## Polycyclic Aromatic Hydrocarbons and Esophageal Squamous Cell Carcinoma

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#### Abstract

Esophageal cancer (EC) is the 8<sup>th</sup> most common cancer and the 6<sup>th</sup> most frequent cause of cancer mortality worldwide. Esophageal squamous cell carcinoma (ESCC) is the most common type of EC. Exposure to polycyclic aromatic hydrocarbons (PAHs) has been suggested as a risk factor for developing ESCC. In this paper we will review different aspects of the relationship between PAH exposure and ESCC.

PAHs are a group of compounds that are formed by incomplete combustion of organic matter. Studies in humans have shown an association between PAH exposure and development of ESCC in many populations. The results of a recent case-control study in a high risk population in northeastern Iran showed a dramatic dose-response relationship between PAH content in non-tumor esophageal tissue (the target tissue for esophageal carcinogenesis) and ESCC case status, consistent with a causal role for PAH exposure in the pathogenesis of ESCC.

Identifying the main sources of exposure to PAHs may be the first and most important step in designing appropriate PAH-reduction interventions for controlling ESCC, especially in high risk areas. Coal smoke and drinking mate have been suggested as important modifiable sources of PAH exposure in China and Brazil, respectively. But the primary source of exposure to PAHs in other high risk areas for ESCC, such as northeastern Iran, has not yet been identified. Thus, environmental studies to determining important sources of PAH exposure should be considered as a high priority in future research projects in these areas.

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## Introduction

E sophageal cancer (EC) is the 8<sup>th</sup> most common cancer and the 6<sup>th</sup> most frequent cause of cancer mortality worldwide, causing over 400,000 deaths in 2008.<sup>1</sup> 80% of esophageal cancers occur in developing countries. Esophageal cancer rates vary greatly between and within countries. There are two large bands of high risk, across central Asia from the Caspian Sea to northern China (the Central Asian Esophageal Cancer Belt) and from eastern to southern Africa.<sup>2</sup> Within these bands, EC incidence rates are 50 – 100/100,000 population/year, whereas in most other areas of the world they are < 10/100,000/year.

At the eastern end of the Central Asian Esophageal Cancer Belt is Iran. The highest rates of EC in Iran occur in Golestan Province, in the northeast of the country. In the 1970s, scientists from Iran and the International Agency for Research on Cancer (IARC) recorded EC incidence rates of 165/100,000/year in men and 195/100,000/year in women,<sup>3</sup> the highest rates of EC ever recorded in the world, and they began initial studies to determine its cause, but sociopolitical events in Iran in 1979 stopped these studies. Since the 1970s, EC rates in Golestan have declined,<sup>4,5</sup> but despite this decline, EC remains the most frequent malignancy in northeastern Iran,<sup>4-6</sup> and the results of the Golestan populationbased Cancer Registry show that the truncated ASR of EC is still very high, especially in the eastern parts of Golestan Province (Gonbad and Kalaleh).<sup>4,5</sup>

Adenocarcinoma and squamous cell carcinoma are the main histologic subtypes of EC. In the 1960s, about 90% of EC cases throughout the world were esophageal squamous cell carcinoma (ESCC).<sup>2</sup> Recent studies, however, have shown a declining trend in the rate of ESCC and a sharply increasing trend in the rate of esophageal adenocarcinoma in the United States and some European countries.<sup>7–9</sup> But the situation seems to be different in the developing world, where all of the highest risk populations are. Reports from developing countries have not shown the changing pattern of EC histologies that has been seen in western countries. For example, ESCC comprises more than 90% of EC cases in northeastern Iran.<sup>6,10</sup> So, ESCC is the most important type of EC in the developing world and a major health problem in high risk areas including northeastern Iran.

ESCC develops by progression from dysplastic lesions within the squamous epithelium of the esophagus <sup>2</sup>. In studies in a highrisk population in China, esophageal squamous dysplasia (ESD) was shown to be the only clinically important precursor lesion for ESCC, and different histologic grades of ESD identified different levels of ESCC risk.<sup>11,12</sup>

Various factors have been shown to increase the risk of ESCC,<sup>13</sup> including tobacco smoking,<sup>14,15</sup> heavy alcohol drinking,<sup>14</sup> opium consumption,<sup>15</sup> nass chewing,<sup>15</sup> hot tea consumption,<sup>16</sup> mate drinking,<sup>17</sup> low intake of fruits and vegetables,<sup>18</sup> nutrient deficiencies,<sup>19–21</sup> tooth loss,<sup>22</sup> and low socioeconomic status.<sup>23</sup> Alcohol and tobacco are the most important risk factors for ESCC in low-risk populations,<sup>24</sup> but its etiology in high risk areas is less clear. Exposure to polycyclic aromatic hydrocarbons (PAHs) has been suggested as an important risk factor for developing ESCC in many

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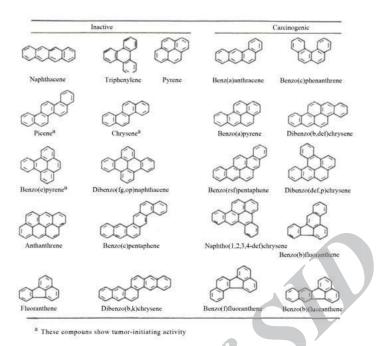


Figure 1. Structures of carcinogenic and noncarcinogenic unsubstituted polycyclic aromatic hydrocarbons (Adopted from: Harvey, 1991).

populations.<sup>13</sup> The aim of this paper is to discuss different aspects of the relationship between PAH exposure and ESCC.

## **Characteristics of PAHs**

PAHs are a group of compounds that are formed by incomplete combustion of organic matter, including vegetation, fossil fuels and oil products.<sup>25,26</sup> They are composed of two or more condensed aromatic rings<sup>27</sup> (figure 1), and they are distributed across gaseous and particulate phases.<sup>28</sup> In the environment, they are found in soil, water, air and food.<sup>29</sup> Exposure to PAHs may occur through inhalation, ingestion or percutaneous penetration.<sup>28,29</sup> They distribute to various organs, especially the liver.<sup>29</sup> PAHs themselves are chemically inert and hydrophobic, but they are metabolized within cells to their active forms, the diol-epoxides. This change results in PAHs binding to cellular macromolecules such as DNA,<sup>27,30</sup> causing the formation of PAH-DNA adducts, which lead to mutations in proto-oncogenes and tumour-suppressor genes.<sup>28</sup> Generating other reactive intermediates by a oneelectron oxidation process is another carcinogenic mechanism of PAHs. This process may cause chemical alkylation of DNA and result in potentially mutagenic depurination.31

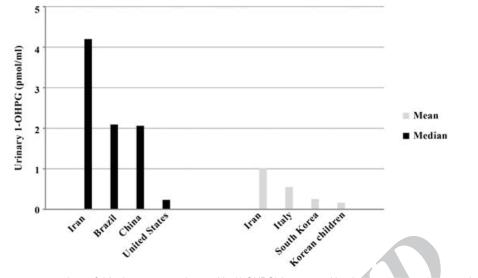
## **Determination of exposure to PAHs**

Two ways to assess the level of exposure to PAHs are measurement of the external dose and measurement of the internal dose. The external dose can be quantified by monitoring the concentration of PAHs in the air and food. The internal dose, reflecting the total uptake of PAHs, may be assessed by monitoring of PAHs or their metabolites in body fluids or tissues.<sup>32</sup> Therefore, biological monitoring, making an accurate estimation of the individual internal dose, has been considered as useful indicator for human exposure to PAHs.<sup>33</sup> Urinary 1-hydroxypyrene concentration is a valid marker for assessing the level of recent exposure to PAHs.<sup>32,34-36</sup> It has also been reported to be a sensitive biomarker for low dose PAH exposure.<sup>36,37</sup> Zhao et al. reported that the maximum concentration of 1-hydroxypyrene in urine samples was found in the evening (21:00 - 23:00), and recommended this as a suitable time for sample collection for evaluation of PAHs.<sup>38</sup> Urinary concentration of 1-hydroxypyrene glucoronide (1-OHPG) has also been shown to be significantly higher in individuals exposed to high levels of PAH.<sup>39,40</sup> In a study of 10 healthy non-smoking men, 24 hours after starting a high-PAH diet (charbroiled beef), the level of urinary 1-OHPG increased 10 – 80 fold above baseline levels. Urinary concentration of this marker decreased to near baseline levels by 1 - 3 days after ending the diet.<sup>40</sup>

Seidel et al. have suggested hydroxylated metabolites of naphthalene, fluorene, phenanthrene, fluoranthene and pyrene as valid biomarkers for monitoring PAH exposure.<sup>41–45</sup> They were also reported to be valid biomarkers of occupational exposure to PAHs,<sup>46</sup> especially lowly condensed volatile PAHs.<sup>43</sup> It has been suggested that measuring hydroxyphenanthrene in addition to 1-hydroxypyrene may provide more complete information about PAH exposure in individuals.<sup>43</sup>

