

Original Article



Identification of VIM and IMP genes and metallo-beta-lactamase enzymes in *Escherichia coli* isolates by molecular and phenotypic methods in shahrekord educational hospitals

Farshad Kakian¹, Kouros Naderi², Mohamad Hosaein Rezaei², Majid Validi³, Behnam Zamanzad^{4*}, Abolfazl Gholipour⁵

¹Department of Bacteriology and Virology, School of Medicine, Shiraz University of Medical Sciences, Shiraz, Iran.

²Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran.

³Department of Laboratory Sciences, School of Allied Medical Sciences, Shahrekord University of Medical Sciences, Shahrekord, Iran.

⁴Professor of Bacteriology, School of Medicine, Shahrekord University of Medical Sciences, Shahrekord, Iran.

⁵Associate Professor of Bacteriology, Department of Microbiology and Immunology, School of Medicine, Shahrekord University of Medical Sciences, Shahrekord, Iran.

*Corresponding Author: Behnam Zamanzad, Department of Microbiology and Immunology, Cellular and Molecular Research Center, Shahrekord University of Medical Sciences, Shahrekord, IR Iran, Tel: +983833346732, Fax: +98383334911, E-mail: bzamanzad@yahoo.com

Abstract

Background and aims: Among urine pathogens, *Escherichia coli* (*E. coli*) causes 80% of urinary tract infections (UTIs). Due to the destructive nature of penicillins, cephalosporins and carbapenems (except for monobactam such as aztreonam) and carbapenemase enzymes have created many problems for treating infectious diseases. Therefore, this study aimed to investigate the phenotypic and molecular characterization of metallo-beta-lactamase (MBL) genes produced by *E. coli* isolates in an educational hospital during 2016-2017.

Methods: This cross-sectional study investigated 80 UTI samples affected by *E. coli*. In addition, antibiotic susceptibility was evaluated by disk diffusion and E-test methods for two antibiotics of meropenem and imipenem. Phenotypic tests containing modified Hodge test, ethylenediaminetetraacetic acid (EDTA) disk synergy test, and AmpC Disk were performed to identify MBL enzyme-producing strains. Finally, the frequency of Verona integron-encoded metallo-β-lactamase (VIM) and imipenemase (IMP) genes was determined by polymerase chain reaction (PCR).

Results: Among 80 *E. coli* samples, 21 (26.25%) isolates were resistant to meropenem and imipenem as detected by the disk-diffusion method and E-test. Further, phenotypic tests including modified Hodge test, EDS test, and AmpC disk test showed the positivity of 15 (18.75%), 15 (18.75%), and 8 (10%) isolates, respectively ($P < 0.001$). Eventually, polymerase chain reaction (PCR) test results for the VIM gene showed 19 (23.75%) positive isolates of *E. coli*, but the IMP gene was observed in none of the isolates ($P < 0.001$).

Conclusion: In general, the emergence of *E. coli* producing MBL enzymes is a serious threat among clinical infections. The findings of this study indicated the presence of *E. coli* producing MBL. These enzymes can degrade carbapenems antibiotics, the last class current treatment of multiple drug-resistance infections.

Keywords: *Escherichia coli*, Drug resistance, VIM, IMP

Received: 8 May 2019, Accepted: 20 June 2019, ePublished: 28 February 2020

Introduction

The family of Enterobacteriaceae, especially *Escherichia coli*, causes urinary tract infections (UTIs) (1). UTI is one of the most important factors in renal scarring, kidney tissue damage, kidney failure, poor growth, kidney stones, and hypertension (2-4). Almost 30%-40% of UTI are nosocomial infections which are considered as the most significant cause of sepsis in hospitalized patients (5). *E. coli* in the Enterobacteriaceae family causes hospital and acquired infections more than the other members of its family (6). In addition, *E. coli* is the most common

etiologic agent of the UTI accounting for 80% of cases (7). Resistance to antibiotic therapy in patients with UTIs represents an example of the increasing problem of antimicrobial resistance (8). This type of resistance has always been a major problem for human health (9) and affects patients in general hospitals worldwide (10). Carbapenem antibiotics family consists of ertapenem, imipenem, doripenem, and meropenem. Nowadays, because of the broad spectrum and non-hydrolysis by beta-lactamases enzymes, this family has become the most important selected drug for treating serious infections in

