

## The Neuroprotective Effect of a Triazine Derivative in an Alzheimer's Rat Model

Fatemeh Alipour<sup>1,2</sup>, Shahrbanoo Oryan<sup>2</sup>, Mohammad Sharifzadeh<sup>3</sup>, Fariba Karimzadeh<sup>1,4</sup>, Laya Kafami<sup>1,5</sup>,  
Hamid Irannejad<sup>6</sup>, Mohsen Amini<sup>7</sup>, and Gholamreza Hassanzadeh<sup>8</sup>

<sup>1</sup> Shafa Neuroscience Research Center, Tehran, Iran

<sup>2</sup> Department of Animal Physiology, Faculty of Biological Sciences, Tehran University of Medical Sciences, Tehran, Iran

<sup>3</sup> Department of Pharmacology and Toxicology, School of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran

<sup>4</sup> Department of Neuroscience, School of Advanced Medical Technology, Tehran University of Medical Sciences, Tehran, Iran

<sup>5</sup> Department of Pathobiology, School of Medicine, Alborz University of Medical Sciences, Alborz, Iran

<sup>6</sup> Department of Medicinal Chemistry, School of Pharmacy, Mazandaran University of Medical Sciences, Mazandaran, Iran

<sup>7</sup> Department of Medicinal Chemistry, Drug Design & Development Research Center, Tehran University of Medical Sciences, Tehran, Iran

<sup>8</sup> Department of Anatomy, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran

Received: 4 Dec. 2013; Accepted: 25 Jan. 2014

**Abstract-** Alzheimer's disease (AD) is the most prevalent neurodegenerative disorder. It is characterized by formation of amyloid plaques and neurofibrillary tangles in the brain, degeneration of the cholinergic neurons and neural cell death. This study was aimed to investigate the effect of a triazine derivative, C16H12Cl2N3S, on learning in an Alzheimer's rat model. Animals were divided into seven groups; each group contained seven animals. Control group: animals received no surgery and treatment; saline group: animals received normal saline after recovery; sham group: animals received 10% DMSO after recovery; STZ group (Alzheimer's model): animals received streptozotocin (STZ) in four and six days after recovery; T5, T10 and T15 groups: animals were treated with triazine derivative, C16H12Cl2N3S, at doses of 5, 10 and 15  $\mu$ M, respectively. All drugs were injected intracerebroventricular. The spatial learning and histological assessment were performed in all groups. Animals in STZ group had more deficits in spatial learning than the control group in Morris water maze. C16H12Cl2N3S improved spatial learning significantly compared to STZ group. The CA1 pyramidal layer thicknesses in STZ group were reduced significantly compared to control group. C16H12Cl2N3S increased the CA1 pyramidal layer thickness in T15 group compared to STZ group. Current findings suggest C16H12Cl2N3S may have a protective effect on learning deficit and hippocampal structure in AD.

© 2015 Tehran University of Medical Sciences. All rights reserved.

*Acta Medica Iranica*, 2015;53(1):8-16.

**Keywords:** Triazine; Learning; Alzheimer's disease

### Introduction

Alzheimer's disease (AD), the most common cause of senile dementia, is a progressive, degenerative disease of the central nervous system (1). The main abnormalities of AD are intracellular twisted strands of the tau protein (neurofibrillary tangles) and deposits of extracellular beta-amyloid (A $\beta$ ) protein (2). Accumulation of A $\beta$  peptide causes an increase in the intracellular reactive oxygen species (ROS) and

apoptotic cell death (3-8). Moreover, oxidative stress has been identified as a major agent in the pathogenesis of AD (9-11). Thus, antioxidants might reduce A $\beta$ -induced neurotoxicity and cell death (12,13). Drugs used to treat AD, such as acetylcholinesterase inhibitors have side effects like hepatotoxicity and gastrointestinal disturbances, and are unable to treat AD completely (14-16). Therefore, developing a novel and effective drug, such as triazine is necessary for AD treatment. Recent studies revealed different derivatives of triazine

**Corresponding Author:** G. Hassanzadeh

Department of Anatomy, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran  
Tel: +98 912 5307515, Fax: +98 21 66419072, e-mail address: hassanzadeh@tums.ac.ir

including 1, 2, 3-triazine, 1, 2, 4-triazine and 1, 3, 5-triazine. These derivatives have different activities in inflammation ROS production, neural damage and cancer (17-24). Intraventricular administration of streptozotocin (STZ) in the brain is widely used as AD model in rats (25,26). Several studies showed various antioxidant agents prevent neural damages in different AD models (12,13). Most of the antioxidants are not permeable through the blood-brain barrier. 3-thiomethyl-5, 6-di-(4-chlorophenyl)-1, 2, 4-triazine (C16H12Cl2N3S), the most active triazine derivative, passes through the blood brain barrier. Moreover, in vitro studies have revealed the antioxidant and neuroprotective effect of C16H12Cl2N3S. Therefore, it seems to be a suitable antioxidant candidate for treatment of neuronal disorders (18). This study was aimed to investigate the effect of C16H12Cl2N3S on spatial learning and structure of CA1 in an AD in vivo model.