Urinary levels of unmetabolised naphthalene and phenanthrene have also been shown to be promising surrogates for assessing PAH exposure.<sup>45</sup> Naphthalene was suggested as a good biomarker for assessing occupational exposure to PAHs.<sup>28</sup>

PAH-DNA adducts are more persistent indicators of PAH exposure than are urinary markers.<sup>36</sup> The levels of PAH-DNA adducts in easily accessible WBCs have been suggested as valid biomarkers in exposure assessment as well as intervention studies.<sup>47</sup> The level of this marker in peripheral WBCs has been reported to reflect both occupational and dietary exposure to PAHs.<sup>48–50</sup> There was a positive association between the level of PAH-DNA ad-



**Figure 2.** Urinary concentrations of 1-hydroxy pyrene glucuronide (1-OHPG) in non-smoking individuals from Golestan, Iran;<sup>84</sup> Rio Grande do Sul, Brazil;<sup>80</sup> Linxian, China;<sup>83</sup> Maryland, United States;<sup>40</sup> Turin, Italy;<sup>116</sup> Pohang, South Korea;<sup>39</sup> and Korean children.<sup>118</sup>

ducts in WBCs and recent exposure to PAHs from consumption of charbroiled food<sup>51</sup> or smoking.<sup>52</sup> But, as Gyorffy et al. showed, the levels of PAH-DNA adducts in WBCs may not appropriately reflect its levels in target tissues.53 So, tissue levels of PAH-DNA adducts are considered a more valid indicator of exposure to PAH in recent studies. van Gijssel et al. detected PAH-DNA adducts in old paraffin-embedded endoscopic biopsies of the esophagus from Linxian, China.54 They observed nuclear PAH-DNA adduct staining in four out of five samples, and concluded that old biopsy samples are appropriate for assessing past exposure to PAHs.54 There are some limitations in using PAH-DNA adducts in epidemiologic studies, including non-linearity at high exposures, lack of sensitivity to low exposures, the large amounts of DNA needed for PAH-DNA adduct measurements, the problem of inter- and intra-lab variability in results using the 32P-postlabeling measurement technique, and the use of surrogate tissues (e.g. WBCs) instead of the target tissues.<sup>13</sup> Nevertheless, these markers have been successfully used in establishing links between PAH exposure and risk of several cancers.

Toriba et al. identified PAHs in human hair samples by HPLC with fluorescence detection. They reported that PAH analysis in hair samples is a good biomarker for assessing PAH exposure.<sup>55</sup> Beskid et al. used fluorescence in situ hybridization (FISH) with whole chromosome painting to assess the levels of exposure to carcinogenic PAHs.<sup>56</sup>

In a recent study from Iran, immunohistochemical staining of target tissue with an anti-PAH antibody (8E11) was successfully used for assessing PAH exposure.<sup>57</sup> It would be interesting to know if such measurements are correlated with the prevalence of genetic mutations or levels of PAH-DNA adducts in the same tissue or in a more accessible tissue such as buccal epithelium.<sup>58</sup> Answers to these questions would be helpful for conducting future epidemiological studies and surveillance programs in high risk areas.

## PAHs and cancers

Some PAH compounds, including benzo-a-anthracene, benzob-fluoranthene, benzo-j-fluoranthene, benzo-k-fluoranthene, benzo-a-pyrene, dibenz-a,h-anthracene, and indeno-1,2,3-c,d-pyrene, are considered to be human carcinogens.<sup>36,59</sup> PAHs have been shown to induce carcinogenic changes in a number of human tissues. The report of Percivall Pott in 1775 was the first evidence of an association between PAH (soot) exposure and cancer (scrotal cancer) in chimney sweeps.<sup>60</sup> PAH exposure has now been documented to increase the risk of cancers of the lung,<sup>61,62</sup> bladder,<sup>61</sup> breast,<sup>63</sup> pancreas<sup>64,65</sup> and prostate.<sup>65</sup> PAH mutagenesis was also found in human epidermal keratinocytes in culture.<sup>66</sup> Benzo-a-pyrene, the most common PAH to cause cancer in human, has been classified as a group 1 carcinogen by IARC.<sup>59</sup> Recently, IARC also reported that diesel engine exhaust (an important source of exposure to PAHs) has a carcinogenic effect in humans and classified it as a group 1 carcinogen.<sup>67</sup> So PAHs have been shown to have carcinogenic effects in a number of human organs.

## **PAH-induced P53 mutations**

P53 is a tumor suppressor gene which is frequently mutated in human cancers. P53 mutations occur in about 40% of lung cancers.<sup>68</sup> Smoking tobacco, which contains many carcinogens, including PAHs, has been shown to cause lung cancer. So, PAHinduced P53 mutation was suggested as a possible mechanism in the pathogenesis of lung cancer. PAH-induced P53 mutations are mostly G:C to T:A transversions,<sup>69</sup> and they are poorly repaired. Puisieux et al. showed the direct role of PAHs in the induction of mutations in the P53 gene in lung cancer for the first time.<sup>69</sup> Recently, Gao et al. suggested that PAHs may have the potential to induce P53 mutations in the cervix of mice.<sup>70</sup> It has been proposed that the most common locations of P53 mutations, known as mutational hotspots, are preferred targets for PAHs.<sup>69</sup> Other studies similarly reported that PAH-DNA adducts preferentially occur at p53 mutational hotspots.<sup>71</sup> Shen et al. reported P53 mutation due to formation of PAH o-quinones, one of the ultimate carcinogens of PAHs, in cigarette smokers.72 It was suggested that PAH o-quinones cause P53 inactivation by generating endogenous reactive oxygen species.73 P53 mutation was reported to be attributed to direct PAH-induced DNA damage rather than selection of pre-existing endogenous mutations.<sup>68</sup> Because of the clear causal relationship between tobacco smoking and lung cancer, most previous investigations about PAH-induced P53 mutations have been conducted on lung cancer. But other malignancies, including ESCC, are also known to have a strong relationship with tobacco smoke carcinogens, especially PAHs. Brennan et al. reported a fourfold increase in the prevalence of P53 mutations in heavy smoking EC patients as compared to nonsmokers.<sup>74</sup> The results of a recent study in northeastern Iran also suggested PAH-induced P53 mutations in ESCC cases. In this study, G:C to A:T transversions were the most common type of mutation, followed by G:C to T:A transversions.<sup>75</sup> Further investigations are needed to clarify if PAH-induced P53 mutations play a major role in the pathogenesis of ESCC, as they do in the case of lung cancer.

## Role of PAHs in the pathogenesis of ESCC

More than half of cancers occur in surface epithelia (such as the esophageal epithelium) that are in direct contact with the external environment.<sup>76</sup> Direct exposure to PAHs has been suggested to contribute significantly to carcinogenesis in these tissues.<sup>66</sup> A dose-response relationship was found between PAH exposure and development of EC in mice.77 Gustavsson et al. reported a significant excess of esophageal cancer in chimney sweeps, and thought that the most probable explanation was a direct action of PAHs on the esophageal mucosa.<sup>61</sup> The results of studies from Canada,<sup>65</sup> Germany,<sup>78</sup> Sweden<sup>79</sup> and Brazil<sup>80</sup> have also shown an excess risk of esophageal cancer in PAH-exposed individuals. Barbecuing and grilling, important sources of exposure to PAHs, have also been associated with an elevated risk of esophageal cancer.<sup>81</sup> Roth et al. found high levels of PAHs in food samples obtained from a high risk area for ESCC in China,<sup>82</sup> and high levels of urine 1-OHPG in the local inhabitants,<sup>83</sup> suggesting a possible role for PAH exposure in ESCC pathogenesis there. Exposure to widespread and very high levels of PAHs was also reported from a high risk area for ESCC in the northeast of Iran<sup>84</sup> (Figure 2). Marjani et al. reported significantly higher levels of PAH-DNA adducts in ESCC tumor tissue than in normal esophageal tissues from these same cases or from control individuals, and concluded that PAHs are risk factors for ESCC in this same population in Golestan Province, Iran.<sup>85</sup> And in another recent study from this area, immunohistochemical staining with an anti-PAH antibody (8E11) was performed on non-tumoral esophageal epithelial tissues obtained from ESCC patients and normal controls.57 There was a striking dose-response relationship between the intensity of 8E11 and ESCC case status, suggesting a causal role for PAH exposure in the pathogenesis of ESCC57 (Table 1). The effects of inherited genetic variation on the levels of such PAH adducts may also be important in this context,86 and need to be further investigated in future studies.