Archive of SID

Enterobacteriaceae resistance to multiple drugs that produce extended broad-spectrum β -lactamases (ESBLs) (11-13). Carbapenemase is an important mechanism of resistance to carbapenem antibiotics. Further, resistance by metallo-beta-lactamases (MBLs) often relies on the plasmid (14,15). According to ambler classification, carbapenemase belongs to molecular classes A, B, and D. Class A carbapenemase includes Guiana-extended-spectrum, imipenem-hydrolyzing β -lactamase, *Serratia marcescens* enzyme, and *Klebsiella pneumonia* carbapenemase (KPC) enzymes that can be inhibited by varying degrees of clavulanate and they hydrolyse penicillins and cephalosporins more than carbapenems. Some of them, including KPC, lack the strong carbapenemase ability, which might be considered as ESBLs. Furthermore, carbapenemase class B is an MBL that is considered as group B of ambler based on the similarity of their amino acid sequences. Moreover, it can be classified into group 3 (Bush's classification) based on the structure of the substrate and the inhibitor component. MBL activity depends on zinc metal, which is capable of hydrolyzing beta-lactam antibiotics and all carbapenems except for aztreonam. MBLs are resistant to beta-lactamase inhibitors but can be prevented by chelating agents such as ethylenediaminetetraacetic acid (EDTA). According to molecular structures, MBLs are divided into 6 categories of Verona integron-encoded metallo- β -lactamase (VIM), imipenemase (IMP), German imipenemase, Seoul imipenemase (SIM), São Paulo metallo-beta-lactamase, and Adelaide imipenemase. The VIM, IMP, and SMP-1 were identified in *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, and other members of the Enterobacteriaceae family (16-20). Considering the above-mentioned explanations, the aim of this study was to determine the frequency of VIM and IMP genes and MBL enzymes in *E. coli* isolates by molecular and phenotypic methods in educational hospitals of Shahrekord, Iran.

Materials and Methods

Bacterial Isolates

In this study, 80 *E. coli* isolates of UTI patients were prepared in several sections of the university hospital (e.g., urology, women, internal, intensive care unit) from May to October 2016. These patients were infected by UTI within 48-72 hours after hospitalization and the primary urine cultures were negative for them.

Antimicrobial susceptibility testing and minimum inhibitory concentration (MIC)

First, the samples were cultured on blood agar and eosin-methylene-blue mediums (Himedia) and then different biochemical identifier tests such as Triple sugar iron agar, Simon citrate, methyl red/Voges-Proskauer, SIM, and oxidase were performed on the isolates. In addition, antibiotic susceptibility was determined by the disk diffusion method (Kerby-Bauer) for meropenem and

imipenem (Mast) and the plates were incubated for 18-24 hours at 37°C. Finally, the Epsilon test (E-test) was used (Liofilchem Company) for two antibiotics of meropenem and imipenem on the Mueller-Hinton agar culture medium in order to determine the MIC.

Phenotypic diagnosis of MBLs

Phenotypic tests such as the modified Hodge test (MHT), EDTA disk synergy test (EDS), and AmpC disk test were performed to detect the strains containing MBL enzymes. Further, *E. coli* ATCC 25922 and *Klebsiella pneumonia* ATCC 1705 were considered as negative and positive controls for running MHT, respectively. Furthermore, bacterial suspension equivalent to 0.5 McFarland was prepared from the fresh cultures of *E. coli* ATCC 25922 and then diluted in a 1:10 ratio and passaged on Mueller-Hinton agar. The plates were maintained at 37°C overnight. The appeared clover leaf shape in the inhibition site of bacterial growth resulted from the production of carbapenemase by the tested bacteria strain (21-23). Moreover, 0.5 molar EDTA solution and meropenem disk were used in the EDS test, followed by passaging sample suspensions equivalent to 0.5 McFarland on Mueller-Hinton agar. A meropenem (10 μ g) disk was placed on the plate and then 10 mL of 0.5 molar EDTA solution was added to a blank disk (filter paper with a diameter of 6 mm) in an aseptic condition. Additionally, the blank disk was put at a 10 mm distance of the meropenem disk and incubated overnight at 37°C. The increased growth inhibition zone between the two disks was the indicator of MBL positive bacteria (21,22). In addition, the AmpC disk test method was used to recognize beta-lactamase AmpC. Next, 0.5 McFarland suspension of *E. coli* ATCC 25922 on the Mueller-Hinton culture medium was passaged and a cefoxitin disk (30 μ g) was placed on the plate near a blank disk (Whatman paper) that was exposed to normal saline. Then, colonies resistant to meropenem were inoculated in the blank disk. A positive result for the production of AmpC β -lactamase was based on the observation of a dentin in the growth area of the cefoxitin disk in the vicinity of the blank disk (21-23).

Determination of MBL genes

The positive isolates for carbapenemase enzymes were used to investigate VIM and IMP genes by the polymerase chain reaction (PCR). Hence, their DNA concentrates were extracted by boiling. In the boiling process, 1 mL of sterile distilled water was poured in a 1.5 mL microtube. Then, 4-5 fresh bacterial colonies were dissolved and the microtube was placed on a heated plate at 95°C for 15 minutes to boil. Next, the samples were centrifuged at 14000 rpm for 5 minutes and the transparent liquid containing bacterial DNA was removed for PCR. In this study, *Acinetobacter baumannii* AC54/97 and *Pseudomonas aeruginosa* PO510 were a positive control for IMP and VIM genes (24). Further, 16srRNA was considered as internal control (25).

The primers used for VIM and IMP genes were designed by Oligo 7 software (Table 1).

