Resveratrol and liver: A systematic review

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Background: Recent studies demonstrated that resveratrol has many therapeutic effects on liver disorders. Resveratrol significantly increased survival after liver transplantation, decreased fat deposition, necrosis, and apoptosis which induced by ischemia in Wistar rats. It provided liver protection against chemical, cholestatic, and alcohol injury. Resveratrol can improve glucose metabolism and lipid profile and decrease liver fibrosis and steatosis. Furthermore, it was able to alter hepatic cell fatty acid composition. According to extension of liver disease around the world and necessity of finding new threat, this review critically examines the current preclinical *in vitro* and *in vivo* studies on the preventive and therapeutic effects of resveratrol in liver disorders. **Materials and Methods:** A search in PubMed, Google Scholar, and Scopus was undertaken to identify relevant literature using search terms, including "liver," "hepatic," and "Resveratrol." Both *in vivo* and *in vitro* studies were included. No time limiting considered for this search. **Results:** A total of 76 articles were eligible for this review. In these articles, resveratrol shows antioxidative properties in different models of hepatitis resulting in reducing of hepatic fibrosis. **Conclusion:** Resveratrol could reduce hepatic steatosis through modulating the insulin resistance and lipid profile in animals. These high quality preclinical studies propose the potential therapeutic implication of resveratrol in liver disorders especially those with hepatic steatosis. Resveratrol can play a pivotal role in prevention and treatment of liver disorders by reducing hepatic fibrosis.

Key words: Antioxidant, fibrosis, liver, liver dysfunction, liver disorders, liver transplantation, nonalcoholic fatty liver, resveratrol, steatosis

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INTRODUCTION

Resveratrol (3,4',5-trihydroxy-trans-stilbene) was first isolated from the white hellebore by Takaoka in 1940 and then isolated from *Polygonum cuspidatum* (Japanese knot-weed) in 1963 by Nonomura.^[1]

Resveratrol, a phytoalexin found in over 300 edible plants, including grapes, berries, and peanuts, is produced by plants as a defense mechanism against microbial injury, fungal infection or environmental stress.^[2] It also found in red wine (0.1-15 mg/l), which proves, in part, the "French paradox" (the comparatively low incidence of cardiovascular disease in the French population despite regular consumption of a high-fat diet [HFD]), and may be responsible

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for many of the health benefits ascribed to red wine consumption.^[3]

Subsequent studies have demonstrated that resveratrol can prevent or slow down the progression of a wide variety of illnesses, including malignancies, neurodegenerative diseases, cardiovascular ailments, ischemic injury, and viral infections.^[4-7]

Resveratrol improved health and survival in obese mice,^[8] and chronic liver diseases.^[9] These findings suggested that resveratrol could therapeutically intervene with liver injury^[10] and polyphenol-rich foods may serve as an adjuvant treatment in chronic liver diseases.^[11]

The mechanisms underlying the beneficial effects of resveratrol are not totally elucidated, but have been related

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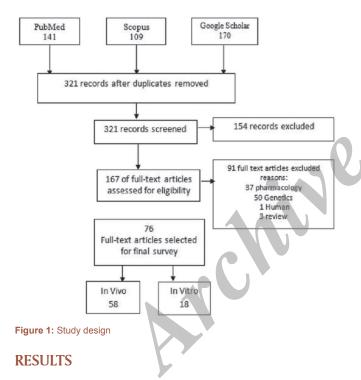
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Address for correspondence: Dr. Peyman Adibi, Department of Medicine, Integrative Functional Gastroenterology Research Center, Isfahan University of Medical Sciences, Isfahan, Iran. E-mail: payman.adibi@gmail.com Received: 01-12-2014; Revised: 27-02-2015; Accepted: 01-09-2015 mainly to its antioxidant activity that has been demonstrated to protect tissues such as liver, kidney, and brain against a variety of damage caused by oxidative stress.^[12]

According to extension of liver disease around the world and necessity of finding new threat, in this review, we studied the effects of resveratrol on liver both *in vivo* and in cell culture to help find novel therapeutic ways.

MATERIALS AND METHODS

We searched "PubMed," "Scopuse," and "Google Scholar" and used the following keywords: "Liver," "hepatic" and "resveratrol" [Figure 1]. Studies should be in English. Both *in vivo* and *in vitro* studies were included. No time limiting considered for this search. So, among 321 articles finally 76 articles were selected for this review. Study diagram is as follow:



Several *in vivo* and *in vitro* studies provide evidence for the potential role of resveratrol as a hepatoprotective agent. The protective effects of resveratrol have been elucidated in animal and in both parenchymal (hepatocyte) as well as nonparenchymal liver cells such as stellate cells and Kupffer cells.

These studies are summarized in Table 1 (*in vitro* studies) and Table 2 (*in vivo* studies).

In vitro studies

Several *in vitro* studies [Table 1] have shown the hepatoprotective effects of resveratrol in different cell cultures.

In human liver myofibroblasts trans-resveratrol markedly reduced proliferation of myofibroblasts in a dose-dependent manner and deactivated human liver myofibroblasts.[13] It also inhibited platelet-derived growth factor signaling via receptor inactivation, whereas it reduced epidermal growth factor-dependent DNA synthesis via inhibition of the phosphatidylinositol 3-kinase/Akt pathway, possibly through inactivity of phospho-inositidedependent kinase-1.[14] Resveratrol inhibited HepG2 cellular triacylglycerol (TG) accumulation (which were incubated with high concentration of glucose and insulin) via activating AMP-activated protein kinase (AMPK) and downregulating sterol regulatory element binding protein 1c and fatty acid synthase gene expressions.^[15] Nah et al.^[16] showed that resveratrol effectively protects HepG2 cells from ethanol and hydrogen peroxide oxidative stress. Reduction in intracellular glutathione (GSH) depletion, cell proliferation, mitotic Cdk, malondialdehyde (MDA), reactive oxygen species (ROS) production, apoptosis, and cell death was also reported. Resveratrol amplifies the profibrogenic activation of human hepatic LX-2 stellate cells,^[11] increases apoptosis and inhibits cell growth in hepatic stellate cells.^[17]Resveratrol pretreatment effectively protected hepatocytes from oxidative stress,^[18,19] increasing the activities of catalase (CAT), superoxide dismutase (SOD), GSH peroxidase (GPx), NADPH quinone oxidoreductase, and GSH-S-transferase.[18] Resveratrol also protected the cells against hydroquinone-induced toxicity through its antioxidant function and possibly a suppressive effect on the expression of cytochrome P450 2E1.^[20] Relatively higher concentrations of resveratrol (≥1 mM) inhibited apoptosis of the mouse primary hepatocytes and increased cell viability in a dose-dependent manner, and in particular the survival rate of the hepatocytes was recovered from 28% to near 100% by 5 mM resveratrol.^[20] In isolated hepatocytes from normal rats a resveratrol-induced short-term inhibition of fatty acids (specially formation of palmitic acid) and TG synthesis occurred.^[21] Cerny et al.^[22] have evaluated the effects of resveratrol as compared to silymarin pretreatments on tert-butylhydroperoxide (tBH) induced apoptotic/necrotic markers in hepatocytes. They found that resveratrol reduced hepatocyte necrotic and apoptotic effects caused by tBH and suggested that resveratrol has significant protective ability against tBH-induced oxidative damage. 10 µM resveratrol showed the similar effect of 500 µM silymarin. The authors concluded that resveratrol may play a chemopreventive role through antiapoptotic and antinecrotic effects via reducing nitric oxide synthase 2 (NOS-2) and hemoxygenase-1 in tBHinduced hepatocyte intoxication.[22] Effects of antioxidants, resveratrol, quercetin, and N-acetylcysteine on the functions of cultured rat hepatic stellate cells and Kupffer cells have been studied; resveratrol could have therapeutic potential against liver injury by regulating functions of hepatic stellate cells and Kupffer cells.^[10] Resveratrol also had

Cell culture	Main result	Mechanisms of action	Concentration	Author/year
			(μ Μ)	
Human liver myofibroblasts	Trans-resveratrol markedly reduced proliferation of myofibroblasts in a dose-dependent manner. Trans-resveratrol can deactivate human liver myofibroblasts	\downarrow expression of α -smooth muscle actin, \downarrow myofibroblast migration, \downarrow mRNA expression of type I collagen. It decreased the \downarrow secretion of matrix metalloproteinase 2	25/50/75/100 μmol	Godichaud <i>et al.</i> ^[13]
Human liver myofibroblasts	Resveratrol inhibits platelet-derived growth factor signaling	\downarrow receptor activation of platelet- derived growth factor signaling		Godichaud <i>et al.</i> ^[14]
Cultured human HepG2 hepatocytes	Resveratrol could protect the liver from NAFLD ^a	↓TG, ↓SREBP-1cʰ, ↓FAS°, ↑phosphorylation of AMPK	50 μmol/L	Shang et al. ^[15]
Human liver adenocarcinoma cell line (HepG2)	Resveratrol effectively protects HepG2 and change liver cells from ethanol and hydrogen peroxide oxidative stress	↓intracellular GSH depletion, ↓cell proliferation, ↓mitotic Cdk, ↓MDA, ↓apoptosis and cell death, blocked ROS generation	50 μmol	Nah <i>et al.</i> ^[16]
HSCs	Resveratrol amplifies the profibrogenic activation of human hepatic LX-2 stellate cells	î activation of human hepatic LX-2stellate cells	15 μM	Bechmann <i>et al.</i> ^[11]
HSCs	Resveratrol inhibits cell growth	↓cell growth, ↑S-phase cell cycle arrest, ↑apoptosis	100 nM or 1 µM	Souza et al. ^[17]
Primary rat hepatocytes	Resveratrol could be a useful drug for the protection of liver cells from oxidative stress induced damage	↑activities of (CAT, SOD, GSH peroxidase, NADPH quinone oxidoreductase and GSH-S-transferase)	25, 50 and 75 μM	Rubiolo et al. ^[18]
Primary rat hepatocytes	Resveratrol protects primary rat hepatocytes from oxidative stress induced cell death	↓necrosis, ↓ROS	25, 50 or 75 μM	Rubiolo and Vega ^[19]
Mouse primary hepatocytes	Resveratrol protected the cells against hydroquinone-induced toxicity	↓apoptosis, ↑cell viability, ↓hydroquinone-induced expression of cytochrome P450 2E1 mRNA	0.5 or 1 or 5 mM	Wang et al. ^{[20}
Rat hepatocyte	Resveratrol has hypolipidemic effect	↓total fatty acid, ↓formation of palmitic acid formation, ↓ACC ^d activity, ↓TGs	0-100 μΜ	Gnoni and Paglialonga ^{[21}
Rat hepatocytes	Resveratrol reduced tBHe-induced hepatocyte toxic effects	↓ALT ^f , NO ^g , ↓ NOS-2 and hemoxygenase-1 gene expression	10 µM	Cerný <i>et al.</i> ^[22]
Rat HSCs and Kupffer cells	Resveratrol may have therapeutic potential against liver injury by regulating functions of HSCs and Kupffer cells	\downarrow NO, \downarrow TNF- α^{h}	100 μmol/L	Kawada et al. ^[10]
Hepatoma G2 cells	Pretreatment with resveratrol blocked the apoptotic biochemical events	\downarrow CTN ⁱ -induced cell apoptosis and oxidative stress, \downarrow ROS, \downarrow CTN-induced activation of caspases-3,-9, and PAK2 ^j	5, 10, 20, 30, 40 μM	Chen and Chan ^[23]
Human hepatocyte- derived cancer cell line HepG2	Resveratrol inhibits the proliferation of HepG2 cells	↓cell proliferation, ↓ROS, ↑apoptosis, ↑iNOS and eNOS expression, ↑NOS activity, ↑NO production	10 ⁻⁷ M	Notas <i>et al.</i> ^[24]
HepG2 cells	Resveratrol inhibits proliferation of HepG2 cells	↓cyclin D1, ↓p38 MAP kinase, ↓Akt ^k , ↓Pak1 expression, ↑ERK ⁱ activity	200 and 225 μM	Parekh et al. ^[25]
Human SK-HEP-1 hepatic cancer cells	Resveratrol causes hepatic cancer cell death	\downarrow expression of antioxidant proteins, \downarrow ROS	225 µM	Choi <i>et al.</i> ^[26]
HepG2 HCC cells	PYB, trans-3,40-dihydroxy-20,30,5 trimethoxystilbene was more effective in inhibiting the growth of HepG2 HCC cells than resveratrol PYB could inhibit the invasion of HCC cells		50 μM	Wang <i>et al.</i> ^[3]
Liver of Wistar rats	Resveratrol plus ethanol counteract the ethanol-induced impairment of energy metabolism	Maintained ATP content, ↑mitochondrial ATP turnover, ↑sn-G3P ^m	20 _mol/L or (4.56 mg/L)	Gallis <i>et al.</i> ^{[27}