### Sources of exposure to PAHs

#### 1. Dietary sources

Foods had been identified as important sources of human exposure to PAHs.<sup>87</sup> Diet is usually the main source of PAH exposure in non-smoking individuals who are not exposed to occupational PAHs.<sup>25,26</sup> Contamination of foods by PAHs may occur during food processing, such as smoking or cooking, or as a result of accidental environmental contamination, such as atmospheric pollution of vegetables.88,89

Agricultural crops may be an important source of exposure to PAHs.<sup>90</sup> By accumulating on the surface of plants exposed to polluted air, PAHs may be present in most crops.<sup>91</sup> Because of the hydrophobic nature of PAHs, it is believed that the contamination of agricultural crops by these compounds comes mainly from the air rather than from the soil.<sup>92</sup>

The Total Human Environmental Exposure Study from the US showed that foods account for more than 96% of the daily intake of carcinogenic PAHs.93 Vyskocil et al. reported no relationship between PAH levels in air and urinary concentration of 1-hydroxypyrene, suggesting that other factors (e.g. dietary intake of PAHs) may be potentially more important sources of internal dose.94 An analysis of UK food samples reported that cereals and oils/fats were the most important source of PAHs in the UK diet, followed by fruits, sugar and vegetables.<sup>89</sup> De Vos et al. assessed Dutch total diet samples and reported that sugar and sweets, cereals, oils, fats and nuts were the major sources of PAHs.95 The results of a study from Italy also showed that food was the main source of PAH exposure, suggesting cereals, milk products, meat, vegetables and fruits as the major contributors to total PAH intake.<sup>96</sup> Falco et al. assessed the dietary intake of PAHs in Catalonia, Spain. They found the highest levels of PAHs in cereals, meat and meat products.97 Chemical analysis of foodstuffs in northeastern Iran in 1970s did not show high levels of PAHs in the major dietary items.98 But in another study from this region, Hakami et al. suggested that diet is a major source of exposure to PAH99 (Table 2).

The level of PAH in food may be affected by methods of food processing and preparation. Some studies reported no change in the PAH content of flour during the bread baking process. For example, Dennis et al. reported similar amounts of PAHs in wheat flour and bread made from this flour.<sup>100</sup> And high levels of PAHs were reported in both raw and cooked food samples from a high risk area of ESCC in China.82 But many other studies showed that some methods of food processing have documented effects on PAH levels in food. High levels of PAHs have been reported in smoked foods.<sup>101</sup> The concentration of PAHs in smoked meat products was shown to be more dangerous when an uncontrolled smoking process (without appropriate technology) is used.<sup>29</sup> Charbroiled foods have been shown to contain some of the highest PAH levels among frequently used foods, suggesting them as a major source of dietary exposure to PAHs in populations which commonly charbroil their food.<sup>51</sup> The results of a study on Italian foods showed the highest levels of PAHs in pizza prepared in a wood-burning oven and in barbecued meat.96 Eating meat prepared by grilling, roasting or broiling was also reported as an important source of exposure to PAHs.<sup>102</sup> The type of heat source (direct versus indirect), the geometry of the grill, grilling duration, the use of marinating sauces, and the fat content of the food were some contributors to PAH formation in grilled and smoked foods.<sup>103</sup> In a recent study from northeastern Iran (unpublished data), there was a significant relationship between cooking method and the level of PAH-DNA adducts in peripheral lymphocytes. The results showed that PAH exposure is significantly higher in individuals using deep frying methods for cooking foods. Meat tikka, whole grilled chicken, meat burgers and grilled vegetables were major dietary sources of exposure to PAHs in Kuwait.103

Drinking mate, a water infusion of the herb yerba mate (*Ilex paraguayensis*), is a common habit in southern South Ameri-

Table 1. Adjusted ORs and 95% CIs for the association between staining intensity of monoclonal antibody 8E11 and esophageal squamous cell
carcinoma in the Golestan Case-Control Study <sup>1</sup>

Quintile	Controls (n)	Cases (n)	Adjusted OR (95% CI) <sup>2</sup>
1	20	2	Reference
2	21	6	2.49 (0.41 – 15.3)
3	21	14	5.59 (1.03 - 30.2)
4	21	19	10.9 (2.07 – 57.4)
5	20	50	26.8 (5.28 - 136)
<sup>1</sup> From Abedi-Ardekani B et al. <i>Gut.</i> 2010: <b>59:</b> 1178–1183: <sup>2</sup> Adjusted for age, sex, education, ethnicity, tobacco use and opium use			

Table 2. Benzo[a]pyrene (BaP	) intako from broad	rice and water in	Colectan and Ear	e provinces Iran1

Daily Benzo(a)pyrene intake (ng/day)			
Cases (n=40)	Golestan controls (n=40)	Fars controls (n=40)	
99.0±39.3*	91.4±31.6	70.6±27.1°	
60.9±31.8ª	48.1±20.4	28.4±14.9 <sup>d</sup>	
24.7±16.8ª	33.5±21.2	34.6±18.9	
13.4±6.7 <sup>b</sup>	9.7±4.3	▲ 7.5±2.9 <sup>e</sup>	
	99.0±39.3* 60.9±31.8 <sup>a</sup> 24.7±16.8 <sup>a</sup>	Cases (n=40) Golestan controls (n=40)   99.0±39.3* 91.4±31.6   60.9±31.8* 48.1±20.4   24.7±16.8* 33.5±21.2	

<sup>1</sup>From Hakami R et al., Nutrition and Cancer 2008;**60**:216–221; \*mean $\pm$ SD;\*P<0.05, <sup>b</sup> P<0.001 between cases and Golestan controls; \*P<0.01, <sup>d</sup> P<0.001, <sup>e</sup> P<0.05 between the two control groups.

Table 3. The main and other sources of PAH exposure in the West and in different populations with a high risk of ESCC

Population	Main sources	Other sources
West	Tobacco smoke	Car exhaust, Coal smoke
China	Coal smoke	Tobacco smoke, Car exhaust
Iran	Unknown	Tobacco smoke, Car exhaust
Brazil	Tobacco smoke, Mate	Barbequed meat, Car exhaust

ca.<sup>104</sup> In previous studies, both the temperature and the quantity of mate consumption have been associated with increased risk of ESCC.17,105 Mate drinking has also been associated with increased risk of other smoking-related cancers, including cancers of the lung,<sup>106</sup> larynx,<sup>107</sup> oral cavity and oropharynx,<sup>108</sup> kidney<sup>109</sup> and bladder,110,111 after adjustment for smoking. A significant positive relationship was reported between drinking mate and urinary level of 1-OHPG in southern Brazil, suggesting that this factor is an important source of PAH exposure in this area.<sup>80</sup> Laboratory measurement of PAHs in commercial yerba mate leaves and in both cold and hot water infusions of these leaves showed very high levels of benzo-a-pyrene and other carcinogenic PAHs, and suggested that drinking a gourd of mate in the traditional manner can expose an individual to as much benzo-a-pyrene as smoking a typical pack of 20 cigarettes.<sup>112</sup> So the PAH content of food and the method of food processing and preparation are major contributors to human exposure to PAHs from dietary sources.

#### 2. Tobacco smoking and opium consumption

Tobacco smoking has been identified as a major source of exposure to PAHs.<sup>113–115</sup> A significant positive relationship has been reported between urinary level of 1-OHPG and smoking.<sup>39,80,116</sup> Van Rooij et al. reported that active smoking and PAH containing food products account for 99% of urinary excretion of PAH biomarkers in individuals who are not occupationally exposed to PAHs.<sup>117</sup> In a study from the US, individuals with recent smoking had 1-OHPG concentrations 10-fold that of non-smokers.<sup>102</sup> Second hand smoke exposure was also correlated with high 1-OHPG concentration.<sup>102</sup> Lee et al. reported significantly higher urinary 1-OHPG levels in South Korean children whose parents smoke at home.<sup>118</sup> The results of studies conducted in the 1970s in northeastern Iran showed high levels of opium consumption in this region and indicated that PAHs are a major active component responsible for its mutagenicity.<sup>98,119</sup> In a recent study, smoking,

nass consumption and opium consumption were all found to be significant sources of PAH exposure in Golestan Province, Iran,<sup>84</sup> and high exposure to PAH may be one explanation for the reported association between opium or tobacco use and higher risk of ESCC in northeastern Iran.<sup>15</sup> The proportion of the excess risk of ESCC due to PAH exposure from smoking and opium consumption has not yet been clearly identified. This is an interesting issue for future studies in this area and other similar populations such as those in central China.

