1. Anti-inflammatory effects of different doses of methanolic extract prepared from aerial parts of *Stachys schtschegleevii* in carrageenan induced paw edema in rat compared to the same points in control group (saline) [* P<0.01, ** P<0.001] and Indometacin 5 mg/kg [† P<0.05, ‡ P<0.01].

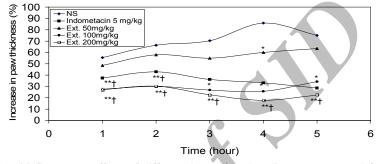


Figure 2. Anti-inflammatory effects of different doses of methanolic extract prepared from aerial parts of *Stachys balansae* in carrageenan induced paw edema in rat compared to the same points in control group (saline) [* P<0.05, ** P<0.01] and indometacin 5 mg/kg [$\uparrow P<0.05$]

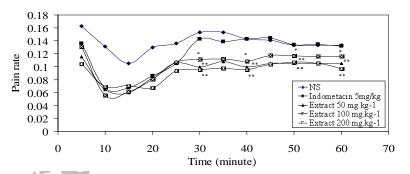


Figure 3. Analgesic effects of different doses of methanolic extract prepared from aerial parts of *Stachys schtschegleevii* and indometacin (5 mg/kg, ip) in formalin-induced paw licking in rat compared to the same points in control group (saline), [* P<0.01, ** P<0.001].

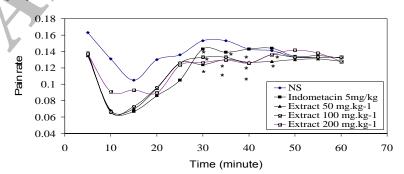


Figure 4. Analgesic effects of different doses of methanolic extract prepared from aerial parts of *Stachys balansae* and Indometacin (5 mg/kg ,ip) in formalin-induced paw licking in rat compared to the same points in control group (saline) (*P<0.05).

Analgesic effects of Stachys schtschegleevii

Intraplantar injection of 5% formalin evoked a characteristic biphasic licking response. All three doses of extract (50, 100 and 200 mg/kg) showed analgesic effects like indometacin (5 mg/kg, intraperitoneal) in the first phase of study (0-5 min) (fig. 3). During the second phase, extracts showed higher analgesic effects (50 mg/kg, P<0.05; 100 and 200 mg/kg, P<0.01).

Analgesic effects of Stachys balansae

All three doses of extracts (50, 100 and 200 mg/kg) showed analgesic effects similar to that of indometacin (5 mg/kg, intraperitoneally) in the first phase and higher effects after the second phase (P<0.05) up to 45 minutes and thereafter no significant differences were observed.

DISCUSSION

Carrageenan injection into the rat paw provokes a local, acute inflammatory reaction that is a suitable criteria for evaluation of antiinflammatory agents (24). The inflammation consists of two phases, early phase which is related to the production of histamine, 5hydroxytryptamin, bradykinins and cyclooxygenase products and delayed phase which has been linked to neutrophil infiltration, as well as production of arachidonic acid metabolites (25, 26 and 27). Extract of flowering aerial parts of Stachys schtschegleevii and Stachys balansae produced anti-inflammatory effects in carrageenan induced inflammation in rats. As shown in Fig.1, S.schtschegleevii had antiinflammatory effects higher than saline at all three doses (50, 100 and 200 mg/kg) (P<0.01) and higher than indometacin in doses of 100 and 200 mg/kg. Doses of 100 and 200 mg/kg of this extract showed higher effect (P<0.001) than saline and indometacin (P < 0.01) and there was no significant differences between these two doses. Extract of the Stachys balansae showed similar anti-inflammatory trends but lower than extract of S. schtschegleevii. In addition, all three doses of the extracts (50, 100 and 200 mg/kg) prepared from both Stachys schtschegleevii and Stachys balansae significantly inhibited the pain associated with the second phase (inflammatory component) of the formalin test, and the effect of