Statistical analysis

Descriptive statistics (i.e., frequency, percentage, mean, and standard deviation) and inferential statistics (Fisher's exact test) were analyzed and $P < 0.05$ was considered as the significance level.

Results

In the disk diffusion method, 21 isolates of *E. coli* were resistant and 5 isolates were intermediate for meropenem and imipenem. Furthermore, 21 isolates of *E. coli* were resistant ($MIC \geq 4 \mu\text{g/mL}$) to meropenem and imipenem antibiotics in the E-test.

Modified Hodge test

Resistant strains to meropenem and imipenem by E-test were used in MHT to detect carbapenemase enzymes and 15 (18.75%) isolates of *E. coli* were positive ($P < 0.001$). Figure 1 shows the positive and negative samples by MHT.

EDTA disk synergy test

Escherichia coli isolates resistant to carbapenemase antibiotics were used in the EDS test to produce carbapenemase enzymes. In this test, 15 (18.75%) isolates were positive ($P < 0.001$). Figure 2 illustrates two separate plates (1 & 2) that are side by side and considered as negative and positive isolates for the EDS test, respectively.

AmpC disk test

The presence of a notch in the side of the disk reflects the production of AmpC β -lactamase by *E. coli*, which is on the blank disk (Figure 3, AmpC positive). In this test, 8 isolates (10%) of *E. coli* were AmpC beta-lactamase ($P < 0.001$). Table 2 presents the results of phenotypic tests for detecting carbapenemase enzymes.

The results were similar for disk diffusion and E-Test. Moreover, among the tests for detecting carbapenemase enzymes, the MHT and EDS demonstrated the same

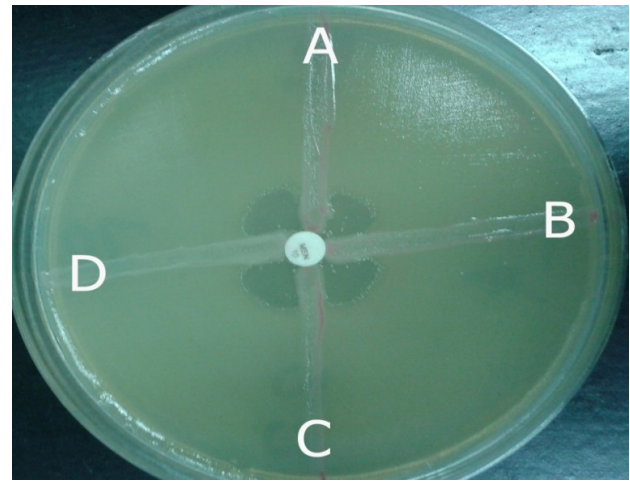


Figure 1. Modified Hodge test. *E. coli* ATCC25922 causes leaf clover halo in the inhibition zone around meropenem as a negative control by the Metallo-beta-lactamase enzyme production. A is the positive control strain of *Klebsiella pneumonia* (ATCC BAA-1705). In addition, D and B are positive *E. coli* strains and C is the negative isolate.

results, while their results differed from that of the AmpC test.

Polymerase chain reaction

The electrophoresis technique was performed on the 8% polyacrylamide gel after completing the PCR steps. After electrophoresis, the poly acryl amide gel was stained with the silver nitrate method (Figure 4). The results of this test are shown in Tables 3 and 4.

Discussion

Today, the resistance of bacteria to various antibiotics has become a global problem due to the uncontrolled administration of different antibiotics for treating bacterial infections. Hence, resistant strains are selected dramatically (26-28). Resistance to antibiotic therapy in patients with UTIs is considered as an example of increased antimicrobial resistance (12). Antimicrobial resistance has always been a major concern for human health (6), affecting patients throughout the world (29). MBL encoding genes are found more frequently on integrons, especially Class 1 integron, which has the genetic mobility of these elements and promotes the release of these enzymes in different parts of a hospital (30). Similar to studies in Columbian, Japanese, and Italian hospitals, VIM and IMP were observed without the use of carbapenem antibiotics, which confirms the

Table 1. Primers Used for VIM and IMP by PCR

Primer Sequence (5' to 3')	Size (bp)
VIM-1F TGGTTGTATACGTCCCCTCA	206
VIM-1R TGTGTGCTGGAGCAAGTCTA	
IMP-1F TAACGGGTGGGGCGTTGTCCT	179
IMP-1R CGCCCGTGCTCGCTATGAAA	

Note. VIM: Verona integron-encoded metallo- β -lactamase; IMP: imipenemase; PCR: polymerase chain reaction.

Table 2. Results of phenotypic tests for the detection of MBL enzymes

Isolates (N)	Disk-diffusion	E-Test	MHT	EDS Test	AmpC Disk Test
80	21 (26.25%)	21 (26.25%)	15 (18.75%)	15 (18.75%)	8 (10%)

Note. MBL: metallo-beta-lactamase; E-test: Epsilon test; MHT: modified Hodge test; EDS: EDTA disk synergy test.

