

## Effects of hydroalcoholic extract of *Cynodon dactylon* (L.) pers. on ischemia/reperfusion-induced arrhythmias

\*<sup>1</sup>Najafi M., <sup>2</sup>Nazemiyeh H., <sup>3</sup>Ghavimi H., <sup>3</sup>Gharakhani A., <sup>3</sup>Garjani A.

<sup>1</sup>Department of Pharmacology, School of Pharmacy, Drug Applied Research Center and Research Center for Pharmaceutical Nanotechnology, <sup>2</sup>Drug Applied Research Center, School of Pharmacy, <sup>3</sup>Department of Pharmacology, School of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran

Received 13 June 2007; Revised 24 June 2008; Accepted 1 Aug 2008

### ABSTRACT

**Background and purpose of the study:** Probable antiarrhythmic effects of *Cynodon dactylon* (L.) pers. (family Poaceae) against ischemia/reperfusion (I/R)-induced arrhythmias were investigated in isolated rat heart.

**Methods:** The hearts were subjected to 30min regional ischemia followed by 30min reperfusion and perfused with hydroalcoholic extract of rhizome of *C. dactylon* (25, 50, 100 and 200µg/ml).

**Results:** During ischemia, the extract produced marked reduction in the number, duration and incidences of ventricular tachycardia (VT) at 25 and 50µg/ml ( $p<0.001$  and  $p<0.01$ , respectively). Total number of ischemic ventricular ectopic beats (VEBs) were lowered by 25-100µg/ml ( $p<0.001$ ,  $p<0.001$  and  $p<0.05$ , respectively). At the reperfusion phase, *C. dactylon* (25 and 50µg/ml) decreased incidence of VT from 100% (control) to 13 and 33% ( $p<0.001$  and  $p<0.05$ ) respectively. Duration and number of VT and total VF incidence were also reduced at the same concentration ( $p<0.05$  for all). Perfusion of the extract (25-100µg/ml) was markedly lowered reversible VF duration from  $218\pm99$ sec to 0 sec, 0 sec and  $10\pm5$ sec ( $p<0.01$ ,  $p<0.01$  and  $p<0.05$ ) respectively. Moreover, *C. dactylon* (25 and 50µg/ml) decreased number of total VEBs from  $349\pm73$  to  $35\pm17$  ( $p<0.001$ ) and  $66\pm26$  ( $p<0.01$ ). In this study, it was also shown that perfusion of the extract produced a marked and concentration-dependent positive inotropic effect.

**Conclusion:** The findings of this study indicate that *C. dactylon* produce protective effects against I/R-induced arrhythmias in isolated rat hearts probably by increase in the myocardial contractility and as a result by improvement of hemodynamic factors.

**Keywords:** *Cynodon dactylon* (L.) pers., ischemia/reperfusion, arrhythmias, isolated heart; rat

### INTRODUCTION

*Cynodon* species are members of the *Cynodonteae* tribe and the *Chloridoideae* sub-family (1). *Cynodon dactylon* (*C. dactylon*, Bermuda grass) is a resilient, perennial grass distributed all over the world and especially it is native to the warm temperate and tropical regions (1-2). Leaf, root and rhizome of the plant have been used in folk medicine of different countries (3, 4), as anti-inflammatory (3, 5), anticystitis (3), antihypertensive (3, 6), antihysteria, antipsychotic (2) antigonorrhoeal infection (4), antiviral, as well as hypolipidemic, hypoglycemic agent (6). In India, the plant is reputed for the treatment of melena, thirst, anorexia, burning sensations of the body, pruritis, miscarriage and erysipelas (6-7), and its leaf juice with a pinch of common salt has

been used orally in stomachache (5). Decoction of whole plant has been given orally to cure menstrual problem (8).

In Iranian folk medicine, the root and rhizome of the plant have been used in the treatment of depression, nausea, cough, epilepsy and hemorrhage (9). In some provinces of Iran (such as Azerbaijan and Kurdistan), *C. dactylon* which is locally named as *Chayer*, has been traditionally used for cardiovascular diseases. Many people of these regions believe that the extract of the rhizome has curative effects in coronary artery diseases and in heart failure. In addition, uncontrolled studies by some Iranian cardiologists have shown cardioprotective effects in the patients who used the plant traditionally. To date, there is no report on protective effects of

the plant on cardiovascular diseases and in the present study the effects of hydroalcoholic extract of *C. dactylon* rhizomes on ischemia/reperfusion (I/R)-induced arrhythmias in isolated rat heart was investigated.

