

A Simple Sample Preparation with HPLC-UV Method for Estimation of Clomipramine from Plasma

Sayed Abolfazl Mostafavi^{a*}, Reza Tahvilian^b, Masoumeh Dehghani Poudeh^a
and Zeinab Rafeepour^a

^aFaculty of Pharmacy and Pharmaceutical Sciences and Isfahan Pharmaceutical Research Center, Isfahan University of Medical Sciences, Isfahan, Iran. ^bSchool of Pharmacy, Kermanshah University of Medical Sciences, Kermanshah, Iran.

Abstract

Clomipramine is a tricyclic antidepressant. Different methods for determination of clomipramine hydrochloride in plasma have been described. Most of these procedures favor the use of acidic back-extraction in extraction procedure and HPLC as the analytical technique. In this study, the clomipramine extraction procedure was modified and a direct injection to the column was performed to shorten the time of sample preparation considerably. Furthermore, the method was applied in bioequivalence study of new formulations of clomipramine in comparison with reference tablets.

The drug and internal standard were extracted from plasma with heptan : isoamyl alcohol (95:5) and re-extracted with 200 μ L of orthophosphoric acid (0.3% v/v). The organic layer was discharged and analysis was performed on C₈ reverse phase ODS2 HPLC column with a mobile phase, acetonitrile : water (75:25) and UV detection set at 215 nm. Additionally, a single dose study was carried out with a two-sequence, crossover block-randomized design for bioequivalence study. Clomipramine tablets (3 \times 25 mg) of either formulations (reference or test products) were administered separately in two occasions to 12 fasting healthy male volunteers. Blood samples were taken prior to and at 9 points within 48 h after dose administration.

The retention time of internal standard (cisapride), clomipramine, and desmethyl clomipramine were 5.6 ± 0.2 , 10.3 ± 0.3 , and 9.5 ± 0.3 min, respectively. The standard curve covering the concentration ranges of 2.5-120 ng/mL was linear ($r^2 = 0.9950$ and 0.9979) for clomipramine and desmethyl clomipramine. The co-efficient of variation for intra-day and inter-day accuracy and precision was less than 18.3%. The pharmacokinetic parameters C_{max} and T_{max} were obtained directly from plasma clomipramine concentrations. K_{el} was estimated by log-linear regression and AUC was calculated by the linear trapezoidal rule. The pharmacokinetic parameters AUC and C_{max} were tested for equivalence after log-transformation of data. The 90% standard confidence intervals of the mean values for the test/reference ratios, AUC, and C_{max} were within the acceptable bioequivalence limits of 0.80-1.20.

These results indicated that the analytical method was linear and accurate. Test and reference formulations were found to be bioequivalent and therefore interchangeable.

Keywords: Clomipramine; Bioequivalency; Anafranil[®]; HPLC method.

* Corresponding author:

E-mail: mostafavi@pharm.mui.ac.ir

Introduction

Clomipramine (3-chloro-5-(3-dimethylamino propyl)-10,11-dihydro-5H-dibenz[b,f]azepine hydrochloride) is a dibenzazepine tricyclic antidepressant with actions and uses similar to those of amitriptyline (Figure 1). It has antimuscarinic properties and is also a potent serotonin reuptake inhibitor (1). Clomipramine (CMI) is one of the most sedating tricyclic (2-4). The drug is recommended in obsessive compulsive disorders (OCD), in cataplexy associated with narcolepsy, and in depression when sedation is required (5-7). CMI is rapidly absorbed from the gastrointestinal tract and extensively desmethylated by first-pass metabolism in the liver to its primary active metabolite, desmethyl clomipramine (DMCMI) (8, 9). The metabolic pathways of both CMI and DMCMI include hydroxylation and N-oxidation. CMI is excreted in the urine, mainly in the form of its metabolites, either free or conjugated form (10-12).

Most studies performed on determination of clomipramine, have measured only CMI. A few of them, however, have simultaneously determined its desmethyl and hydroxy-metabolites. Furthermore, these methods have accomplished a complex, expensive, and time-consuming three-step liquid-liquid extraction which requires more sophisticated equipment (13). These methods are time-consuming; therefore, they are cumbersome for bioavailability and bioequivalence studies.