## Materials and Methods

### Animals

A total of 49 male adult Wistar rats (200-260 g) were housed in the animal room with 24-26 °C and 12 hours dark / light cycle (lights on from 06:00 to 18:00) with food and water ad libitum. The behavioral experiments were performed from 8 AM to 4 PM. All the experiments were carried out according to the protocol approved by the Animal Ethics Committee of Tehran University of Medical Science, Tehran, Iran.

### Stereotaxic surgery

Animals were anesthetized with ketamine (80 mg/kg) and xylazine (15 mg/kg) intraperitoneally, and their head were fixed into stereotaxic instrument (Stoelting Instruments, USA). Two stainless steel, 23-gauge guide cannulae were implanted in the lateral ventricles bilaterally. Stereotaxic coordinates were based on Paxinos and Watson atlas of the rat brain as following:

AP = -0.8 mm, ML =  $\pm$ 1.5 mm, and DV = -3 mm down from the skull surface (27). Animals were kept in cages for six days to recover.

### Experimental protocol

Animals were divided into seven groups (seven in each group). Control group: animals in this group got no surgery and treatment; saline group: animals received normal saline after recovery; sham group: animals received 10% DMSO after recovery; STZ group (Alzheimer's model): animals received streptozotocin (3 mg/kg) in fourth and sixth days after recovery; T5, T10

and T15 groups: STZ was injected in days four and six and animals were treated with C16H12Cl2N3S at doses of 5, 10 and 15  $\mu$ M, respectively in 1st, 2nd, 3rd and 5th days after recovery.

All drugs were injected intracerebroventricular (i.c.v.) in a total volume of 10  $\mu$ L in the rate of 1  $\mu$ L/min. The 27-gauge injection needle was inserted into the guide cannula. The injection needle was attached to a 10  $\mu$ L Hamilton syringe by a polyethylene tube.

### Morris water maze test

Morris water maze consists of a circular water tank with 160 cm diameter and 60 cm height, filled with non-toxic water (25  $\pm$  2 °C) to a depth of 25 cm. The pool was divided into four quadrants (North, South, East, and West) which were used as start points. An escape platform (10 cm in diameter) made of Plexiglass was placed in the middle of one of the randomly selected quadrants of the pool, 1 cm below the surface of water and kept in the same position throughout the entire experiment (north-west for this study).

Spatial learning of animals was tested 14 days after STZ infusion in Morris water maze. All animals in each group were tested (one animal at a time). Rats were trained for four days. Each animal was subjected to four consecutive trials on each day with an interval of 1 min. Each trial was started by placing the animal randomly in one of the four starting points. Animals were allowed to swim in the pool during a period of 90 second to find the hidden platform. If an animal did not find the hidden platform within this period, it was manually guided to the platform by the researcher. Then they were allowed to remain on the platform for 30 second (s). All trials were performed at about the same time in the morning (28).

Directions of the rats were recorded by a video camera located above the center of the maze that was linked to a computer. Spatial acquisition was evaluated by measuring escape latency (time to find the platform), traveled distance (path length to reach the platform), and swimming speed using the EthoVision tracking system (Noldus Information Technology, Wageningen, The Netherlands), as explained previously (29, 30). The data obtained from rats with visual impairment was excluded.

### Histological assessment

After the behavioral test, animals were deeply anesthetized with chloral hydrate (350 mg/kg; Sigma-Aldrich) and perfused transcardially with 200 ml of saline followed by 600 ml of 4% paraformaldehyde (PFA) solution. The brains were removed and kept in

## The neuroprotective effect of a triazine in AD

4% PFA for at least four days at 4 °C and the serial (8 µm) coronal sections were prepared.

Some sections were selected by random systemic sampling from each animal and stained by cresyl violet. CA1 area was studied under a light microscope (BX51, Olympus, Japan) linked to a digital camera. Digital photographs were taken using a 40X objective lens (Olympus, Japan).

### Drugs

The triazine derivative, 3-thiomethyl-5, 6-di-(4-chlorophenyl)-1, 2,4-triazine (C<sub>16</sub>H<sub>12</sub>Cl<sub>2</sub>N<sub>3</sub>S), was prepared in our laboratory as reported previously (18). The purification of the above compound was carried out using column chromatography. Crystallization from methanol- water gave the pure compound (Mp = 137-139 °C). STZ was purchased from Sigma–Aldrich, USA. DMSO was used as a vehicle for C<sub>16</sub>H<sub>12</sub>Cl<sub>2</sub>N<sub>3</sub>S dilution.

### Statistical analysis

Analysis of variance (ANOVA) was used for comparison of behavioral and histological data. A Tukey multiple comparison post-test was performed to assess differences between groups. Significance was established when the probability values were less than or equal to 0.05.