*NAFLD = Nonalcoholic fatty liver disease; *SREBP-1c = Sterol regulatory element binding proteins 1c; *FAS = Fatty acid synthase; TG = Triacylglycerol; AMPK = AMP-activated protein kinase; GSH = Glutathione; ^dACC = Acetyl-CoA carboxylase; ^etBH = Tert-butylhydroperoxide; ^fALT = Alanine aminotransferase; ^eNO = Nitric oxide; ^hTNF-a = Tumor necrosis factor alpha; CTN = Citrinin; PAK = Activated kinase; Akt = Protein kinase B; ERK = Extracellular signal-regulated kinases; msn-G3P = sn-glycerol-3-phosphate; NOS-2 = Nitric oxide synthase 2; ROS = Reactive oxygen species; iNOS = Inducible initic oxide synthase; eNOS = Endothelial nitric oxide synthase; HSCs = Hepatic stellate cells; PYB = Phoyunbene B; CAT = Catalase; SOD = Superoxide dismutase; HCC = Hepatocellular carcinoma protective effects in carcinoma HepG2 cells. Pretreatment with resveratrol in these cells blocked the apoptotic biochemical events.^[23] It also inhibited the proliferation of HepG2 cells.^[24,25] Resveratrol causes hepatic cancer cell death by suppressing the expression of antioxidant proteins and reduction of ROS.^[26] Wang *et al.* compared the effects of phoyunbene B (PYB) and resveratrol; they found that PYB, trans-3,40-dihydroxy-20,30,5 trimethoxystilbene) was more effective in inhibiting the growth of HepG2 hepatocellular carcinoma cells than resveratrol.^[3] Resveratrol can also counteract the ethanol-induced impairment of energy metabolism by maintaining ATP content and increasing mitochondrial ATP turnover.^[27]

In vivo studies [Table 2] *Liver transplantation*

To study the effect of resveratrol on transplantation Wu et al. used Sprague-Dawley rats as donors and Wistar rats as recipients then 25, 50, and 100 mg/kg resveratrol was given once a day to the recipient mice. Increase in survival period and decrease in severity of rejection was seen in a larger dose of resveratrol as compared to the others.^[28] Another study also reported an increase in survival period, decrease in the expression of protein kinase C Θ , \downarrow I κ B kinase- β protein in lymphocytes.^[29] Combination use of resveratrol (100 mg/Kg) and cyclosporine (20 mg/Kg) had better outcomes than cyclosporine alone.^[30] This combination increased survival period and albumin and decreased serum total bile acid, alanine aminotransferase (ALT), serum interleukin-2 (IL-2) and interferon-y (INF-y) levels, activation of transcription factor nuclear factor-kB (NF-kB) peripheral blood T-lymphocytes and the severity of rejection. Resveratrol alone (100 mg/Kg) was able to increase survival period and albumin, and decrease serum total bile acid, ALT, IL-2, INF-γ, activation of transcription factor NF-κB and severity of rejection.^[31]

Cancer and metastasis

Resveratrol could inhibit the growth of murine hepatoma 22, after the mice bearing H22 tumor were treated with 10 mg/kg or 15 mg/kg resveratrol for 10 days.[32] It also induced the S phase arrest of H22 cells and enhanced the antitumor effect of 5-FU on murine hepatoma 22.[32] Yu et al. ^[33] used murine transplanted hepatoma H22 model to evaluate the in vivo antitumor activity of resveratrol and reported that resveratrol exhibits anti-tumor activities on murine hepatoma H22. The underlying anti-tumor mechanism of resveratrol might involve the inhibition of the cell cycle progression by decreasing the expression of cyclin B1 and p34cdc2 protein.[33] Bishayee et al.[34] have examined the underlying mechanisms of resveratrol chemoprevention of hepatocarcinogenesis by investigating the effects of resveratrol on oxidative damage and inflammatory markers during diethylnitrosamine (DENA)- initiated rat liver carcinogenesis. In this study, resveratrol elevated the protein and mRNA expression of hepatic NF E2-related factor 2 (Nrf2). Resveratrol also reduced the hepatic iNOS protein expression and thiobarbituric acid reactive substances (TBARS) in these animals. It has been shown that resveratrol (50, 100 and 300 mg/kg) inhibits DENA-induced hepatocyte nodules in Sprague–Dawley rats in a dose-responsive manner.^[35] Resveratrol exerts chemoprevention of hepatocarcinogenesis possibly through anti-inflammatory effects during DENA-evoked rat liver carcinogenesis by suppressing elevated levels of heat shock protein (HSP)70, COX-2 as well as NF-KB.[35] DENA-initiated hepatocarcinogenesis was suppressed by resveratrol administration for 20 weeks.[36] Resveratrol treatment reversed the DENA-induced alteration of the level and expression of hepatic tumor necrosis factor-alpha (TNF- α) and IL-1 β and IL-6.^[36] Luther *et al.*^[2] also showed that long-term dietary administration of resveratrol dose-dependently suppressed hepatic tumor multiplicity.

Another study^[37] investigated anti-inflammatory properties of resveratrol and reported that 1 mg/Kg resveratrol remarkably inhibited hepatic retention and metastatic growth of melanoma cells by 50% and 75%, respectively. The mechanism involved IL-18 blockade at three levels: First, resveratrol prevented IL-18 augmentation in the blood of melanoma cell-infiltrated livers. Second, resveratrol inhibited IL-18-dependent expression of VCAM-1 by tumoractivated hepatic sinusoidal endothelium, preventing melanoma cell adhesion to the microvasculature. Third, resveratrol inhibited adhesion and proliferation-stimulating effects of IL-18 on metastatic melanoma.

Hepatic glucose metabolism

Burgess *et al.*^[38] studied the effect of resveratrol (100 mg/kg) on glucose metabolism in a Swine Model of Metabolic Syndrome induced by hypercholesterolemic diet. Increase in insulin receptor substrate 1, phosphorylated AKT, glucose transporter type 4 (Glut 4), peroxisome proliferating activation receptor- γ coactivator 1 α , peroxisome proliferatoractivated receptor- α , peroxisome proliferator-activated receptor- γ , phosphorylated AKT at threonine 308 expression and reduction in retinol binding protein 4 was seen according to resveratrol consumption.

Hepatic ischemia

Liver ischemia-reperfusion (I/R) injury occurs in many clinical conditions, including liver surgery and transplantation. Oxygen free radicals generated during I/R reduce endogenous antioxidant systems and contribute to hepatic injury. As resveratrol has antioxidant properties, different studies have conducted to examine the possible effect of resveratrol.

