

EFFECT OF SALICYLIC ACID & CHITOSAN ON INDUCTION OF RESISTANCE IN CHICKPEA AGAINST FUSARIAL WILT & ROOT ROT *

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Abstract

There are many reports that *Fusarium* species causing wilt (*F. oxysporum* f. sp. *ciceri*) and root rot (*F. solani*) are among the main causes of yield loss in chickpea production. Since chemical treatment against these fungi are not so effective, it seems that application of non-toxic chemicals to induce defense mechanism in plant host is desirable. Induction of host resistance in chickpea against these diseases using salicylic acid (SA) and chitosan were investigated. Foliage spray of solutions in 0, 200 and 400 ppm concentration were applied. Plants were grown in growth chamber; at 24 °C and 16:8 (L:D) photoperiod for forty days. Application of 400 ppm of SA had a significant effect on reduction of wilt symptoms, but other applied concentrations of SA had no significant effects on root rot symptoms. Application of 400 ppm concentration of chitosan had a significant effect on wilt symptoms reduction, whereas application of 200 ppm of chitosan had partial effect on reduction of root rot symptoms. An *in vitro* experiment carried out, application of different SA & chitosan concentrations had a direct effect on mycelium growth of both fungal species on PDA. The level of free SA variations in plant tissues treated by 400 ppm concentration of SA was detected by using HPLC method in different post inoculation intervals. The results indicated that the free SA levels decreased after 168 h post inoculation, and wilt disease symptoms appeared gradually afterwards. Chitinase and β -1,4 glucanase enzymes activity and total phenol content in chickpea leaf tissues were evaluated as induced resistance indicators (among others) at 0, 48, 96, and 168 h after inoculation. Increased levels of enzymes activities were observed in 200 ppm of SA however, no significant effects on root rot disease control were obtained in this treatment. The results would suggest that the application of this elicitor (SA) could have partial effects on chickpea wilt symptoms reduction. Maximum levels of indicators activities were observed in 400 ppm of chitosan however, no significant effects on root rot disease control were obtained in this treatment. The results would support the idea that application of this elicitor (chitosan) can play an important role in inducing systemic resistance in chickpea plants against Fusarial wilt disease.

Keywords: Chitinase, β -1,4-glucanase, Total phenolic compounds, HPLC, Chickpea.

See Persian text for figures and tables (Pages ۱۸۱-۱۹۹).