#### 3. Occupational exposure

Occupation is also an important source of exposure to PAHs. The level of exposure to PAHs varies between different occupations.<sup>120</sup> High levels of PAH exposure have been reported in foundry workers,<sup>37</sup> chimney sweeps,<sup>61</sup> blast furnace and cokeoven workers,<sup>39</sup> vendors of broiled food,<sup>121</sup> and steel plant and waste incineration workers.<sup>39,122,123</sup> Coke-oven exposure is a major predictor of urinary 1-OHP levels.115 The results of a study from the UK showed that coke-oven workers had the highest levels of exposure to benzo[a]pyrene.<sup>124</sup> The highest levels of urinary 1-OHPG were reported in timber impregnators using creosote and workers using coal tar. Both dermal and inhalation exposure were suggested as possible explanations for high PAH exposure in the above mentioned occupations.<sup>124</sup> In a study from the US, the highest levels of exposure to PAHs were detected in coke-oven workers, asphalt workers and diesel-exposed workers.<sup>45</sup> Job is an important indicator of exposure to PAH, and should be considered in future related studies.

#### 4. Indoor air pollution

Combustion of solid fuels for cooking and heating is the major source of indoor air levels of PAHs.<sup>125</sup> China is the largest producer and consumer of coal throughout the world. A substantial portion of domestic energy needs in China are supplied by coal, a known source of PAH.<sup>125</sup> Thus, domestic coal, especially "smoky coal", combustion is known as a main source of high PAH exposure in China.<sup>126</sup> Mumford et al. showed very high PAH levels in indoor air samples from Chinese homes, especially ones which used smoky coal in unvented stoves for heating and cooking.<sup>127</sup> Similar findings were reported in other studies from China.<sup>125</sup> High PAH concentrations were also reported in soot deposits from coal-burning stoves in Chinese homes.<sup>128</sup> Gevao et al. also reported high levels of indoor PAHs in indoor air and dust in Kuwait.<sup>129</sup>

Housewives and children usually spend a lot of time at home. This may lead to high levels of exposure to PAHs from indoor PAH sources in these groups. Therefore, housewives, children and all individuals spending a lot of time in homes with significant indoor air pollution may be considered as high risk groups in studies related to the effects of PAH exposure on human health.

## 5. Environmental air pollution

There are many sources of PAHs in environmental air. Unknown sources of environmental PAHs make it difficult to assess the exact level of PAH exposure from the environment. Various methods may be used to estimate the concentration of PAHs in the environment. For example, the level of PAHs in the environment is reflected by their presence in some uncooked foods or fruits.<sup>25</sup> PAH levels in olive fruits have been considered one measure of air pollution.<sup>91</sup>

It has been suggested that constituents of car exhaust, industrial emissions and smoke are the main sources of outdoor air PAHs. In a study from the US, outdoor concentrations of PAHs were higher in highly urbanized areas (containing many cars and industries) than in the Great Lakes region.93 The results of another study from an area near a large city bus terminal showed higher air levels of PAHs on weekdays and during morning rush hours, probably related to road traffic.<sup>130</sup> They also reported significantly higher PAH levels in areas closer to the bus terminal.<sup>130</sup> Marr et al. reported high levels of PAHs along Mexico City's roadways, suggesting vehicle traffic as a source of human exposure to PAHs.<sup>131</sup> Significantly higher 1-OHPG concentrations have also been reported in individuals who recently traveled in automobiles for more than 3 hours.<sup>102</sup> The results of a study from Shenzhen, China, showed significantly higher levels of PAHs in areas with many roads than in forested land.<sup>132</sup> Individuals living in the vicinity of an aluminum plant in Canada were exposed to high levels of PAHs.<sup>133</sup> Sediments from certain commercial and residential ponds have also been suggested as potential sources of exposure to PAHs.<sup>134</sup>

Identifying the nature and characteristics of environmental sources of exposure to PAHs as well as the levels of PAH exposure from these sources are interesting issues for future studies. These investigations are especially important in high risk areas of PAH-related cancers such as northeastern Iran.

# Effects of genetic changes on an individual's response to PAH exposure

Genetic factors may affect an individual's response to PAH exposure.<sup>86</sup> Usually, p450 enzymes (CYP1A1, CYP1A2, CYP 1B1) metabolize PAHs (such as benzo-a-pyrene) to biologically active intermediates (such as benzo-a-pyrene diol expoxide) and detoxification enzymes, including glutathione S-transferases (GSTM1, GSTP1, GSTT1), UDP-glucuronosyltransferases, and sulphotransferases, conjugate these PAH metabolites and de-

toxify them.26 But genetic polymorphisms may affect the activity of these enzymes and make individuals more or less susceptible to the carcinogenic effects the same PAH exposure.<sup>86</sup> Roth et al. showed an association between the inactive polymorphism of the GSTM1 gene and the presence of esophageal squamous dysplasia, the precursor of ESCC, in a PAH-exposed population in China.<sup>135</sup> Also, a significantly higher expression of the aryl hydrocarbon receptor (AhR) gene, which codes for the cell membrane receptor for PAHs, was reported in subjects from the same population who had a positive family history of upper gastrointestinal cancers, suggesting that these individuals might be more susceptible to PAH-related carcinogenesis.<sup>136</sup> And Abnet et al. reported a strong association between CYP1B1 and urinary 1-OHPG concentrations in a Brazilian population with a high risk of ESCC.137 So, in addition to sources of exposure, genetic susceptibility to PAH-related carcinogenesis may also influence the relationship between PAH exposure and ESCC pathogenesis.

## Identifying the main scirces of PAH exposure

PAH exposure has now been documented to be an important risk factor for developing ESCC. Thus, reducing the levels of exposure to PAHs may be an effective strategy for controlling ESCC, especially in high risk areas. Appropriate PAH-reduction interventions may differ, depending on the main sources of PAH exposure in each area. For example, Li et al. showed that replacement of open pit stoves by closed stoves equipped with a chimney significantly reduced PAH exposure in Peru.<sup>138</sup> Identifying the main sources of exposure to PAHs will result in maximum effectiveness of such PAH-reduction interventions. The major sources of PAH exposure have been identified in some populations with high ESCC rates (Table 3). But the situations in other high risk populations such as the northeast of Iran remain unclear. Previous studies conducted in the 1970s in northeastern Iran showed low levels of PAHs in food samples,<sup>139,140</sup> and the investigations by the joint Iran-IARC study group on major dietary items in northern Iran showed no unusual levels of carcinogens such as PAHs in food samples.141 These studies suggested that food is not the main source of PAH exposure in this area. On the other hand, Hakami et al. reported that food may be an important source of exposure to PAHs in Golestan Province.99 As reported by Kamangar et al. smoking and opium consumption are probably not the main sources of exposure to PAHs in this area.<sup>84</sup> Further evaluations, including environmental studies, are needed to identify the major sources of PAH exposure in this region as well as other high risk populations.

## Conclusion

An association has been found between PAH exposure and development of ESCC in many populations. Identifying the main sources of exposure to PAHs is the first and most important step for conducting appropriate PAH-reduction interventions to control ESCC, especially in high risk areas. Coal smoke and drinking mate have been suggested as important modifiable sources of PAH exposure in China and Brazil, respectively. But the primary source of exposure to PAHs in other high risk areas for ESCC, such as northeastern Iran, has not yet been identified. Thus, environmental studies to determining important sources of PAH exposure should be considered as a high priority in future research projects in these areas.