The pharmacokinetics of clomipramine has been documented elsewhere (14, 15), however, bioavailability issues have been an increasing concern to drug regulatory authorities once assessing the safety and efficacy of drug products. The increasing number of synonym drug products requires special attention terms of bioavailability issues. Hence, drug regulatory authorities have issued guidelines to ensure adequate bioavailability studies in new drug applications for synonym drugs (16). In this study, a simple and sensitive HPLC method using cisapride as internal standard (IS) with capability of simultaneously determining CMI and its N-desmethylated metabolites in human plasma were evaluated. Furthermore, the possible bioequivalency of a generic tablet

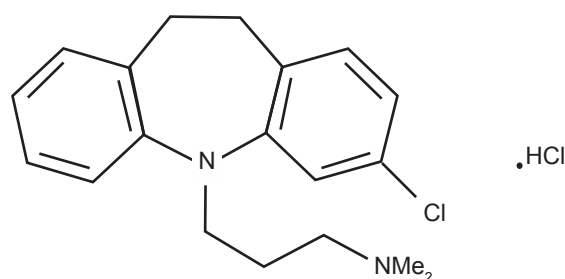


Figure 1. Chemical structure of clomipramine.

of clomipramine (25 mg) made by an Iranian company in comparison to reference formulation was evaluated.

Experimental

Reagents and solutions

Clomipramine and desmethyl clomipramine powder were obtained from Farmaceutici (Italy) and cisapride from Jenson Co. (Belgium). Acetonitrile (HPLC grade), *n*-heptane (analytical grade, distilled before use), isoamyl-alcohol, sulfuric acid (all from Merck, Germany) were obtained from the local market. All other reagents and solutions were either HPLC or analytical grade.

Clomipramine HCL tablets were obtained from a local pharmaceutical company. Anafranil® (25 mg) tablets were from Novartis.

Chromatographic conditions

A reversed phase HPLC method was developed to quantitate plasma levels of clomipramine. The apparatus used was a Jasco HPLC system (Japan), consisting of a model 980-PU intelligent solvent delivery pump, 7125-Rheodyne injector, a computerized system controller (with the Borwin software), and a UV-975 detector set at 215 nm. Chromatographic separation was performed using a Perkin Elmer C-8 reverse phase ODS2 HPLC column. The mobile phase consisted of 75% acetonitrile, 25% water, and 0.01% triethylamine. The apparent pH of the mixed solvent system was adjusted to 4 ± 0.1 with a dilute solution of orthophosphoric acid. The aqueous phase was eluted at a flow rate of 1 mL/min, and effluent was monitored at

215 nm, at attenuation of 0.0005 and gain $\times 10$. Quantitation was achieved by measurement of the peak area ratios of the drug to the internal standard.

Sample preparation

To 1 mL plasma in a 10 mL test tube, 0.5 mL NaOH (1 M) and 100 μL of IS (1 $\mu\text{g}/\text{mL}$) was added and extracted twice with 3 mL of extracted solvent [heptane: isoamylalcohol (95:5)]. Vortexed for 1 min and centrifuged at 2000 g for 5 min. Upper layer was separated in a tube and back extracted with 200 μL of 0.3% orthophosphoric acid. The organic layer was aspirated and 100 μL of the residue was injected to the column.

Calibration procedure

In order to make CMI standard concentrations, to 1 mL of blank plasma, 100 μL of CMI standard solution of CMT and DMCMCMI at concentrations of 2.5, 5, 10, 20, 40, 80 and 120 ng/100 μL plus 100 μL of IS 1 $\mu\text{g}/\text{mL}$ was added. All calibration samples were taken through the extraction procedure. The calibration curve was plotted using peak ratios of CMI/IS versus CMI concentrations. Final sample concentrations were calculated by determining the peak area ratio of CMI related to IS and comparing the ratio with the standard curve obtained after analysis of calibration samples.

Extraction efficiency

Recoveries of CMI from spiked samples were determined by comparing the peak areas obtained by extraction of freshly prepared plasma at concentrations of 2.5-120 ng/mL with those found by direct injection of an aqueous standard solution at equivalent concentrations.