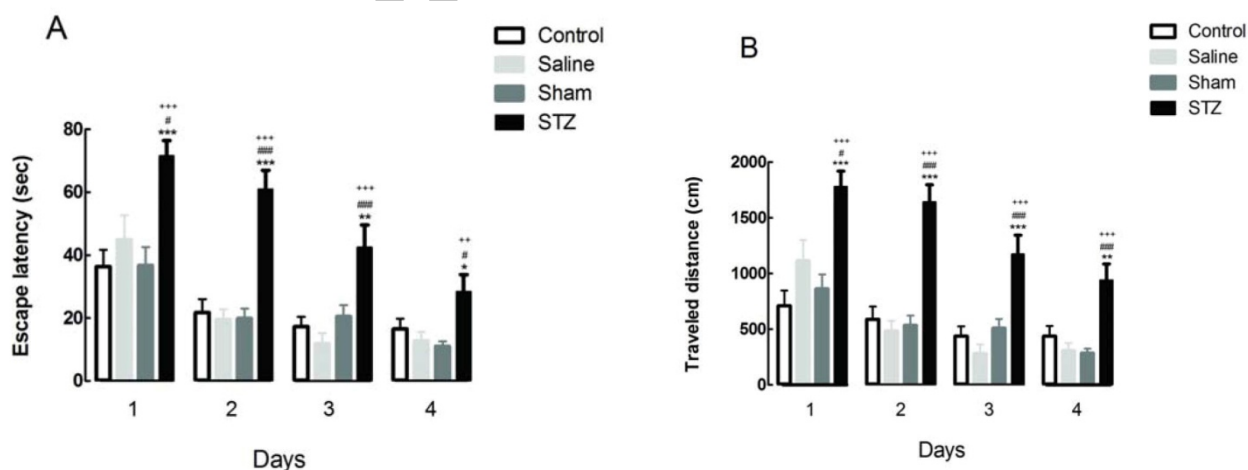
## Results

### The effect of STZ on spatial learning

The spatial learning was analyzed in control, saline, sham, and STZ groups. There were no significant differences in escape latency and traveled distance parameters between the same training days in control, saline and sham groups ( $p = 0.99$ ; Figures 1A and 1B). STZ injection significantly increased escape latency and traveled distance in the same training days compared to other groups (Table 1).

**Table 1. The mean ± S.E.M escape latency and traveled distance in every day of training in control, saline, sham, and STZ group**

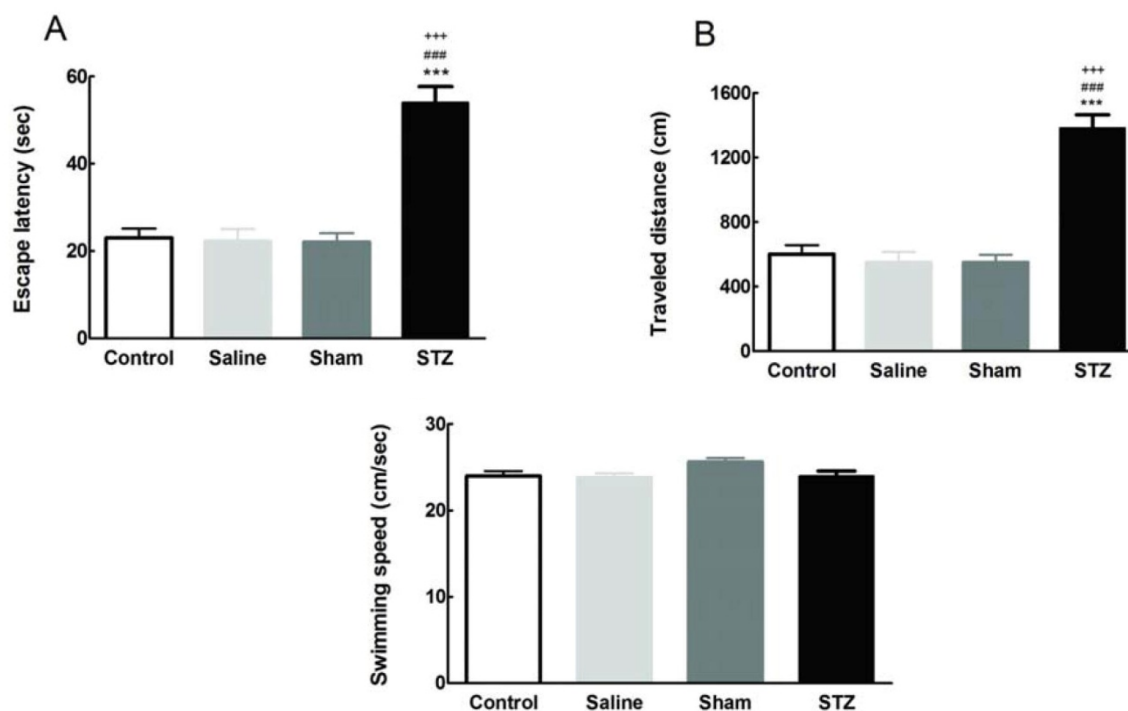
	Control	Saline	Sham	STZ
<b>Escape latency (sec)</b>				
Day 1	63.39±5.36	44.97±7.70	36.78±5.83	71.51±4.95
Day 2	21.80±4.23	19.47±3.36	19.87±3.13	60.87±6.11
Day 3	17.30±3.09	11.91±3.36	20.51±3.55	42.45±7.14
Day 4	16.59±3.21	12.80±2.81	11.04±1.55	28.39±5.42
<b>Travelled distance (cm)</b>				
Day 1	712.25±122	1117.68±183	864.89±127	1780.08±160
Day 2	590.46±113	487.57±89.12	536.09±86.54	1640.91±174
Day 3	439.25±87.33	281.9±81.95	513.31±79.93	1171.92±183
Day 4	438.76±91.85	306.93±69.72	285.28±39.86	941.37±165