## MATERIALS AND METHODS

### *Plant materials*

Rhizomes of *C. dactylon* were freshly collected from the field (Maragheh- East Azerbaijan, Iran) in November, 2005 and a voucher specimen was deposited at herbarium of School of Pharmacy, Tabriz University of Medical Sciences. The collected rhizomes were washed and dried at room temperature in shade.

### *Preparation of the plant extracts*

Two hundred grams of the dried and powdered rhizomes of *C. dactylon* were extracted three times by maceration with 1 L of a mixture of methanol-water (70:30) each time for 12 h. The combined extracts were filtered and evaporated under reduced pressure and temperature (40 °C) to dryness. Dried residue (15 g) was kept at 4 °C until used.

### *Chemical tests*

The crude extract of *C. dactylon* was screened for the presence of different classes of compounds by some modifications in standard methods (10-12). Thin Layer Chromatography procedures using precoated silicagel plates (Merck; GF<sub>254</sub>, 0.25 mm) were performed to confirm the results of screenings (14). The following spray reagents were used for detection of respective classes of compounds: NEU (for polyphenols/flavonoids), Antimony trichloride in chloroform (for steroidal saponins/sterols), Kedd reagent (for cardiac glycosides), Dragendorff's reagent (for alkaloids), 5% Ethanolic sodium hydroxide (for anthraquinones) (13-14). Since the dried extract contained mainly flavonoid glycosides, it was standardized in its flavonoid glycosides content (4.6%) by the reported method (15).

### *Free radical scavenging activity (DPPH assay)*

2,2-Diphenyl-1-picrylhydrazyl (DPPH), molecular formula of C<sub>18</sub>H<sub>12</sub>N<sub>5</sub>O<sub>6</sub>, was obtained from Fluka Chemie AG, Bucks. Quercetin (a well-known natural antioxidant) was obtained from Merck Company. The antioxidant activity of the extract was determined by a reported method (16,17).

### *Quantitative assay*

The extract was dissolved in MeOH to obtain a concentration of 0.5 mg/mL. Dilutions were made

to obtain concentrations of 5x10<sup>-2</sup>, 5x10<sup>-3</sup>, 5x10<sup>-4</sup>, 5x10<sup>-5</sup>, 5x10<sup>-6</sup>, 5x10<sup>-7</sup>, 5x10<sup>-8</sup>, 5x10<sup>-9</sup>, 5x10<sup>-10</sup> mg/mL. Diluted solutions (1.00 mL each) were mixed with DPPH (1.00 mL) and allowed to stand for 30 min for completion of reactions. The UV absorbance was recorded at 517 nm. The experiment was performed in triplicates and the average absorption was recorded for each concentration. The same procedure was followed for quercetin in MeOH as positive controls.

### *Animals and surgical procedure*

The hearts of male Sprague-Dawley rats (280-320g) were rapidly excised and mounted via the aorta on a standard Langendorff perfusion apparatus with a perfusion pressure of 100 cm H<sub>2</sub>O. Modified Krebs-Henseleit buffer solution containing (mM): NaCl (118.5), NaHCO<sub>3</sub> (25.0), KCl (4.8), MgSO<sub>4</sub> (1.2), KH<sub>2</sub>PO<sub>4</sub> (1.2), D-glucose (12.0) and CaCl<sub>2</sub> (1.7) of pH of 7.4 as the perfusion medium was gassed with 95% O<sub>2</sub> 5% CO<sub>2</sub>, pH 7.4 at 37°C throughout the experiment (18-21). An epicardial ECG was recorded by a polygraph during the experiment using two silver electrodes attached directly to the heart. A fluid filled balloon was introduced into the left ventricle and inflated to give a pre-load of 8–10 mmHg (22). Hemodynamic factors including heart contractility force, left ventricular developed pressure (LVDP), rate pressure product (RPP) and coronary flow rate (CFR) were measured. CFR was measured by a time collection of the coronary perfusate that dripped from the heart. RPP was calculated by multiplying LVDP by HR.