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### References

- Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Globocan 2008, cancer incidence and mortality worldwide: Iarc cancerbase no. 10. Lyon: International Agency for Research on Cancer; 2010 [03/02/2011]; Available from: URL: http://globocan.iarc.fr.
- Das A. Tumors of the esophagus. In: Feldman M, Friedman L, Brandt LJ, eds. Sleisenger and fordtran's gastrointestinal and liver disease: Pathophysiology, diagnosis, management. 9th ed. Philadelphia: Saunders Elsevier; 2010. 745 – 770.
- Mahboubi E, Kmet J, Cook PJ, Day NE, Ghadirian P, Salmasizadeh S. Oesophageal cancer studies in the caspian littoral of iran: The caspian cancer registry. *Br J Cancer*. 1973; 28: 197 – 214.
- Semnani S, Sadjadi A, Fahimi S, Nouraie M, Naeimi M, Kabir J, et al. Declining incidence of esophageal cancer in the turkmen plain, eastern part of the caspian littoral of iran: A retrospective cancer surveillance. *Cancer Detect Prev.* 2006; **30:** 14 – 19.
- Roshandel G, Sadjadi A, Aarabi M, Keshtkar A, Sedaghat S, Nouraie S, et al. Cancer incidence in golestan province: Report of an ongoing population-based cancer registry in iran between 2004 and 2008. *Arch Iran Med.* 2012; 15: 196.
- Islami F, Kamangar F, Aghcheli K, Fahimi S, Semnani S, Taghavi N, et al. Epidemiologic features of upper gastrointestinal tract cancers in northeastern iran. *Br J Cancer*. 2004; **90:** 1402 – 1406.
- Brown LM, Devesa SS, Chow W-H. Incidence of adenocarcinoma of the esophagus among white americans by sex, stage, and age. *J Natl Cancer Inst.* 2008; **100:** 1184 – 1187.
- Cook MB, Chow WH, Devesa SS. Oesophageal cancer incidence in the united states by race, sex, and histologic type, 1977-2005. Br J Cancer. 2009; 101: 855 – 859.
- 9. Lepage C, Rachet B, Jooste V, Faivre J, Coleman MP. Continuing rapid increase in esophageal adenocarcinoma in england and wales. *Am J Gastroenterol.* 2008; **103**: 2694 2699.
- Islami F, Kamangar F, Nasrollahzadeh D, Moller H, Boffeta P, Malekzadeh R. Oesophageal cancer in golestan province, a highincidence area in northern iran - a review. *Eur J Cancer*. 2009; 45: 3156-3165.
- Dawsey SM, Lewin KJ, Wang GQ, Liu FS, Nieberg RK, Yu Y, et al. Squamous esophageal histology and subsequent risk of squamous cell carcinoma of the esophagus. A prospective follow-up study from linxian, china. Cancer. 1994; 74: 1686 – 1692.
- Wang GQ, Abnet CC, Shen Q, Lewin KJ, Sun XD, Roth MJ, et al. Histological precursors of oesophageal squamous cell carcinoma: Results from a 13 year prospective follow up study in a high risk population. *Gut.* 2005; 54: 187–192.
- Kamangar F, Chow W-H, C. Abnet CM, Dawsey S. Environmental causes of esophageal cancer. *Gastroenterol Clin North Am.* 2009; 38: 27 – 57.
- Freedman ND, Abnet CC, Leitzmann MF, Mouw T, Subar AF, Hollenbeck AR, et al. A prospective study of tobacco, alcohol, and the risk of esophageal and gastric cancer subtypes. *Am J Epidemiol*. 2007; 165: 1424.
- Nasrollahzadeh D, Kamangar F, Aghcheli K, Sotoudeh M, Islami F, Abnet CC, et al. Opium, tobacco, and alcohol use in relation to oesophageal squamous cell carcinoma in a high-risk area of iran. *Br J Cancer*. 2008; **98:** 1857 – 1863.
- Islami F, Pourshams A, Nasrollahzadeh D, Kamangar F, Fahimi S, Shakeri R, et al. Tea drinking habits and oesophageal cancer in a high risk area in northern iran: Population based case-control study. *Br Med J*. 2009; **338**: b929.
- Castellsague X, Munoz N, De Stefani E, Victora CG, Castelletto R, Rolon PA. Influence of mate drinking, hot beverages and diet on esophageal cancer risk in south america. *Int J Cancer*. 2000; 88: 658 – 664.
- Freedman ND, Park Y, Subar AF, Hollenbeck AR, Leitzmann MF, Schatzkin A, et al. Fruit and vegetable intake and esophageal cancer in a large prospective cohort study. *Int J Cancer*. 2007; **121**: 2753 – 2760.

- Mark SD, Qiao YL, Dawsey SM, Wu YP, Katki H, Gunter EW, et al. Prospective study of serum selenium levels and incident esophageal and gastric cancers. *J Natl Cancer Inst.* 2000; 92: 1753 – 1763.
- Taylor PR, Qiao YL, Abnet CC, Dawsey SM, Yang CS, Gunter EW, et al. Prospective study of serum vitamin e levels and esophageal and gastric cancers. *J Natl Cancer Inst.* 2003; 95: 1414 – 1416.
- Wei WQ, Abnet CC, Qiao YL, Dawsey SM, Dong ZW, Sun XD, et al. Prospective study of serum selenium concentrations and esophageal and gastric cardia cancer, heart disease, stroke, and total death. *Am J Clin Nutr.* 2004; **79:** 80 – 85.
- Abnet CC, Kamangar F, Islami F, Nasrollahzadeh D, Brennan P, Aghcheli K, et al. Tooth loss and lack of regular oral hygiene are associated with higher risk of esophageal squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev.* 2008; **17:** 3062 – 3068.
- Islami F, Kamangar F, Nasrollahzadeh D, Aghcheli K, Sotoudeh M, Abedi-Ardekani B, et al. Socio-economic status and oesophageal cancer: Results from a population-based case-control study in a high-risk area. Int J Epidemiol. 2009; 38: 978 – 988.
- Engel LS, Chow WH, Vaughan TL, Gammon MD, Risch HA, Stanford JL, et al. Population attributable risks of esophageal and gastric cancers. *J Natl Cancer Inst.* 2003; 95: 1404 – 1413.
- Phillips DH. Polycyclic aromatic hydrocarbons in the diet. *Mutat Res.* 1999; 443: 139 – 147.
- Straif K, Baan R, Grosse Y, Secretan B, El Ghissassi F, Cogliano V. Carcinogenicity of polycyclic aromatic hydrocarbons. *Lancet Oncol.* 2005; 6: 931.
- Harvey RG. Polycyclic aromatic hydrocarbons: Chemistry and carcinogenicity. Cambridge: Cambridge University Press; 1991.
- Rappaport SM, Waidyanatha S, Serdar B. Naphthalene and its biomarkers as measures of occupational exposure to polycyclic aromatic hydrocarbons. *J Environ Monit*. 2004; 6: 413 – 416.
- Simko P. Determination of polycyclic aromatic hydrocarbons in smoked meat products and smoke flavouring food additives. J Chromatogr B Analyt Technol Biomed Life Sci. 2002; 770: 3 – 18.
- Phillips DH. Fifty years of benzo (a) pyrene. *Nature*. 1983; 303: 468 472.
- Rogan EG, Devanesan PD, Ramakrishna NVS, Higginbotham S, Padmavathi NS, Chapman K, et al. Identification and quantitation of benzo [a] pyrene-DNA adducts formed in mouse skin. *Chem Res Toxi*col. 1993; 6: 356 – 363.
- 32. Jongeneelen FJ, Bos RP, Anzion RB, Theuws JL, Henderson PT. Biological monitoring of polycyclic aromatic hydrocarbons. Metabolites in urine. *Scand J Work Environ Health.* 1986; **12:** 137 – 143.
- Becher G, Bjorseth A. Determination of exposure to polycyclic aromatic hydrocarbons by analysis of human urine. *Cancer Lett.* 1983; 17: 301-311.
- Clonfero E, Jongeneelen F, Zordan M, Levis AG. Biological monitoring of human exposure to coal tar. Urinary mutagenicity assays and analytical determination of polycyclic aromatic hydrocarbon metabolites in urine. *IARC Sci Publ.* 1990; 215 – 222.
- Zhao ZH, Quan WY, Tian DH. Urinary 1-hydroxypyrene as an indicator of human exposure to ambient polycyclic aromatic hydrocarbons in a coal-burning environment. *Sci Total Environ*. 1990; 92: 145 – 154.
- Castano-Vinyals G, D'errico A, Malats N, Kogevinas M. Biomarkers of exposure to polycyclic aromatic hydrocarbons from environmental air pollution. *Occup Environ Med.* 2004; 61: e12.
- Omland O, Sherson D, Hansen AM, Sigsgaard T, Autrup H, Overgaard E. Exposure of iron foundry workers to polycyclic aromatic hydrocarbons: Benzo(a)pyrene-albumin adducts and 1-hydroxypyrene as biomarkers for exposure. *Occup Environ Med.* 1994; 51: 513-518.
- Zhao ZH, Quan WY, Tian DH. Experiments on the effects of several factors on the 1-hydroxypyrene level in human urine as an indicator of exposure to polycyclic aromatic hydrocarbons. *Sci Total Environ*. 1992; **113**: 197 – 207.
- Kang D, Rothman N, Cho SH, Lim HS, Kwon HJ, Kim SM, et al. Association of exposure to polycyclic aromatic hydrocarbons (estimated from job category) with concentration of 1-hydroxypyrene glucuronide in urine from workers at a steel plant. *Occup Environ Med.* 1995; 52: 593 – 599.
- Kang DH, Rothman N, Poirier MC, Greenberg A, Hsu CH, Schwartz BS, et al. Interindividual differences in the concentration of 1-hydroxypyrene-glucuronide in urine and polycyclic aromatic hydrocarbon-DNA adducts in peripheral white blood cells after charbroiled beef consumption. *Carcinogenesis.* 1995; 16: 1079 – 1085.
- 41. Buratti M, Campo L, Fustinoni S, Cirla PE, Martinotti I, Cavallo D, et al. Urinary hydroxylated metabolites of polycyclic aromatic hydro-

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carbons as biomarkers of exposure in asphalt workers. *Biomarkers*. 2007; **12**: 221-239.