**Figure 1.** Morris water maze assessment in the same training days in control, saline, sham, and STZ groups. A) The bar graphs show the quantitative results (mean ± S.E.M) of escape latency in the same training days. Escape latency of STZ group was increased significantly in the same training days compared to control, saline, and sham groups. B) The bar graphs show the quantitative results (mean ± S.E.M) of traveled distance in the same training days. Traveled distance of STZ group was increased significantly in the same training days compared to control, saline, and sham groups. \*, \*\* and \*\*\* indicate  $P < 0.05$ ,  $P < 0.01$ , and  $P < 0.001$ . \*, #, and + indicate comparing of STZ group with the control, saline, and sham groups, respectively

The average of escape latency and traveled distance in all trials during four training days were not significantly different among control, saline and sham groups ( $p = 0.96$ ; Figures 2A and 2B). The mean average of escape latency and traveled distance in all trials during four training days in STZ group increased significantly compared to the other groups ( $p < 0.001$ ). The mean escape latency was  $23.02 \pm 2.16$  s in control group,  $22.28 \pm 2.76$  s in saline group,  $22.05 \pm 2.06$  s in

sham group and  $53.93 \pm 3.39$  s in STZ group ( $p < 0.001$ ). The mean traveled distance was  $602.47 \pm 55.65$  cm in the control group,  $548.52 \pm 68.20$  cm in the saline group,  $549.89 \pm 47.73$  cm in the sham group, and  $1353.57 \pm 89.83$  cm in STZ group. The swimming speed in different experimental groups was not significantly different, indicated there was no motor activity disturbance in tested animals ( $p = 0.96$ ; Figure 2C).



**Figure 2.** The average of escape latency, traveled distance and swimming speed in all trials during four training days in control, saline, sham, and STZ groups. A) The bar graph shows the quantitative results (mean  $\pm$  S.E.M) of escape latency. The mean escape latency in STZ group was increased significantly compared to the other groups. B) The bar graph shows the quantitative results (mean  $\pm$  S.E.M) of traveled distance. The mean traveled distance in STZ group was increased significantly compared to the other groups ( $***P < 0.001$ ). Swimming speed was not significantly different between all groups. \*, \*\* and \*\*\* indicate  $P < 0.05$ ,  $P < 0.01$ , and  $P < 0.001$ . \*, #, and + indicate comparing of STZ group with the control, saline, and sham groups, respectively

### The effect of triazine on spatial learning

The effect of triazine on escape latency and travel distance in the same training days of different groups were illustrated in figures 3A and 3B, respectively. Escape latency in the first training day decreased significantly in T5 and T15 groups compared to STZ group ( $P < 0.05$ ,  $P < 0.01$ ). Escape latency in the second training day decreased in all treated groups compared to STZ group ( $P < 0.001$ ). In the third training day, escape latency decreased in T10 and T15 groups compared to STZ group ( $P < 0.05$ ,  $P < 0.01$ ).

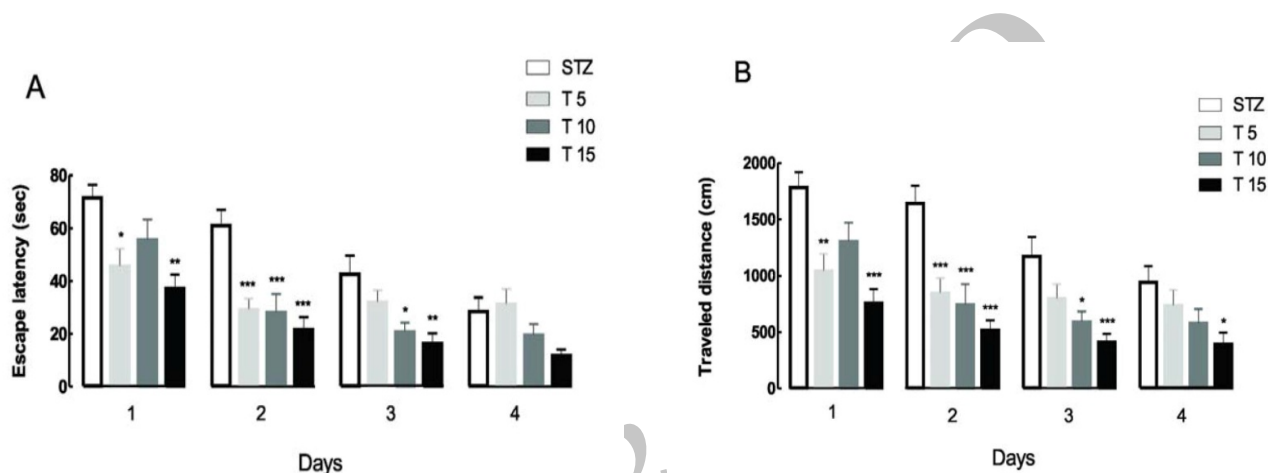
Traveled distance in the first training day decreased

in T5 and T15 groups compared to STZ group ( $P < 0.01$ ,  $P < 0.001$ ). Traveled distance in the second training day decreased in all treated groups compared to STZ group ( $P < 0.001$ ). In the third training day, traveled distance decreased in T10 and T15 groups compared to STZ group ( $P < 0.05$ ,  $P < 0.001$ ). In the fourth training day, traveled distance decreased in T15 group compared to STZ group ( $P < 0.05$ ; Table 2).