Precision

Within-day variability

The within-day variability of the assay was determined by repeated analysis of quality control samples at concentrations ranging from 2.5 to 120 ng/mL on the same day.

Between-day variability

The between-day variability of the assay was determined by repeated analysis of quality control samples at concentrations ranging from

2.5 to 120 ng/mL on 3 different days.

In-vivo study design

Two separate groups consisted of twelve healthy non-smoking male volunteers weighing from 60-85 kg completed the studies (aged 21 to 27). The study was performed on the basis of medical history, clinical examinations, and laboratory tests including hematology, blood and biochemistry, and urine analysis. No subject had a history or evidence of hepatic, renal, gastrointestinal or hematological deviations or any acute or chronic disease or drug allergy. All volunteers were instructed to abstain from taking any medication and xanthin containing foods for at least 2 weeks prior to and during the study. No milk or dairy products were served during the study. Informed consent was obtained from the subjects after explaining the nature and purpose of the study. The ethics committee of the Iranian ministry of health approved the study and was conducted in accordance with good clinical practice guidelines.

The protocols used were the conventional, two-sequence, crossover block-randomized design for bioequivalence study with six subjects in each of the treatment group. After an overnight fasting, all subjects received a single dose of three units of 25 mg tablets (reference or test product) randomly, with 250 mL of water. Food and drinks were not allowed until 2 h after ingestion of tablets. The same lunch and dinner was served at 5 and 12 h after dosing for all volunteers.

Clinical protocol

Approximately 10 mL of blood samples were drawn into heparinized tube through an indwelling cannula before (0 h) and at 1, 2, 4, 5, 6, 8, 12, 24 and 48 h after dosing. The blood samples were centrifuged at 2000 rpm for 15 min and plasma was separated and kept frozen at -20°C in coded glass tubes.

After a period of 14 days the study was repeated in the same manner to complete the crossover design.

Pharmacokinetic analysis

Estimation and calculation of pharmacokinetic parameters were performed using MS Excel software. The maximum CMI concentration

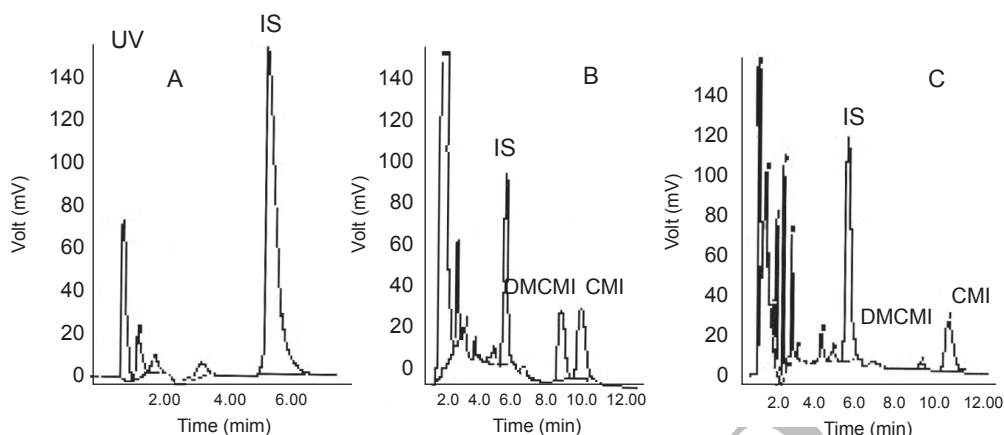


Figure 2. Chromatograms of blank human plasma and the internal standard (A), control plasma spiked with clomipramine and the internal standard (B), and plasma from a healthy subject 6 hours after ingestion of a clomipramine tablet (C).

(C_{max}) and the corresponding peak time (T_{max}) were determined by inspection of the individual drug plasma concentration-time profiles. The elimination rate constant (K_{el}) was obtained from the least square fitted terminal log-linear portion of the plasma concentration-time profile. The elimination half-life ($T_{1/2}$) was calculated as $0.693/K_{el}$. The area under the curve to the last measurable concentration (AUC_{0-t}) was calculated by the linear trapezoidal rule. The area under the curve extrapolated to infinity ($AUC_{0-\infty}$) was calculated by equation $AUC_{0-t} + C_t / K_{el}$ where C_t is the last measurable concentration.