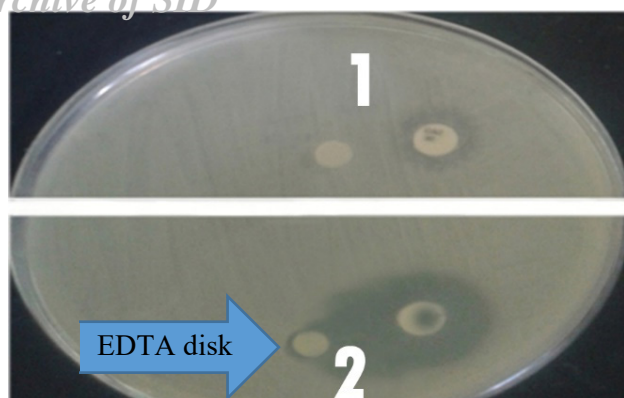


Figure 2. Ethylenediaminetetraacetic acid (EDTA) disk synergy test. At plate 2, increasing the diameter of the inhibition zone around the disk containing EDTA (blank) and between two disks of meropenem (right) and blank (left) represents Metallo-beta-lactamase production by bacteria in the presence of EDTA, inhibits the production of enzymes, and increases the diameter of the halo between the two disks.

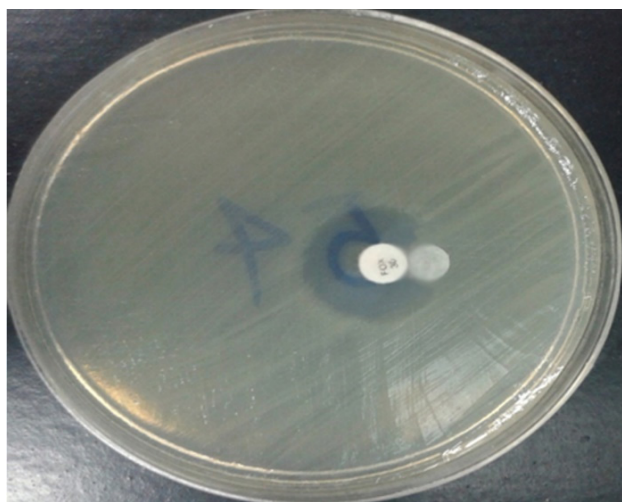


Figure 3. AmpC disk test. The notches on the side of the disk represent AmpC β -lactamase producing *E. coli*, which prevented the creation of a complete halation around the ceftiofur disk by *E. coli* ATCC 25922.

transfer of these genes between different species of bacteria (31,32). In this study, the prevalence of resistance to meropenem and imipenem in *E. coli* strains was 26%. In a study by Sahin et al, the production of MBL enzymes in 43 isolates of *E. coli* and *Klebsiella* (collected from various parts of the hospital) was evaluated during which 35 strains were carbapenem-resistant and produced MBL enzymes (33). The Center for Disease Control has supplied the MHT for MBL-producing Enterobacteriaceae isolates in order to detect MBL enzymes. Although this test has a high potential for detecting carbapenemase enzymes, the results are sometimes false positive by CTX-M and AmpC due to the partial hydrolysis of the carbapenemase (34). In

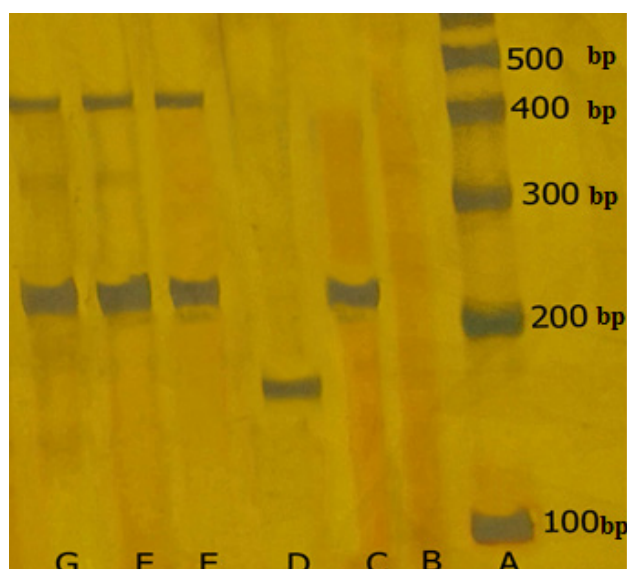


Figure 4. Colored gel of polyacrylamide. Column A: Ladder bp; B: Negative Control; C: Positive control of Verona integron-encoded Metallo- β -lactamase (VIM) gene (*P. aeruginosa* PO510); D: Positive control of imipenemase (IMP) gene (*A. baumannii* AC54 / 97); E, F, and G columns of VIM gene and internal control gene 16srRNA without IMP.