A 4/0 braided silk suture was placed around the left anterior descending coronary artery. Following 20min stabilization, coronary occlusion (30min) was achieved by threading the loose ends of the ligature through a polyethylene occluder and clamping in place. Release of the clamp allowed reperfusion of the previously ischemic tissue (30min). Based on the Lambeth conventions, the ECGs were analyzed to determine the total number of ventricular ectopic beats (VEBs), the number of beats occurring as ventricular tachycardia (VT), and the incidence and duration of VT and ventricular fibrillation (VF) during both ischemia and reperfusion phases (23). The isolated hearts were allocated randomly to one of the following 5 groups (n=8-12 in each group): (1) control; (2-5) in which the hearts were perfused with rhizome hydroalcoholic extract of *C. dactylon* (25, 50, 100 and 200µg/ml, respectively). Selection of concentrations for the experiment was based on the results of a previous pilot study and for the preparation of the required concentrations, different amounts of the dried extract was completely dissolved in fresh Krebs-

**Table 1** Effects of total extract of rhizome of *C. dactylon* (25-200µg/ml) on cardiac arrhythmias during 30min ischemia and 30min reperfusion in isolated rat hearts.

Groups	Ischemia time			Reperfusion time		
	VT Counts	VT Duration (sec)	Reversible VF Duration (sec)	VT Counts	VT Duration (sec)	Reversible VF Duration (sec)
Control	280±89	58±19	6±4	154±29	23±5	218±99
<i>C. dactylon</i> (25µg/ml)	10±4 ***	2±1 ***	0±0	17±10 ***	3±2 **	0±0 **
<i>C. dactylon</i> (50µg/ml)	22±18 **	7±4 **	0±0	42±20 *	7±4 *	0±0 **
<i>C. dactylon</i> (100µg/ml)	71±35	14±6	3±3	85±31	17±7	10±5 *
<i>C. dactylon</i> (200µg/ml)	154±40	20±7	5±4	161±46	29±8	43±15

Total VEBs is sum of arrhythmias occurring as single, salvos and VT. \*\*\* p<0.001, \*\* p<0.01, \* p<0.05 compared to the control value. n= 8-12 rats in each group.

Henseleit solution then filtered and the hearts were perfused with enriched solution for the whole period of ischemia and reperfusion. To determine the isolated hearts' contractility force, LVDP as an index of contractility force was measured. Krebs buffer solution containing different concentrations of the extract was perfused for 1min at 20min intervals and maximum responses were recorded and compared to pre-perfusion values. The experiments were carried out in accordance with regulations of Tabriz University of Medical Sciences guideline for the care of laboratory animals.

#### Statistical analyses

Except for the incidences of VT and VF, all results are expressed as mean±SEM. One-way ANOVA with LSD post hoc test was carried out to test any differences between the mean values of hemodynamic factors and heart contractility force. To compare the number of VEBs and duration of VT and VF between groups, the Mann-Whitney non-parametric U test was employed. For the analysis of the incidences of VT and VF Fisher's exact test with Yates correction was used. Differences between groups were considered significant at a level of p<0.05.

## RESULTS

#### Phytochemistry results

Phytochemical screenings showed that the rhizomes total extract of *C. dactylon* have significant amounts of sugars, flavonoids, sterols, steroidal saponins and trace amount of alkaloids.