Subject	Author	Animal model	Drug dose	Duration	Main result
Transplantation	Wu <i>et al.</i> ^[28]	Sprague-Dawley rats as donors/Wistar rats as recipients			$\hat{1}$ survival period, $\hat{1}$ longivity
	Wu <i>et al.</i> ^[29]	Sprague-Dawley rats as donors and Wistar rats as recipients			↑survival period ↓expression of PKCΘ', ↓IKKβ° protein in lymphocytes
	Wu <i>et al.</i> ^[30]	Sprague-Dawley rats as donors and Wistar rats as recipients			↑survival period ↑albumin, ↓serum total bile acid, ↓ALTº,↓serum IL-2 and INF-γ levels, ↓activation of transcription factor NF-kBª n peripheral blood T lymphocytes, ↓ the severity of rejection
	Wu <i>et al.</i> ^[31]	Sprague-Dawley rats as donors and Wistar rats as recipients			Tsurvival period, Talbumin, Userum total bile acid, UALT, UL-2',UINF-y, Lactivation of transcription factor NF-kB, Useverity of rejection
Cancer and metastasis	Wu <i>et al.</i> ^[32]	Balb/c mice	1	10 days	Induce the S phase arrest of H22 cells and enhance the anti-tumor effect of 5-FU on murine hepatoma 22
	Yu <i>et al.</i> ^[33]	H22 tumour bearing mice		10 days	Anti-tumour activities, ↓expression of cyclinB1 and p34cdc2 protein. Blocking of S stage of tumour cells
	Bishayee et al. ^[34]	Sprague-Dawley rats		20 weeks	$ar{}$ TBARS $^{\circ}$, $ar{}$ Nrf2, $ar{}$ hepatic iNOS protein expression
	Bishayee <i>et al.</i> ^[35]	Sprague-Dawley rats	50, 100 or 300 mg/kg	24 weeks	UNF-kB, UHSP70;
	NIDIMDA <i>et al.</i>	sprague-Dawley rats Sprague-Dawley rats	50, 100 and 500 mg/kg	20 WEEKS	¢nepauc INF-α, ↓L-1p, ↓L-0 hometin turmer multiplicity
	Salado <i>et al.</i> ^[37]		1 mg/kg	12 days	↓the tastatic growth of melanoma, ↓IL-18, ↓expression of VCAM-1
Hepatic glucose metabolism	Burgess <i>et al.</i> ^[38]	Pigs/3 groups, control/ hypercholesterolemic diet (HCC)/hypercholesterolemic diet with supplemental resveratrol (HCRV)	100 mg/kg	11 weeks	Tinsulin receptor substrate 1, Tphosphorylated Akt, TGlut 4, Tperoxisome proliferating activation receptor γ coactivator 10, Tperoxisome proliferator-activated receptor α , Tperoxisome proliferator-activated receptor α , Tperoxisome 908 expression, \downarrow retinol binding protein 4
Hepatic ischemia	Gedik <i>et al.</i> ^[39]	Sprague-Dawley rats	10 µmol/L		<code><code> <code>tactivity</code> of aminotransferase. <code>UMDA, Uhepatic</code> injury score, while <code><code>TSOD, <code>TCAT, TGSH, Uhistopathological</code> changes in livers</code></code></code></code>
	Hassan-Khabbar <i>et al.</i> ^[40] Plin <i>et al.</i> ^[41]	Sprague-Dawley rats Wistar rats	0.02 or 20 mg/kg		↓ALT, ↓AST, ↑Cu/Zn-SOD and CAT Activities, ↑GSH, ↑GSH reductase, ↑Cu/Zn-SOD, and ↑ CAT activities ↓lipid peroxidation, protects mitochondrial functions, ↑respiratory chain activity, ↓activation of the cellular markers of necrosis and apoptosis
Hepatic Chemical injury	Rivera et al. ^[42]	Wistar rats	10 mg/kg		JGSH, JGSH/GSSG ratio were decreased by CCI,; both effects were completely prevented by any of the compounds tested Jlipid peroxidation, Jactivity of Y-glutam/l transpeptidase, JALT
	Fan <i>et al.</i> ^[43]	Swiss albino mice	30 mg/kg	Single dose	<code>JAST, <code>JALT, <code>JSALP, LDH, <code>Jy-GT</code> and <code>JCK, <code>Jtotal</code> bilirubin, <code>Jurea</code> and <code>Juric</code> acid, <code>Jhepatic</code> peroxidative stress, <code>TGSH, JGSSG</code></code></code></code></code>
	Lodovici et al.[44]	Fisher 344 rats	50 mg/kg	14 days	↓sod, ↓xov, ↑gsh/gssg
	Tunali-Akbay <i>et al.</i> [^{45]}	Wistar albino rats	20 mg/kg	5 days	↓hepatic MDA, ↓ALT, ↓AST, ↓TNF-α, ↓LDH, ↑liver GSH
	Roy <i>et al.</i> ^[46]	Wistar rats	100 or 200 mg/kg	8 weeks	\downarrow TNF- $lpha$, \downarrow IL-6, \downarrow liver MDA, \downarrow hepatocyte nodules, \downarrow apoptosis
SID.	Vitaglione <i>et al.</i> ^[47]	Sprague-Dawley rats	3 mg/kg	14 days	↓lipid peroxidation (↓MDA)

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Subject	Author	Animal model	Drug dose	Duration	Main result
Hepatic toxicity	EI-Agamy ^[48]	Fischer rats	10 mg/kg BW	90 days	No effect on aflatoxin B1-induced liver injury
	Hamadi <i>et al.</i> ^[49]	Sprague-Dawley rats	10 mg/kg	15 days	↓blood glucose, ↓mortality, ↓AST, ↓ALT, ↓total bilirubin, ↓TC ^w , ↓TG, ↓LDL×, ↓TC/HDL ^v , ↑SOD, ↑CAT, ↑GST ² , ↑QR*
Hepatic cholestatic injury	Chan <i>et al.</i> ^[50]	C57BL/6 mice	4 mg/kg	3 days or 7 days	↓ALT, ↓AST, ↓TNF-a, ↓IL-6 mRNA, ↓number of Kupffer cells (CD681) in the injured liver, ↓hepatic fibrosis, ↓reduced collagen la1, ↓TIMP-1 mRNA ↓inflammation, ↓activation of Kupffer cells, ↓fibrosis, ↑hepatocyte regeneration
Irradiation-induced hepatic injury	Velioglu-Ogünç et al. ^[51]	Sprague-Dawley rats	10 mg/kg	20 days	↑GSH level, ↓MDA, ↓myeloperoxidase activity, ↓collagen content, ↓plasma LDH, ↓pro-inflammatory cytokine levels, ↓leukocyte apoptosis
Hepatic injury after trauma-hemorrhage	Yu <i>et al.</i> ^[52]	Sprague-Dawley rats	30 mg/kg	Single dose	↓myeloperoxidase activity, ↓ICAM-1, ↓IL-6, ↓ALT, ↓AST
Heoatic alcohol injury	Bujanda <i>et al.</i> ^[53]	Balb/c mice/four groups (control, resveratrol- treated control, alcohol and resveratrol-treated alcohol)	10 mg/ml in drinking water	6 weeks	↓ALT, ↓AST, ↓IL-1, ↓severity of liver lesions, ↓mortality
	Kasdallah-Grissa <i>et al.</i> [^{54]}	Wistar rats	5 g/kg food	6 weeks	$\downarrow hepatic lipid peroxidation, \uparrow SOD^{**}, \uparrow GPx and \uparrow CAT activities in the liver$
	Jinghui and Yingbao ^[55]		25, 50 or 100 mg/kg	7 days	\downarrow TNF- α , \downarrow IL-6, \downarrow ICAM-1, \downarrow Iiver NFkB p65, \downarrow phosphorylating IkB- α
Hepatic oxidative damage	Schirmer <i>et al.</i> ^[56]	Zebrafish	5 or 50 mg/l	30 or 60 min	$\downarrow expression$ levels of the SIRT3, SIRT4, and NAMPT genes. $\uparrow NADH$ levels, $\downarrow NAD^{+}/H$
)	Schmatz <i>et al.</i> ^[12]	Streptozotocin-induced	10 or 20 mg/kg	30 days	↓TBARS, ↑activities of CAT, SOD and d-ALA-D and the levels of NPSH
		UIADELIC I ALS			anu vuannin C ↓ALT, ↓AST and ↓g-glutamiltransferase (g-GT) activities ,↓urea, ↓creatinine, ↓cholesterol, ↓triglycerides, ↓lipid peroxidation
	Chang <i>et al.</i> ^[57]	Long-Evans rats	0.1 or 1 mg/kg	7 days	\downarrow superoxide anion, PCL and Mn-SOD protein expression in the hepatic tissues, \downarrow activated form of NF+kB, \downarrow hepatic IL-1 β
	Sebai <i>et al.</i> ^[58]	Wistar rats	20mg/kg	7 days	$\mbox{LPS-induced lipoperoxidation, \mbox{TSOD}, \mbox{TCAT}, \mbox{Tglutathione} peroxidase (GPx) activity, \mbox{UNO}, Lendotoxemia-induced hepatic tissue injury, UNO, there iron$
	Rocha <i>et al.</i> ^[59]	Wistar rats	1 mg/kg	30 days	Jelucose level, ↑SOD, ↓ox-LDL, ↓hepatic oxidative stress. In standard- fed-rats reduced ↓GSH-antioxidant defense system and ↑hepatic lipid hydroperoxide
	Bagul <i>et al.</i> ^[60]	Sprague-Dawley rats	10 mg/kg	8 weeks	<pre>↓blood glucose, ↓triglyceride, ↓uric acid, ↓NO, ↓TBARS</pre>
	Palsamy <i>et al.</i> ^[61]	Wistar rats	30 days	5 mg/kg b.w.	JTNF-ox, JIL-1B, JIL-6, JNF-kB, Jliver NO, Jlipid peroxides, Uhydroperoxides. Uprotein carbonyls, Tactivity of (SOD, CAT, GPx, GST and GR)
	Upadhyay <i>et al.</i> ^[62] Farghali <i>et al.</i> ^[63]	Swiss albino mice Wistar rats	10 mg/kg 2.3 mg/kg i.p (10 Imol/ bai	1-4 weeks Single dose	<code>LALT, LAST, Ulipid peroxidation</code> <code>LALT, LAST, Ulipid peroxidation</code> <code>LALT, LAST, $\downarrow \alpha$-GST, <code>TCAT, JTBARS, \downarrowconjugated dienes</code></code>
	Kirimlioglu et al. ^[64]	Wistar albino rats	30 mg/kg	7 days	\downarrow PH-induced lipid peroxidation, \downarrow hepatic GSH, \downarrow NO
v.SIL	Sengottuvelan and Nalini ^[65]	Wistar rats	8 mg/kg	30 weeks	↓activity of SOD, CAT, Gpx, GST and GR, ↓GSH, Vitamin C, Vitamin E levels
					(Contined)