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References

- AGRAWAL, G. K., RAKWAL, R., TAMOGAMI, M., YONEKURA, A. and SAJI, H. 2002. Chitosan active defense/stress response in the leaves of *Oryza sativa* seedlings. **Plant Physiol. Biochem.** 40: 1061-1069.
- AMBORABE, B. E., FLEURAT-LESSARD, P., CHOLLET, J. F. and ROBLIN, G. 2002. Antifungal effects of salicylic acid and other benzoic acid derivatives towards *Eutypa lata*: structure activity relationship. **Plant Physiol. Biochem.** 40: 1051-1060.
- AMBORABE, B-E., BONMORT, J., FLEURAT-LESSARD, P. and ROBLIN, G. 2008. Early events induced by chitosan on plant cells. **J. Exp. Bot.** 59(9): 2317-2324.
- BANSODE, Y. and BAJEKALS, S. 2006. Characterization of chitinase from microorganisms isolated from Lonar Lake. **Indian J. Biotechnol.** 5: 357-363.
- BEN-SHALOM, N. and FALLIK, E. 2003. Further suppression of *Botrytis cinerea* disease in cucumber seedling by chitosan-copper complex as compared with chitosan alone. **Phytoparasitica** 31: 99-102.
- BHATTACHARYA, A., SOOD, P. and CITOVSKEY, V. 2010. The roles of plant phenolics in defence & communication during *Agrobacterium* & *Rhizobium* infection. **Plant Pathol.** 115:705-19
- BRADFORD, M. M. 1976. A rapid and sensitive method for quantification of protein utilizing the principle of protein-dye binding. **Anal. Biochem.** 72: 248-254.
- DEMPSEY, D. A., SHAH, J. and KLESSIG, D. F. 1999. Salicylic acid and disease resistance in plants. **Crit. Rev. Plant Sci.** 18: 547-575.
- DURRANT, W. E. and DONG, X. 2004. Systemic Acquired Resistance. **Ann. Rev. Phytopathol.** 42: 185-209.
- ELAD, Y. and FREEMAN, S. 2002. Biological control of fungal plant pathogens. Pp. 3-109, *In*: F. Kempken (Eds.), **The Mycota, A Comprehensive Treatise on Fungi as Experimental Systems for Basic and Applied Research.** Vol. XI, Agric. Appl.
- GERLACH, W. and ERSHAD, D. 1970. Beitrag zur kenntnis der *Fusarium* and *Cylandrocarpon* arten in Iran. **Nov. Hedwigia** 20: 725-784.
- GONG, M., LI, C., XU, L. and WEI, G. 2011. Isolation of endophytic bacteria from nodule of *Sophora alopecuroides* and effect of biological control against *Fusarium* wilt. **Microbiol.** 38: 865-870.
- GOPALAKRISHNAN, S. 2000. **Toxigenicity of Fusarium species causing wilt of chickpea. Presented for the degree of doctor of philosophy,** University of London. 219 p.
- HAHLBROCK, K. and SCHEEL, D. 1989. Physiology and molecular biology of phenylpropanoid metabolism. **Ann. Rev. Plant Physiol. Plant Mol. Biol.** 40:347-369.
- HUMMERSCHMID, T. R., METRAUX, J-P. and VAN LOON, L.C. 2001. Inducing resistance: a summary of papers presented at the first international symposium on induced resistance to plant diseases, Corfu, May 2000. **Eur. J. Plant Pathol.** 107:1-6.
- JAYARAJ, J., RAHMAN, M., WAN, A. and PUNJA, Z. K. 2008. Enhanced resistance to foliar fungal pathogens in carrot by application of elicitors. **Ann. Appl. Biol.** 155: 71-80.
- MALAMY, J., HENNIG, J. and KLESSIG, D. F. 1992. Temperature – dependent induction of salicylic acid and its conjugates during the resistance response to tobacco mosaic virus infection. **Plant Cell.** 4: 359-366.
- MALICK, C. P. and SINGH, M. B. 1980. **Plant Enzymology and Histo-Enzymology.** Kalyani Pub., New Delhi, 280 pp.
- MANDAL, S., MALLICK, N. and MITRA, A. 2009. Salicylic acid-induced resistance to *Fusarium oxysporum* f. sp. *lycopersici* in tomato. **Plant Physiol. Biochem.** 47: 642-649.
- MAUCH-MANI, B. and METRAUX, J. P. 1998. Salicylic acid and Systemic Acquired Resistance to pathogen attack. **Ann. Bot.** 82:535-540.
- NANDEESHKUMAR, P., SUDISHA, J., RAMACHANDRA, K. K., PRAKASH, H. S., NIRANJANA, S. R. and SHEKAR, S. H. 2008. Chitosan induced resistance to downy mildew in sunflower caused by *Plasmopara halstedii*. **Physiol. Mol. Plant Pathol.** 72: 188-194.
- NELSON, P. E., TOUSSOUN, T. A. and MARASAS, W. F. O. 1983. **Fusarium species, An illustrated manual for identification.** The Pennsylvania State University Press, Univa. Park and London. 193 pp.

- RAJU, S. , JAYALAKSHMI, S. K. and SREERAMULU, K. 2008. Comparative study on the induction of defense related enzymes in two different cultivar of chickpea genotypes by alicyclic acid, spermin and *F. oxysporum* f. sp. *ciceri*. **Aust. J. Crop Sci.** 2(3): 121-140.
- REIGNAULT, P. and WALTERS, D. 2007. Topical application of inducers for disease control. Pp. 179-200, *In*: D. Walters, A. Newton, G. Lyon (Eds.), **Induced Resistance for Plant Defence**. Oxford, Blackwell Pub., UK.
- RODRIGUEZ, A. T. , RAMIREZ, M. A. , CARDENAS, R. M. , HERNANDEZ, A. N. , VELAZQUEZ, M. G. and BAUTISTA, S. 2007. Induction of defense response of *Oryza sativa* against *Pyricularia grisea* by treating seeds with chitosan and hydrolyzed chitosan. **Pest. Biochem. Physiol.** 89: 206-215.
- SAFAIE, H. , and MANSOORI, B. 2007. Reactions of some chickpea genotypes to black root rot pathogen in Fars province. **Seed & Plant** 23: 31-41(In Farsi).
- SAREMI, H. 2000. Plant disease caused by Fusarium species. Jahade daneshgahi press mashhad, Iran.160p.
- WHITE, R. F. 1979. Acetylsalicylic acid (aspirin) induces resistance to tobacco mosaic virus in tobacco. **Virology** 99: 410-412.
- YAO, K. , DE LUCA, V. and BRISSON, N. 1995. Creation of a metabolic sink for tryptophan alters the phenylpropanoid pathway and the susceptibility of potato to *Phytophthora infestans*. **Plant Cell** 7:1787-99.
- YEDIDIA, I. , BENHAMOU, N. , KAPULNIK, Y. and CHET, I. 2000. Induction and accumulation of PR-proteins activity during early stages of root colonization by the mycoparasite *Trichoderma harzianum* strain T-203. **Plant Physiol. Biochem.** 38: 863-873.

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