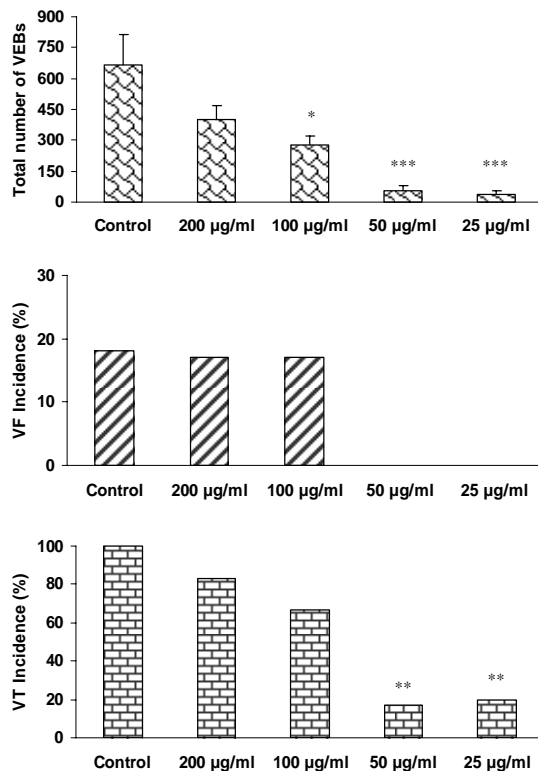
#### DPPH assay

RC<sub>50</sub> values (the concentration of the extract that reduces 50% of DPPH) for quercetin (standard)

and the extract were  $2.88 \times 10^{-5}$  and 0.346 mg/ml, respectively.

#### Antiarrhythmic effects

Effects of *C. dactylon* on numbers, duration and incidences of arrhythmias during 30min ischemia followed by 30min reperfusion are summarized in Table 1 and Figures 1 and 2. In the ischemic time, perfusion of the hearts with 25 and 50µg/ml hydroalcoholic extract of *C. dactylon* produced significant reduction in the number and duration of VT (p<0.001 and p<0.01, respectively). The total number of ischemic VEBs were lowered from 667±148 in the control to 38±17 and to 56±22 in the treated group by the same concentrations (p<0.001 for both). The incidence of ischemic VT was also significantly decreased from 100% in the control group to 20 and 17% by *C. dactylon* (25 and 50µg/ml, respectively). Perfusion of the hearts at the dose of 100µg/ml produced significant reduction in the number of ischemic VEBs (p<0.05). Similar to the ischemic phase, *C. dactylon* at the concentration of 25 and 50µg/ml, significantly reduced the number and duration of reperfusion induced VT (Table 1). During reperfusion, incidences of VT was also lowered from 100% in the control group to 13 and 33% by the same concentrations (p<0.001 and p<0.05, respectively). At the same time, total VF incidences was decreased by 25 and 50µg/ml (p<0.05 for both). *C. dactylon* (25 and 50µg/ml) decreased the total number of VEBs from 349±73 (control) to 35 ± 17 (p<0.001) and 66 ± 26 (p<0.01). Perfusion of the extract (25-100µg/ml) markedly lowered reversible VF duration from 218±99sec in the control group to 0 sec, 0 sec and 10±5sec (p<0.01, p<0.01 and p<0.05, respectively).



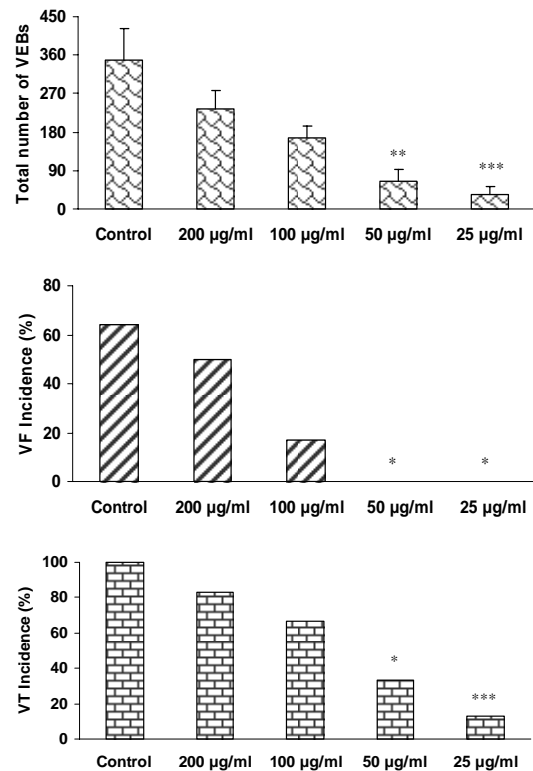
**Figure 1.** Effects of hydroalcoholic extract of rhizomes of *C. dactylon* (25-200µg/ml) on ischemic phase arrhythmias in isolated rat hearts. Total VEBs is sum of arrhythmias occurring as single, salvos and ventricular tachycardia (VT). \*\*\* p<0.001, \*\* p<0.01 and \* p<0.05 compared to the control, respectively. n= 8-12 in each group.