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 ^[79] Sprague-Dawley rats 10 mg/kg Z days ^{0]} Sprague-Dawley rats 20 mg/kg 4 weeks Wistar rats 25,50 or 100 mg/kg 6 weeks 	Hong et al.P91 Sprague-Dawley rats 10 mg/kg 7 days Unfiltration of inflammatory cells and fibrosis of liver tissue. JM Pipe 1, and pipe 1, and appression of inflammatory cells and fibrosis of liver tissue. JM Pipe 1, and appression of inflammatory cells and interleukin-blet Lee et al.P01 Sprague-Dawley rats 20 mg/kg 4 weeks Jaranine transaminase, Jaspartate transaminase, Jakanine Lee et al.P01 Wistar rats 25,50 or 100 mg/kg 6 weeks Jaranine transaminase, Jalanine Lv et al.P01 Wistar rats 25,50 or 100 mg/kg 6 weeks Jaranine transaminase, Jalanine PROFO = Protein kinase CO; %IKG = IkB kinase-β; PALT = Alanine arminotransferase; %IF-kB = nuclear factor-kBi mg oveh factor-βi mRNA expression Jivet hydroxyproline, Lipe to collagen mRNA expression HSP70 = Heat shock protein 70; "COX-2 eCycloxoxgenase-2; YO = Xuathine oxidase; "TC = Total cholesterol; "IL2 = Interleukin-2; TBARS = Thiobarbituric acid reactive substant HSP70 = Heat shock protein ; ikt = Protein kinase B; Glut 4 = Glucose transforming growth factor-βi mRNA expression HSP70 = Heat shock protein 70; "COX-2 eCycloxoxgenase-2; YO = Suproxide dismutase; "TC = Interleukin-2; TL2 = Interleukin-2; TBARS = Thiobarbituric acid reactive substant HSP70 = Heat shock protein ; ikt = Protein kinase B; Glut 4 = Glucose transformase; WIDA = Miondialderydes; IL-1 = Interleukin-1; IL-1 = Interleukin-1; ICAM = High density lipoprotein; 'GSG = Glutathione; GSG = Glutathione; GLucose transfore Ase, SIR1 = Struins; 4ALP = Ention facid vight ecol; C		Liver Fibrosis	Di Pascoli <i>et al.</i> ^{[78}		10 and 20 mg/kg	2 weeks	↓portal pressure, improved vasodilatory response to acetylcholine, ↓TXA2 production, ↑eNO, ↓liver fibrosis, ↓collagen-1, TGF-β, ↓NF-κB mRNA expression, ↓desmin and α-SMA protein expression	
^{ol} Sprague-Dawley rats 20 mg/kg 4 weeks Wistar rats 25,50 or 100 mg/kg 6 weeks	Lee <i>et al.</i> ^[80] Sprague-Dawley rats 20 mg/kg 4 weeks Jalanine transaminase, Jalkaline Phosphatase, Jbilitrubin, Talbumin, Thepatic glutathione, JHSC activ JHydroxyproline, Jtype I collagen mRNA expression Lv <i>et al.</i> ^[81] Wistar rats 25,50 or 100 mg/kg 6 weeks JALT, Jliver hydroxyproline, JmDA, Jliver fibrogensis "PKCØ = Protein kinase CØ; °IKKβ = IkB kinase-β; °ALT = Alanine aminotransferase; %IF-kB = nuclear factor-kB; °IL2 = Interleukin-2; °TBARS = Thiobarbituric acid reactive substant "PKCØ = Protein kinase CØ; °IKKβ = IkB kinase-β; °ALT = Alanine aminotransferase; %IF-kB = nuclear factor-kB; °IL2 = Interleukin-2; °TBARS = Thiobarbituric acid reactive substant "PKCØ = Protein kinase CØ; °IKKβ = IkB kinase-β; °ALT = Alanine aminotransferase; %IF-kB = nuclear factor-kB; °IL2 = Interleukin-2; °TBARS = Thiobarbituric acid reactive substant "HSP70 = Heat shock protein 70; "COX-2 = Cyclooxygenase-2; "YO = Xanthine oxidase; wTC = Total cholesterol; *LDL = Low density lipoprotein; "GS Super oxide dismutase; VAH = Protein kinase B; Glut 4 = Glucose transporter type 4; MDA = Malondialehyde; IL-1 = Interleukin-1; IL-1β = Interleukin-2; SALP = Serum alkaline phosphatase; IDH = Lactate deitydrase; NPSH = Norprotein thiols; NO = Nitris oxide synthase; TN+α = Tumor necrois SALP = Serum alkaline phosphatase; IDH = Lactate dismutase. NIH-1 = Interleukin-2; IL-1 = Interleukin-1; IL-1β = Interleukin-2; IL-1 = Interleukin-2; IL-2 = Interleukin-2; IL-1 = Interleukin-1; IL-1β = Interleukin-2; IL-1 = Interleukin-2; IL-1 = Interleukin-2; IL-2 = Int			Hong <i>et al.</i> ^[79]	Sprague-Dawley rats	10 mg/kg	7 days	Unfiltration of inflammatory cells and fibrosis of liver tissue. \downarrow MDA, \uparrow glutathione peroxidase, \uparrow SOD. \downarrow mRNA expression of inflammatory mediators (iNO, tumor necrosis factor-alpha and interleukin-1beta) \downarrow mRNA expression of fibrosis-related genes such as TGF- β 1, collagen type 1, and alpha-smooth muscle actin, \downarrow hydroxyproline	
Wistar rats 25,50 or 100 mg/kg 6 weeks	Lv <i>et al.</i> ^[81] Wistar rats 25,50 or 100 mg/kg 6 weeks JALT, Jliver hydroxyproline, JMDA, Jliver fibrogensis P KC Θ = Protein kinase C Θ ; O (K β = IkB kinase- β ; P LT = Alanine aminotransferase; N F-kB = nuclear factor-kB; I L2 = Interleukin-2; T PBARS = Thiobarbituric acid reactive substant H KC Θ = Protein kinase C Θ ; O (K β = IkB kinase- β ; P LT = Alanine aminotransferase; N TC = Total cholesterol; I LL = Low density lipoprotein; V HDL = High density lipoprotein; 2 GS Super oxide dismutase; * QR = Quinone reductase; ** SOD = Superoxide dismutase. Nrf2 = NF E2-related factor 2; iNOS = Inducible nitric oxide synthase; TINF- α = Tumor necrosis alpha; VCAM-1 = Vascular cell adhesion molecule 1; Akt = Protein kinase B; Glut 4 = Glucose transporter type 4; MDA = Malondialdehyde; IL-1 = Interleukin-1; IL-1 β = Interleukin-1 SALP = Serum alkaline phosphatase; LDH = Lactate dehydrogenase; NF-4 = Interferon- * ; AST = Aspartate aminotransferase; GSH = Glutathione; GSG = Glutathione disulfide; CO ₁ , tetrachoride; CK = Creatine kinase; TG = Triacylglycerol; CAT = Catalase; TIMP-1 = Tissue inhibitor of metalloproteinases-1; ICAM-1 = Intercellular adhesion molecule 1; IkB- α = Inh kappa B; SIRTs = Sirtuins; d-ALA-D = Aminolevulinic acid dehydratase; NPSH = Nonprotein thiols; NO = Nitric oxide; ox-LDL = Oxidized low-density lipoprotein; GR = Glutathione rep PH = Partial hepatectomy; FAS = Fatty acid synthase; LDL = Low density lipoprotein receptor; SR-BI = Scavenger receptor class B type I; CPI/ α = Camilite palmitoyl transferase-I Acyl-coenzyme A oxidase; ACC = Acetyl-CoA carboxylase; AMPK = AMP-activated protein kinase; PGC-1 α = Peroxisome proliferator-activated receptor- α ; SREBP-1 = S			Lee <i>et al.</i> ⁽⁸⁰⁾	Sprague-Dawley rats	20 mg/kg	4 weeks	↓ Latanine transaminase, ↓aspartate transaminase, ↓alkaline phosphatase, ↓bilirubin, ↑albumin, ↑hepatic glutathione, ↓hepatic MDA, ↓hydroxyproline, ↓type \ collagen mRNA expression, ↓HSC activation, ↓transforming growth factor-β1 mRNA expression	
	[¬] PKCΘ = Protein kinase CΘ; ^o IKKβ = IkB kinase-β; ^p ALT = Alanine aminotransferase; ^o NF-kB = nuclear factor-kB; ^r 1L-2 = Interleukin-2; ^s TBARS = Thiobarbituric acid reactive substant ⁺ HSP70 = Heat shock protein 70; ^u COX-2 = Cyclooxygenase-2; ^v XO = Xanthine oxidase, ^w TC = Total cholesterol; ^v 1DL = Low density lipoprotein; ^v HDL = High density lipoprotein; ^z GS ² Super oxide dismutase, [*] OR = Quinone reductase; ^{**} SOD = Superoxide dismutase. Nrf2 = NF E2-related factor 2; iNOS = Inducible nitric oxide synthase; TNF-α = Tumor necrosis ³ alpha; VCAM-1 = Vascular cell adhesion molecule 1; Akt = Protein kinase B; Glut 4 = Glucose transporter type 4; MDA = Malondialdehyde; IL-1 = Interleukin-1; IL-1β = Interleukin-1 SALP = Serum alkaline phosphatase; LDH = Lactate dehydrogenase; INF-y = Interferon-y; AST = Aspartate aminotransferase; GSH = Glutathione; GSSG = Glutathione disulfide; CCl ₄ tetrachoride; CK = Creatine kinase; TG = Triacylglycerol; CAT = Catalase; TIMP-1 = Tissue inhibitor of metalloproteinases-1; ICAM-1 = Interleukin-1; IL-1β = Interleukin-1 kappa B; SIRTs = Sirtuins; d-ALA-D = Aminolevulinic acid dehydrogenase; NPSH = Nonprotein thiols; NO = Nitric oxide; ox-LDL = Oxidized low-density lipoprotein; GR = Glutathione rei PH = Partial hepatectomy; FAS = Fatty acid synthase; LDLr = Low density lipoprotein receptor; SR-BI = Scavenger receptor class B type I; CPT-Iα = Camitine palmitoyI transferase-I Acyl-coenzyme A oxidase; ACC = Acetyl-CoA carboxylase; AMPK = AMP-activated protein kinase; PGC-Iα = Peroxisome proliferator-activated receptor-γ coactivator α; SREBP-I = S			Lv <i>et al.</i> ^[81]	Wistar rats	25,50 or 100 mg/kg	6 weeks	JALT, Uliver hydroxyproline, JMDA, Uliver fibrogensis	

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www.SID.ir | August 2015 | Gedik *et al.*^[39] investigated the effects of resveratrol (10 µmol/L) on the liver I/R injury in rats. Rats underwent liver ischemia for 45 min followed by reperfusion for 45 min. They reported that the activity of aminotransferase increased by resveratrol supplementation, while significant decrease in MDA, hepatic injury score and histopathological changes in livers and elevation in SOD, CAT, GSH resveratrol were occured.

Another study by Hassan-Khabbar *et al.*^[40] showed decrease in aminotransferase levels by about 40% and improvement of sinusoidal dilatation. In this study, trans-resveratrol preserved antioxidant defense by preventing total and reduced GSH depletion caused by I/R. At 0.2 mg/kg, trans-resveratrol significantly increased GSH reductase, Cu/Zn–SOD, and CAT activities. However, at a high dose (20 mg/kg), trans-resveratrol became prooxidant with an aggravation of liver injury evaluated by aminotransferase release and histological analysis and associated with a depletion of total and reduced GSH levels and a decrease of antioxidant enzyme activities.

Plin *et al.*^[41] investigated the effect of the natural phytoalexin resveratrol on the prevention of liver injuries induced by 40-h cold preservation followed by a warm reperfusion. They concluded that resveratrol inhibits lipid peroxidation and protects mitochondrial functions. It improves respiratory chain activity and prevents opening of the permeability transition pore, allowing better recovery of ATP energetic charge. Resveratrol also limits the activation of the cellular markers of necrosis and apoptosis.