- 42. Chetiyanukornkul T, Toriba A, Kameda T, Tang N, Hayakawa K. Simultaneous determination of urinary hydroxylated metabolites of naphthalene, fluorene, phenanthrene, fluoranthene and pyrene as multiple biomarkers of exposure to polycyclic aromatic hydrocarbons. *Anal Bioanal Chem.* 2006; **386:** 712 – 718.
- Rossbach B, Preuss R, Letzel S, Drexler H, Angerer J. Biological monitoring of occupational exposure to polycyclic aromatic hydrocarbons (pah) by determination of monohydroxylated metabolites of phenanthrene and pyrene in urine. *Int Arch Occup Environ Health*. 2007; 81: 221 – 229.
- Seidel A, Spickenheuer A, Straif K, Rihs HP, Marczynski B, Scherenberg M, et al. New biomarkers of occupational exposure to polycyclic aromatic hydrocarbons. *J Toxicol Environ Health A*. 2008; **71:** 734 745.
- Sobus JR, Waidyanatha S, Mcclean MD, Herrick RF, Smith TJ, Garshick E, et al. Urinary naphthalene and phenanthrene as biomarkers of occupational exposure to polycyclic aromatic hydrocarbons. *Occup Environ Med.* 2009; 66: 99 – 104.
- Pesch B, Kappler M, Straif K, Marczynski B, Preuss R, Rossbach B, et al. Dose-response modeling of occupational exposure to polycyclic aromatic hydrocarbons with biomarkers of exposure and effect. *Cancer Epidemiol Biomarkers Prev.* 2007; 16: 1863 – 1873.
- Godschalk RW, Van Schooten FJ, Bartsch H. A critical evaluation of DNA adducts as biological markers for human exposure to polycyclic aromatic compounds. *J Biochem Mol Biol.* 2003; 36: 1–11.
- Rothman N, Poirier M, Baser M, Hansen J, Gentile C, Bowman ED, et al. Formation of polycyclic aromatic hydrocarbon-DNA adducts in peripheral white blood cells during consumption of charcoal-broiled beef. *Carcinogenesis*. 1990; **11**: 1241 – 1243.
- Perera FP, Hemminki K, Young TL, Brenner D, Kelly G, Santella RM. Detection of polycyclic aromatic hydrocarbon-DNA adducts in white blood cells of foundry workers. *Cancer Res.* 1988; 48: 2288 – 2291.
- Kriek E, Rojas M, Alexandrov K, Bartsch H. Polycyclic aromatic hydrocarbon-DNA adducts in humans: Relevance as biomarkers for exposure and cancer risk. *Mutat Res.* 1998; 400: 215 – 231.
- Rothman N, Poirier MC, Haas RA, Correa-Villasenor A, Ford P, Hansen JA, et al. Association of pah-DNA adducts in peripheral white blood cells with dietary exposure to polyaromatic hydrocarbons. *Environ Health Perspect.* 1993; 99: 265 – 267.
- Taioli E, Sram RJ, Binkova B, Kalina I, Popov TA, Garte S, et al. Biomarkers of exposure to carcinogenic pahs and their relationship with environmental factors. *Mutat Res*. 2007; 620: 16–21.
- Gyorffy E, Anna L, Gyori Z, Segesdi J, Minarovits J, Soltesz I, et al. DNA adducts in tumour, normal peripheral lung and bronchus, and peripheral blood lymphocytes from smoking and non-smoking lung cancer patients: Correlations between tissues and detection by 32ppostlabelling and immunoassay. *Carcinogenesis*. 2004; 25: 1201 – 1209.
- Van Gijssel HE, Schild LJ, Watt DL, Roth MJ, Wang GQ, Dawsey SM, et al. Polycyclic a omatic hydrocarbon-DNA adducts determined by semiquantitative immunohistochemistry in human esophageal biopsies taken in 1985. *Mutat Res.* 2004; 547: 55 – 62.
- Toriba A, Kuramae Y, Chetiyanukornkul T, Kizu R, Makino T, Nakazawa H, et al. Quantification of polycyclic aromatic hydrocarbons (pahs) in human hair by hplc with fluorescence detection: A biological monitoring method to evaluate the exposure to pahs. *Biomed Chromatogr.* 2003; 17: 126 – 132.
- Beskid O, Binkova B, Dusek Z, Rossner P, Solansky I, Kalina I, et al. Chromosomal aberrations by fluorescence in situ hybridization (fish)biomarker of exposure to carcinogenic pahs. *Mutat Res.* 2007; 620: 62 – 70.
- Abedi-Ardekani B, Kamangar F, Hewitt SM, Hainaut P, Sotoudeh M, Abnet CC, et al. Polycyclic aromatic hydrocarbon exposure in oesophageal tissue and risk of oesophageal squamous cell carcinoma in north-eastern iran. Gut. 2010; 59: 1178 – 1183.
- Ponder B. Polycyclic aromatic hydrocarbon exposure in oesophageal tissue and risk of oesophageal squamous cell carcinoma in north-eastern iran. *Arch Iran Med.* 2010; 13: 457 – 458.
- Iarc. Iarc monographs on the evaluation of carcinogenic risks to humans, no 92: Some non-heterocyclic polycyclic aromatic hydrocarbons and some related exposures. Lyon: International Agency for Research on Cancer; 2010.
- Pott P. Chirurgical observations. *Natl Cancer Inst Monogr.* 1963; 10: 7.

- 61. Gustavsson P, Gustavsson A, Hogstedt C. Excess of cancer in swedish chimney sweeps. *Br J Ind Med.* 1988; **45:** 777 781.
- Armstrong BG, Gibbs G. Exposure-response relationship between lung cancer and polycyclic aromatic hydrocarbons (pahs). *Occup Environ Med.* 2009; 66: 740 – 746.
- Petralia SA, Vena JE, Freudenheim JL, Dosemeci M, Michalek A, Goldberg MS, et al. Risk of premenopausal breast cancer in association with occupational exposure to polycyclic aromatic hydrocarbons and benzene. *Scand J Work Environ Health*. 1999; 25: 215 – 221.
- Carta P, Aru G, Cadeddu C, Gigli G, Papi G, Carta F, et al. Mortality for pancreatic cancer among aluminium smelter workers in sardinia, italy. *G Ital Med Lav Ergon*. 2004; 26: 83 – 89.
- Nadon L, Siemiatycki J, Dewar R, Krewski D, Gerin M. Cancer risk due to occupational exposure to polycyclic aromatic hydrocarbons. *Am J Ind Med.* 1995; 28: 303 – 324.
- Allen-Hoffmann BL, Rheinwald JG. Polycyclic aromatic hydrocarbon mutagenesis of human epidermal keratinocytes in culture. *Proc Natl Acad Sci USA*. 1984; 81: 7802 – 7806.
- Benbrahim-Tallaa L, Baan RA, Grosse Y, Lauby-Secretan B, El Ghissassi F, Bouvard V, et al. Carcinogenicity of diesel-engine and gasoline-engine exhausts and some nitroarenes. *Lancet Oncol.* 2012; DOI. 10.1016/S1470-2045(12)70280-2.
- Pfeifer GP, Denissenko MF, Olivier M, Tretyakova N, Hecht SS, Hainaut P. Tobacco smoke carcinogens, DNA damage and p 53 mutations in smoking-associated cancers. *Oncogene*, 2002; 21: 7435 – 7451.
- Puisieux A, Lim S, Groopman J, Ozturk M. Selective targeting of p53 gene mutational hotspots in human cancers by etiologically defined carcinogens. *Cancer Res.* 1991; 51: 6185 – 6189.
- Gao M, Li Y, Sun Y, Long J, Kong Y, Yang S, et al. A common carcinogen benzo [a] pyrene causes p53 overexpression in mouse cervix via DNA damage. *Mutat Res.* 2011; 725: 69 – 75.
- Denissenko MF, Pao A, Tang M-S, Pfeifer GP. Preferential formation of benzo[a]pyrene adducts at lung cancer mutational hotspots in p53. *Science*. 1996; 274: 430 – 432.
- 72. Shen YM, Troxel AB, Vedantam S, Trevor M, Field J. Comparison of p53 mutations induced by pah o-quinones with those caused by anti-benzo [a] pyrene diol epoxide in vitro: Role of reactive oxygen and biological selection. *Chem Res Toxicol*. 2006; **19**: 1441 1450.
- Yu D, Jesse A, Trevor M, Field J. Reactive oxygen species generated by pah o-quinones cause change-in-function mutations in p53. *Chem Res Toxicol.* 2002; 15: 832 – 842.
- Brennan JA, Boyle JO, Koch WM, Goodman SN, Hruban RH, Eby YJ, et al. N Engl J Med. 1995; 332: 712 – 717.
- Abedi-Ardekani B, Kamangar F, Sotoudeh M, Villar S, Islami F, Aghcheli K, et al. Extremely high tp53 mutation load in esophageal squamous cell carcinoma in golestan province, iran. *PLoS One*. 2011; 6: e29488.
- 76. Cairns J, editor. Cancer and society. San Francisco: Freeman; 1978.
- Culp SJ, Gaylor DW, Sheldon WG, Goldstein LS, Beland FA. A comparison of the tumors induced by coal tar and benzo[a]pyrene in a 2-year bioassay. *Carcinogenesis*. 1998; 19: 117 – 124.
- Vieth M, Stolte M. [do polycyclic aromatic hydrocarbons (pah) cause squamous cell carcinoma of the esophagus?]. *Z Gastroenterol.* 1999; 37: 579 – 580.
- Gustavsson P, Jakobsson R, Johansson H, Lewin F, Norell S, Rutkvist LE. Occupational exposures and squamous cell carcinoma of the oral cavity, pharynx, larynx, and oesophagus: A case-control study in sweden. *Occup Environ Med.* 1998; 55: 393 – 400.
- Fagundes RB, Abnet CC, Strickland PT, Kamangar F, Roth MJ, Taylor PR, et al. Higher urine 1-hydroxy pyrene glucuronide (1-ohpg) is associated with tobacco smoke exposure and drinking mate in healthy subjects from rio grande do sul, brazil. *BMC Cancer*. 2006; 6: 139.
- Ward MH, Sinha R, Heineman EF, Rothman N, Markin R, Weisenburger DD, et al. Risk of adenocarcinoma of the stomach and esophagus with meat cooking method and doneness preference. *Int J Cancer*. 1997; **71**: 14 19.
- Roth MJ, Strickland KL, Wang GQ, Rothman N, Greenberg A, Dawsey SM. High levels of carcinogenic polycyclic aromatic hydrocarbons present within food from linxian, china may contribute to that region's high incidence of oesophageal cancer. *Eur J Cancer*. 1998; 34: 757 – 758.
- Roth M, Qiao YL, Rothman N, Tangrea J, Dawsey S, Wang GQ, et al. High urine 1-hydroxypyrene glucuronide concentrations in linxian, china, an area of high risk for squamous oesophageal cancer. *Biomark*ers. 2001; 6: 381 – 386.
- 84. Kamangar F, Strickland PT, Pourshams A, Malekzadeh R, Boffetta P,