#### Effects on cardiac hemodynamic functions

During ischemia, LVDP was significantly increased in the groups who received 25 and 50µg/ml of the extract (p<0.05 for both) compared to the control (Table 2). However, the effect was not significant at higher concentrations of the extract. Similarly, RPP was significantly increased only in concentrations of 25 and 50µg/ml in comparison with the control group. CFR was not significantly changed throughout the ischemia. At the reperfusion phase, *C. dactylon* (25 and 50µg/ml) elevated LVDP phase (p<0.05 for both) but at higher concentrations failed to increase this parameter. While RPP in comparison to the control was not changed by *C. dactylon*, perfusion of the hearts by 25 and 50µg/ml of *C. dactylon* produced significant improvement in RPP versus other concentrations. At the same time, CFR was lowered by all concentrations of *C. dactylon* (Table 2).

#### Effects of the extract on contractility force

Perfusion of the hearts with hydroalcoholic extract of rhizomes of *C. dactylon* (25, 50, 100



**Figure 2.** Effects of hydroalcoholic extract of rhizomes of *C. dactylon* (25-200µg/ml) on reperfusion phase arrhythmias in isolated rat hearts. Total VEBs is sum of arrhythmias occurring as single, salvos and ventricular tachycardia (VT). Total ventricular reversible and irreversible fibrillation (VF) incidence was recorded. \*\*\* p<0.001, \*\* p<0.01 and \* p<0.05 compared to the control, respectively. n= 8-12 in each group.

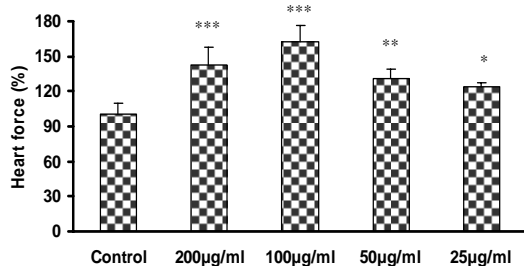
and 200µg/ml) produced potent and clear positive inotropic effects in isolated rat hearts. Compared to the control value (100), contractility force was increased by the above concentrations to 124±3% (p<0.05), 131±8% (p<0.01), 163±14% (p<0.001) and 143±15% (p<0.001), respectively (Figure 3).

## DISCUSSION

Results of present study clearly show that hydroalcoholic extract of the rhizome of *C. dactylon* produces antiarrhythmic effects against I/R-induced arrhythmias when it was used during 30min ischemia and 30min reperfusion.

Perfusion of low concentrations of the extract produced significant reduction in the number, duration and incidences of VT and the number of VEBs during I/R. In addition, *C. dactylon* produced marked reduction in reversible VF duration and incidence of total VF at reperfusion time. During both ischemia and reperfusion phases, antiarrhythmic effects of *C. dactylon* were reversely dependent on the extract concentration, where lower concentrations showed greater effects. The discrepancies between the inhibitory

effects upon low concentrations and ineffectiveness at higher concentrations of the extract might be explained by this hypothesis that some of the active constituent(s) of *C. dactylon* at high concentration may probably exhibit pro-arrhythmic properties. It is also likely that the total extract may have components with both anti- and pro-arrhythmic effects.



**Figure 3.** Effects of hydroalcoholic extract of rhizomes of *C. dactylon* (25-200 µg/ml) on heart force in isolated rat hearts. \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$  and \*  $p < 0.05$  compared to control value.  $n = 8-12$  in each group.