Postischemic treatment with resveratrol (0.02 and 0.2 mg/kg) resulted in a significant decrease in aminotransferase, IL-1b and IL-6 plasma levels by about 40%, 60%, and 40%, respectively, compared to the vehicle I/R group.^[82]

Hepatic chemical injury

Rivera *et al.*^[42] examined the effect of resveratrol and trimethylated resveratrol on hepatotoxicity induced by carbon tetrachloride (CCl₄). The GSH/GSSG (oxidized GSH) ratio decreased in the groups receiving CCl₄ and resveratrol associated with an increase in GSSG. In blood GSH and the GSH/GSSG ratio were decreased by CCl₄; both effects were completely prevented by any of the compounds tested. Lipid peroxidation and the activity of γ -glutamyl transpeptidase were increased significantly after CCl₄. Resveratrol partially prevented these increases and surprisingly, trimethylated resveratrol completely prevented the increase of these markers. Both compounds partially but significantly prevented the increase in the activity of alanine aminotransferase.

In another study, resveratrol was able to mitigate hepatic damage induced by acute intoxication of CCl_4 and showed pronounced curative effect against lipid peroxidation and deviated serum enzymatic variables as well as maintained GSH status toward control.^[43]

In a model of chemically induced acute hepatic injury, resveratrol at dose of 50 mg/kg/day administered for 14 days long could ameliorate hepatic injury by its antioxidant and scavenging properties, through a reduction of xanthine oxidase activity, a partial restoration of GSH/GSSG ratio in addition to its capacity to inhibit apoptosis.^[44]

To investigate the possible protective effect of resveratrol on some liver and blood parameters in methotrexate (MTX) induced toxicity in rats, Tunali-Akbay *et al.*^[45] administered resveratrol (10 mg/kg, orally) for 5 days after a single dose of MTX. The results showed that MTX administration increased the hepatic enzymes, TNF- α , MDA, myeloperoxidase and tissue factor activities and collagen contents and decreased GSH, while all of these alterations were reversed by resveratrol.

Roy *et al.*^[46] also examined the chemopreventive potential of resveratrol in rat hepatic injury model by CCl_4 . Resveratrol (100 mg/kg, or 200 mg/kg body weight) was given orally for 8 weeks. They reported that resveratrol decreased the immunopositivity of TNF- α and IL-6 and restored the altered architectural structure of challenged hepatic tissue. Resveratrol also protected hepatic cells by suppressing oxidative stress and apoptosis.

Vitaglione *et al.*^[47] investigated the effect of resveratrol on an animal model of CCl_4 -induced liver lipid peroxidation. Grape-stalk extract determining a daily resveratrol dosage of 3 mg/kg was given to the rats for 14 days. Resveratrol could reduce liver concentration of MDA after 24 h and 1-week by 38% and a 63%, respectively.

Hepatic toxicity

Aflatoxin B1 is a potent hepatotoxic and hepatocarcinogenic mycotoxin. Lipid peroxidation and oxidative DNA damage are the principal manifestations of afatoxin B1-induced toxicity that could be counteracted by antioxidants. El-Agamy^[48] compared the effect of curcumin (polyphenolic antioxidant purifed from turmeric) and resveratrol for possible protection against liver injury induced by afatoxin B1 in rats. Results showed that curcumin showed a significant hepatoprotective activity by lowering the levels of serum marker enzymes, lipid peroxidation and elevating the levels of GSH, SOD, CAT, and GSH-Px. However, resveratrol failed to protect from the afatoxin B1-induced liver injury. Resveratrol (10 mg/kg) for 15 days in streptozotocin (STZ)-induced diabetic rats has shown hepatoprotective effects against diabetes-induced liver damage via reduction of serum glucose level and oxidative damage and improving serum lipid profile.^[49]

Hepatic cholestatic injury

Liver injuries can trigger a cascade of inflammatory responses and as a result, initiate the process of hepatic regeneration and fibrogenesis. To study the potential protective effects and mechanism of resveratrol on cholestatic liver injury, resveratrol was given (4 mg/kg/day) for either 3 days or 7 days after bile duct ligation (BDL) injury. Resveratrol significantly reduced serum ALT, aspartate aminotransferase (AST) but not bilirubin on day 3. At this early stage of injury, resveratrol significantly reduced TNF- α and IL-6 mRNA and decreased the number of Kupffer cells (CD681) recruited in the injured liver. It also decreased hepatic fibrosis. Totally resveratrol attenuated inflammation and reduced Kupffer cells activation leading to decrease in fibrosis and promotion of hepatocyte regeneration, which increased the survival of BDL mice.[50]

Irradiation-induced hepatic injury

Velioğlu-Öğünç *et al.*^[51] studied the effects of resveratrol on ionizing radiation-induced oxidative injury. After 10 days pretreatment with resveratrol (10 mg/kg), rats were exposed to whole-body IR (800 cGy) and the resveratrol treatment was continued for 10 more days after the irradiation. Resveratrol caused a significant increase in GSH level, while MDA levels, myeloperoxidase activity and collagen content decreased in hepatic tissues. Similarly, plasma lactate dehydrogenase and pro-inflammatory cytokine levels, 8-hydroxy-2'-deoxyguanosine and leukocyte apoptosis decreased, while antioxidant-capacity increased in the irradiated rats with resvratrol as compared with the control group. Resveratrol treatment reversed histopathological alterations induced by irradiation.^[51]

Hepatic injury after trauma-hemorrhage

Resveratrol administration after trauma-hemorrhage attenuated hepatic injury, likely through reduction of proinflammatory mediators. Resveratrol-mediated hepatic preservation seemed to progress via an estrogen receptor (ER) related pathway.^[52]

ALCOHOLIC LIVER INJURY

Bujanda *et al.*^[53] investigated the effect of resveratrol on alcohol-induced mortality and liver lesions in mice. They compared the effect of consuming alcohol, alcohol and resveratrol, and resveratrol alone. Transaminase concentration was significantly higher in the alcohol group than in the other groups. IL-1 levels were significantly reduced in the alcohol plus resveratrol group compared with the alcohol group. Histologically, the liver lesions were more severe in the alcohol group, though no significant differences between groups were observed. Mortality in the alcohol group was 78% in the 7th week, versus 22% in the alcohol plus resveratrol group. The results of this article suggest that resveratrol reduces mortality and liver damage in mice consuming alcohol. Kasdallah-Grissa et al.[54] investigated whether resveratrol has a preventive effect on the main indicators of hepatic oxidative status as an expression of the cellular damage caused by free radicals, and on antioxidant defense mechanism during chronic ethanol treatment. They prescribed 5 g/kg resveratrol to Wistar rats for 6 weeks and observed that dietary supplementation with resveratrol during ethanol treatment inhibited hepatic lipid peroxidation and ameliorated SOD, GPx and CAT activities in the liver so that resveratrol could have a beneficial effect on inhibiting the oxidative damage induced by chronic ethanol administration. Jinghui et al.[55] investigated the effect of resveratrol (25, 50, or 100 mg/kg) on alcohol-induced hepatic injury and reported significant reduction in the concentrations of serum TNF- α , IL-6, ICAM-1 and the contents of liver NF κ B p65, phosphorylating IkB- α compared with the model group consumed resveratrol for 7 days.

Hepatic oxidative damage

The effects of resveratrol on oxidative damage was examine by analyzing it's effects on sirtuins (SIRTs). SIRTs are NAD+dependent deacetylases that catalyze the hydrolysis of acetyl-lysine residues. They play an important role in many physiological and pathophysiological processes, such as the regulation of lifespan and the prevention of metabolic diseases. Schirmer et al.[56] in their study analyzed the effect of resveratrol on the gene expression levels of SIRT1, SIRT3, SIRT4, PGC1a, and NAMPT, as well as its effect on NAD+ and NADH levels, in the liver of nonstressed or nonimpaired wild-type zebra fish. They concluded that resveratrol plays a modulatory role in the transcription of the NAMPT, SIRT3, and SIRT4 genes. Schmatz et al.[12] investigated the effects of resveratrol on oxidative stress parameters in liver and serum biochemical parameters of STZ-induced diabetic rats. Animals were treated with 10 or 20 mg/kg resveratrol for 30 days. They reported the elevation in serum ALT, AST and γ -glutamiltransferase (GGT) activities as well as in levels of urea, creatinine, cholesterol, and TGs observed in the nonresveratrol group were reverted to levels close to normal by the administration of resveratrol. Reduction in TBARS and elevation in activities of CAT, SOD and aminolevulinic acid dehydratase, nonprotein thiols, and Vitamin C were also seen. Another study investigated the effect of resveratrol (0.1 or 1 mg/kg) on oxidative stress and inflammatory response in the liver and spleen of STZinduced type 1 diabetic animal models.[57] The experimental results indicated that resveratrol significantly decreased oxidative stress (superoxide anion content, protein

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carbonyl level and Mn-SOD expression) in both tissues and hepatic inflammation (NF- κ B and IL-1 β), but implicated proinflammatory potential of resveratrol in diabetic spleen (TNF- α and IL-6). Lipopolysaccharide (LPS) is a glycolipid component of the cell wall of Gram-negative bacteria inducing deleterious effects on several organs including the liver. Sebai et al.^[58] investigated the effect of pre-treatment with resveratrol on LPS-induced hepatotoxicity in rat. Resveratrol counteracted LPS-induced lipoperoxidation and depletion of antioxidant enzyme activities as SOD and CAT but slightly GPx activity. It also abrogated LPS-induced liver and plasma nitric oxide (NO) elevation and attenuated endotoxemia-induced hepatic tissue injury. Importantly resveratrol treatment abolished LPS-induced iron sequestration from plasma to liver compartment. In another study, 1 mg/kg/day resveratrol accompanied by standard diet and HFD in Wistar rats for 30 days;^[59] in HFD, resveratrol improved lipid profile and glucose level, enhanced SOD, and reduced oxidized low-density lipoprotein (ox-LDL) and hepatic oxidative stress. While in standard-fed-rats it reduced GSH-antioxidant defense system and enhanced hepatic lipid hydroperoxide. It may be concluded that resveratrol may have beneficial effects in high-fat diets (e.g., ox-LDL, decreased serum and hepatic oxidative stress), but not in standard-fed diets (effects produced include enhanced hepatic oxidative stress). Bagul et al.^[60] compared the effect of metformin (300 mg/kg) and resveratrol (10 mg/kg) on hepatic oxidative stress in fructose-fed rats. Administration of metformin or resveratrol normalized all the altered metabolic parameters. However, a marked insulin sensitizing action was only observed in the resveratrol group. Similarly, metformin administration failed to normalize the increased TBARS levels and decreased SOD activity, while resveratrol showed a more promising effect of all oxidative stress parameters measured in this study. This study demonstrates that resveratrol is more effective than metformin in improving insulin sensitivity, and attenuating metabolic syndrome and hepatic oxidative stress in fructose-fed rats. To investigate the hepatoprotective nature of resveratrol in averting hyperglycemia-mediated oxidative stress in the hepatic tissues of STZ-nicotinamide-induced diabetic rats, 5 mg/kg resveratrol was administered for 30 days.^[61] In this study, resveratrol could decline hepatic proinflammatory cytokines and hepatic lipid peroxides, hydroperoxides and protein carbonyls. In addition, diminished activities of hepatic enzymic antioxidants as well as the decreased levels of hepatic nonenzymic antioxidants of diabetic rats were reverted to near normal by resveratrol administration. Moreover, the histological and ultrastructural observations evidenced that resveratrol effectively rescues the hepatocytes from hyperglycemiamediated oxidative damage without affecting its cellular function and structural integrity. Pyrogallol, an