Stachys schtschegleevii was more pronounced. Doses of 100 and 200 mg/kg showed similar potency which were high than those of dose of 50 mg/kg. In the formalin test, the initial nociceptive scores normally peaked 5 min (first phase) and then 15-30 min after injection (second phase), which represent the neurogenic and inflammatory pain responses, respectively (28). Phenylethanoid glycosides, triterpenoids and flavonoids have been considered as active components responsible for the biological actions of the Stachys genus (10, 11, 8, and 6). However, the anti-inflammatory effects of Stachys, or its components, have not been elucidated completely so far. It has been reported that acteoside, a phenylethanoid glycoside of Stachys sieboldii, has a suppressive effect on the accumulation of leukocytes in the nephritic glomeruli through prevention of the upregulation of adhesion molecules (17). The discrepancy between the inhibitory effect upon low doses and the lack of increase in potency phenomena at higher doses of the extracts might be explained by hypotheses that some of the active constituent(s) of these two Stachys genus at high concentrations may exhibit pro-inflammatory properties. It is also likely that the extracts may have components with different anti- and proinflammatory effects. The co-existence of both anti-nociceptive and anti-inflammatory effects which was observed with this extract is welldefined for various non-steroidal antiinflammatory drugs (NSAIDS) particularly salicylates. It is therefore interesting that the extract behaved liked NSAIDS in this study which correlates well with the traditional application of the plant. The results presented in this study should be taken as a basis for further investigation for determination of the exact mode of action of individual constituents of the extracts.

ACKNOWLEDGMENT

This study was supported by a grant from the research council of Tehran University of Medical Sciences; also we are grateful of Dr. V. Mozaffarian (research institute of forest and rangeland of Tehran) for identification of the plants.

REFRENCES

- 1. Zargari A. Medicinal Plants, Vol.4, Tehran: Tehran University Publication; 1990. p. 123.
- 2. Evans WC. Trease and Evans' Pharmacognosy, 13th ed, Bailliere Tindall, London; 2002. p. 33.
- 3. Mozaffarian V. A Dictionary of Iranian Plant Names, Farhang Moaser, Tehran;1996. p. 522.
- 4. Rechinger KH. *Stachys L.* In: Rechinger KH. (Ed) *Flora Iranica*. Akademische Druck-U. Graz, 1982. Vol. 150, 354-396.
- 5. Nishimura H, Sasaki H, Inagaki, N, Chin M, Mitsuhashi H. Nine phenethyl alcohol glycosides from *Stachys sieboldii*. Phytochemistry 1991; 30, 965–969.