The mean average of escape latency and traveled distance in all trials during four training days were significantly reduced in T5, T10 and T15 groups compared to STZ group ( $P < 0.001$ ; Figures 4A and 4B).

**Table 2. The mean ± S.E.M escape latency and traveled distance in every day of training in STZ, T5, T10, and T15 group**

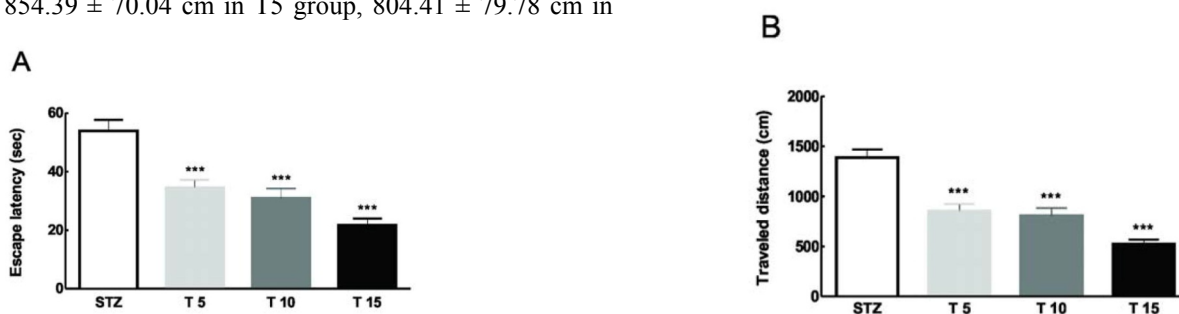
	STZ	T 5	T 10
<b>Escape latency (sec)</b>			
Day 1	71.15±4.95	45.53±6.68	55.55±7.71
Day 2	60.87±6.11	29.17±4.14	28.05±7.06
Day 3	42.45±7.14	31.85±4.66	20.60±3.70
Day 4	28.39±5.42	31.10±5.92	19.39±4.22
<b>Traveled distance (cm)</b>			
Day 1	1780.08±160	1041.58±154	1304.56±168
Day 2	1640.91±174	845.13±135	742.94±184
Day 3	1171.92±183	796.22±129	590.74±93.40
Day 4	941.37±165	734.62±141	579.39±127

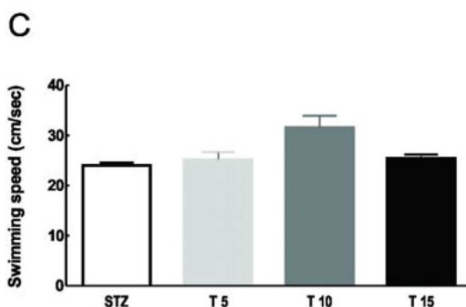


**Figure 3.** The effect of triazine on spatial learning in the same training days. A) The bar graphs show the quantitative results (mean ± S.E.M) of escape latency in the same training days. Escape latency in the first day of training decreased significantly in T5 and T15 groups compared to STZ group. Escape latency in the second day of training decreased in all of the treated groups compared to STZ group. In the third day of training, escape latency decreased in T10 and T15 groups compared to STZ group. There were no significant differences between all treated groups and STZ group in the fourth day of training. B) The bar graphs show the quantitative results (mean ± S.E.M) of traveled distance in the same training days. Traveled distance in the first day of training decreased in T5 and T15 groups compared to STZ group. Traveled distance in the second day of training decreased in all of the treated groups compared to STZ group. In the third day of training, traveled distance decreased in T10 and T15 groups compared to STZ group. In the fourth day of training, traveled distance decreased in T15 group compared to STZ group \*, \*\* and \*\*\* indicate  $P < 0.05$ ,  $p < 0.01$ , and  $P < 0.001$ .

The mean escape latency time was  $53.93 \pm 3.39$  s in STZ group,  $34.41 \pm 2.76$  s in T5 group,  $30.89 \pm 3.34$  s in T10 group, and  $21.67 \pm 2.27$  s in T15 group. The mean traveled distance was  $1383.57 \pm 89.83$  cm in STZ group,  $854.39 \pm 70.04$  cm in T5 group,  $804.41 \pm 79.78$  cm in

T10 group and  $520.37 \pm 55.41$  cm in T15 group. The mean swimming speed was not different among all tested groups, indicated there was no motor activity disturbances in tested animals ( $P = 0.96$ ; Figure 4C).