anti-psoriatic agent, causes hepatotoxicity in experimental animals. Resveratrol has hepatoprotective properties against pyrogallol.^[62] In an animal study, administration of 10 mg/kg resveratrol modulated pyrogallol-induced changes in hepatic toxicity markers, xenobiotic metabolizing enzymes and oxidative stress. Reduction in ALT, AST, and lipid peroxidation were also seen. Farghali et al.[63] investigated the effects of resveratrol pretreatment on the enhancing action of D-galactosamine (D-GalN; 800 mg/kg) on (LPS; 0.5 l g/kg) inducing liver failure in rats. Reduction in ALT, AST, α -GST, TBARS, conjugated dienes and increase in CAT was seen according to resveratrol consumption. Kirimlioglu et al.[83] compared the antioxidant effects of resveratrol (30 mg/kg) and melatonin (10/kg) after 70% partial hepatectomy (PH). Resveratrol and melatonin suppressed PH-induced oxidative damage, attenuated proliferation, and stimulated apoptosis. When resveratrol and melatonin were compared, resveratrol showed more potent antioxidative effects and was morphologically more protective to hepatocytes. Antiproliferative effects of melatonin were more potent.^[64,83] 1,2-dimethylhydrazine induced oxidative stress was suppressed by administration of resveratrol (20 mg/kg) for 30 weeks.^[65] Resveratrol supplementation attenuated the liver lipid peroxidation with concomitant up-regulation of enzymic and nonenzymic antioxidant reserves.

Hepatic heat stress

The effects of dietary resveratrol on the induction of Hsp, transcription factors and antioxidative enzyme system in liver of quails under heat stress were investigated by Sahin *et al.*^[66] They reported a linear increase in food intake, egg production, hepatic SOD, CAT, and GSH-Px activities as well as Nrf2 expression, but linear decrease in hepatic MDA concentrations and Hsp 70, Hsp90, and NF- κ B expressions with increasing supplemental resveratrol level.

Hepatic steatosis

Xin et al. concluded that Alleviative effects of resveratrol on nonalcoholic fatty liver disease (NAFLD) are associated with up regulation of hepatic LDL receptor and scavenger receptor class B type I gene expressions.^[67] Unexpectedly, increase in hepatic FASgene expressions was seen.^[67] The effect of long-term administration (4 months) of resveratrol with high fructose (HF) diet (63%) on selected biochemical parameters and lipids content in different tissues of rats was evaluated by Kopec et al.[68] The concentration of lipids was significantly lower in the liver of animals fed with HFD by addition of 0.04% or 0.06% resveratrol. The concentration of TBARS (MDA equivalents) was significantly lower (P < 0.05) in serum of rats fed HFD with the addition of 0.04% and 0.06% of resveratrol as compared to the rodents fed with negative fructose control diet and HFD.^[68] Poulsen et al.^[69] showed that an increased number of mitochondria and, particularly, an increase in hepatic uncoupling protein 2 expression might be involved in normalizing the hepatic fat content due to resveratrol supplementation in rodents fed a high-fat diet. To analyze the influence of resveratrol on hepatic TG metabolism, 30 mg/kg resveratrol was given to Sprague-Dawley rats for 6 weeks; resveratrol decreased liver fat accumulation, increased carnitine palmitoyl transferase-Ia and acyl-coenzyme A oxydase, and decreased ACC activities.^[70] Male C57BL/6J mice were fed a normal diet or a HFD (20% fat, w/w) combined with 0.005 or 0.02 % (w/w) resveratrol for 10 weeks.^[71] Resveratrol significantly reduced TAG and cholesterol, as well as lipid droplet number and size. A low dose of resveratrol (0.005%) appeared to be more effective than a higher dose of resveratrol (0.02 %) for suppressing adiposity and hepatic steatosis development with a significant decrease in body weight gain, plasma TAG and total cholesterol levels. These changes were seemingly attributable to a suppression of the fatty acid (FA) synthase, glucose-6-phosphate dehydrogenase, and phosphatidate phosphohydrolase and/ or an activation of FA oxidation in the liver and epididymal adipose tissue.^[71] Bujanda et al.^[72] investigated whether resveratrol decreases hepatic steatosis in an animal model of steatosis and found that Fat deposition was decreased in the resveratrol group as compared to the control group. TNF- α and MDA levels were significantly more increased in the control group. This was accompanied by increased SOD, GPx and CAT and decreased NOS in the liver of resveratrol group significantly. Glucose levels were also decreased in the group of rats given resveratrol. Resveratrol treatment led to reduced lipid synthesis and increased rates of fatty acid oxidation and prevented alcoholic liver steatosis. The protective action of resveratrol is in whole or in part mediated through the upregulation of a SIRT1-AMPK signaling system in the liver of ethanol-fed mice.[73] Rats fed a high-fat diet developed abdominal obesity, NAFLD, and insulin resistance (IR), which were markedly improved by 10 weeks of resveratrol administration. Resveratrol treatment prevented the development of abdominal obesity and IR. Chronic resveratrol administration stimulated AMPK phosphorylation and downregulated SREBP-1c and FAS gene expressions in HFD rats.^[15] Resveratrol (0.04%) for 20 weeks in senescence-accelerated mouse P10 hepatocytes caused the following results;^[74] lipid droplets were reduced and mitochondria were increased in number in hepatocytes. Phosphorylation of acetyl-CoA carboxylase and the expression of both the mitochondrial ATP synthase β subunit and Mn SOD2 were increased. Mitochondria, expressing more SOD2, were more tightly associated with lipid droplets, suggesting the enhancement of lipolysis through the activation of mitochondrial functions. Cathepsin D expression was less in hepatocytes but enhanced in Kupffer cells, which were increased in number and size with more numerous lysosome-related profiles.^[74]

The antioxidant activity of resveratrol in cholesterol-fed rats, along with its hypolipidemic effects was determined by Zhu et al.[75] Resveratrol significantly lowered serum lipid, hepatic cholesterol (TC), and TG levels compared to the control. Excretion of bile acids was significantly enhanced by resveratrol. The overall potential of the antioxidant system was significantly enhanced by the resveratrol as plasma and hepatic TBARS levels were lowered while serum SOD, GPx, and CAT activities were increased in nonresveratrol group. These findings suggest that resveratrol maintains an antioxidant efficacy as well as its anti-hyperlipidemic effect. Resveratrol (200 mg/kg/day) reduced body weight and liver weight gains, improved serum lipid parameters, reduced hepatic cholesterol accumulation and increased the bile acid pool size in mice fed an HFD for 8 weeks.^[76] By feeding 10 or 50 ppm resveratrol in the diet to hepatomabearing rats for 20 days, solid tumor growth and metastasis tended to be suppressed dose-dependently.[77] Resveratrol (50 ppm) significantly suppressed the serum lipid peroxide level, indicating its antioxidative properties or those of its metabolite (s) in vivo. Resveratrol dose-dependently suppressed both the serum TG and very low-density lipoprotein + LDL-cholesterol levels.[77]

Liver fibrosis

Oxidative stress in liver injury is a major pathogenetic factor in progress of liver fibrosis.

Resveratrol (10 and 20 mg/kg/day) or its vehicle was administered to cirrhotic rats for 2 weeks and results were as follow:^[78] Reduction in portal pressure compared to vehicle without significant changes in systemic hemodynamics. Reduction in portal pressure was associated with an improved vasodilatory response to acetylcholine, with decreased TXA2 production, increased endothelial NO, and with a significant reduction in liver fibrosis. The decrease in hepatic fibrosis was associated with a reduced collagen-1, TGF- β , NFjB mRNA expression and desmin and α -SMA protein expression.

Hong *et al.*^[79] investigated the protective effects of Resveratrol on dimethylnitrosamine (DMN)-induced liver fibrosis in rats and observed that resveratrol remarkably recovered body and liver weight loss due to DMN induced liver fibrosis. Liver histology showed that Resveratrol alleviated the infiltration of inflammatory cells and fibrosis of hepatic tissue. Resveratrol decreased the level of MDA and increased the levels of GPx and SOD. Also, It significantly inhibited the mRNA expression of inflammatory mediators including inducible NO, TNF-alpha and IL-1beta. In addition, Resveratrol showed not only a reduction in mRNA expression of fibrosis-related genes such as transforming growth factor beta 1, collagen type I, and alpha-smooth muscle actin, but also a significant decrease of hydroxyproline in rats with DMN-induced liver fibrosis.

Another study^[80] has shown that resveratrol (20 mg/ kg daily for 4 weeks) remarkably prevented the DMNinduced loss in body and liver weight, and inhibited the elevation of serum alanine transaminase, aspartate transaminase, alkaline phosphatase, and bilirubin levels. Resveratrol also increased serum albumin and hepatic GSH levels and reduced the hepatic level of MDA due to its antioxidant effect. Furthermore, DMN-induced elevation of hydroxyprolinecontent was reduced in the Resveratrol treated rats, the result of which was consistent with the reduction in type I collagen mRNA expression. The reduction in hepatic stellate cell activation, as assessed by α -smooth muscle actin staining, and the reduction in transforming growth factor-β1 mRNA expression were associated with resveratrol treatment indicating the in vivo hepatoprotective and antifibrogenic effects of resveratrol against DMN-induced liver injury.

Lv *et al.*^[81] assessed the effects of resveratrol (25, 50, or 100 mg/kg, for 6 weeks) on chronic liver fibrosis induced by CCl_4 in rats. Rats fed with 50 and 100 mg/kg of resveratrol had significant reductions in levels of the serum ALT, and liver hydroxyproline and MDA and relieving liver fibrogensis, compared with the control group.

CONCLUSION

Resveratrol can play a pivotal role in prevention and treatment of liver disorders. Previous studies confirmed its antioxidative properties in different models of hepatitis resulting in reducing of hepatic fibrosis; on the other hand, Resveratrol could reduce hepatic steatosis through modulating the insulin resistance and lipid profile in animals. These high quality preclinical studies propose the potential therapeutic implication of Resveratrol in liver disorders especially those with hepatic steatosis. Additional carefully designed, mechanistic based, laboratory, and clinical studies need to be undertaken to provide scientific evidence for the efficacy of it in treatment of liver disorders especially those with hepatic steatosis and fibrosis.