the MHT, the results are more reliable when used with the imipenem disk with 0.5 molar EDTA or 50 mM ZNSO₄ (35). The results of previous studies showed that MHT has less positive outcomes compared to the other tests, ranging from 14.8% to 51.16% (35-37), which is consistent with our results. In this study, 21 and 15 isolates of *E. coli* were positive by E-test and MHT method, respectively. In the research by Kumar et al, 28 isolates resistant of *E. coli* to meropenem (carbapenem) were reported and 6 isolates were positive by the MHT test, which indicates the low accuracy of MHT in detecting MBL strains (38). In addition, Sahin et al studied 43 strains of *Klebsiella* and *E. coli* and found that 35 strains were positive for the MHT, which were not reported in any of the VIM and IMP genes after the PCR (33). In this study, 15 isolates were positive in producing MBLs by the EDS test. Further, Kanchanadevi and Sekaran concluded that between 76 isolates of *E. coli*, 25 were resistant to imipenem using the E-test and 12 isolates were carbapenemase producing by the EDS test (39). AmpC is a cephalosporinase enzyme that leads to the hydrolysis of cefamycins (ceftiofur and cefotetan), which is a component of broad-spectrum beta-lactamase. To detect the AmpC beta-lactamase enzyme, AmpC disk testing can routinely be used in clinical laboratories. These laboratories have difficulty to appropriately detect this enzyme and there is little knowledge about the importance of this enzyme, which has led to the outbreak of these bacterial strains and even to treatment failure. In a study conducted in India, this enzyme was present in hospitalized strains, which

Table 3. Abundance VIM and IMP Genes

Isolates (N)	VIM-1		P value	IMP-1		P value
	Positive	Negative		Positive	Negative	
80	19 (23.75%)	61 (76.2%)	< 0.001	0	80 (100%)	< 0.001

Note. VIM-1: Verona integron–encoded metallo-β-lactamase 1; IMP-1: imipenemase 1.

Table 4. The relationship between resistance to meropenem and imipenem and the presence of VIM and IMP genes in *E. coli*

Gene	Meropenem and Imipenem Resistant (N=21)	
	Positive	Negative
VIM	19	2
IMP	0	21
Total	19	23

Note. VIM: Verona integron–encoded Metallo-β-lactamase; IMP: Imipenemase.

was more common in the *Acinetobacter* spp. Further, the prevalence was 6.97% for *E. coli* (39, 40). In our research, 8 isolates produced AmpC beta-lactamase among *E. coli* isolates, which is consistent with the results of the study of Koshesh et al in Kerman. In total, 2% AmpC β-lactamase was positive among the 105 samples of *E. coli* isolated from UTI (41).

The presence of VIM and IMP genes with PCR was also evaluated in our study. The frequency of the VIM gene in *E. coli* isolates was 23.75% and the IMP gene was not found in *E. coli* strains. Based on the reports of Mosavian and Koraei, the frequency of the IMP gene in the strains of *Klebsiella*, *E. coli*, and *Enterobacter* was zero (42). In the study conducted by Tawfik et al, 15 strains of MBL-producing bacteria were reported by phenotypic methods. All the strains were VIM positive, but the IMP gene was not found in any of these strains. The results of the above-mentioned studies are in line with those of our study, showing a low prevalence of IMP genes in *E. coli* (43). Furthermore, Zeighami et al evaluated the molecular prevalence of VIM and IMP genes among 100 isolates and found that the prevalence was 41% and zero for *Klebsiella* and *E. coli* samples, respectively, indicating the different outbreaks of these genes in bacteria and regions (44). Similarly, Mushi et al investigated 227 bacterial samples including *Klebsiella*, *E. coli*, and *Pseudomonas aeruginosa*. After PCR, the highest frequency was attributed to IMP and VIM genes with 49 and 28 cases of IMP and VIM, respectively, which is also consistent with our study results representing a lower abundance of VIM and IMP genes in *E. coli* compared to the other bacteria (45).

Conclusion

Due to the failure of antibiotic treatment and increased therapeutic charges, especially in UTIs, all microbiology laboratories should now be able to identify MBLs-producing bacteria. In the absence of new antibiotics for treating infections caused by Gram-negative bacteria

resistant to several antibiotics, the carbapenem will be ineffective in the near future. In addition, the treatment failure of these infections is predictable since these antibiotics are the preferred drug in treating multi-drug resistant bacteria. Resistance to carbapenem antibiotics is a growing problem and they might be replaced by novel antibiotics in the near future. Eventually, to control the emission of MBL-producing strains and the resistance of these microorganisms, it is necessary to come up with an appropriate treatment for these patients.

Conflict of Interests

None.

Acknowledgments

This study is a research project approved by Shahrekord University of Medical Sciences (approval No. 2057) on March 6, 2016. We, hereby, gratefully thank all the people who assisted us in conducting this work. In addition, the authors wish to thank Mr. H. Argasi at the Research Consultation Center of Shiraz University of Medical Sciences for his invaluable assistance in editing this manuscript.

Ethical Statement

The study protocol was obtained by the ethics code of IR.SKUMS. REC.1394.282 from Shahrekord University of Medical Sciences.