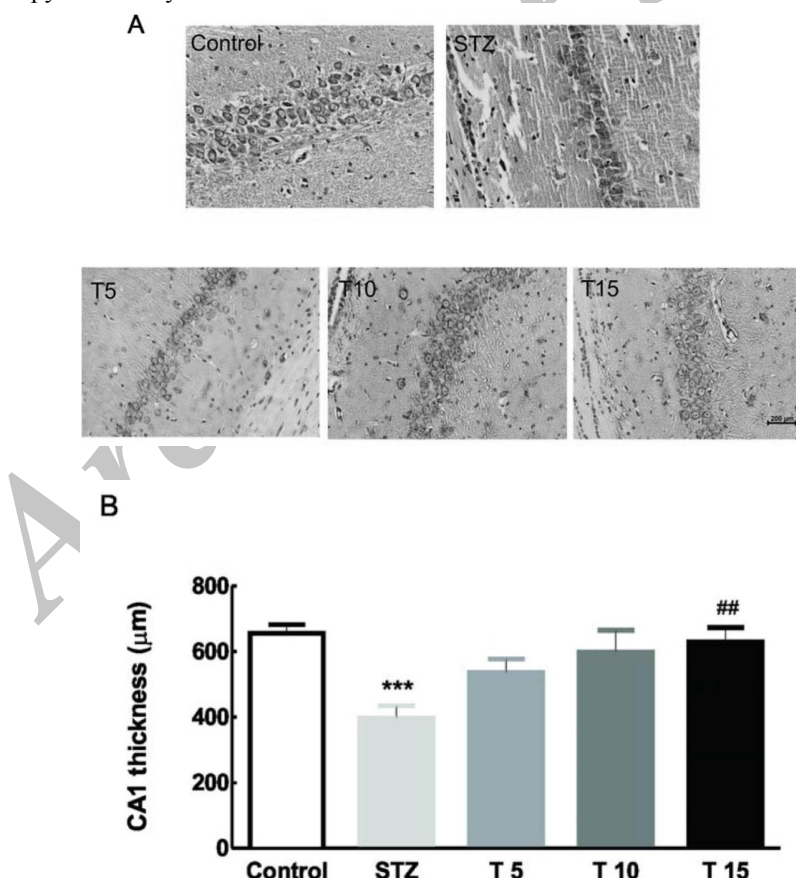
**Figure 4.** The effect of triazine on the average of escape latency, traveled distance and swimming speed in all trials during four training days.

A) The bar graph shows the quantitative results (mean ± S.E.M) of escape latency. The mean escape latency reduced significantly in T5, T10 and T15 groups compared to STZ group. B) The bar graph shows the quantitative results (mean ± S.E.M) of traveled distance. The mean traveled distance reduced significantly in T5, T10 and T15 groups compared to STZ group. \*, \*\* and \*\*\* indicate  $P < 0.05$ ,  $P < 0.01$ , and  $P < 0.001$ . Swimming speed was not significantly different among all groups

**Histological results**

To analyze the histological results, we measured the hippocampal CA1 pyramidal layer thickness in all tested groups. The CA1 pyramidal layer thickness in the STZ group was significantly less than the control group ( $P < 0.001$ ). The mean CA1 pyramidal layer thicknesses were

$654.81 \pm 27.01 \mu\text{m}$  in the control group and  $396.32 \pm 37.63 \mu\text{m}$  in STZ group ( $P < 0.001$ ). The mean CA1 pyramidal layer thickness in the T15 group was  $629.44 \pm 57.80 \mu\text{m}$  which shows a significant difference to STZ group ( $P < 0.01$ , Figures 5A and 5B).



**Figure 5.** Analysis of the CA1 pyramidal layer thickness. A) Photomicrographs of the CA1 pyramidal layer thickness. B) The bar graph shows the quantitative results (mean ± S.E.M) of the CA1 pyramidal layer thickness. The CA1 pyramidal layer thickness in STZ group was significantly less than the control group ( $***P < 0.001$ ). The CA1 pyramidal layer thickness in the T15 group was higher compared to STZ group ( $**P < 0.01$ )

## Discussion

The results of this study indicate that C16H12Cl2N3S reduced the escape latency and traveled distance parameters in Morris water maze in the animal model of AD. These results indicate that C16H12Cl2N3S can improve spatial learning in this experimental model. Moreover, it prevents the thickness reduction of CA1 pyramidal cell layer. This finding along with the previous in vitro studies revealed the neuroprotective effect of triazine derivatives in the experimental model (18-22).

STZ administration in rats induces oxidative stress in the brain, A $\beta$  plaques aggregation, Tau protein hyperphosphorylation, neuroinflammation, and apoptosis. All these pathological changes impair memory and learning in the AD animal models (26,31,32). Confirming these findings, current data shows that STZ injection causes defect of spatial memory.

Oxidative stress is the most important hypothesis involved in the pathophysiology of AD (33). Excess free radicals of oxygen lead to cellular damage, a progressive cognition and memory loss (34-36). The brain is highly sensitive to the harmful effect of oxidative stress, moreover the antioxidants play a crucial role to remove these toxic agents (36). C16H12Cl2N3S working as free radical scavenger inhibits free radical products (18).

A $\beta$  plaque is the proteolytic product of the amyloid precursor protein (34). A $\beta$  can generate the ROS especially hydrogen peroxide, which may cause cell death in the neuronal cultures and toxicity in hippocampal neurons (37,38). The reciprocal effects of oxidative stress and A $\beta$  aggregation intensify this effect (22,39-41).