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AUTHOR'S CONTRIBUTION

All authors contributed in the study design, conducting the systematic review, and drafting the manuscript. All authors

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REFERENCES

- 1. Brown L, Kroon PA, Das DK, Das S, Tosaki A, Chan V, *et al.* The biological responses to resveratrol and other polyphenols from alcoholic beverages. Alcohol Clin Exp Res 2009;33:1513-23.
- 2. Luther DJ, Ohanyan V, Shamhart PE, Hodnichak CM, Sisakian H, Booth TD, *et al.* Chemopreventive doses of resveratrol do not produce cardiotoxicity in a rodent model of hepatocellular carcinoma. Invest New Drugs 2011;29:380-91.
- 3. Wang G, Guo X, Chen H, Lin T, Xu Y, Chen Q, *et al.* A resveratrol analog, phoyunbene B, induces G2/M cell cycle arrest and apoptosis in HepG2 liver cancer cells. Bioorg Med Chem Lett 2012;22:2114-8.
- 4. Baur JA, Sinclair DA. Therapeutic potential of resveratrol: The *in vivo* evidence. Nat Rev Drug Discov 2006;5:493-506.
- Shankar S, Singh G, Srivastava RK. Chemoprevention by resveratrol: Molecular mechanisms and therapeutic potential. Front Biosci 2007;12:4839-54.
- 6. Saiko P, Szakmary A, Jaeger W, Szekeres T. Resveratrol and its analogs: Defense against cancer, coronary disease and neurodegenerative maladies or just a fad? Mutat Res 2008;658: 68-94.
- 7. Shakibaei M, Harikumar KB, Aggarwal BB. Resveratrol addiction: To die or not to die. Mol Nutr Food Res 2009;53:115-28.
- 8. Baur JA, Pearson KJ, Price NL, Jamieson HA, Lerin C, Kalra A, *et al.* Resveratrol improves health and survival of mice on a high-calorie diet. Nature 2006;444:337-42.
- 9. Muriel P, Rivera-Espinoza Y. Beneficial drugs for liver diseases. J Appl Toxicol 2008;28:93-103.
- Kawada N, Seki S, Inoue M, Kuroki T. Effect of antioxidants, resveratrol, quercetin, and N-acetylcysteine, on the functions of cultured rat hepatic stellate cells and Kupffer cells. Hepatology 1998;27:1265-74.
- 11. Bechmann LP, Zahn D, Gieseler RK, Fingas CD, Marquitan G, Jochum C, *et al.* Resveratrol amplifies profibrogenic effects of free fatty acids on human hepatic stellate cells. Hepatol Res 2009;39:601-8.
- 12. Schmatz R, Perreira LB, Stefanello N, Mazzanti C, Spanevello R, Gutierres J, *et al.* Effects of resveratrol on biomarkers of oxidative stress and on the activity of delta aminolevulinic acid dehydratase in liver and kidney of streptozotocin-induced diabetic rats. Biochimie 2012;94:374-83.
- Godichaud S, Krisa S, Couronné B, Dubuisson L, Mérillon JM, Desmoulière A, *et al.* Deactivation of cultured human liver myofibroblasts by trans-resveratrol, a grapevine-derived polyphenol. Hepatology 2000;31:922-31.
- Godichaud S, Si-Tayeb K, Augé N, Desmoulière A, Balabaud C, Payrastre B, *et al.* The grape-derived polyphenol resveratrol differentially affects epidermal and platelet-derived growth factor signaling in human liver myofibroblasts. Int J Biochem Cell Biol 2006;38:629-37.
- Shang J, Chen LL, Xiao FX, Sun H, Ding HC, Xiao H. Resveratrol improves non-alcoholic fatty liver disease by activating AMP-activated protein kinase. Acta Pharmacol Sin 2008;29:698-706.
- 16. Nah HY, Lee WS, Joo YE, Kim HS, Choi SK, Rew JS, *et al.* Resveratrol protects HepG2 and chang liver cells from oxidative stress. Chonnam Med J 2005;41:243-52.
- 17. Souza IC, Martins LA, Coelho BP, Grivicich I, Guaragna RM, Gottfried C, *et al.* Resveratrol inhibits cell growth by inducing cell

cycle arrest in activated hepatic stellate cells. Mol Cell Biochem 2008;315:1-7.

- Rubiolo JA, Mithieux G, Vega FV. Resveratrol protects primary rat hepatocytes against oxidative stress damage: Activation of the Nrf2 transcription factor and augmented activities of antioxidant enzymes. Eur J Pharmacol 2008;591:66-72.
- 19. Rubiolo JA, Vega FV. Resveratrol protects primary rat hepatocytes against necrosis induced by reactive oxygen species. Biomed Pharmacother 2008;62:606-12.
- 20. Wang DH, Ootsuki Y, Fujita H, Miyazaki M, Yie Q, Tsutsui K, *et al.* Resveratrol inhibited hydroquinone-induced cytotoxicity in mouse primary hepatocytes. Int J Environ Res Public Health 2012;9:3354-64.
- 21. Gnoni GV, Paglialonga G. Resveratrol inhibits fatty acid and triacylglycerol synthesis in rat hepatocytes. Eur J Clin Invest 2009;39:211-8.
- 22. Cerný D, Canová NK, Martínek J, Horínek A, Kmonícková E, Zídek Z, et al. Effects of resveratrol pretreatment on tert-butylhydroperoxide induced hepatocyte toxicity in immobilized perifused hepatocytes: Involvement of inducible nitric oxide synthase and hemoxygenase-1. Nitric Oxide 2009;20:1-8.
- Chen CC, Chan WH. Inhibition of citrinin-induced apoptotic biochemical signaling in human hepatoma G2 cells by resveratrol. Int J Mol Sci 2009;10:3338-57.
- 24. Notas G, Nifli AP, Kampa M, Vercauteren J, Kouroumalis E, Castanas E. Resveratrol exerts its antiproliferative effect on HepG2 hepatocellular carcinoma cells, by inducing cell cycle arrest, and NOS activation. Biochim Biophys Acta 2006;1760:1657-66.
- 25. Parekh P, Motiwale L, Naik N, Rao KV. Downregulation of cyclin D1 is associated with decreased levels of p38 MAP kinases, Akt/ PKB and Pak1 during chemopreventive effects of resveratrol in liver cancer cells. Exp Toxicol Pathol 2011;63:167-73.
- 26. Choi HY, Chong SA, Nam MJ. Resveratrol induces apoptosis in human SK-HEP-1 hepatic cancer cells. Cancer Genomics Proteomics 2009;6:263-8.
- Gallis JL, Serhan N, Gin H, Couzigou P, Beauvieux MC. Resveratrol plus ethanol counteract the ethanol-induced impairment of energy metabolism: 31P NMR study of ATP and sn-glycerol-3-phosphate on isolated and perfused rat liver. Pharmacol Res 2012;65:387-95.
- Wu SL, Yu L, Pan CE, Jiao XY, Lv Y, Fu J, et al. Apoptosis of lymphocytes in allograft in a rat liver transplantation model induced by resveratrol. Pharmacol Res 2006;54:19-23.
- 29. Wu SL, Yu L, Jiao XY, Meng KW, Pan CE. The suppressive effect of resveratrol on protein kinase C theta in peripheral blood T lymphocytes in a rat liver transplantation model. Transplant Proc 2006;38:3052-4.
- Wu SL, Pan CE, Yu L, Meng KW. Immunosuppression by combined use of cyclosporine and resveratrol in a rat liver transplantation model. Transplant Proc 2005;37:2354-9.
- Wu SL, Yu L, Meng KW, Ma ZH, Pan CE. Resveratrol prolongs allograft survival after liver transplantation in rats. World J Gastroenterol 2005;11:4745-9.
- Wu SL, Sun ZJ, Yu L, Meng KW, Qin XL, Pan CE. Effect of resveratrol and in combination with 5-FU on murine liver cancer. World J Gastroenterol 2004;10:3048-52.
- Yu L, Sun ZJ, Wu SL, Pan CE. Effect of resveratrol on cell cycle proteins in murine transplantable liver cancer. World J Gastroenterol 2003;9:2341-3.
- Bishayee A, Barnes KF, Bhatia D, Darvesh AS, Carroll RT. Resveratrol suppresses oxidative stress and inflammatory response in diethylnitrosamine-initiated rat hepatocarcinogenesis. Cancer Prev Res (Phila) 2010;3:753-63.
- 35. Bishayee A, Waghray A, Barnes KF, Mbimba T, Bhatia D, Chatterjee M, *et al.* Suppression of the inflammatory cascade

is implicated in resveratrol chemoprevention of experimental hepatocarcinogenesis. Pharm Res 2010;27:1080-91.