Roth MJ, et al. High exposure to polycyclic aromatic hydrocarbons may contribute to high risk of esophageal cancer in northeastern iran. *Anticancer Res.* 2005; **25:** 425 – 428.

- Marjani HA, Biramijamal F, Rakhshani N, Hossein-Nezhad A, Malekzadeh R. Investigation of nqo1 genetic polymorphism, nqo1 gene expression and pah-DNA adducts in escc. A case-control study from iran. *Genet Mol Res.* 2010; **9**: 239 – 249.
- Garte S, Taioli E, Raimondi S, Paracchini V, Binkova B, Sram RJ, et al. Effects of metabolic genotypes on intermediary biomarkers in subjects exposed to pahs: Results from the expah study. *Mutat Res.* 2007; 620: 7 – 15.
- Santodonato J, Howard P, Basu D. Health and ecological assessment of polynuclear aromatic hydrocarbons. *J Environ Pathol Toxicol*. 1981; 5: 1.
- Larsson BK. Polycyclic aromatic hydrocarbons in smoked fish. Z Lebensm Unters Forsch. 1982; 174: 101.
- Dennis MJ, Massey RC, Mcweeny DJ, Knowles ME, Watson D. Analysis of polycyclic aromatic hydrocarbons in uk total diets. *Food Chem Toxicol.* 1983; 21: 569 – 574.
- Kobayashi R, Okamoto RA, Maddalena RL, Kado NY. Polycyclic aromatic hydrocarbons in edible grain: A pilot study of agricultural crops as a human exposure pathway for environmental contaminants using wheat as a model crop. *Environ Res.* 2008; **107**: 145 – 151.
- Ignesti G, Lodovici M, Dolara P, Lucia P, Grechi D. Polycyclic aromatic hydrocarbons in olive fruits as a measure of air pollution in the valley of florence (italy). *Bull Environ Contam Toxicol*. 1992; 48: 809 814.
- Wild SR, Berrow ML, Mcgrath SP, Jones KC. Polynuclear aromatic hydrocarbons in crops from long-term field experiments amended with sewage sludge. *Environ Pollut*. 1992; **76**: 25 – 32.
- Menzie CA, Potocki BB, Santodonato J. Exposure to carcinogenic pahs in the environment. *Environ Sci Technol*. 1992; 26: 1278 – 1284.
- Vyskocil A, Fiala Z, Fialova D, Krajak V, Viau C. Environmental exposure to polycyclic aromatic hydrocarbons in czech republic. *Hum Exp Toxicol.* 1997; 16: 589 595.
- De Vos RH, Van Dokkum W, Schouten A, De Jong-Berkhout P. Polycyclic aromatic hydrocarbons in dutch total diet samples (1984-1986). *Food Chem Toxicol.* 1990; 28: 263 – 268.
- Lodovici M, Dolara P, Casalini C, Ciappellano S, Testolin G. Polycyclic aromatic hydrocarbon contamination in the italian diet. *Food Addit Contam.* 1995; 12: 703 – 713.
- Falco G, Domingo JL, Llobet JM, Teixido A, Casas C, Muller L. Polycyclic aromatic hydrocarbons in foods: Human exposure through the diet in catalonia, spain. *J Food Prot.* 2003; 66: 2325 – 2331.
- Mahboubi EO, Aramesh B. Epidemiology of esophageal cancer in iran, with special reference to nutritional and cultural aspects. *Prev Med.* 1980; 9: 613 – 621.
- Hakami R, Mohtadinia J, Etemadi A, Kamangar F, Nemati M, Pourshams A, et al. Dietary intake of benzo(a)pyrene and risk of esophageal cancer in north of iran. *Nutr Cancer*. 2008; 60: 216 221.
- Dennis MJ, Massey RC, Cripps G, Venn I, Howarth N, Lee G. Factors affecting the polycyclic aromatic hydrocarbon content of cereals, fats and other food products. *Food Addit Contam.* 1991; 8: 517–530.
- Gomaa EA, Gray JI, Rabie S, Lopez-Bote C, Booren AM. Polycyclic aromatic hydrocarbors in smoked food products and commercial liquid smoke flavourings. *Food Addit Contam.* 1993; 10: 503 – 521.
- 102. Gunier RB, Reynolds P, Hurley SE, Yerabati S, Hertz A, Strickland P, et al. Estimating exposure to polycyclic aromatic hydrocarbons: A comparison of survey, biological monitoring, and geographic information system-based methods. *Cancer Epidemiol Biomarkers Prev.* 2006; 15: 1376 1381.
- Alomirah H, Al-Zenki S, Al-Hooti S, Zaghloul S, Sawaya W, Ahmed N, et al. Concentrations and dietary exposure to polycyclic aromatic hydrocarbons (pahs) from grilled and smoked foods. *Food Control*. 2011; 22: 2028 – 2035.
- Victora CG, Munoz N, Horta BL, Ramos EO. Patterns of mate drinking in a brazilian city. *Cancer Res.* 1990; 50: 7112 – 7115.
- Sewram V, De Stefani E, Brennan P, Boffetta P. Mate consumption and the risk of squamous cell esophageal cancer in uruguay. *Cancer Epidemiol Biomarkers Prev.* 2003; **12:** 508 – 513.
- De Stefani E, Fierro L, Correa P, Fontham E, Ronco A, Larrinaga M, et al. Mate drinking and risk of lung cancer in males: A case-control study from uruguay. *Cancer Epidemiol Biomarkers Prev.* 1996; 5: 515 – 519.
- Pintos J, Franco EL, Oliveira BV, Kowalski LP, Curado MP, Dewar R. Mate, coffee, and tea consumption and risk of cancers of the upper

aerodigestive tract in southern brazil. *Epidemiology*. 1994; **5:** 583 – 590.