Statistical analysis

For the purpose of bioequivalence analysis, AUC_{0-t} , $AUC_{0-\infty}$, and C_{max} were considered as primary variables. The two-way ANOVA for crossover design was used to assess the effect of formulations, periods, sequences, and subjects on these parameters. A difference between two related parameters was considered statistically significant for a P-value equal to or less than 0.05. The 90% confidence interval of the ratio of pharmacokinetic parameters of logarithmically transformed were also estimated. All statistical analysis was performed using SPSS version 12.

Results

No interfering peaks near the retention

times of CMI, DMCMI, and IS were present in the chromatograms of blank or basal (time 0) plasma samples. Representative chromatograms are shown in Figure 2.

Linearity

Calibration standard containing 2.5-120 ng/mL was prepared from working solutions of CMI, DCMI, and blank plasma. The calibration curve was constructed by plotting the peak area ratio of CMI and DCMI to IS against the CMI and DCMI concentration in plasma. The ratios of the areas of the peaks of each compound versus IS were linearly related to the concentrations in the range encountered in drug monitoring. The calibration curves of CMI and DCMI after six-time replication are shown in Figure 3 which showed a good linearity within the examined concentration range.

Accuracy and precision

The intra-day coefficients of variation (CV%) of CMI and its metabolite (DCMI) were between 3.09 and 18.33% and the inter-day coefficients of variation were between 7.25 and 14.51% for all compounds (Table 1 and 2). These results, therefore, validate the calibration curves used for each set of samples.

Sensitivity and recovery

The limit of quantitation (sensitivity) and detection limit of the assay for CMI and DCMI

Table 1. The intra-day variability of clomipramine and its major metabolite (desmethyl clomipramine) in plasma (n = 6).

Clomipramine				Desmethyl clomipramine			
Added concentrations (ng/mL)	Experimental concentrations (ng/mL)	SD	CV (%)	Added concentrations (ng/mL)	Experimental concentrations (ng/mL)	SD	CV (%)
5	6.45	1.12	17	5	5.80	0.20	4
10	10.24	1.13	11	10	10.32	1.10	11
20	19.96	2.45	12	20	18.21	1.59	9
40	35.28	3.46	10	40	38.28	4.33	11
80	80.43	8.33	10	80	77.63	11	15
120	115.48	3.58	3	120	98.82	17	18

SD: Standard deviation; CV: Coefficient of variation.

was found to be less than 5 and 1 ng/mL, respectively.

The absolute recovery of CMI and its metabolite, which was determined by comparing the areas of the peaks of not-extracted standards with those of extracted standards according to the procedure described above, over the 5-80° ng/mL plasma concentration ranges, was between 62-72%.

Pharmacokinetic parameters

The mean concentration-time profile for two brands of clomipramine is shown in Figure 4. The pharmacokinetic parameters of clomipramine 25 mg in comparison to the reference product is shown in Table 3. For bioequivalence evaluation, various statistical modules were applied to AUC_{0-t} , $AUC_{0-\infty}$, and C_{max} in accordance with current US Food and Drug Administration (FDA) guidelines (16). The 90% confidence intervals for AUC_{0-t} , $AUC_{0-\infty}$, and C_{max} were within the range of 0.8-1.2 required by authorities. According

to the mean plasma levels of the 12 subjects completing the study, the relative bioavailability of clomipramine tablet is shown in Table 3. The mean and standard deviation of both parameters for the two brands were found to be very close, indicating that the plasma profiles generated by clomipramine 3×25 mg is comparable to the reference standard (Anafranil®). Analysis of variance (ANOVA) for these parameters, after log-transformation of the data, showed no statistically significant differences ($P > 0.05$) between the two brands. In addition, 90% confidence interval demonstrated that the ratio of the AUC_{0-t} or $AUC_{0-\infty}$ of the two brands lie within the FDA accepted range 80-125%. For bioequivalence evaluation, t-student statistical procedure was performed on C_{max} values. This analysis showed that the two formulations of clomipramine 25 are not statistically different ($P > 0.05$) from each other. Moreover, 90% confidence interval also demonstrated that the ratio of the C_{max} lie within the FDA accepted range of 80-125%.