- Mbimba T, Awale P, Bhatia D, Geldenhuys WJ, Darvesh AS, Carroll RT, *et al.* Alteration of hepatic proinflammatory cytokines is involved in the resveratrol-mediated chemoprevention of chemically-induced hepatocarcinogenesis. Curr Pharm Biotechnol 2012;13:229-34.
- Salado C, Olaso E, Gallot N, Valcarcel M, Egilegor E, Mendoza L, et al. Resveratrol prevents inflammation-dependent hepatic melanoma metastasis by inhibiting the secretion and effects of interleukin-18. J Transl Med 2011;9:59.
- Burgess TA, Robich MP, Chu LM, Bianchi C, Sellke FW. Improving glucose metabolism with resveratrol in a swine model of metabolic syndrome through alteration of signaling pathways in the liver and skeletal muscle. Arch Surg 2011;146:556-64.
- Gedik E, Girgin S, Ozturk H, Obay BD, Ozturk H, Buyukbayram H. Resveratrol attenuates oxidative stress and histological alterations induced by liver ischemia/reperfusion in rats. World J Gastroenterol 2008;14:7101-6.
- 40. Hassan-Khabbar S, Cottart CH, Wendum D, Vibert F, Clot JP, Savouret JF, *et al.* Postischemic treatment by trans-resveratrol in rat liver ischemia-reperfusion: A possible strategy in liver surgery. Liver Transpl 2008;14:451-9.
- Plin C, Tillement JP, Berdeaux A, Morin D. Resveratrol protects against cold ischemia-warm reoxygenation-induced damages to mitochondria and cells in rat liver. Eur J Pharmacol 2005;528:162-8.
- Rivera H, Shibayama M, Tsutsumi V, Perez-Alvarez V, Muriel P. Resveratrol and trimethylated resveratrol protect from acute liver damage induced by CCl4 in the rat. J Appl Toxicol 2008;28:147-55.
- 43. Fan G, Tang JJ, Bhadauria M, Nirala SK, Dai F, Zhou B, *et al.* Resveratrol ameliorates carbon tetrachloride-induced acute liver injury in mice. Environ Toxicol Pharmacol 2009;28:350-6.
- Lodovici M, Bigagli E, Luceri C, Manni EM, Zaid M. Protective effect of resveratrol against oxidation stress induced by 2-nitropropane in rat liver. Pharmacology 2011;2:127.
- 45. Tunali-Akbay T, Sehirli O, Ercan F, Sener G. Resveratrol protects against methotrexate-induced hepatic injury in rats. J Pharm Pharm Sci 2010;13:303-10.
- 46. Roy S, Sannigrahi S, Majumdar S, Ghosh B, Sarkar B. Resveratrol regulates antioxidant status, inhibits cytokine expression and restricts apoptosis in carbon tetrachloride induced rat hepatic injury. Oxid Med Cell Longev 2011;2011:703676.
- Vitaglione P, Ottanelli B, Milani S, Morisco F, Caporaso N, Fogliano V. Dietary trans-resveratrol bioavailability and effect on CCl4-induced liver lipid peroxidation. J Gastroenterol Hepatol 2009;24:618-22.
- 48. El-Agamy DS. Comparative effects of curcumin and resveratrol on aflatoxin B(1)-induced liver injury in rats. Arch Toxicol 2010;84:389-96.
- 49. Hamadi N, Mansour A, Hassan MH, Khalifi-Touhami F, Badary O. Ameliorative effects of resveratrol on liver injury in streptozotocin-induced diabetic rats. J Biochem Mol Toxicol 2012;26:384-92.
- 50. Chan CC, Cheng LY, Lin CL, Huang YH, Lin HC, Lee FY. The protective role of natural phytoalexin resveratrol on inflammation, fibrosis and regeneration in cholestatic liver injury. Mol Nutr Food Res 2011;55:1841-9.
- 51. Velioglu-Ogünç A, Sehirli O, Toklu HZ, Ozyurt H, Mayadagli A, Eksioglu-Demiralp E, *et al.* Resveratrol protects against irradiation-induced hepatic and ileal damage via its anti-oxidative activity. Free Radic Res 2009;43:1060-71.
- 52. Yu HP, Hsu JC, Hwang TL, Yen CH, Lau YT. Resveratrol attenuates hepatic injury after trauma-hemorrhage via estrogen receptorrelated pathway. Shock 2008;30:324-8.

- Bujanda L, García-Barcina M, Gutiérrez-de Juan V, Bidaurrazaga J, de Luco MF, Gutiérrez-Stampa M, *et al.* Effect of resveratrol on alcohol-induced mortality and liver lesions in mice. BMC Gastroenterol 2006;6:35.
- 54. Kasdallah-Grissa A, Mornagui B, Aouani E, Hammami M, El May M, Gharbi N, *et al.* Resveratrol, a red wine polyphenol, attenuates ethanol-induced oxidative stress in rat liver. Life Sci 2007;80:1033-9.
- 55. Jinghui L, Yingbao Y. Effects of resveratrol on serum TNF-α, IL-6, ICAM-1 in alcohol-induced hepatic injury in rats. Pharmacol Clin Chin Mater Med 2010;4:006.
- 56. Schirmer H, Pereira TC, Rico EP, Rosemberg DB, Bonan CD, Bogo MR, et al. Modulatory effect of resveratrol on SIRT1, SIRT3, SIRT4, PGC1a and NAMPT gene expression profiles in wild-type adult zebrafish liver. Mol Biol Rep 2012;39:3281-9.
- 57. Chang CC, Chang CY, Huang JP, Hung LM. Effect of resveratrol on oxidative and inflammatory stress in liver and spleen of streptozotocin-induced type 1 diabetic rats. Chin J Physiol 2012; 55:192-201.
- Sebai H, Sani M, Yacoubi MT, Aouani E, Ghanem-Boughanmi N, Ben-Attia M. Resveratrol, a red wine polyphenol, attenuates lipopolysaccharide-induced oxidative stress in rat liver. Ecotoxicol Environ Saf 2010;73:1078-83.
- Rocha KK, Souza GA, Ebaid GX, Seiva FR, Cataneo AC, Novelli EL. Resveratrol toxicity: Effects on risk factors for atherosclerosis and hepatic oxidative stress in standard and high-fat diets. Food Chem Toxicol 2009;47:1362-7.
- 60. Bagul PK, Middela H, Matapally S, Padiya R, Bastia T, Madhusudana K, *et al.* Attenuation of insulin resistance, metabolic syndrome and hepatic oxidative stress by resveratrol in fructosefed rats. Pharmacol Res 2012;66:260-8.
- Palsamy P, Sivakumar S, Subramanian S. Resveratrol attenuates hyperglycemia-mediated oxidative stress, proinflammatory cytokines and protects hepatocytes ultrastructure in streptozotocinnicotinamide-induced experimental diabetic rats. Chem Biol Interact 2010;186:200-10.
- 62. Upadhyay G, Singh AK, Kumar A, Prakash O, Singh MP. Resveratrol modulates pyrogallol-induced changes in hepatic toxicity markers, xenobiotic metabolizing enzymes and oxidative stress. Eur J Pharmacol 2008;596:146-52.
- 63. Farghali H, Cerný D, Kameníková L, Martínek J, Horínek A, Kmonícková E, et al. Resveratrol attenuates lipopolysaccharideinduced hepatitis in D-galactosamine sensitized rats: Role of nitric oxide synthase 2 and heme oxygenase-1. Nitric Oxide 2009;21:216-25.
- 64. Kirimlioglu V, Karakayali H, Turkoglu S, Haberal M. Effect of resveratrol on oxidative stress enzymes in rats subjected to 70% partial hepatectomy. Transplant Proc 2008;40:293-6.
- 65. Sengottuvelan M, Nalini N. Resveratrol, a phytoalexin enhances hepatic antioxidant defense in 1,2-dimethylhydrazine-induced colon carcinogenesis. Int J Pharmacal 2006;2:335-40.
- 66. Sahin K, Orhan C, Akdemir F, Tuzcu M, Iben C, Sahin N. Resveratrol protects quail hepatocytes against heat stress: Modulation of the Nrf2 transcription factor and heat shock proteins. J Anim Physiol Anim Nutr (Berl) 2012;96:66-74.
- 67. Xin P, Han H, Gao D, Cui W, Yang X, Ying C, *et al.* Alleviative effects of resveratrol on nonalcoholic fatty liver disease are associated with up regulation of hepatic low density lipoprotein receptor and

scavenger receptor class B type I gene expressions in rats. Food Chem Toxicol 2013;52:12-8.

- Kopeć A, Piatkowska E, Leszczyńska T, Koronowicz A. Effect of long term administration of resveratrol on lipid concentration in selected organs and liver's histology in rats fed high fructose diet. J Funct Foods 2013;5:299-305.
- 69. Poulsen MM, Larsen JØ, Hamilton-Dutoit S, Clasen BF, Jessen N, Paulsen SK, *et al.* Resveratrol up-regulates hepatic uncoupling protein 2 and prevents development of nonalcoholic fatty liver disease in rats fed a high-fat diet. Nutr Res 2012;32:701-8.
- Alberdi G, Rodríguez VM, Macarulla MT, Miranda J, Churruca I, Portillo MP. Hepatic lipid metabolic pathways modified by resveratrol in rats fed an obesogenic diet. Nutrition 2013;29:562-7.
- Cho SJ, Jung UJ, Choi MS. Differential effects of low-dose resveratrol on adiposity and hepatic steatosis in diet-induced obese mice. Br J Nutr 2012;108:2166-75.
- 72. Bujanda L, Hijona E, Larzabal M, Beraza M, Aldazabal P, García-Urkia N, *et al.* Resveratrol inhibits nonalcoholic fatty liver disease in rats. BMC Gastroenterol 2008;8:40.
- Ajmo JM, Liang X, Rogers CQ, Pennock B, You M. Resveratrol alleviates alcoholic fatty liver in mice. Am J Physiol Gastrointest Liver Physiol 2008;295:G833-42.
- 74. Shiozaki M, Hayakawa N, Shibata M, Koike M, Uchiyama Y, Gotow T. Closer association of mitochondria with lipid droplets in hepatocytes and activation of Kupffer cells in resveratroltreated senescence-accelerated mice. Histochem Cell Biol 2011;136:475-89.
- 75. Zhu L, Luo X, Jin Z. Effect of resveratrol on serum and liver lipid profile and antioxidant activity in hyperlipidemia rats. Asian Australas J Anim Sci 2008;21:890-5.
- 76. Chen Q, Wang E, Ma L, Zhai P. Dietary resveratrol increases the expression of hepatic 7a-hydroxylase and ameliorates hypercholesterolemia in high-fat fed C57BL/6J mice. Lipids Health Dis 2012;11:56.
- Miura D, Miura Y, Yagasaki K. Hypolipidemic action of dietary resveratrol, a phytoalexin in grapes and red wine, in hepatomabearing rats. Life Sci 2003;73:1393-400.
- Di Pascoli M, Diví M, Rodríguez-Vilarrupla A, Rosado E, Gracia-Sancho J, Vilaseca M, *et al.* Resveratrol improves intrahepatic endothelial dysfunction and reduces hepatic fibrosis and portal pressure in cirrhotic rats. J Hepatol 2013;58:904-10.
- 79. Hong SW, Jung KH, Zheng HM, Lee HS, Suh JK, Park IS, *et al.* The protective effect of resveratrol on dimethylnitrosamine-induced liver fibrosis in rats. Arch Pharm Res 2010;33:601-9.
- 80. Lee ES, Shin MO, Yoon S, Moon JO. Resveratrol inhibits dimethylnitrosamine-induced hepatic fibrosis in rats. Arch Pharm Res 2010;33:925-32.
- Lv QJ, Xie JQ, Wen LQ, Chen YY, Zhang M, Ye QN. Effects of resveratrol on chronic liver fibrosis in rats. Chin New Drugs J 2005;14:855.
- 82. Hassan-Khabbar S, Vamy M, Cottart CH, Wendum D, Vibert F, Savouret JF, *et al.* Protective effect of post-ischemic treatment with trans-resveratrol on cytokine production and neutrophil recruitment by rat liver. Biochimie 2010;92:405-10.
- Kirimlioglu H, Ecevit A, Yilmaz S, Kirimlioglu V, Karabulut AB. Effect of resveratrol and melatonin on oxidative stress enzymes, regeneration, and hepatocyte ultrastructure in rats subjected to 70% partial hepatectomy. Transplant Proc 2008;40:285-9.