Authors Contribution

FK, AG and BZ: design of study; KN, AG, BZ, FK and MV: acquisition of data; AG and MR: evaluation of data and preparation of the manuscript; AG, FK and MV assessment of data.

Funding/Support

This research was supported by the grant of the research project of the Shahrekord University of Medical Sciences (Shahrekord, Iran).

References

- Yang J, Ye L, Wang W, Luo Y, Zhang Y, Han L. Diverse prevalence of 16S rRNA methylase genes *armA* and *rmtB* amongst clinical multidrug-resistant *Escherichia coli* and *Klebsiella pneumoniae* isolates. *Int J Antimicrob Agents*. 2011;38(4):348-51. doi: 10.1016/j.ijantimicag.2011.04.021.
- Hodson CJ, Maling TM, McManamon PJ, Lewis MG. The pathogenesis of reflux nephropathy (chronic atrophic pyelonephritis). *Br J Radiol*. 1975;Suppl 13:1-26.
- Gordon I, Barkovics M, Pindoria S, Cole TJ, Woolf AS. Primary vesicoureteric reflux as a predictor of renal damage in children hospitalized with urinary tract infection: a systematic review and meta-analysis. *J Am Soc Nephrol*. 2003;14(3):739-44. doi: 10.1097/01.asn.0000053416.93518.63.
- Roussey-Kesler G, Gadjos V, Idres N, Horen B, Ichay L, Leclair MD, et al. Antibiotic prophylaxis for the prevention of recurrent urinary tract infection in children with low grade vesicoureteral reflux: results from a prospective randomized study. *J Urol*. 2008;179(2):674-9. doi: 10.1016/j.juro.2007.09.090.
- Karchmer TB, Giannetta ET, Muto CA, Strain BA, Farr BM. A randomized crossover study of silver-coated urinary catheters