In addition, the low concentrations of the total extract of *C. dactylon* produced significant improvement in LVDP and RPP (as a marker for heart performance). Similar to the antiarrhythmic effects, changes in hemodynamic factors was reversely dependent on the concentration of *C. dactylon*. Consistent with the above results, our unpublished data have shown that infusion of low dose infusion of the extract improved hemodynamic factors in isolated rat hearts while higher doses produced marked but sudden elevation in the heart force followed by severe arrhythmias and heart death. Although antiarrhythmic effects of *C. dactylon* extract has not been reported, but application of similar methods for effects of other medicinal plants against I/R-induced arrhythmias are not uncommon. In another study infusion of a hydroalcoholic extract of the flowering tops of *Crataegus meyeri* in anaesthetized male rats, resulted in a significant decrease in the total number of VEBs, mainly by reduction in the

number of beats occurring as VT. Also a significant reduction in the time for VF without significant changes in the heart rate and blood pressure during the infusion of hydroalcohol and ethyl acetate extracts have been reported (24). However it has been reported that long-term application of *Crataegus oxyacantha* on I/R-induced arrhythmias did not show any cardioprotective effects neither in the heart in situ nor in the Langendorff preparations. (25). In agreement with results of this study, Nasa et al. demonstrated that *Crataegus* extract (0.05%) had cardioprotective effects on the ischemic-reperfused heart (26).

Results of this study showed marked positive inotropic effects of the extract at all concentrations. Phytochemical analysis have shown that the extract contain flavonoids, sterols and steroidal saponin (27,28). It has been reported that some plant's saponin improve cardiac function in the early stage after myocardial infraction in rats (29). Although it has been shown that different classes of flavonoids scavenge oxygen free radicals (30), since free radical scavenging activity of the extract was not significant in comparison with the standard it is unlikely that the antiarrhythmic action of the extract is directly related to its antioxidant effect.

From the results of this study, it seems that total extract of the rhizome of *C. dactylon* has some important effects on hemodynamic factors such as heart contractility, LVDP and could RPP could recover ischemic-reperfused isolated rat hearts and consequently has antiarrhythmic activity. It is also likely that *C. dactylon* has direct antiarrhythmic effects against I/R-induced arrhythmias. Future studies are required to determine the exact cardioprotective mechanism(s) of action of the extract.

#### ACKNOWLEDGEMENTS

This study was supported by the Research Affairs of Tabriz University of Medical Sciences, Tabriz, Iran.

#### REFERENCES

- Bethel CM, Sciara EB, Estill JC, Bowers JE, Hanna W, Paterson AH. A framework linkage map of bermudagrass (*Cynodon dactylon* X *transvaalensis*) based on single-dose restriction fragments. *Theor Appl genet* 2006; 112: 727-737.
- Auddy B, Ferreira M, Blasina F. Screening of antioxidant activity of three Indian medicinal plants, traditionally used for the management of neurodegenerative diseases. *J Ethnopharmacol* 2003; 84: 131-138.
- Uncini Manganelli RE, Tomei PE. Ethnopharmacobotanical studies of the Tuscan Archipelago. *J Ethnopharmacol* 1999; 65: 181-202.
- Yesilada E, Sezik E, Honda G, Takaishi Y, Takeda, Y, Tanaka T. Traditional medicine in Turkey IX: Folk medicine in north-west Anatolia. *J Ethnopharmacol* 1999; 64: 195-210.