Table 2. The inter-day variability of clomipramine and its major metabolite (desmethyl clomipramine) in plasma (n = 9).

Clomipramine				Desmethyl clomipramine			
Added concentrations (ng/mL)	Experimental concentrations (ng/mL)	SD	CV (%)	Added concentrations (ng/mL)	Experimental concentrations (ng/mL)	SD	CV (%)
5	5.52	0.63	12	5	5.52	0.48	9
10	10.29	1.49	15	10	10.04	1.39	14
20	19.68	2.66	14	20	18.74	1.58	8
40	39.92	3.87	10	40	40.34	5.03	12
80	87.29	7.30	8	80	83.37	9.85	12
120	118.63	8.60	7	120	121.30	12.9	11

SD: Standard deviation; CV: Coefficient of variation.

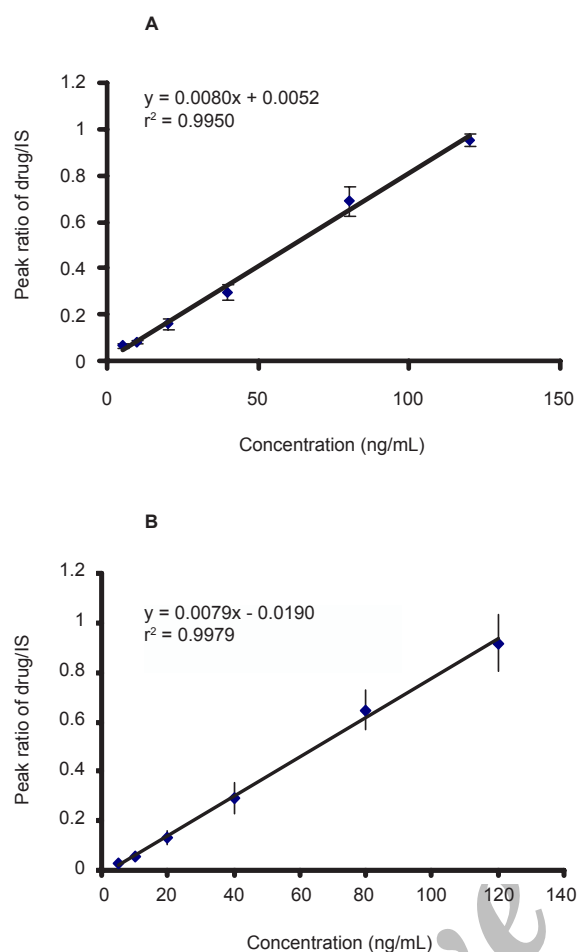


Figure 3. The calibration curve of A: clomipramine and B: desmethyl clomipramine in plasma.

Discussion

Different methods for the determination of clomipramine hydrochloride in plasma have been described. Most of these procedures favor the use of acidic back-extraction in extraction procedure and HPLC as the analytical technique (13, 17, 18). In our experience, the acidic back-extraction and drying under nitrogen has been avoided and a modification was performed in extraction procedure. Extraction with 200 μ L of 0.3% solution of ortho-phosphoric acid and direct injection to the column shortened the time of sample preparation significantly.

Regulatory authorities usually require that one single dosing form to be administered. Exception is allowed if the plasma/serum levels are extremely low for the assay of the

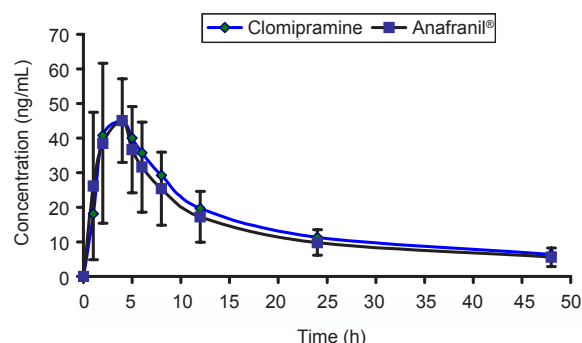


Figure 4. The mean concentration-time profiles of 3 \times 25 mg clomipramine and Anafranil[®] tablets.

analytes. In those situations, a multiple of the dosage units are allowed to be administered for analytical reason. As reported by Pirola et al, CMI concentration is very low in plasma (19). Therefore, three units of CMI 25 mg dose were administered. During the experiment, the performance of measuring the concentration of the drug by HPLC could be increased. The blood was taken from volunteers by giving three units of 25 mg CMI from volunteers we used them for this protocol.