- in hospitalized patients. *Arch Intern Med*. 2000;160(21):3294-8. doi: 10.1001/archinte.160.21.3294.
6. Baroud M, Dandache I, Araj GF, Wakim R, Kanj S, Kanafani Z, et al. Underlying mechanisms of carbapenem resistance in extended-spectrum beta-lactamase-producing *Klebsiella pneumoniae* and *Escherichia coli* isolates at a tertiary care centre in Lebanon: role of OXA-48 and NDM-1 carbapenemases. *Int J Antimicrob Agents*. 2013;41(1):75-9. doi: 10.1016/j.ijantimicag.2012.08.010.
 7. Sahm DF, Thornsberry C, Mayfield DC, Jones ME, Karlowsky JA. Multidrug-resistant urinary tract isolates of *Escherichia coli*: prevalence and patient demographics in the United States in 2000. *Antimicrob Agents Chemother*. 2001;45(5):1402-6. doi: 10.1128/aac.45.5.1402-1406.2001.
 8. Ko KS, Suh JY, Peck KR, Lee MY, Oh WS, Kwon KT, et al. In vitro activity of fosfomycin against ciprofloxacin-resistant or extended-spectrum beta-lactamase-producing *Escherichia coli* isolated from urine and blood. *Diagn Microbiol Infect Dis*. 2007;58(1):111-5. doi: 10.1016/j.diagmicrobio.2006.11.015.
 9. Green VL, Verma A, Owens RJ, Phillips SE, Carr SB. Structure of New Delhi metallo-beta-lactamase 1 (NDM-1). *Acta Crystallogr Sect F Struct Biol Cryst Commun*. 2011;67(Pt 10):1160-4. doi: 10.1107/s1744309111029654.
 10. Fernández A, Pereira MJ, Suárez JM, Poza M, Treviño M, Villalón P, et al. Emergence in Spain of a multidrug-resistant *Enterobacter cloacae* clinical isolate producing SFO-1 extended-spectrum beta-lactamase. *J Clin Microbiol*. 2011;49(3):822-8. doi: 10.1128/jcm.01872-10.
 11. Meletis G, Tzampaz E, Protonotariou E, Sofianou D. Emergence of *Klebsiella pneumoniae* carrying bla(VIM) and bla(KPC) genes. *Hippokratia*. 2010;14(2):139-40.
 12. Cagnacci S, Gualco L, Roveta S, Mannelli S, Borgianni L, Docquier JD, et al. Bloodstream infections caused by multidrug-resistant *Klebsiella pneumoniae* producing the carbapenem-hydrolysing VIM-1 metallo-beta-lactamase: first Italian outbreak. *J Antimicrob Chemother*. 2008;61(2):296-300. doi: 10.1093/jac/dkm471.
 13. Zhang F, Zhu D, Xie L, Guo X, Ni Y, Sun J. Molecular epidemiology of carbapenemase-producing *Escherichia coli* and the prevalence of ST131 subclone H30 in Shanghai, China. *Eur J Clin Microbiol Infect Dis*. 2015;34(6):1263-9. doi: 10.1007/s10096-015-2356-3.
 14. Goudarzi H, Taherpour A, Fallah F, Pourkaveh B, Erfanimesh S, Hashemi A. Laboratory detection of carbapenemases in gram-negative bacteria. *Arch Clin Infect Dis*. 2016;11(2):e32816. doi: 10.5812/archcid.32816.
 15. Giske CG, Gezelius L, Samuelsen Ø, Warner M, Sundsfjord A, Woodford N. A sensitive and specific phenotypic assay for detection of metallo-beta-lactamases and KPC in *Klebsiella pneumoniae* with the use of meropenem disks supplemented with aminophenylboronic acid, dipicolinic acid and cloxacillin. *Clin Microbiol Infect*. 2011;17(4):552-6. doi: 10.1111/j.1469-0691.2010.03294.x.
 16. Gholipour A, Soleimani N, Shokri D, Mobasherizadeh S, Kardi M, Baradaran A. Phenotypic and molecular characterization of extended-spectrum beta-lactamase produced by *Escherichia coli*, and *Klebsiella pneumoniae* isolates in an educational hospital. *Jundishapur J Microbiol*. 2014;7(10):e11758. doi: 10.5812/jjm.11758.
 17. Thomson KS. Extended-spectrum-beta-lactamase, AmpC, and Carbapenemase issues. *J Clin Microbiol*. 2010;48(4):1019-25. doi: 10.1128/jcm.00219-10.
 18. Khalil MAF, Elgaml A, El-Mowafy M. Emergence of multidrug-resistant New Delhi metallo-beta-lactamase-1-producing *Klebsiella pneumoniae* in Egypt. *Microb Drug Resist*. 2017;23(4):480-7. doi: 10.1089/mdr.2016.0003.
 19. Mlynarcik P, Roderova M, Kolar M. Primer evaluation for PCR and its application for detection of carbapenemases in Enterobacteriaceae. *Jundishapur J Microbiol*. 2016;9(1):e29314. doi: 10.5812/jjm.29314.
 20. Adwan G, Bourinee H, Othman S. Prevalence of metallo-beta-lactamases producing *Escherichia coli* isolated from north of Palestine. *J Microbiol Antimicrob Agents*. 2016;2(1):9-15.
 21. Amjad A, Mirza I, Abbasi S, Farwa U, Malik N, Zia F. Modified Hodge test: a simple and effective test for detection of carbapenemase production. *Iran J Microbiol*. 2011;3(4):189-93.
 22. Lee K, Chong Y, Shin HB, Kim YA, Yong D, Yum JH. Modified Hodge and EDTA-disk synergy tests to screen metallo-beta-lactamase-producing strains of *Pseudomonas* and *Acinetobacter* species. *Clin Microbiol Infect*. 2001;7(2):88-91. doi: 10.1046/j.1469-0691.2001.00204.x.
 23. Goel V, Hogade S, Karadesai S. Prevalence of extended-spectrum beta-lactamases, AmpC beta-lactamase, and metallo-beta-lactamase producing *Pseudomonas aeruginosa* and *Acinetobacter baumannii* in an intensive care unit in a tertiary care hospital. *Journal of the Scientific Society*. 2013;40(1):28-31. doi: 10.4103/0974-5009.109691.
 24. Shahcheraghi F, Abbasalipour M, Feizabadi M, Ebrahimipour G, Akbari N. Isolation and genetic characterization of metallo-beta-lactamase and carbapenemase producing strains of *Acinetobacter baumannii* from patients at Tehran hospitals. *Iran J Microbiol*. 2011;3(2):68-74.
 25. Damavandi MS, Gholipour A, Latif Pour M. Prevalence of class D carbapenemases among extended-spectrum beta-lactamases producing *Escherichia coli* isolates from educational hospitals in Shahrekord. *J Clin Diagn Res*. 2016;10(5):DC01-5. doi: 10.7860/jcdr/2016/17722.7739.
 26. Zowawi HM, Sartor AL, Balkhy HH, Walsh TR, Al Johani SM, Aljindan RY, et al. Molecular characterization of carbapenemase-producing *Escherichia coli* and *Klebsiella pneumoniae* in the countries of the Gulf cooperation council: dominance of OXA-48 and NDM producers. *Antimicrob Agents Chemother*. 2014;58(6):3085-90. doi: 10.1128/aac.02050-13.
 27. Peleg AY, Hooper DC. Hospital-acquired infections due to gram-negative bacteria. *N Engl J Med*. 2010;362(19):1804-13. doi: 10.1056/NEJMra0904124.
 28. Podschun R, Ullmann U. *Klebsiella* spp. as nosocomial pathogens: epidemiology, taxonomy, typing methods, and pathogenicity factors. *Clin Microbiol Rev*. 1998;11(4):589-603.
 29. Essack SY. Treatment options for extended-spectrum beta-lactamase-producers. *FEMS Microbiol Lett*. 2000;190(2):181-4. doi: 10.1111/j.1574-6968.2000.tb09283.x.
 30. Kil KS, Darouiche RO, Hull RA, Mansouri MD, Musher DM. Identification of a *Klebsiella pneumoniae* strain associated with nosocomial urinary tract infection. *J Clin Microbiol*. 1997;35(9):2370-4.
 31. Khorsi K, Messai Y, Hamidi M, Ammari H, Bakour R. High prevalence of multidrug-resistance in *Acinetobacter baumannii* and dissemination of carbapenemase-encoding genes blaOXA-23-like, blaOXA-24-like and blaNDM-1 in Algiers hospitals. *Asian Pac J Trop Med*. 2015;8(6):438-46. doi: 10.1016/j.apjtm.2015.05.011.
 32. Yatsuyanagi J, Saito S, Harata S, Suzuki N, Ito Y, Amano K, et al. Class 1 integron containing metallo-beta-lactamase gene blaVIM-2 in *Pseudomonas aeruginosa* clinical strains isolated in Japan. *Antimicrob Agents Chemother*. 2004;48(2):626-8. doi: 10.1128/aac.48.2.626-628.2004.
 33. Sahin K, Tekin A, Ozdas S, Akin D, Yapislir H, Dilek AR, et al. Evaluation of carbapenem resistance using phenotypic and