It has been shown that ROS modulators, such as vitamin E, melatonin, and estrogens, reduce cellular injury and neurotoxicity during A $\beta$  exposure (42). Antioxidants and neuroprotective agents decrease the risk of memory deficit and AD progression (43,44). Current findings indicate that C16H12Cl2N3S improve spatial learning and prevents the thickness reduction of CA1 pyramidal cell layer in AD model. This effect might be due to antioxidative characteristic of this agent. C16H12Cl2N3S as a potent antioxidant might inhibit free radical products (37) and disperse A $\beta$  plaques (20,33). C16H12Cl2N3S can be a suitable therapeutic candidate for neural disorders such as AD, therefore, should be considered in the experimental and clinical trials.

## Acknowledgment

This study was supported by Tehran University of Medical Sciences and Health Services, Tehran, Iran (Grant No. 90-02-30-13162).

## References

1. Cohen E, Bieschke J, Perciavalle RM, et al. Opposing activities protect against age-onset proteotoxicity. *Science* 2006;313(5793):1604-10.
2. Alzheimer's Association. 2009 Alzheimer's disease facts and figures. *Alzheimer's Dement* 2009;5(3):234-70.
3. Marcus DL, Thomas C, Rodriguez C, et al. Increased peroxidation and reduced antioxidant enzyme activity in Alzheimer's disease. *Exp Neurol* 1998;150(1):40-4.
4. Zheng L, Kågedal K, Dehvari N, et al. Oxidative stress induces macroautophagy of amyloid  $\beta$ -protein and ensuing apoptosis. *Free Radic Biol Med* 2009;46(3):422-9.
5. Behl C, Holsboer F. Oxidative stress in the pathogenesis of Alzheimer's disease and antioxidant neuroprotection. *Fortschr Neurol Psychiatr* 1998;66(3):113-21.
6. Peng QL, Buz'Zard AR, Lau BH. Pycnogenol protects neurons from amyloid- $\beta$  peptide-induced apoptosis. *Brain Res Mol Brain Res* 2002;104(1):55-65.
7. Tamagno E, Parola M, Guglielmotto M, et al. Multiple signaling events in amyloid beta-induced, oxidative stress-dependent neuronal apoptosis. *Free Radic Biol Med* 2003;35(1):45-58.
8. Xiao XQ, Zhang HY, Tang XC. Huperzine A attenuates amyloid  $\beta$ -peptide fragment 25-35-induced apoptosis in rat cortical neurons via inhibiting reactive oxygen species formation and caspase-3 activation. *J Neurosci Res* 2002;67(1):30-6.
9. Andersen JK. Oxidative stress in neurodegeneration: cause or consequence? *Nat Med* 2004;10(Suppl):S18-25.
10. Butterfield DA, Perluigi M, Sultana R. Oxidative stress in Alzheimer's disease brain: new insights from redox proteomics. *Eur J Pharmacol* 2006;545(1):39-50.
11. Nunomura A, Perry G, Aliev G, et al. Oxidative damage is the earliest event in Alzheimer disease. *J NeuropatholExp Neurol* 2001;60(8):759-67.
12. Aliev G, Obrenovich ME, Reddy VP, et al. Antioxidant therapy in Alzheimer's disease: theory and practice. *Mini Rev Med Chem* 2008;8(13):1395-406.
13. Lin YH, Liu AH, Wu HL, et al. Salvianolic acid B, an antioxidant from *Salvia miltiorrhiza*, prevents Abeta(25-35)-induced reduction in BPRP in PC12 cells. *Biochem Biophys Res Commun* 2006;348(2):593-9.