- Goldenberg D, Golz A, Joachims HZ. The beverage mate: A risk factor for cancer of the head and neck. *Head Neck*. 2003; 25: 595 – 601.
- De Stefani E, Fierro L, Mendilaharsu M, Ronco A, Larrinaga MT, Balbi JC, et al. Meat intake, 'mate' drinking and renal cell cancer in uruguay: A case-control study. *Br J Cancer*. 1998; **78**: 1239 – 1243.
- De Stefani E, Boffetta P, Deneo-Pellegrini H, Correa P, Ronco AL, Brennan P, et al. Non-alcoholic beverages and risk of bladder cancer in uruguay. *BMC Cancer*. 2007; 7: 57.
- Bates MN, Hopenhayn C, Rey OA, Moore LE. Bladder cancer and mate consumption in argentina: A case-control study. *Cancer Lett.* 2007; 246: 268 – 273.
- Kamangar F, Schantz MM, Abnet CC, Fagundes RB, Dawsey SM. High levels of carcinogenic polycyclic aromatic hydrocarbons in mate drinks. *Cancer Epidemiol Biomarkers Prev.* 2008; 17: 1262 – 1268.
- Lioy PJ, Greenberg A. Factors associated with human exposures to polycyclic aromatic hydrocarbons. *Toxicol Ind Health*. 1990; 6: 209 - 223.
- Pufulete M, Battershill J, Boobis A, Fielder R. Approaches to carcinogenic risk assessment for polycyclic aromatic hydrocarbons: A uk perspective. *Regul Toxicol Pharmacol.* 2004; 40: 54 – 66.
- 115. Qiu L, Leng S, Wang Z, Dai Y, Zheng Y, Wang Z. Path analysis of biomarkers of exposure and early biological effects among coke-oven workers exposed to polycyclic aromatic hydrocarbons. *Cancer Epidemiol Biomarkers Prev.* 2007; 16: 1193–1199.
- 116. Vineis P, Talaska G, Malaveille C, Bartsch H, Martone T, Sithisarankul P, et al. DNA adducts in urothelial cells: Relationship with biomarkers of exposure to arylamines and polycyclic aromatic hydrocarbons from tobacco smoke. *Int J Cancer.* 1996; 65: 314 316.
- 117. Van Rooij JG, Veeger MM, Bodelier-Bade MM, Scheepers PT, Jongeneelen FJ. Smoking and dietary intake of polycyclic aromatic hydrocarbons as sources of interindividual variability in the baseline excretion of 1-hydroxypyrene in urine. *Int Arch Occup Environ Health*. 1994; 66: 55 – 65.
- Lee KH, Vermeulen R, Lenters V, Cho SH, Strickland PT, Kang D. Determinants of urinary 1-hydroxypyrene glucuronide in south korean children. *Int Arch Occup Environ Health.* 2009; 82: 961–968.
- Dowlatshahi K, Miller RJ. Role of opium in esophageal cancer: A hypothesis. *Cancer Res.* 1985; 45: 1906 1907.
- Taioli E, Sram RJ, Garte S, Kalina I, Popov TA, Farmer PB. Effects of polycyclic aromatic hydrocarbons (pahs) in environmental pollution on exogenous and oxidative DNA damage (expah project): Description of the population under study. *Mutat Res.* 2007; 620: 1 – 6.
- Kuo CY, Chang SH, Chien YC, Chiang FY, Wei YC. Exposure to carcinogenic pahs for the vendors of broiled food. *J Expo Sci Environ Epidemiol.* 2006; 16: 410 – 416.
- 122. Lee J, Kang D, Lee KH, Ichiba M, Zhang J, Tomokuni K, et al. Influence of gstm1 genotype on association between aromatic DNA adducts and urinary pah metabolites in incineration workers. *Mutat Res.* 2002; **514**: 213 221.
- 123. Yilmazer M, Ada AO, Suzen S, Demiroglu C, Demirbag AE, Efe S, et al. Biological monitoring of environmental exposure to polycyclic aromatic hydrocarbons: 1-hydroxypyrene in urine of turkish coke oven workers. *Bull Environ Contam Toxicol*. 2006; **76:** 559 – 565.
- Unwin J, Cocker J, Scobbie E, Chambers H. An assessment of occupational exposure to polycyclic aromatic hydrocarbons in the uk. *Ann Occup Hyg.* 2006; 50: 395 – 403.
- 125. Peabody JW, Riddell TJ, Smith KR, Liu Y, Zhao Y, Gong J, et al. Indoor air pollution in rural china: Cooking fuels, stoves, and health status. *Arch Environ Occup Health*. 2005; **60**: 86 – 95.
- Finkelman RB, Belkin HE, Zheng B. Health impacts of domestic coal use in china. Proc Natl Acad Sci U S A. 1999; 96: 3427 – 3431.
- Mumford JL, He XZ, Chapman RS, Cao SR, Harris DB, Li XM, et al. Lung cancer and indoorair pollution in xuan wei, china. *Science*. 1987; 235: 217 – 220.
- Wornat MJ, Ledesma EB, Sandrowitz AK, Roth MJ, Dawsey SM, Qiao YL, et al. Polycyclic aromatic hydrocarbons identified in soot extracts from domestic coal-burning stoves of henan province, china. *Environ Sci Technol.* 2001; 35: 1943 – 1952.
- 129. Gevao B, Al-Bahloul M, Zafar J, Al-Matrouk K, Helaleh M. Polycyclic aromatic hydrocarbons in indoor air and dust in kuwait: Implications for sources and nondietary human exposure. *Arch Environ Contam Toxicol.* 2007; **53**: 503 – 512.
- Levy JI, Houseman EA, Spengler JD, Loh P, Ryan L. Fine particulate matter and polycyclic aromatic hydrocarbon concentration patterns

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in roxbury, massachusetts: A community-based gis analysis. Environ Health Perspect. 2001; **109**: 341 – 347.

- 131. Marr LC, Grogan LA, Wohrnschimmel H, Molina LT, Molina MJ, Smith TJ, et al. Vehicle traffic as a source of particulate polycyclic aromatic hydrocarbon exposure in the mexico city metropolitan area. *Environ Sci Technol.* 2004; **38:** 2584 – 2592.
- Cao SP, Ni HG, Qin PH, Zeng H. Occurrence and human non-dietary exposure of polycyclic aromatic hydrocarbons in soils from shenzhen, china. *J Environ Monit.* 2010; 12: 1445 – 1450.
- Bouchard M, Normandin L, Gagnon F, Viau C, Dumas P, Gaudreau E, et al. Repeated measures of validated and novel biomarkers of exposure to polycyclic aromatic hydrocarbons in individuals living near an aluminum plant in quebec, canada. *J Toxicol Environ Health A*. 2009; 72: 1534 – 1549.
- Weinstein JE, Crawford KD, Garner TR, Flemming AJ. Screeninglevel ecological and human health risk assessment of polycyclic aromatic hydrocarbons in stormwater detention pond sediments of coastal south carolina, USA. *J Hazard Mater*, 2010; **178**: 906 – 916.
- 135. Roth MJ, Dawsey SM, Wang G, Tangrea JA, Zhou B, Ratnasinghe D, et al. Association between gstm1\*0 and squamous dysplasia of the esophagus in the high risk region of linxian, china. *Cancer Lett.* 2000; 156: 73 81.
- 136. Roth MJ, Wei WQ, Baer J, Abnet CC, Wang GQ, Sternberg LR, et

al. Aryl hydrocarbon receptor expression is associated with a family history of upper gastrointestinal tract cancer in a high-risk population exposed to aromatic hydrocarbons. *Cancer Epidemiol Biomarkers Prev.* 2009; **18**: 2391 – 2396.

- 137. Abnet CC, Fagundes RB, Strickland PT, Kamangar F, Roth MJ, Taylor PR, et al. The influence of genetic polymorphisms in ahr, cyp1a1, cyp1a2, cyp1b1, gst m1, gst t1 and ugt1a1 on urine 1-hydroxypyrene glucuronide concentrations in healthy subjects from rio grande do sul, brazil. *Carcinogenesis*. 2007; 28: 112 – 117.
- Li Z, SjDin A, Romanoff LC, Horton K, Fitzgerald CL, Eppler A, et al. Evaluation of exposure reduction to indoor air pollution in stove intervention projects in peru by urinary biomonitoring of polycyclic aromatic hydrocarbon metabolites. *Environ Int.* 2011; 37: 1157 – 1163.
- Cook-Mozaffari PJ, Azordegan F, Day NE, Ressicaud A, Sabai C, Aramesh B. Oesophageal cancer studies in the caspian littoral of iran: Results of a case-control study. *Br J Cancer*. 1979; **39:** 293 – 309.
- Hormozdiari H, Day NE, Aramesh B, Mahboubi E. Dietary factors and esophageal cancer in the caspian littoral of iran. *Cancer Res.* 1975; 35: 3493 – 3498.
- Joint Iran-Iarc Study Group. Esophageal cancer studies in the caspian littoral of iran: Results of population studies: A prodrome. *J Natl Cancer Inst.* 1977; 54: 1127 – 1138.