- genotypic techniques in Enterobacteriaceae isolates. *Ann Clin Microbiol Antimicrob.* 2015;14:44. doi: 10.1186/s12941-015-0105-1.
34. Swain B, Ota S, Rout S. Resistance pattern of *Klebsiella* from urinary tract infections in a tertiary care hospital. *Indian Pract.* 2015;68(2):18-22.
 35. Peleg AY, Seifert H, Paterson DL. *Acinetobacter baumannii*: emergence of a successful pathogen. *Clin Microbiol Rev.* 2008;21(3):538-82. doi: 10.1128/cmr.00058-07.
 36. Lee K, Lim YS, Yong D, Yum JH, Chong Y. Evaluation of the Hodge test and the imipenem-EDTA double-disk synergy test for differentiating metallo-beta-lactamase-producing isolates of *Pseudomonas* spp. and *Acinetobacter* spp. *J Clin Microbiol.* 2003;41(10):4623-9. doi: 10.1128/jcm.41.10.4623-4629.2003.
 37. John S, Balagurunathan R. Metallo beta lactamase producing *Pseudomonas aeruginosa* and *Acinetobacter baumannii*. *Indian J Med Microbiol.* 2011;29(3):302-4. doi: 10.4103/0255-0857.83918.
 38. Kumar S, Mehra SK. Performance of modified Hodge test and combined disc test for detection of carbapenemases in clinical isolates of enterobacteriaceae. *Int J Curr Microbiol Appl Sci.* 2015;4(5):255-61.
 39. Kanchanadevi P, Sekaran SC. Importance of EDTA in the detection of metallo beta lactamase from imipenem resistant gram-negative bacilli. *Int J Curr Microbiol Appl Sci.* 2016;5(11):702-6. doi: 10.20546/ijcmas.2016.511.081.
 40. Singhal S, Mathur T, Khan S, Upadhyay DJ, Chugh S, Gaiind R, et al. Evaluation of methods for AmpC beta-lactamase in gram negative clinical isolates from tertiary care hospitals. *Indian J Med Microbiol.* 2005;23(2):120-4. doi: 10.4103/0255-0857.16053.
 41. Koshesh M, Mansouri S, Hashemizadeh Z, Kalantar-Neyestanaki D. Identification of extended-spectrum beta-lactamase genes and AmpC-beta-lactamase in clinical isolates of *Escherichia coli* recovered from patients with urinary tract infections in Kerman, Iran. *Arch Pediatr Infect Dis.* 2017;5(2):e37968. doi: 10.5812/pedinfect.37968.
 42. Mosavian M, Koraei D. Molecular detection of IMP carbapenemase-producing gram-negative bacteria isolated from clinical specimens in Ahvaz, Iran. *Jentashapir J Cell Mol Biol.* 2016;7(6):e36394. doi: 10.17795/jjhr-36394.
 43. Tawfik AF, Shibl AM, Aljohi MA, Altammami MA, Al-Agamy MH. Distribution of Ambler class A, B and D beta-lactamases among *Pseudomonas aeruginosa* isolates. *Burns.* 2012;38(6):855-60. doi: 10.1016/j.burns.2012.01.005.
 44. Zeighami H, Haghi F, Hajiahmadi F. Molecular characterization of integrons in clinical isolates of betalactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* in Iran. *J Chemother.* 2015;27(3):145-51. doi: 10.1179/1973947814y.0000000180.
 45. Mushi MF, Mshana SE, Imirzalioglu C, Bwanga F. Carbapenemase genes among multidrug resistant gram negative clinical isolates from a tertiary hospital in Mwanza, Tanzania. *Biomed Res Int.* 2014;2014:303104. doi: 10.1155/2014/303104.