All calculated pharmacokinetic parameter values were in good agreement with the previously reported values (9-11, 14, 15). Clomipramine was well tolerated by the volunteers in both phases of the study. All volunteers who started the study continued to the end and were discharged in good health. Unexpected incidents that could have influenced the outcome of the study did not occur. All formulations were readily absorbed from the gastrointestinal tract and clomipramine was measurable at the first sampling time (1 h) in almost all volunteers.

The method was validated (20) using a linearity range of 2.5-120 ng/mL with a limit of detection less than 1 ng/mL and recovery ranged between 61.53-71.73%. The mean intra-day and inter-day CV were 10.68% (3.09-18.33%) and 10.82% (7.25-14.51%).

The most important objective of bioequivalence testing is to assure the safety and efficacy of generic formulations. When two formulations of the same drug are equivalent

Table 3. Pharmacokinetic parameters of clomipramine hydrochloride after administration of 3 × 25 mg tablets.

Pharmacokinetic parameters	Clomipramine	Anafranil®	Relative bioavailability (%)
C _{max} (ng/mL)	46.42 ± 17.45	47.57 ± 18.79	1.00 ± 0.30
T _{max} (h)	3.75 ± 1.21	3.91 ± 1.08	NA
K _{el} (h ⁻¹)	0.040 ± 0.014	0.044 ± 0.021	NA
T _{1/2} (h)	19.04 ± 6.59	18.08 ± 6.06	NA
AUC ₀₋₄₈ (ng.h/mL)	678.79 ± 171.43	618.17 ± 196.31	1.10 ± 0.30
AUC _{0-∞} (ng.h/mL)	836.31 ± 267.56	754.61 ± 283.01	1.10 ± 0.30

NA: Not applicable

in the rate and extent to which the active drug becomes available to the site of drug action, the two formulations are bioequivalent and thus considered therapeutically equivalent (21). In order to demonstrate bioequivalence, certain limits should be set depending on the nature of drug, patient population, and clinical end points. It is generally accepted that for basic pharmacokinetic characteristics, such as AUC and C_{max}, the standard equivalence range is 0.8-1.25 (16). Pharmacokinetic and statistical results founded in our study showed Anafranil® (3 × 25 mg) and clomipramine (3 × 25 mg) tablets are bioequivalent.

In conclusion, the HPLC method with modification in extraction procedure described above for the quantification of clomipramine was fast, sensitive, reliable, and reduced the time of clomipramine determination in plasma comparing to other methods. The statistical comparison of AUC₀₋₄₈, AUC_{0-∞}, and C_{max}, obviously indicated no significant difference between the two brands of clomipramine tablets. In addition, 90% confidence intervals for the mean ratios (T/R) of AUC₀₋₄₈, AUC_{0-∞}, and C_{max} indicated that the reported values were entirely within the bioequivalence acceptance range of 80-125% (using log-transformed data). Based on the above mentioned pharmacokinetic and statistical results of this study, we can conclude that Anafranil® (3 × 25 mg) and clomipramine (3 × 25 mg) tablets are bioequivalent. Thus, these products can be considered interchangeable in medical practice.