- 6. Miyase T, Yamamoto R, Ueno A. Phenylethanoid glycosides from *Stachys officinalis*. Phytochemistry 1996; 43: 475–479.
- 7. Ross SA, Zinchenko TV. Study of triterpenoids and steroids of *Stachys palustris* L. Farm Zh (Kiev) 1975; 30: 91–92.
- 8. Yamamoto R, Miyase T, Ueno A. *Stachys* saponins I –VIII, new oleanane-type triterpene saponins from *Stachys riederi* Chamisso. Chem Pharm Bull 1994; (Jun) 42: 1291–1296.
- Piozzi F, Savona G, Hanson JR. Kaurenoid diterpenes from *Stachys lanata* Phytochemistry 1980; 19: 1237.
- 10. Zinchenko TV. Flavonoid glycosides of Stachys neglecta. Farm Zh (Kiev) 1970; 25(4): 81-82.
- 11. EL-Ansari MA, Barron D, Abdalla M.F, Saleh NAM, LE Quere JL. Flavonoid constituents of *Stachys aegyptica*. Phytochemistry 1991; 30: 1169–1173.
- 12. Skaltsa HD, Lazari DM, Chinou IB, Loukis AE. Composition and antibacterial activity of the essential oils of *Stachys candida* and S. *chrysantha* from southern Greece [letter]. Planta Med 1999; 65: 255–256.
- 13. CËakir A, Duru ME, Harmandar M, Izumi S, Hirata T. The volatile constituents of *Stachys balansae* L. from Turkey.Flavour Fragr J 1997; 12: 215-218.
- Maleki N, Garjani A, Nazemiyeh H, Nilfouroushan N, Eftekharsadat AT, Allameh Z, Hasannia N. Potent anti-inflammatory activities of hydroalcoholic extract from aerial parts of *Stachys inflata* on rats. J Ethnopharmacol 2001; 75: 213–218
- 15. Zinchenko TV, Voitenko GN, Lipkan GN. Anti-inflammatory, antitoxic and hypoazotemic effect of a *Stachys recta* preparation, stachyrene. Farmakol Toksikol 1981; 44: 191–194.
- Hayashi K, Nagamatsu T, Ito M, Hattori T, Suzuki Y. Acteoside, a component of *Stachys sieboldii* MIQ, may be a promising antinephritic agent (1): Effects of acteoside on crescentic-type anti-GBM nephritis in rats. Jpn J Pharmacol 1994; 65: 143–151.
- 17. Hayashi K, Nagamatsu T, Ito M, Hattori T, Suzuki Y. Acteoside, a component of *Stachys sieboldii* MIQ, may be a promising antinephritic agent (2): Effects of acteoside on leukocyte accumulation in the glomeruli of nephritic rats. Jpn J Pharmacol 1994; 66: 47–52.
- Savchenko VM, Khvorostinka VM. Effects of a preparation from *Stachys inflata* on the course of experimental hepatitis in rats. Farm Zh (Kiev) 1978; 33: 50–53.
- 19. Yamahara J, Kitani T, Kobayashi H, Kawahara Y. Studies on *Stachys sieboldii* MIQ.II. Antianoxia action and the active constituents. Yakugaku Zasshi 1990; 110 : 932–935.
- 20. Khaksa G, Zolfaghari ME, Dehpour AR, Samadian T. Anti-inflammatory and antinociceptive activity of disodium glycyrrhetinic acid hemiphthalate. Planta Med 1996; 62: 326–328.
- Carvalho JCT, Sertie JAA, Barbosa MVJ, Patricio KCM, Caputo LRG, Sartio SJ, Ferreira LP, Bastos JK. Anti-inflammatory activity of the crude extract from the fruits of *Pterodon emarginatus* Vog. J Ethnopharmacol 1999; 64: 127-133.
- Al-Haboubi HA., Zeitlin IJ. Re-appraisal of the role of histamine in carrageenan-induced oedema. Eur J Pharmacol 1983;88: 160–176.
- 23. Dubuisson D, Dennis SG. The formalin test: a quantitative study of the analgesic effects of morphine, meperidine and brain stem stimulation in rats and cats. Pain 1977; 4: 167-174.
- 24. Winter CA, Ristey EA, Nuss GW. Carrageenan-induced oedema in hind paw of the rat as an assay for anti-inflammatory drugs. Proc Soc Exp Biol Med 1962; 111: 544–547.
- 25. Dawson J, Sedgwick AD, Edwards JC, Lees PA. Comparative study of the cellular, exudative and histological responses to carrageenan, dextran and zymosan in the mouse. Int J Tissue React 1991; 13: 171–185.
- Salvemini D, Wang ZQ, Wyatt DM, Bourdon MH, Marino PT, Currie MG. Nitric oxide: A key mediator in and late phase and late phase of carrageenan-induced rat paw inflammation. Br J Pharmacol 1996; 118: 829–838.
- 27. Boughton-Smith NK, Deakin AM, Follenfant RL, Whittle BJR, Coarland LG. Role of oxygen radicals and arachidonic acid metabolites in the reverse passive arthus reaction and carrageenan paw oedema in the rat. Br J Pharmacol 1999; 110, 896–902.
- 28. Hunskaar S, Hole K. 1987. The formalin test in mice: dissociation between inflammatory and non-inflammatory pain. Pain 1987; 30: 103–114.