5. Biswas TK, Mukherjee B. Plant medicines of Indian origin for wound healing activity. *Int J Low Extrem Wounds* 2003; 2: 25-39.
6. Leporatti ML, Corradi L. Ethnopharmacobotanical remarks on the Province of Chieti town (Abruzzo, Central Italy). *J Ethnopharmacol* 2001; 74: 17-40.
7. Shinwari MI, Khan MA. Folk use of medicinal herbs of Margalla Hills National Park, Islamabad. *J Ethnopharmacol* 2000; 69: 45-56.
8. Katewa SS, Guria BD, Jain A. Ethnomedicinal and obnoxious grasses of Rajasthan, India. *J Ethnopharmacol* 2001; 76: 293-297.
9. Miraldi E, Ferri S, Mostaghimi V. Botanical drugs and preparations in the traditional medicine of West Azerbaijan (Iran). *J Ethnopharmacol* 2001; 75: 77-87.
10. Mojab F, Kamalinejad M, Ghaderi N, Vahidipour HR. Phytochemical Screening of Some Species of Iranian Plants. *Iranian J Pharm Res* 2003; 2: 77-82.
11. Harborne J. *Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis*, 3<sup>rd</sup> ed. Chapman & Hall, London; 1998.
12. Nazemiyeh H. Study of the Polyphenoles of *Crataegus meyeri* A. pojark, *C. atrosanguinea* A.Pojark. and *C. curvisepala* Lindman. The PhD Thesis; 1998.
13. Wagner H, Bladt S. *Plant drug analysis: A Thin Layer Chromatography Atlas*, 2<sup>nd</sup> ed., Springer-Verlag, Berlin; 1996.
14. Stahl E. *Thin Layer Chromatography*. Springer-Verlag, Berlin; 1969.
15. Lamaison JL, Carant A. The amount of main flavonoids in flowers and leaves of *Crataegus monogyna* Jacq. and *Crataegus laevigata* (Poiret) DC.(Rosacea). *Pharm Acta Helv* 1996; 65: 315-320.
16. Takao T, Watanabe N, Yagi I, Sakata K. A simple screening method for antioxidants and isolation of several antioxidants produced by marine bacteria from fish and shellfish. *Biosci Biotechnol Biochem* 1994; 58: 1780-1783.
17. Delazar A, Shoeb M, Kumarasamy Y, Byres M, Nahar L, Modarresi M, Sarker SD. Two bioactive ferulic acid derivatives from *Eremostachys Glabra* DARU 2004; 12: 49-53.
18. Tosaki A, Engelman DT, Engelman RM, Das DK. The evolution of diabetic response to ischemia/reperfusion and preconditioning in isolated working rat hearts. *Cardiovasc Res* 1996; 31, 526-536.
19. Almeida AP, Cortes SF, Ferreira AJ, Lemos VS. Increase on the coronary flow induced by dioclein in isolated rat heart. *Life Sci* 2002; 70: 1121-1128.
20. Hausenloy DJ, Duchon MR, Yellon DM. Inhibiting mitochondrial permeability transition pore opening at reperfusion protects against ischemia-reperfusion injury. *Cardiovasc Res* 2003; 60: 617-625.
21. Trueblood NA, Ramasamy R, Wang LF, Schaefer S. Niacin protects the isolated heart from ischemia-reperfusion injury. *Am J Physiol: Heart Circ Physiol* 2000; 279: H764-771.
22. Cui J, D. Das K, Bertelli A, Tosaki A. Effects of L-carnitine and its derivatives on postischemic cardiac function, ventricular fibrillation and necrotic and apoptotic cardiomyocyte death in isolated rat hearts. *Mol Cell Biochem* 2003; 254: 227-234.
23. Walker MJA, Curtis MJ, Hearse DJ. The Lambeth conventions: Guidelines for the study of arrhythmia in ischemia, infarction and reperfusion. *Cardiovasc Res* 1988; 22: 447-455.
24. Garjani A, Nazemiyeh H, Maleki N, Valizadeh H. Effects of extracts from flowering tops of *Crataegus meyeri* A. Pojark on ischaemic arrhythmias in anaesthetized rats. *Phytother Res* 2000; 14: 428-431.
25. Nasa Y, Hashizume H, Hoque AN, Abiko Y. Protective effect of crataegus extract on the cardiac mechanical dysfunction in isolated perfused working rat heart. *Arzneimittelforschung* 1993; 43: 945-949.
26. Rothfuss MA, Pascht U, Kissling G. Effect of long-term application of *Crataegus oxyacantha* on ischemia and reperfusion induced arrhythmias in rats. *Arzneimittelforschung* 2001; 51: 24-28.
27. Aishah HS, Amri AMM, Ramlan MF, Mamat AS. Organic materials and nitrogen-potassium ratios for Bermuds tifdwarf (*Cynodon dactylon*). *Acta Hort* 1997; 450: 505-510.
28. Patil MB, Jalalpure SS, Prakash NS, Kokate CK. Antiulcer properties of alcoholic extract of *Cynodon dactylon* in rats. *Acta Hort* 2005; 680: 115-118.
29. Guo Y, Shi DZ, Yin HJ, Chen KJ. Effects of Tribuli saponins on ventricular remodeling after myocardial infarction in hyperlipidemic rats. *Am J Chin Med* 2007; 35: 309-316.
30. Middleton E Jr, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease and cancer. *Pharmacol Rev* 2000; 52: 673-751.