References

- Herrera D, Mayet L, Galindo MC and Jung H. Pharmacokinetics of a sustained-release dosage form of clomipramine. *J. Clin. Pharmacol.* (2000) 40: 1488-1493.
- Kelly D. Physiological measurements in phobic and obsession disorders. *Scott. Med. J.* (1975) 20: 7-11.
- McTavish D and Benfield P. Clomipramine. An overview of its pharmacological properties and a review of its therapeutic use in obsessive compulsive disorder and panic disorder. *Drugs* (1990) 39: 136-153.
- Papp LA, Schneier FR, Fyer AJ, Leibowitz MR, Gorman JM, Coplan JD, Campeas R, Fallon BA and Klein DF. Clomipramine treatment of panic disorder: pros and cons. *J. Clin. Psychiatry* (1997) 58: 423-425.
- Houghton WC, Scammell TE and Thorpy M. Pharmacotherapy for cataplexy. *Sleep Med. Rev.* (2004) 8: 355-366.
- Vythilingum B, Cartwright C and Hollander E. Pharmacotherapy of obsessive-compulsive disorder: experience with the selective serotonin reuptake inhibitors. *Int. Clin. Psychopharmacol.* (2000) 2: S7-S13.
- Fineberg NA and Craig KJ. Pharmacological treatment for obsessive-compulsive disorder. *Psychiatry* (2007) 6: 234-239.
- Glazener CM, Evans JH and Peto RE. Tricyclic and related drugs for nocturnal enuresis in children. *Cochrane Database Syst. Rev.* (2003) 3: CD002117.
- Muller FO, Schall R, Mogilnicka EM, Groenewoud G, Hundt HK, Luus HG, Middle MV, Swart KJ and DeVaal AC. Relative bioavailability of four clomipramine hydrochloride tablet products. *Biopharm. Drug Dispos.* (1996) 17: 81-90.
- Westenberg HG. Developments in the drug treatment of panic disorder: what is the place of the selective serotonin reuptake inhibitors? *J. Affect. Disord.* (1996) 40: 85-93.
- Nielsen KK, Brosen K, Hansen MG and Gram LF. Single-dose kinetics of clomipramine: relationship to the sparteine and S-mephenytoin oxidation polymorphisms. *Clin. Pharmacol. Ther.* (1994) 55: 518-527.
- Balant-Gorgia AE, Gex-Fabry M and Balant LP. Clinical pharmacokinetics of clomipramine. *Clin. Pharmacokinet.* (1991) 20: 447-462.
- Godbillon J and Gauron S. Determination of

- clomipramine or imipramine and their mono-demethylated metabolites in human blood or plasma by high-performance liquid chromatography. *J. Chromatogr. A* (1981) 204: 303-311.
- (14) Balant-Gorgia AE, Gex-Fabry M and Balant LP. Clinical Pharmacokinetics of clomipramine. *Clin. Pharmacokinet.* (1991) 20: 447-462.
- (15) Muller FO, Schall R, Mogilnicka M, Groenewoud G, Hundt HKL, Luus HG, Middle MV, Swart KJ and DeVall AC. Relative bioavailability of four clomipramine hydrochloride tablet products. *Biopharm. Drug Dispos.* (1996) 17: 81-90.
- (16) FDA Guidelines. Rockville, MD (1992) Bioequivalence Food and Drug Administration, Division 4 on of Bioequivalence, Office of Generic Drugs.
- (17) Coudore F, Hourcade F, Molinier-Manoukian C, Eschaliere A and Lavarenne J. Application of HPLC with silica-phase and reversed-phase eluents for the determination of clomipramine and demethylated and 8-hydroxylated metabolites. *J. Anal. Toxicol.* (1996) 20: 101-105.
- (18) Westenberg HG, Drenth BF, De Zeeuw RA, De Cuyper H, van Praag HM and Korf J. Determination of clomipramine and desmethyl clomipramine in plasma by means of liquid chromatography. *J. Chromatogr. A* (1977) 142: 725-733.
- (19) Pirola R, Mundo E, Bellodi L and Bareggi SR. Simultaneous determination of clomipramine and its desmethyl and hydroxy metabolites in plasma of patients by high-performance liquid chromatography after solid-phase extraction. *J. Chromatogr. B* (2002) 772: 205-210
- (20) Shah VP, Midha KK and Dighe S. Analytical method validation: bioavailability, bioequivalence and pharmacokinetic studies. *Eur. J. Drug Metabol. Pharmacokin.* (1992) 16: 249-255.
- (21) Chow CS and Liu JP. *Design and Analysis of Bioavailability and Bioequivalence Studies*. Marcel Dekker, New York (1992) 55.

This article is available online at <http://www.ijpr-online.com>

Archive of SID