14. Mukherjee PK, Kumar V, Mal M, et al. Acetylcholinesterase inhibitors from plants. *Phytomedicine* 2007;14(4):289-300.
15. Knapp MJ, Knopman DS, Solomon PR, et al. A 30-week randomized controlled trial of high-dose tacrine in patients with Alzheimer's disease. *JAMA* 1994;271(13):985-91.
16. Schulz V. Ginkgo extract or cholinesterase inhibitors in patients with dementia: what clinical trials and guidelines fail to consider. *Phytomedicine* 2003;10(Suppl 4):74-9.
17. Saxena S, Verma M, Saxena AK, et al. Triazines as anti-inflammatory agents. *Arzneimittelforschung* 1994;44(6):766-9.
18. Irannejad H, Amini M, Khodagholi F, et al. Synthesis and in vitro evaluation of novel 1, 2, 4-triazine derivatives as neuroprotective agents. *Bioorg Med Chem* 2010;18(12):4224-30.
19. Shaykhalishahi H, Taghizadeh M, Yazdanparast R, et al. Anti-amyloidogenic effect of AA3E2 attenuates  $\beta$ -amyloid induced toxicity in SK-N-MC cells. *Chem Biol Interact* 2010;186(1):16-23.
20. Shaykhalishahi H, Yazdanparast R, Ha HH, et al. Inhibition of H<sub>2</sub>O<sub>2</sub>-induced neuroblastoma cell cytotoxicity by a triazine derivative, AA3E2. *Eur J Pharmacol* 2009;622(1-3):1-6.
21. Tusi SK, Ansari N, Amini M, et al. Attenuation of NF- $\kappa$ B and activation of Nrf2 signaling by 1, 2, 4-triazine derivatives, protect neuron-like PC12 cells against apoptosis. *Apoptosis* 2010;15(6):738-51.
22. Yazdanparast R, Shaykhalishahi H. Protective effect of a triazine-derivative (AA3E2) on  $\beta$ -amyloid-induced damages in SK-N-MC cells. *Toxicol Vitro* 2009;23(7):1277-83.
23. Bekircan O, K x k M, Kahveci B, et al. Convenient Synthesis of Fused Heterocyclic 1, 3, 5-Triazines from Some N-Acyl Imidates and Heterocyclic Amines as Anticancer and Antioxidant Agents. *Archiv Pharm* 2005;338(8):365-72.
24. Brzozowski Z, S czewski F. Synthesis and antitumor activity of novel 2-amino-4-(3, 5, 5-trimethyl-2-pyrazolino)-1, 3, 5-triazine derivatives. *Eur J Med Chem* 2002;37(9):709-20.
25. Veerendra Kumar MH, Gupta YK. Effect of *Centella asiatica* on cognition and oxidative stress in an intracerebroventricular streptozotocin model of Alzheimer's disease in rats. *Clin Exp Pharm Physiol* 2003;30(5-6):336-42.
26. Grunblatt E, Salkovic-Petrisic M, Osmanovic J, et al. Brain insulin system dysfunction in streptozotocin intracerebroventricularly treated rats generates hyperphosphorylated tau protein. *J Neurochem* 2007;101(3):757-70.
27. Paxinos G, Watson C, editors. *The rat brain in stereotaxic coordinates*. 6th ed. San Diego: Academic Press; 2007.
28. Saffarzadeh F, Eslamizade MJ, Nemati Karimooy HA, et al. The effect of L-arginine on Morris water maze tasks of ovariectomized rats. *Acta Physiol Hung* 2010;97(2):216-23.
29. Sharifzadeh M, Naghdi N, Khosrovani S, et al. Post-training intrahippocampal infusion of the COX-2 inhibitor celecoxib impaired spatial memory retention in rats. *Eur J Pharm* 2005;511(2-3):159-66.
30. Tabrizian K, Najafi S, Belaran M, et al. Effects of Selective iNOS Inhibitor on Spatial Memory in Recovered and Non-recovered Ketamine Induced-anesthesia in Wistar Rats. *Iran J Pharm Res* 2010;9(3):313-20.
31. Sharma VK. Streptozotocin, An experimental tool in diabetes and alzhemir's disease. *Int J Pharm Res Deveopment* 2010;2(1):1-7.
32. Salkovic-Petrisic M, Hoyer S. Central insulin resistance as a trigger for sporadic Alzheimer-like pathology: an experimental approach. *J Neural Transm Suppl* 2007;(72):217-33.
33. Selkoe DJ. Alzheimer's disease: genes, proteins, and therapy. *Physiol Rev* 2001;81(2):741-66.
34. Maiese K, Chong ZZ. Insights into oxidative stress and potential novel therapeutic targets for Alzheimer disease. *Restor Neurol Neurosci* 2004;22(2):87-104.
35. Mattson MP. Pathways towards and away from Alzheimer's disease. *Nature* 2004;430(7000):631-9.
36. Maiese K, editor. *Organic brain disease*. Encyclopedia of the human brain. 1st ed. Oxford: Elsevier Inc; 2002: p. 509-27.
37. Behl C, Davis J, Lesley R, et al. Hydrogen peroxide mediates amyloid beta protein toxicity. *Cell* 1994;77(6):817-27.
38. Varadarajan S, Yatin S, Kanski J, et al. Methionine residue 35 is important in amyloid beta-peptide-associated free radical oxidative stress. *Brain Res Bull* 1999;50(2):133-41.
39. Behl C, Holsboer F. Oxidative stress in the pathogenesis of Alzheimer's disease and antioxidant neuroprotection. *Fortschr Neurol Psychiatr* 1998;66(3):113-21.
40. Misonou H, Morishima-Kawashima M, Ihara Y. Oxidative stress induces intracellular accumulation of amyloid beta-protein (A $\beta$ ) in human neuroblastoma cells. *Biochemistry* 2000;39(23):6951-9.
41. Zheng L, Kagedal K, Dehvari N, et al. Oxidative stress induces macroautophagy of amyloid beta-protein and ensuing apoptosis. *Free Radic Biol Med* 2009;46(3):422-9.
42. Subramaniam R, Koppal T, Green M, et al. The free radical antioxidant vitamin E protects cortical synaptosomal membranes from amyloid beta-peptide(25-35) toxicity but not from hydroxynonenal toxicity: relevance to the free radical hypothesis of Alzheimer's disease. *Neurochem Res* 1998;23(11):1403-10.



### The neuroprotective effect of a triazine in AD

43. Rasoolijazi H, Joghataie MT, Roghani M, et al. The beneficial effect of (-)-epigallocatechin-3-gallate in an experimental model of Alzheimer's disease in rat: A behavioral analysis. *Iran Biomed J* 2007;11(4):237-43.
44. Gilgun-Sherki Y, Rosenbaum Z, Melamed E, et al. Antioxidant therapy in acute central nervous system injury: current state. *Pharmacological reviews*. 2002;54(2):271-84.

Archive of SID